

## **Supplemental Materials and Methods**

### **Fractionation of mitochondrial versus cytosolic proteins**

U343 cells were seeded at 1.5 million cells per plate and reverse transfected in two 10-cm Petri plates per condition. After 96 hours, the control or eIF5B-depleted cells were pooled, lysed in 1.5 mL of mitochondrial isolation buffer (250 mM sucrose, 10 mM HEPES, 1 mM EGTA, 0.5 mM PMSF, pH 7.4) by Dounce homogenization, and fractionated by differential centrifugation as previously described<sup>1</sup>. The mitochondrial pellets were resuspended in 50  $\mu$ L of RIPA lysis buffer supplemented with protease inhibitors. Western blotting was performed as described in the main text.

**Table S1. Genetic markers in glioblastoma lines used in this study.**

Cell line	p53	PTEN	MGMT	DR4/5	EGFR
U343	WT <sup>2</sup>	Mut <sup>3</sup>	Neg <sup>4</sup>	Pos <sup>5</sup>	WT <sup>3</sup>
U251N	Mut <sup>2</sup>	Mut <sup>6</sup>	Neg <sup>7</sup>	Pos <sup>8</sup>	WT <sup>9</sup>
A172	WT <sup>10</sup>	Mut <sup>11</sup>	Neg <sup>12</sup>	Pos <sup>13</sup>	Active mut <sup>14</sup>
U87MG	WT <sup>2</sup>	Mut <sup>3</sup>	Pos <sup>4</sup>	Pos <sup>8</sup>	WT <sup>15</sup>
U373	Mut <sup>16</sup>	Mut <sup>3</sup>	Neg <sup>7</sup>	Pos <sup>17</sup>	WT <sup>3</sup>

**Table S2. qPCR Primers and antibodies used in this study.**

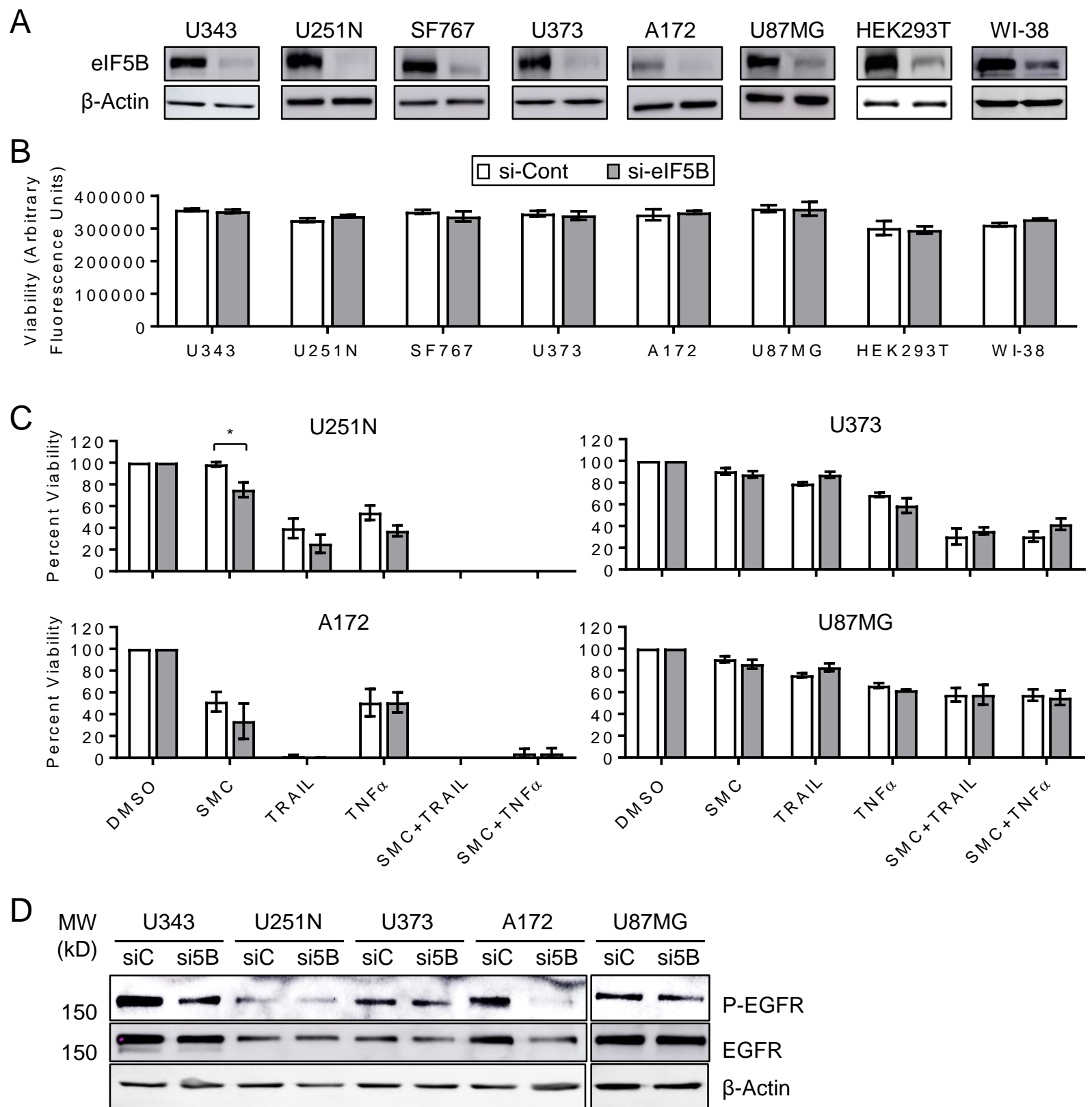
Target	Primer	Sequence (5' to 3')	Annealing Temp (°C)	Reference or source
XIAP	-	QuantiTect primer assay, QT00042854	55	Qiagen
Bcl-xL	-	QuantiTect primer assay, QT00236712	55	
Nrf2	-	QuantiTect primer assay, QT00027384	55	
p21	-	QuantiTect primer assay, QT00062090	55	
cIAP1	Fw Rv	TCTGGAGATGATCCATGGGTAGA TGGCCTTTCATTCGTATCAAGA	51	18
c-FLIP <sub>S</sub>	Fw Rv-S	GATGTTGCTATAGATGTGGTTC ATTTCCAAGAATTTTCAGATCAGGA	47	19
c-FLIP <sub>L</sub>	Rv-L	CCTGAATGGATTTCTTCACTGG	48	
β-actin	Fw Rv	TCACCCACACTGTGCCCATCTACGA TGAGGTAGTCAGTCAGGTCCC	55	20
Company	Antibody Target		Catalogue number	
Abcam	Secondary: Goat anti-rabbit-HRP conjugate		ab97051	
ProteinTech	eIF5B		13527-1-AP	
Bio-Rad	β-actin (hFAB Rhodamine)		12004163	
Abcam	cIAP1		ab108361	
	cIAP2		ab137393	
	eIF1A		ab177939	
	Mdm2		ab178938	
	Nrf2		ab71890	
	tBid		ab10640	
SantaCruz Biotech.	p27		SC-521	
Calbiochem	p53 (C-terminus)		pAB421	
Thermo Fisher	Phospho PERK		PA5-40294	
Cell Signalling Tech.	Akt		9272	
	Phospho Akt		9271	
	Bak		12105	
	Bax		2772	
	Bcl-xL		2762	
	Bid		2002	
	Bim		2933	

