

Supporting Information for : A biosensor-based approach reveals links between efflux pump expression and cell cycle regulation in pleiotropic drug resistance of yeast

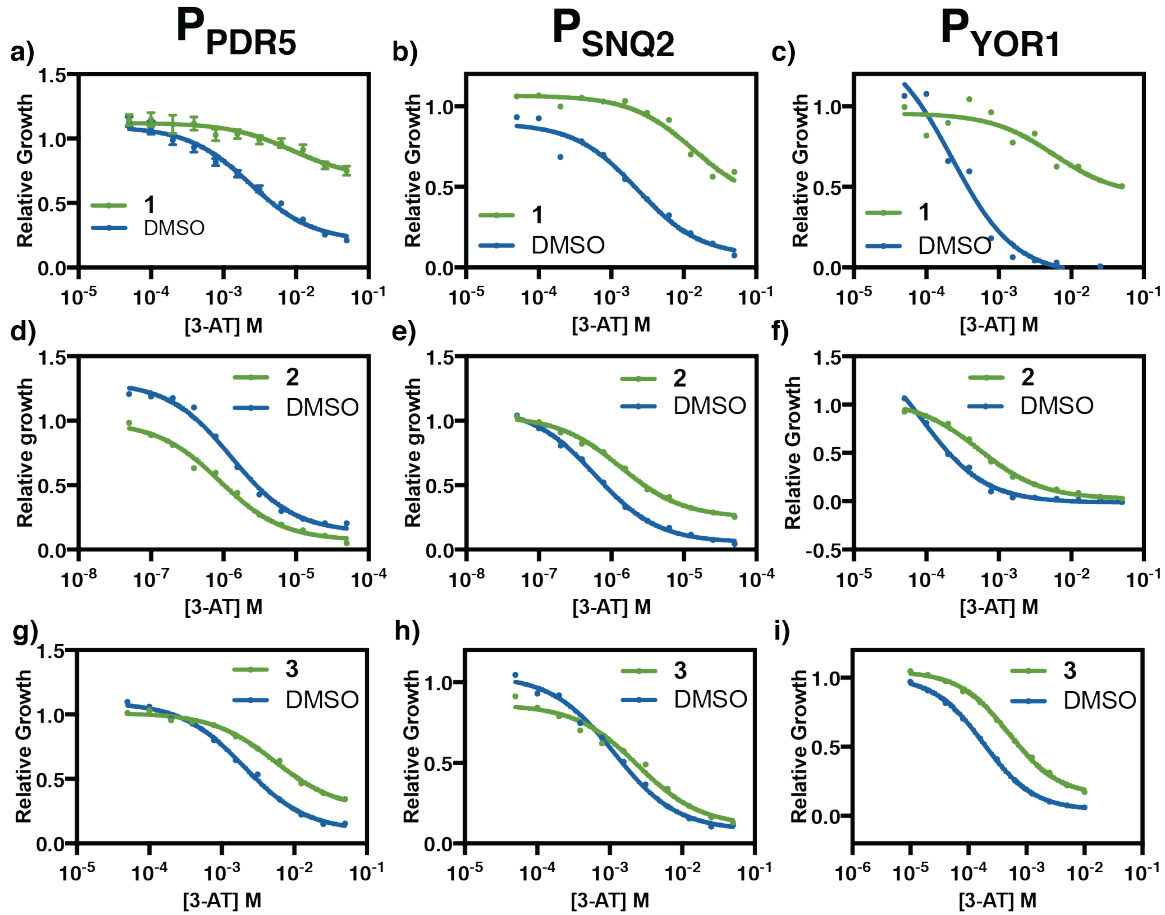
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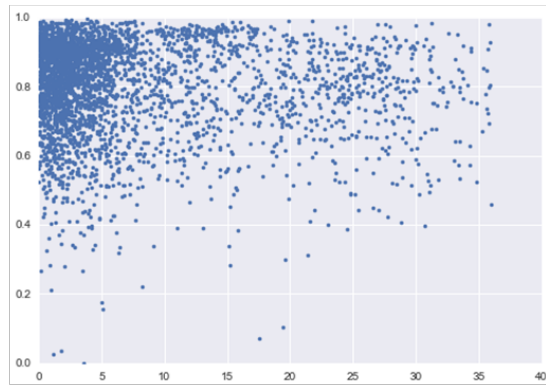
Supplementary Figures:



