Current Biology, Volume 29

## **Supplemental Information**

## The Genomic Footprints of the Fall

### and Recovery of the Crested Ibis

Shaohong Feng, Qi Fang, Ross Barnett, Cai Li, Sojung Han, Martin Kuhlwilm, Long Zhou, Hailin Pan, Yuan Deng, Guangji Chen, Anita Gamauf, Friederike Woog, Robert Prys-Jones, Tomas Marques-Bonet, M. Thomas P. Gilbert, and Guojie Zhang



Figure S1. Demographic history estimation based on the pairwise sequentially Markovian coalescent (PSMC) model, Related to Figure 2. Population sizes inferred from the three Northeast (NE) sub groups, as well as the East (EA), Northwest (NW), and Contemporary group (MC). g, generation time;  $\mu$ , mutation rate.



### Figure S2. Patterns of deleterious mutations across populations, Related to Figure 3.

(A)-(D) Site-Frequency-Spectrum for each category: deleterious, benign and synonymous, for each group. All sequenced samples were used for this analysis. The analyses were applied to four groups as follows: East (EA, A), Northeast (NE, B), Northwest (NW, C), and Contemporary groups (D). Comparison of the ratio of homo- to homo- and heterozygous sites. (E) Deleterious missense mutations based on Grantham Scores. (F) Loss of function mutations based on the SnpEff algorithm.



## Figure S3. Haplotype structures and length distribution of Class II and Extended Regions in the historical and contemporary groups, Related to Figure 4.

(Left) Haplotype blocks are detected based on the SNP datasets for historical and contemporary groups, respectively. Colored circles show the haplotype structure (each box indicates the start and end SNPs of one haplotype) and the diversity along the regions (color indicates the allelic richness). The allelic richness of historical groups is estimated in the random way (see Method). Gaps in the colored circles are the intervals between two neighboring haplotype blocks. Genomic structures of the regions are drawn as grey bars, with different sizes showing different gene loci of varied sizes. (Right) Length distribution of haplotype blocks. Haplotype blocks with lengths larger than 2000 bp are grouped into the last bar. The plotting area for the bin of 250-2000bp is enlarged in the top right corner. (A) Haplotype blocks and length distribution in the Class II region. (B) Haplotype blocks and length distribution in the Extended region.

# Table S1. Effective population sizes (*Ne*) of each group at the sampling time point estimated by NeEstimator, Related to Figure 2.

Four general geographic groups are the East (EA), Northeast (NE), Northwest (NW), and Contemporary groups. Following the insights shown by the NJ tree, the *Ne* of three NE subgroups was calculated respectively.

Group	Number of individuals	Number of SNPs <sup>a</sup>	Ne <sup>b</sup>
NE-1	8	1,527,932	72.7(71.2-74.2)
NE-2	8	1,652,489	144.5(139.2-150.2)
NE-3	6	1,687,450	94.7(91.3-98.4)
EA	11	1,339,682	356.2(335.7-379.4)
NW	23 <sup>c</sup>	1,522,027	163.2(161.3-165.1)
Contemporary	8	1,124,530	10.7(10.7-10.8)

a: Only SNP loci without missing data were used in this analysis.

b: Data are represented as mean (95% Cls).

c: CHKSEI14060220 is not involved.

### Table S2. Summary of estimated inbreeding coefficient (FUNI) and Ne, Related to Figure 3.

SNP VCF files for *Pan paniscus, Pongo abelii, Pongo pygmaeus, Pan troglodytes* and *Gorilla gorilla* are from [S1]. SNP VCF file for *Vulpes Vulpes* is from [S2]. SNP VCF files for *Ursus maritimus* and *Ursus arctos* are from [S3]. SNP VCF file for *Anas platyrhynchos* is from [S4]. SNP VCF file for Tibetan fowl from Qinghai is from [S5]. Raw sequence read FASTQ files for *Platyspiza crassirostris* are from [S6]. SNP BAM files for *Ficedula albicollis* are from [S7]. Raw sequence read FASTQ files for *Ectopistes migratorius* are from [S9].

