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Supplemental Information

**Rbf Activates the Myogenic Transcriptional Program
to Promote Skeletal Muscle Differentiation**

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SUPPLEMENTAL INFORMATION

SUPPLEMENTAL FIGURES:

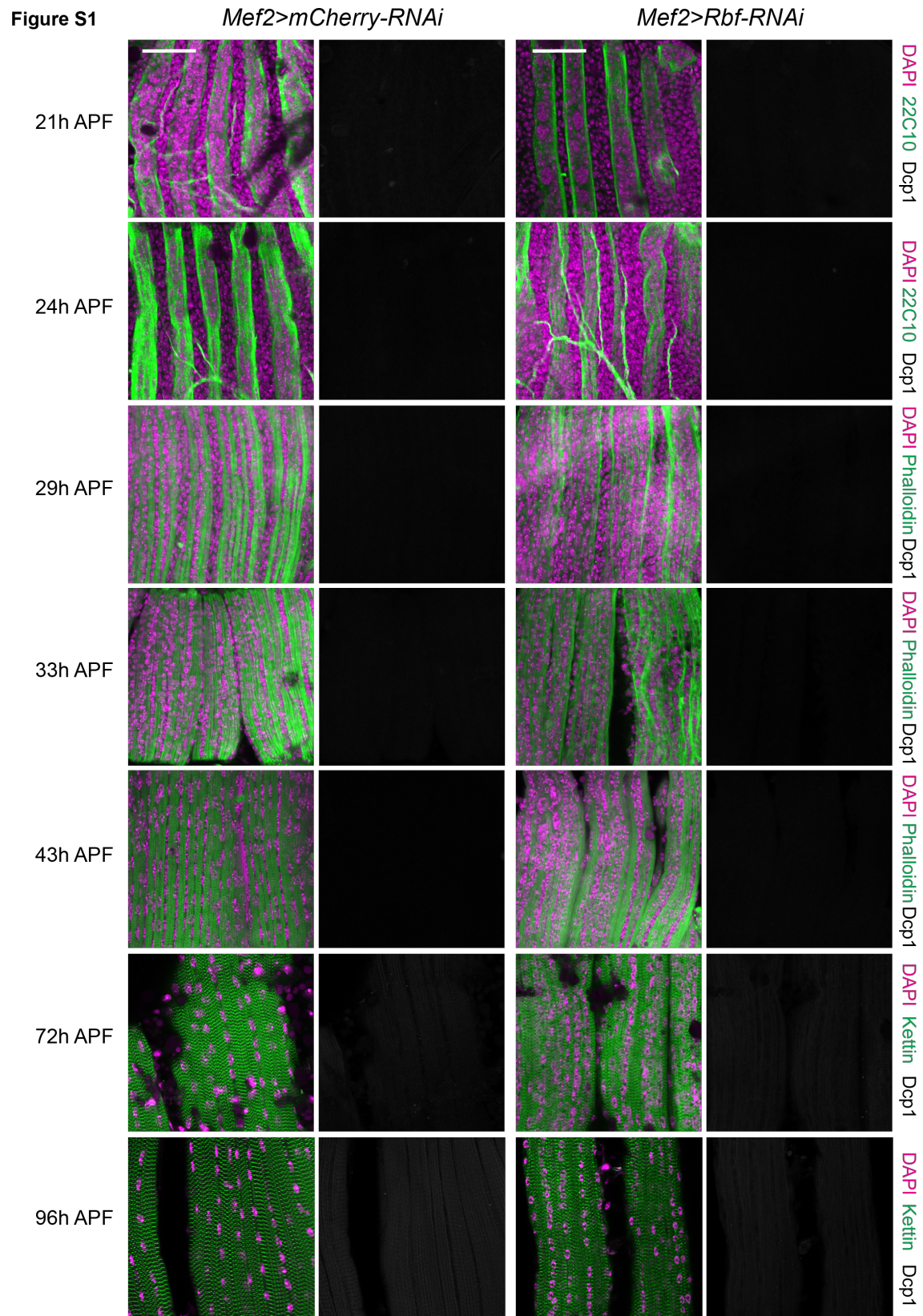


Figure S1: The loss of Rbf does not induce apoptosis mediated by dcp1 in developing indirect flight muscles. Related to Figure 2.