Cas-3	9662
Cas-7	9492
Cas-8	9746
Cas-9	9502
COX IV	4844
Cytochrome c	11940
DR4	42533
DR5	3696
EGFR	2232
Phospho EGFR	3777
FADD	2782
Phospho FADD	2781
c-FLIP	56343
JNK	9252
Phospho JNK	9251
Livin	5471
PARP (total)	9542
PARP (cleaved)	9541
p21	2947
p53 (total)	2527
PERK	3192
p65	4764
Phospho p65	3033
Survivin	2808
S6	2317
Phospho S6	5364
XIAP	2045

## Supplemental References

1. Jin Hyun Jun JK, Kyung Tae Kim and Ho Joong Sung. Application of an Efficient Method for Isolation of Mitochondria from Biological Samples. *Biomedical Science Letters* 2014, **20**(3): 180-184.
2. Asai A, Miyagi Y, Sugiyama A, Gamanuma M, Hong SH, Takamoto S, *et al.* Negative effects of wild-type p53 and s-Myc on cellular growth and tumorigenicity of glioma cells. Implication of the tumor suppressor genes for gene therapy. *Journal of neuro-oncology* 1994, **19**(3): 259-268.
3. Sauvageot CM, Weatherbee JL, Kesari S, Winters SE, Barnes J, Dellagatta J, *et al.* Efficacy of the HSP90 inhibitor 17-AAG in human glioma cell lines and tumorigenic glioma stem cells. *Neuro-oncology* 2009, **11**(2): 109-121.
4. Voss V, Senft C, Lang V, Ronellenfitsch MW, Steinbach JP, Seifert V, *et al.* The pan-Bcl-2 inhibitor (-)-gossypol triggers autophagic cell death in malignant glioma. *Molecular cancer research : MCR* 2010, **8**(7): 1002-1016.
5. Cheung HH, Mahoney DJ, Lacasse EC, Korneluk RG. Down-regulation of c-FLIP Enhances death of cancer cells by smac mimetic compound. *Cancer Res* 2009, **69**(19): 7729-7738.
6. Ren Y, Zhou X, Mei M, Yuan XB, Han L, Wang GX, *et al.* MicroRNA-21 inhibitor sensitizes human glioblastoma cells U251 (PTEN-mutant) and LN229 (PTEN-wild type) to taxol. *BMC cancer* 2010, **10**: 27.
7. Chahal M, Abdulkarim B, Xu Y, Guiot MC, Easaw JC, Stifani N, *et al.* O6-Methylguanine-DNA methyltransferase is a novel negative effector of invasion in glioblastoma multiforme. *Molecular cancer therapeutics* 2012, **11**(11): 2440-2450.
8. Kang YJ, Kim IY, Kim EH, Yoon MJ, Kim SU, Kwon TK, *et al.* Paxilline enhances TRAIL-mediated apoptosis of glioma cells via modulation of c-FLIP, survivin and DR5. *Experimental & molecular medicine* 2011, **43**(1): 24-34.
9. Torsvik A, Stieber D, Enger PO, Golebiewska A, Molven A, Svendsen A, *et al.* U-251 revisited: genetic drift and phenotypic consequences of long-term cultures of glioblastoma cells. *Cancer medicine* 2014, **3**(4): 812-824.
10. Mittelbronn M, Platten M, Zeiner P, Dombrowski Y, Frank B, Zachskorn C, *et al.* Macrophage migration inhibitory factor (MIF) expression in human malignant gliomas contributes to immune escape and tumour progression. *Acta neuropathologica* 2011, **122**(3): 353-365.
11. Zhang R, Banik NL, Ray SK. Differential sensitivity of human glioblastoma LN18 (PTEN-positive) and A172 (PTEN-negative) cells to Taxol for apoptosis. *Brain research* 2008, **1239**: 216-225.
12. Gaspar N, Marshall L, Perryman L, Bax DA, Little SE, Viana-Pereira M, *et al.* MGMT-independent temozolomide resistance in pediatric glioblastoma cells associated with a PI3-kinase-mediated HOX/stem cell gene signature. *Cancer research* 2010, **70**(22): 9243-9252.

