Supporting Information

Incorporation of amniotic membrane as an immunomodulatory design element in collagen scaffolds for tendon repair

Rebecca A. Hortensius¹, Jill H. Ebens², Marley Dewey³, Brendan A.C. Harley^{2,4}

 ¹ Dept. of Bioengineering
² Dept. of Chemical and Biomolecular Engineering
³ Dept. of Materials Science and Engineering
⁴ Carl R. Woese Institute for Genomic Biology University of Illinois at Urbana-Champaign Urbana, IL 61801

Contains:

5 pages

3 figures

Supporting Methods: Lyophilized amniotic membrane sheets were cut and homogenized in 0.5M acetic acid. The diluted particles were placed on a glass slide and the acetic acid was allowed to evaporate overnight. The slide was sputter coated and imaged with a Philips XL30 ESEM-FEG scanning electron microscope under high vacuum. A minimum of 25 fields of view were analyzed to obtain the particle size distribution found in Fig. S1.



Figure S1. Particle size distribution of amniotic membrane post-homogenization. (**A**) Representative ESEM image of amniotic membrane particles seen in dark grey. Black arrow indicates larger amniotic membrane particle and white arrow displays smaller amniotic membrane particle. (**B**) Particle size distribution of amniotic membrane particles based on particle measurement of ESEM images.



Figure S2. Human mesenchymal stem cell metabolic activity in 3D scaffolds cultured in noninflammatory control media. C:CS = collagen-chondroitin sulfate scaffolds, C:AM = collagenamniotic membrane scaffolds, C:CS+AM = collagen-chondroitin sulfate scaffolds (core) wrapped with a layer of amniotic membrane (shell).



Figure S3. Cross-sectional SEM images of (**A**) C:CS and (**B**) C:AM scaffold microstructure. Scale bar = 250 microns.