Developing DLMS stained over time from 21 to 96 h APF. Apoptotic cells were marked using anti-dcp1 antibody (right panel). Overall structure was analyzed using DAPI and Phalloidin or 22c10 to mark developing IFM or Kettin to stain sarcomere depending on the developmental stages (left panel). Single confocal sections are shown. Scale 40 μm . Full genotypes are *w*-, *UAS-Dicer2*; +; *Mef2-GAL4/UAS-mCherry-RNAi* (left panel) and *w*-, *UAS-Dicer2*; +; *Mef2-GAL4/UAS-Rbf-RNAi* (right panel)

Figure S2

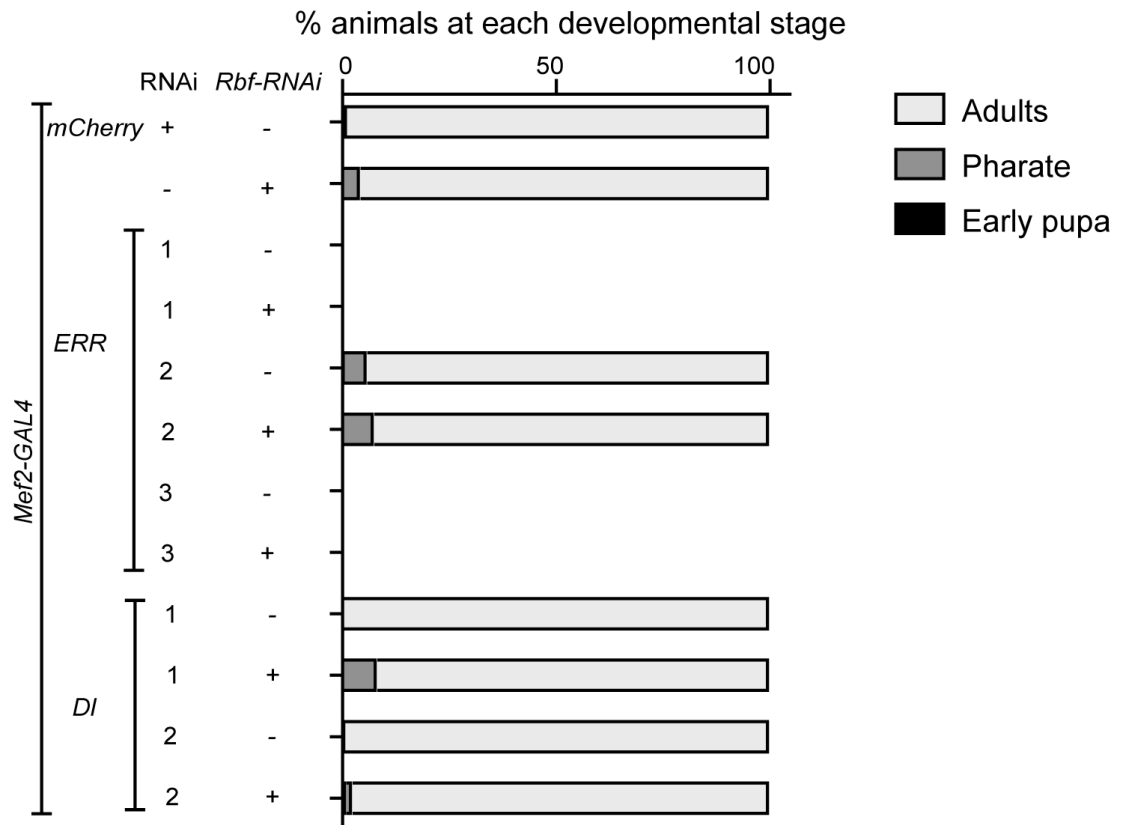


Figure S2: Rbf interacts with other transcription factors. Related to Figure 7.

Genetic interactions between Rbf and candidates were analyzed by knocking down the expression of each gene with or without *Rbf-RNAi* in muscles using *Mef2-GAL4* driver. The candidates tested are estrogen-related receptor (ERR) and Delta (DI). Lethality during pupa development was analyzed as % animals scored at each developmental stage. Adults: light grey, Pharate: grey, Early pupa: dark grey. N= 94 - 319 flies per genotype. Two independent experiments. Genotypes are *Mef2-GAL4/UAS-mCherry-RNAi*, *UAS-Rbf-RNAi/Mef2-GAL4*, three RNAi lines were used for ERR (1: *UAS-ERR-RNAi^{HMJ23520}*, 2: *UAS-ERR-RNAi^{JF02431}* and 3: *UAS-ERR-RNAi^{HMC03087}*), and two for DI (1: *UAS-DI-RNAi^{GD1238}* and 2: *UAS-DI-RNAi^{JF02825}*).