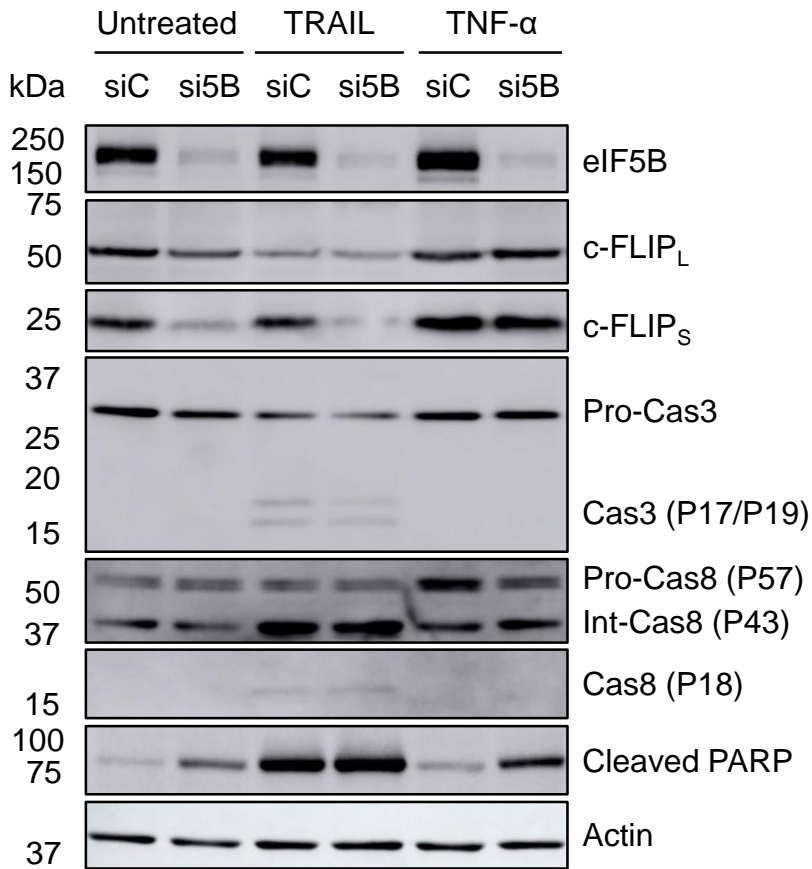
13. van Roosmalen IAM, Reis CR, Setroikromo R, Yuvaraj S, Joseph JV, Tepper PG, *et al.* The ER stress inducer DMC enhances TRAIL-induced apoptosis in glioblastoma. *SpringerPlus* 2014, **3**: 495.
14. Fenstermaker RA, Ciesielski MJ. EGFR Intron Recombination in Human Gliomas: Inappropriate Diversion of V(D)J Recombination? *Current genomics* 2007, **8**(3): 163-170.
15. Fontana AO, Piffaretti D, Marchi F, Burgio F, Faia-Torres AB, Paganetti P, *et al.* Epithelial growth factor receptor expression influences 5-ALA induced glioblastoma fluorescence. *Journal of neuro-oncology* 2017, **133**(3): 497-507.
16. Gomez-Manzano C, Fueyo J, Kyritsis AP, McDonnell TJ, Steck PA, Levin VA, *et al.* Characterization of p53 and p21 functional interactions in glioma cells en route to apoptosis. *Journal of the National Cancer Institute* 1997, **89**(14): 1036-1044.
17. Hetschko H, Voss V, Horn S, Seifert V, Prehn JH, Kogel D. Pharmacological inhibition of Bcl-2 family members reactivates TRAIL-induced apoptosis in malignant glioma. *Journal of neuro-oncology* 2008, **86**(3): 265-272.
18. Graber TE, Baird SD, Kao PN, Mathews MB, Holcik M. NF45 functions as an IRES trans-acting factor that is required for translation of cIAP1 during the unfolded protein response. *Cell Death Differ* 2010, **17**(4): 719-729.
19. Ueffing N, Singh KK, Christians A, Thorns C, Feller AC, Nagl F, *et al.* A single nucleotide polymorphism determines protein isoform production of the human c-FLIP protein. *Blood* 2009, **114**(3): 572-579.
20. Khan D, Katoch A, Das A, Sharathchandra A, Lal R, Roy P, *et al.* Reversible induction of translational isoforms of p53 in glucose deprivation. *Cell death and differentiation* 2015, **22**(7): 1203-1218.



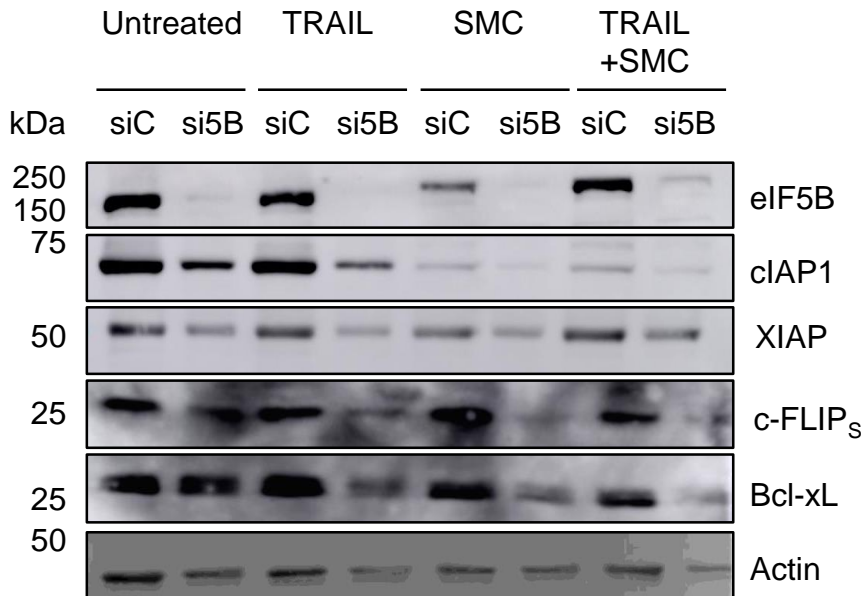
**Figure S1. Effects of eIF5B depletion on various cell lines.**

**(A)** Immunoblots confirming eIF5B silencing in various cell lines. **(B)** Depletion of eIF5B has little effect on alamarBlue activity in the tested cell lines. **(C)** Various cell lines were treated as in Figure 1. Data are expressed as mean  $\pm$  SEM for three independent biological replicates. \*,  $p < 0.05$ . **(D)** Effects of eIF5B depletion on total or phospho-EGFR (Tyr1068) in various cell lines.