Species	Conservation status <sup>a</sup>	F <sub>UNI</sub>	Ne
Pan paniscus	EN	0.027804	40.5
Pongo abelii	CR	-0.003456	223.8
Pongo pygmaeus	CR	0.001124	100.3
Pan troglodytes	EN	0.112602	5.8
Gorilla gorilla	CR	0.073231	26.6
Vulpes vulpes	LC	-0.075651	370.0
Ursus maritimus	VU	-0.062079	238.7
Ursus arctos	LC	0.093783	7.2
Anas platyrhynchos	LC	0.015845	161.9
Tibetan fowl from Qinghai	LC	0.059132	35.0
Platyspiza crassirostris	LC	0.086422	5.0
Ficedula albicollis	LC	0.001172	272.0
Ailuropoda melanoleuca	VU	0.085511	29.0
Ectopistes migratorius	EX	-0.006270	64.0
Historical groups of	EX	0.044317	72.7-356.2
Nipponia nippon			
Contemporary group of	EN	0.172716	10.7
Nipponia nippon			

a: Five conservation status are listed in the table: Least concern (LC), Vulnerable (VU), Endangered (EN), Critically endangered (CR), and Extinct (EX).

### Supplemental references

- S1. Prado-Martinez, J., Sudmant, P.H., Kidd, J.M., Li, H., Kelley, J.L., Lorente-Galdos, B., Veeramah,
  K.R., Woerner, A.E., O'Connor, T.D., Santpere, G., et al. (2013). Great ape genetic diversity and
  population history. Nature 499, 471.
- S2. Kukekova, A.V., Johnson, J.L., Xiang, X., Feng, S., Liu, S., Rando, H.M., Kharlamova, A.V., Herbeck,
  Y., Serdyukova, N.A., Xiong, Z., et al. (2018). Red fox genome assembly identifies genomic regions associated with tame and aggressive behaviours. Nat. Ecol. Evol. 2, 1479-1491.
- Liu, S., Lorenzen, E.D., Fumagalli, M., Li, B., Harris, K., Xiong, Z., Zhou, L., Korneliussen, T.S., Somel, M., Babbitt, C., et al. (2014). Population Genomics Reveal Recent Speciation and Rapid Evolutionary Adaptation in Polar Bears. Cell 157, 785-794.
- S4. Zhang, Z., Jia, Y., Almeida, P., Mank, J.E., van Tuinen, M., Wang, Q., Jiang, Z., Chen, Y., Zhan, K., and Hou, S. (2018). Whole-genome resequencing reveals signatures of selection and timing of duck domestication. GigaScience 7, giy027.
- S5. Li, D., Che, T., Chen, B., Tian, S., Zhou, X., Zhang, G., Li, M., Gaur, U., Li, Y., Luo, M., et al. (2017). Genomic data for 78 chickens from 14 populations. GigaScience 6, 1-5.
- S6. Lamichhaney, S., Berglund, J., Almén, M.S., Maqbool, K., Grabherr, M., Martinez-Barrio, A., Promerová, M., Rubin, C.J., Wang, C., and Zamani, N. (2015). Evolution of Darwin's finches and their beaks revealed by genome sequencing. Nature 518, 371.
- S7. Nadachowska-Brzyska, K., Burri, R., Smeds, L., and Ellegren, H. (2016). PSMC analysis of effective population sizes in molecular ecology and its application to black-and-white Ficedula flycatchers. Mol. Ecol. 25, 1058-1072.
- Zhao, S., Zheng, P., Dong, S., Zhan, X., Wu, Q., Guo, X., Hu, Y., He, W., Zhang, S., Fan, W., et al. (2012). Whole-genome sequencing of giant pandas provides insights into demographic history and local adaptation. Nat. Genet. 45, 67.
- S9. Murray, G.G., Soares, A.E., Novak, B.J., Schaefer, N.K., Cahill, J.A., Baker, A.J., Demboski, J.R., Doll, A., Da Fonseca, R.R., and Fulton, T.L. (2017). Natural selection shaped the rise and fall of passenger pigeon genomic diversity. Science 358, 951-954.