SUPPLEMENTAL TABLES:

Table S7: Primer sequence. Related to STAR Methods

Gene	Forward primer sequence	Reverse primer sequence	Application
<i>twi</i>	AAGCTCAGCAAGATCCAGACC	GAGCTGGCCGATCCATACG	RT-qPCR
<i>Mef2</i>	CGGATATCATGAGCCTCAACAC	CGTGGAACCATTGCTATTCTGC	RT-qPCR
<i>htl</i>	AACGCATCGAAACCGTTCAC	TGGTGCCTGTTTCTGTATC	RT-qPCR
<i>stumps</i>	AACAAGGTGGTTGCTCTGCT	ACTGCAGGGTGTAGGGATTG	RT-qPCR
<i>Act88F</i>	ATGGTGGGTATGGGTCAGAA	CTTCTCCATGTCGTCCCAGT	RT-qPCR
<i>Mhc</i>	AAGAACGACCTCGAGAACCAG	TCGGCCTTCTTCTTGCTG	RT-qPCR
<i>fln</i>	GGCAAAGAGGGACAAACAAC	ACTACGAGTGCTCATCCGTTT	RT-qPCR
<i>RpL32</i>	TACAGGCCCAAGATCGTGAAG	GACGCACTCTGTTGTCGATACC	RT-qPCR
<i>RpL30</i>	GCAAATACTGCCTGGGCTAC	ACTTCAGTCTTGGCCAGCAT	RT-qPCR
<i>Lmpt-transcript K</i>	AGTGGCTGCCCTAAAACAAC	ATTGCTCCTCTGCTGCGATAG	RT-qPCR
<i>sals</i>	GCAAGCCATGAAGAAGAACCAG	TCGTCTTCGTCTAGCTTCATGG	RT-qPCR
<i>Tm1-transcript H</i>	ATAAGCAGCCGCAGCAAAAAG	TTTCTGCTGCCTCTGTTTTCG	RT-qPCR
<i>Tm2</i>	TCCAAGATCATGGAGCTGGAG	TTCATCTCGCGCTTGAAGTC	RT-qPCR
<i>Mlc2</i>	TTCTCTGTGTTCTCCAGAAGC	TTGTCCGCATCCATGAGTTG	RT-qPCR
<i>how</i>	ATCTGTCCGATGACCTGCATG	TTAGCTCATCTTCGCCTTCGG	RT-qPCR
<i>salm</i>	ACTACAGGAGCCACACCAAAG	ATGTGTTGCTTCAGGTTGCC	RT-qPCR
<i>Rbf</i>	AACAACGTGAAGCAGCTGAG	ACTTGGCGAGGAATTTCTGC	RT-qPCR
<i>P5CDh1 157/40</i>	TATTCGGCGCTTTTCCACTG	ACAAAGGCACACCACACTTC	ChIP-qPCR
<i>Cyt-c-p -496/-372</i>	AAAGATGGTGTTCGCAGGTC	TGGCCGATCTTGTTGTTTCG	ChIP-qPCR
<i>Zasp66 -847/-712</i>	TCGAAAACCTCTGCACATCGG	GGGCCACAGGATTTAATTGACC	ChIP-qPCR
<i>Negative site</i>	TGTGTATGCCTTGCTTGCAC	TCTATGCACACGCTCTACTGAG	ChIP-qPCR
<i>Rbf</i>	<u>CTAATACGACTCACTATAGGGAGCG</u> AATTTCGACGAACGCATTC	<u>CTAATACGACTCACTATAGGGA</u> GCAGCCAAGCAGGTACACCTTG	dsRNA
<i>GFP</i>	<u>CTAATACGACTCACTATAGGGAGCG</u> TAAACGGCCACAAGTTCAG	<u>CTAATACGACTCACTATAGGGA</u> GACGAACTCCAGCAGGACCATG	dsRNA

* T7 sequence is underlined

Table S7: Primer sequences. Sequence of primers used to measure gene expression (RT-qPCR), genomic occupancy (ChIP-qPCR) and primers used to make dsRNA.