A

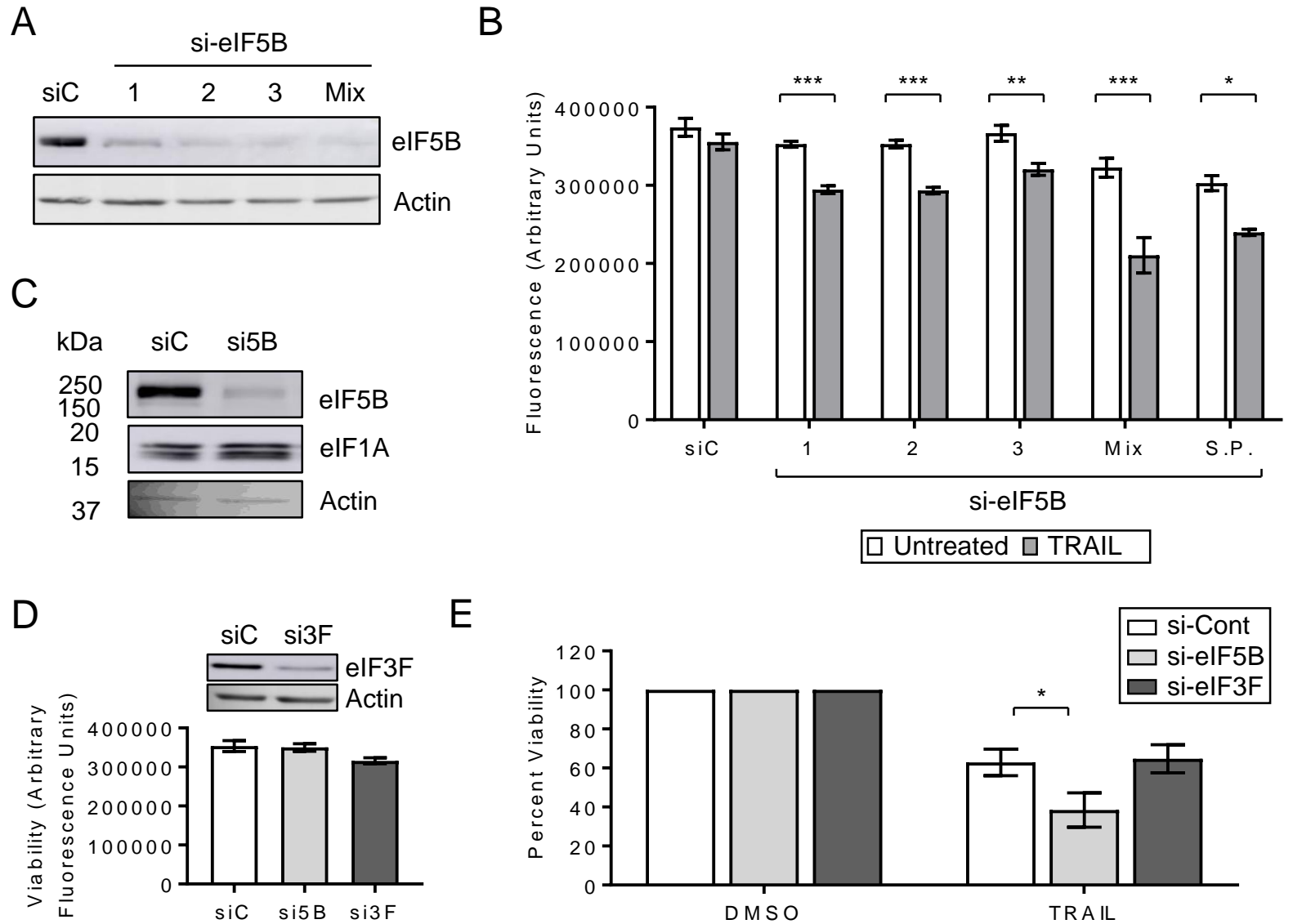


B



**Figure S2. Effects of eIF5B depletion on the levels of anti-apoptotic proteins and caspase activation in TNF- $\alpha$ - and SMC-treated U343 cells.**

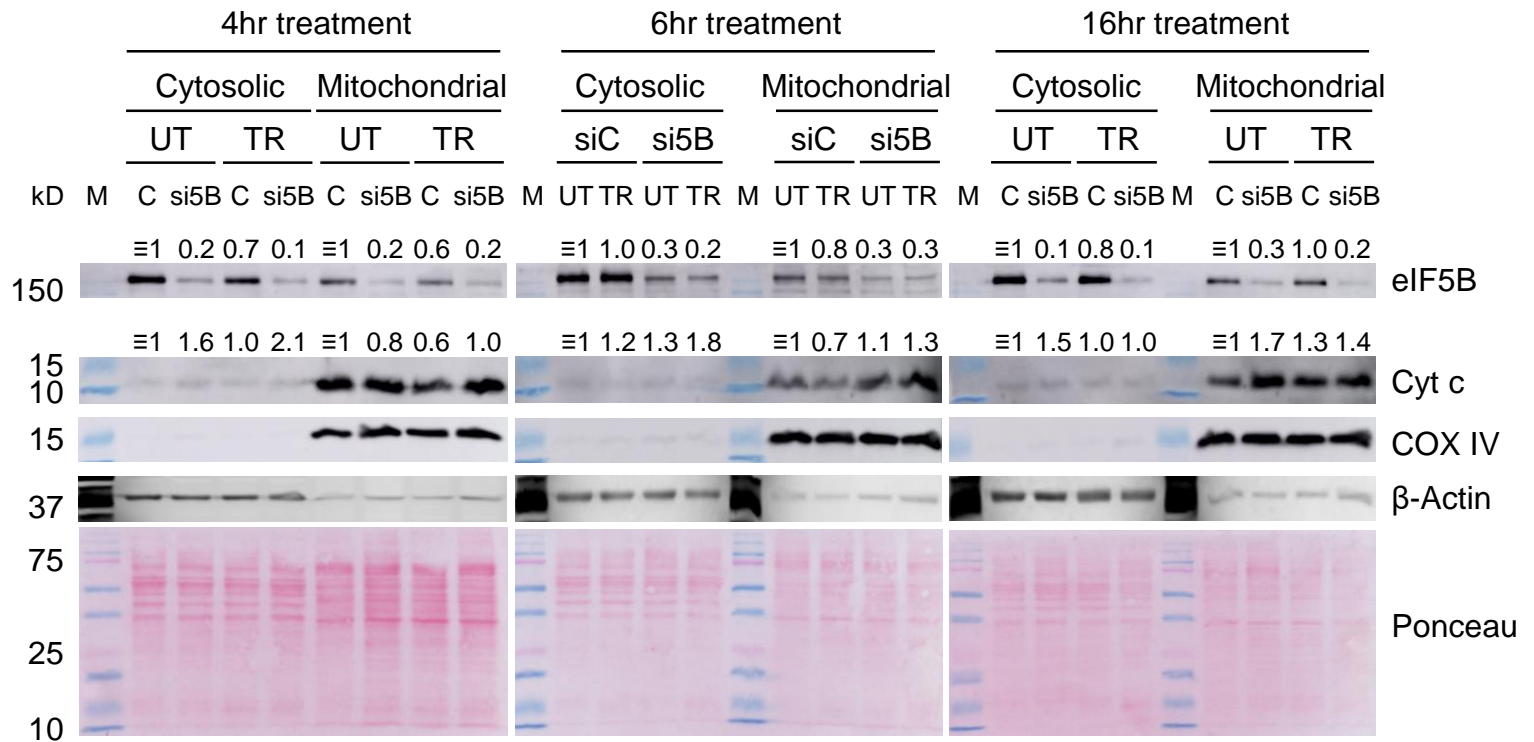
**(A)** TNF- $\alpha$  prevents down-regulation of c-FLIP<sub>L</sub> levels upon eIF5B depletion and enhances c-FLIP<sub>S</sub> levels. Unlike TRAIL, TNF- $\alpha$  causes no activation of caspase-8 or -3 and no further cleavage of PARP than does eIF5B depletion alone. Lanes 1-4 for eIF5B, Cas-3, cFLIP<sub>L</sub>, and cFLIP<sub>S</sub> are also shown in Figures 4B and 5A, respectively. Here we have shown the exact same lanes to compare them with TNF $\alpha$  lanes. **(B)** SMC decreases levels of cIAP1 but not XIAP, c-FLIP<sub>S</sub> or Bcl-xL. Depletion of eIF5B causes a decrease in cIAP1, XIAP, c-FLIP<sub>S</sub>, and Bcl-xL in the presence of TRAIL, SMC, or TRAIL+SMC.



**Figure S3. Controls related to Figure 1.**

**(A)** Immunoblot confirming eIF5B silencing by the individual siRNAs or the mix of three siRNAs used throughout this study, relative to the non-specific control siRNA (siC). **(B)** Each siRNA, as well as the mix and a SMARTpool from Dharmacon (S.P.), yields increased sensitivity of U434 cells to TRAIL. **(C)** Immunoblot showing no obvious effect of eIF5B-depletion on eIF1A levels. **(D)** Like eIF5B, depletion of eIF3F alone has no significant effect on the viability of U434 cells. **(E)** Unlike eIF5B, silencing of eIF3F does not sensitize U434 cells to TRAIL. Data are expressed as mean  $\pm$  SEM for 3 independent biological replicates. \*,  $p < 0.05$ ; \*\*,  $p < 0.01$ ; \*\*\*,  $p < 0.001$ .

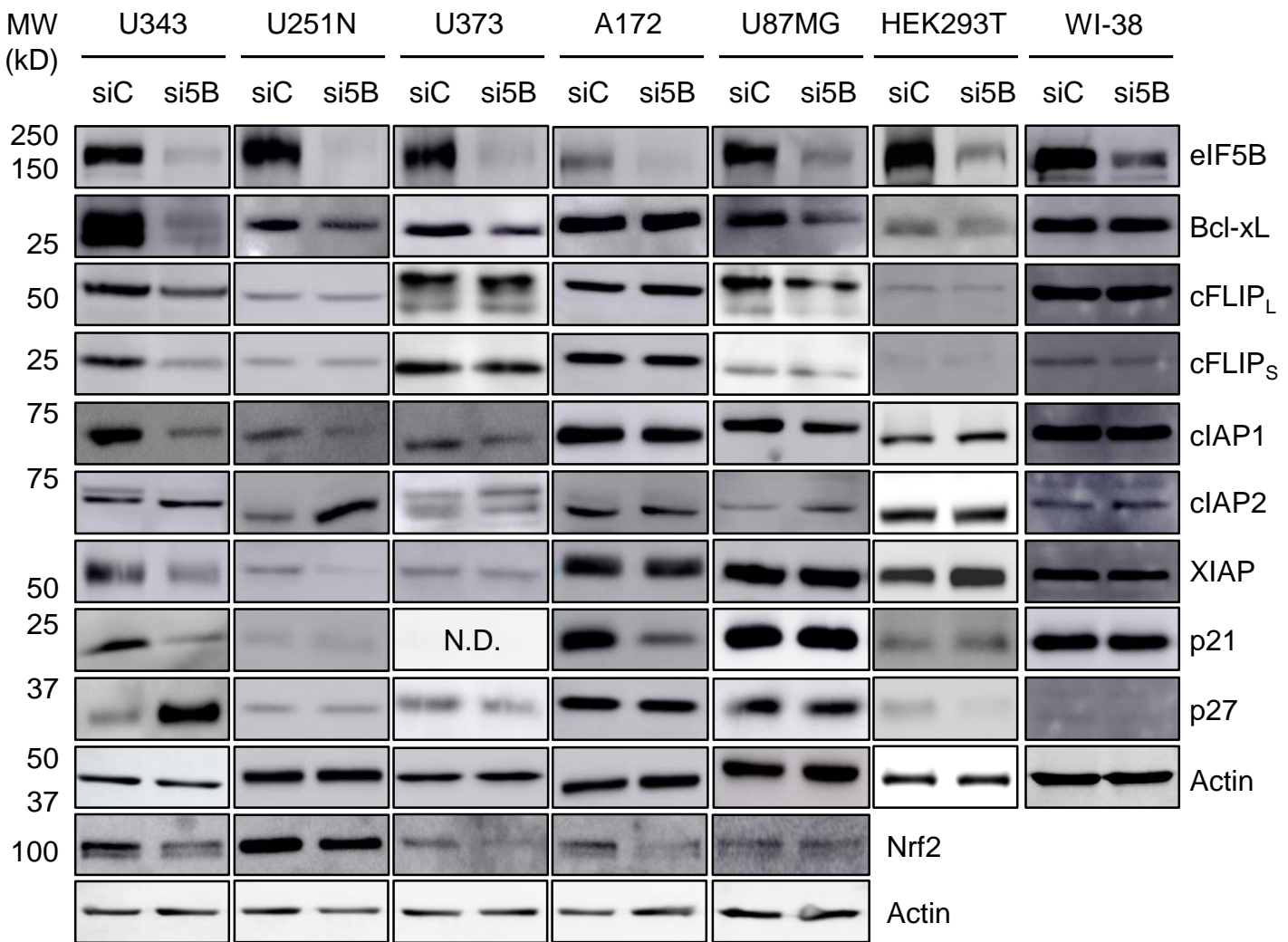




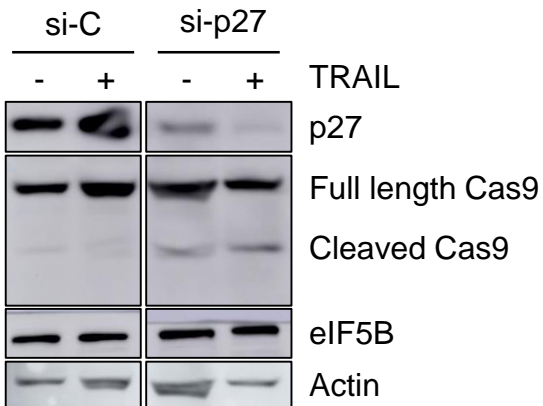
**Figure S4. Immunoblots of cytosolic versus mitochondrial proteins in control versus eIF5B-depleted U343 cells.**

Control or eIF5B-depleted U343 cells were harvested after a total of 96 hours of incubation; TRAIL was added to 100 ng/mL for the last 4, 6, or 16 hours of incubation, as indicated. Immunoblotting was performed for eIF5B, cytochrome c (Cyt c),  $\beta$ -actin (internal standard for cytosolic fractions), or cytochrome oxidase (COX IV; internal standard for mitochondrial fractions). A representative image is shown for a total protein stain of the membranes (Ponceau). The numbers above the blots represent a quantitation of eIF5B or cytochrome c, normalized to  $\beta$ -actin (in the case of the cytosolic fractions) or COX IV (in the case of the mitochondrial fractions).

A



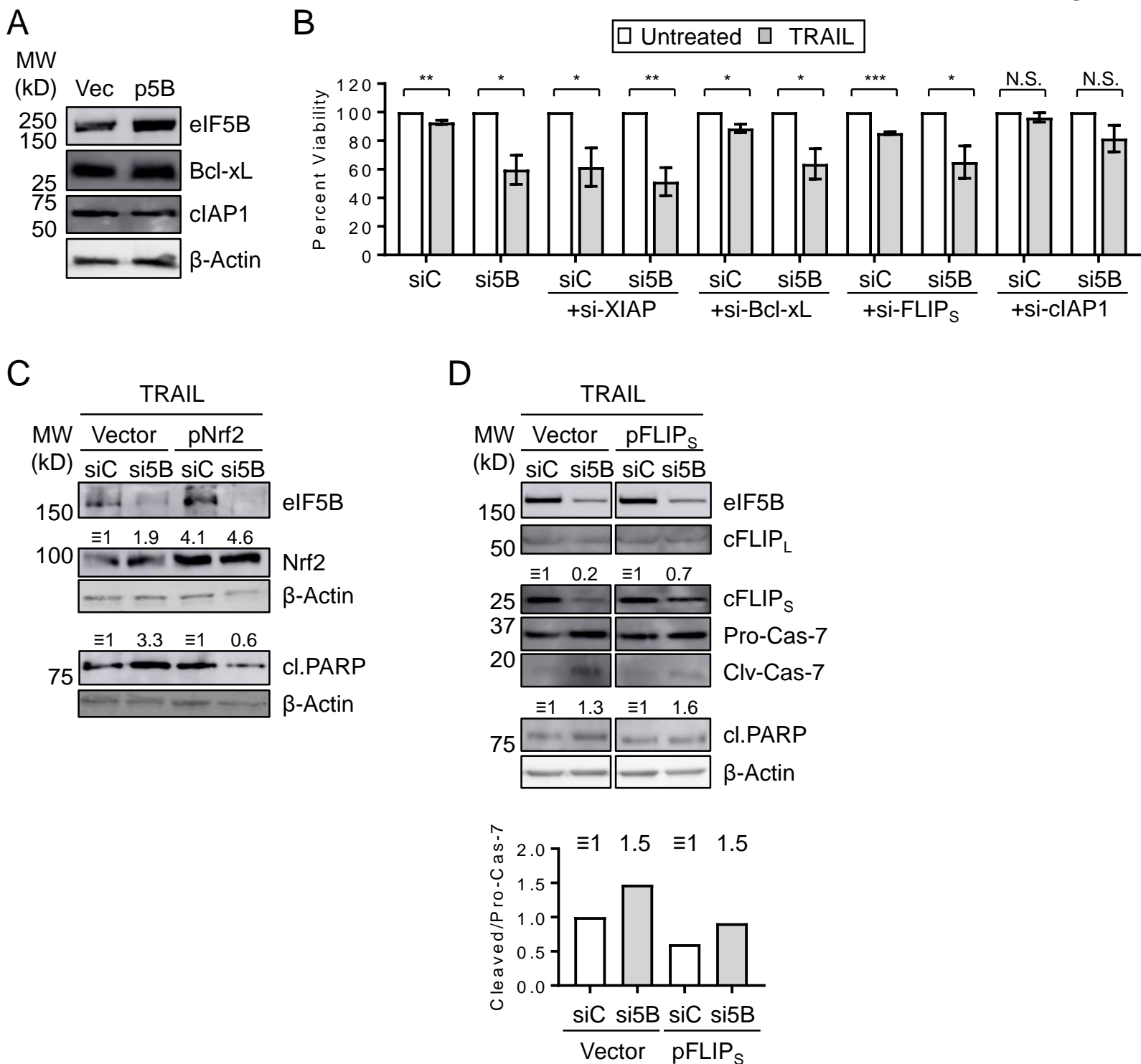
B



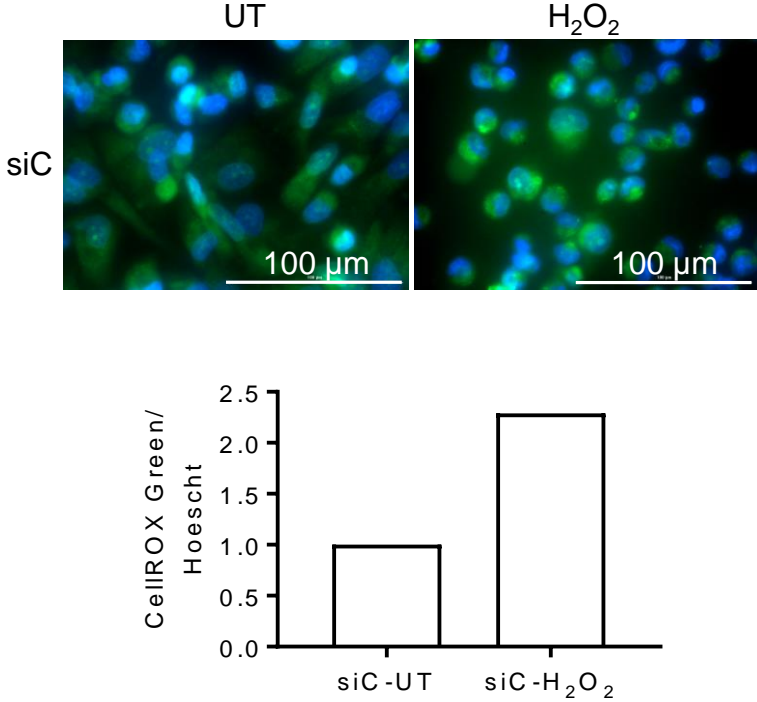
### Figure S5. Effects of eIF5B on pro-survival proteins.

(A) Effects of eIF5B depletion on anti-apoptotic proteins, p21, p27, and Nrf2 in various cell lines.

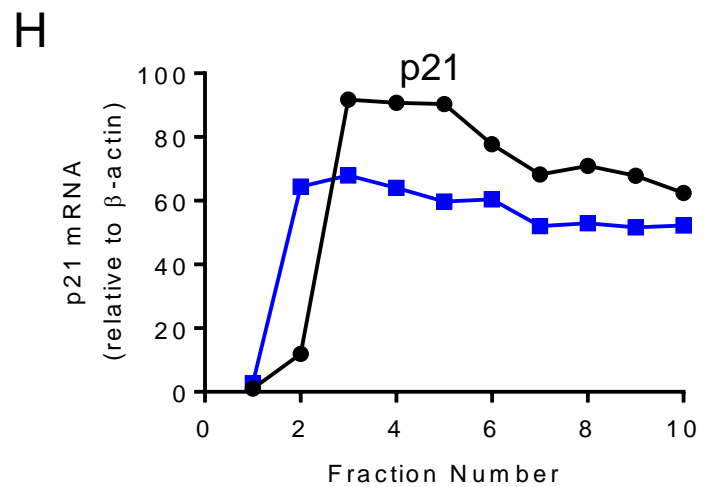
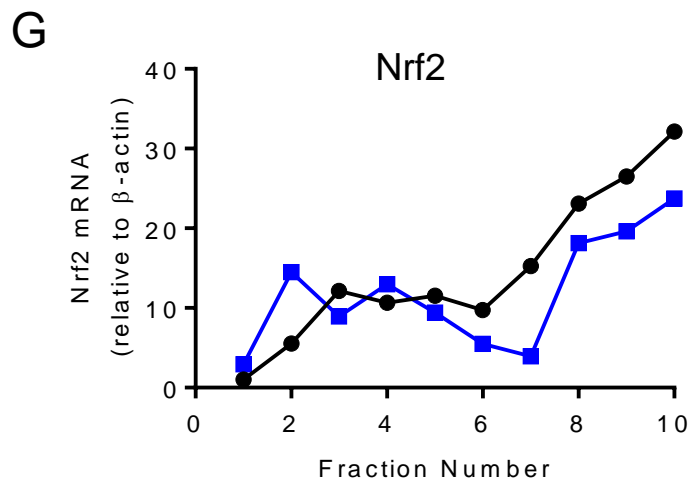
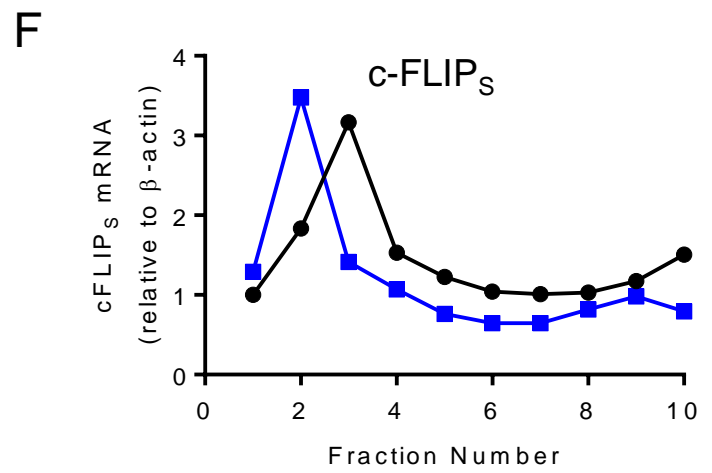
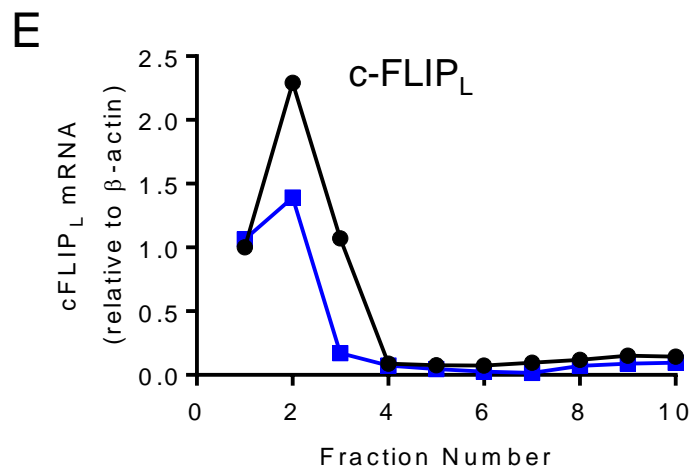
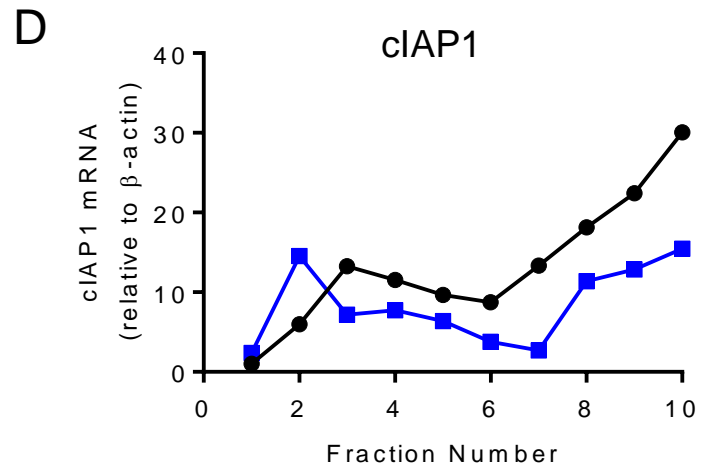
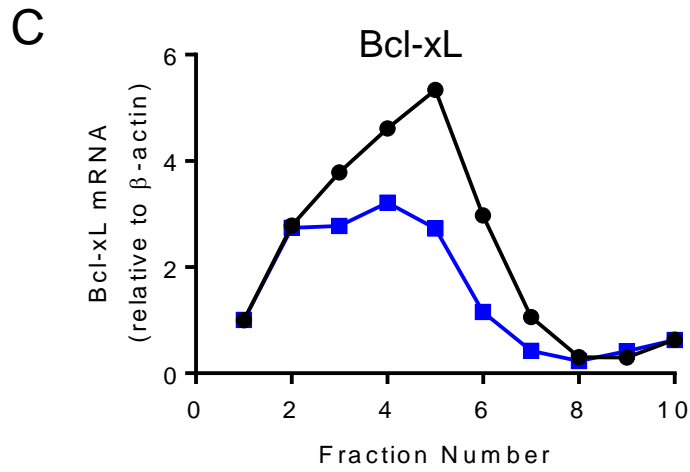
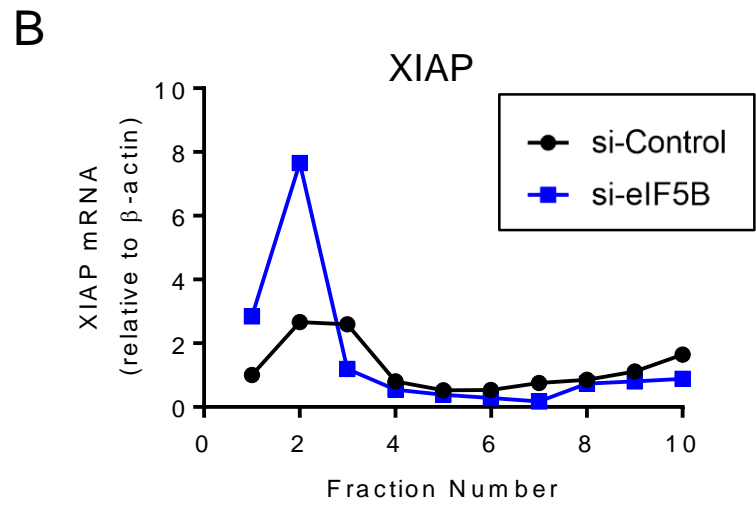
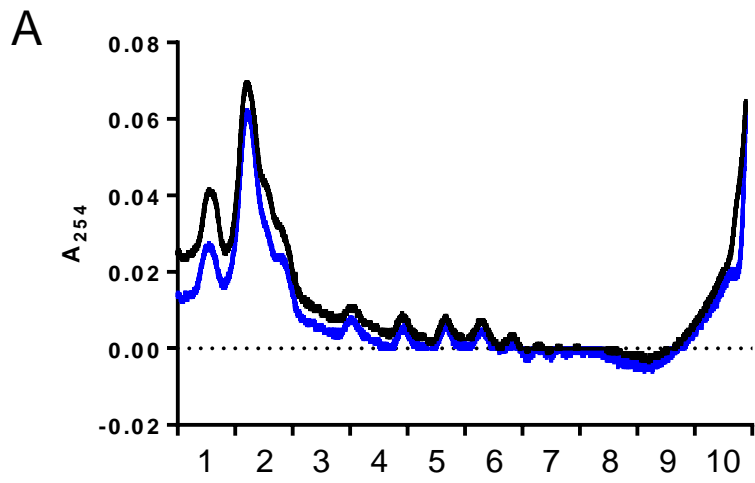
N.D., not detected. (B) Depletion of p27 leads to increased caspase-9 activation in U343 cells.



**Figure S6: The effect of eIF5B depletion on TRAIL sensitivity is exerted via several IAPs and Nrf2. (A)** Modest over-expression of eIF5B from a plasmid (p5B), obtained from Dharmacon (OHS6085-213579952), had little effect on Bcl-xL or cIAP1 levels. **(B)** The effect of eIF5B depletion on TRAIL sensitivity is epistatic with depletion of XIAP, Bcl-xL, and c-FLIP<sub>S</sub>. U434 cells were treated essentially as in Figure 1, except that eIF5B was depleted either alone or in combination with XIAP, Bcl-xL, c-FLIP<sub>S</sub> or cIAP1. Data are expressed as mean  $\pm$  SEM for 3 independent biological replicates. \*,  $p < 0.05$ ; \*\*,  $p < 0.01$ ; \*\*\*,  $p < 0.001$ . **(C)** Depletion of eIF5B causes increased PARP cleavage, which is reversed by expression of Nrf2 from a plasmid (pNrf2), obtained from AddGene (NC16 pCDNA3.1 FLAG NRF2). **(D)** Depletion of eIF5B causes increased cas-7 activation and PARP cleavage, neither of which is reversed by expression of c-FLIP<sub>S</sub> from a plasmid (pFLIP<sub>S</sub>), a gift of Christian Schwerk. For eIF5B depletion coupled with Nrf2 or c-FLIP<sub>S</sub> overexpression, 420,000 U434 cells were seeded per well in a 6-well plate and reverse transfected with control or eIF5B-specific siRNAs. After 48 hours of incubation, the cells were transfected with the indicated plasmid. After a further 42 hours of incubation, the cells were treated with 100 ng/mL TRAIL for 4 hours before being harvested for western blotting.



**Figure S7. CellROX green staining of untreated and H<sub>2</sub>O<sub>2</sub>-treated U343 cells (related to Figure 6E).** U43 cells were treated with siC as in Figure 6A. Instead of TRAIL, cells were treated for 4 hours with 200 μM H<sub>2</sub>O<sub>2</sub> or left untreated, before staining with CellROX green and processing as in Figure 6A.



**Figure S8. Representative polysome profiles for control or eIF5B-depleted U343 cells (related to Figure 7).**

**(A)** Representative polysome profiles for control and eIF5B-depleted U343 cells (black and blue curves, respectively). **(B-H)** RNA was purified from the sucrose fractions indicated in panel (A) and subjected to RT-qPCR analysis. The proportion of mRNAs encoding XIAP **(B)**, Bcl-xL **(C)**, cIAP1 **(D)**, c-FLIP<sub>L</sub> **(E)**, c-FLIP<sub>S</sub> **(F)**, Nrf2 **(G)** and p21 **(H)**, all relative to  $\beta$ -actin mRNA, were quantified for each fraction.