**Figure S1: Does dependent response of strains JLY31-33 to 3-AT in the presence of compounds 1-3.** All strains were grown in YNB media with 10  $\mu$ M drug or an equivalent amount of DMSO. Line represents best fit dose-response curve, and error bars represent SD (n=4). **a)** JLY31 treated with **1** (reused from Figure 1c and included here for context). **b)** JLY32 treated with **1**. **c)** JLY33 treated with **1**. **d)** JLY31 treated with **2**. **e)** JLY32 treated with **2**. **f)** JLY33 treated with **2**. **g)** JLY31 treated with **3**. **h)** JLY32 treated with **3**. **i)** JLY33 treated with **3**.

**A**

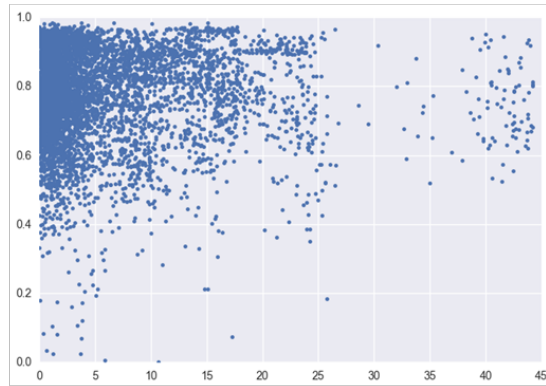
jaccard distance  
between pairs of  
structures



euclidean distance between pairs of  
expression Z-scores

**B**

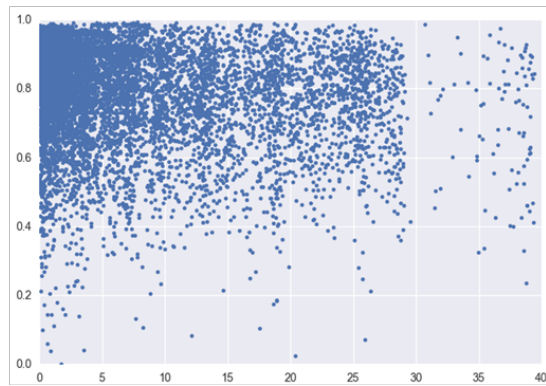
jaccard distance  
between pairs of  
structures



euclidean distance between pairs of  
expression Z-scores

**C**

jaccard distance  
between pairs of  
structures



euclidean distance between pairs of  
expression Z-scores

**Figure S2: Similarity in structure of microsource natural products does not correlate with similarities in transporter activations.** For drugs that had a Z-score  $>3$  for each promoter, jaccard distance were calculated between pairs of the structures, and euclidean distance were calculated between the difference in Z-scores. **a)** JLY31 induction. **b)** JLY32 induction. **c)** JLY33 induction.



Supplementary Tables

Table S1: Sequences of the promoters used in this study

Promoter	Sequence
P <sub>PDR5</sub>	CGTTAACGTAAATATGCTTCCTCTTTGATTCCAACAATGATATTATAAGGAAATTATCGTGCACA ATCTAATCAACAAGGACAAAAGAAAAGAAAAGTTGAGAGAGAGAACAAAAGTTCAAATCAAAG AAAAAAAAAGAACATCAAATTACCTATTACAATAAACAAATTAAGCCATACTCGCAACAATTGCC ATAATAGAAAAGCAGCACCTCGTTGGCGCAGTCCCTTACATAGTACACAACATTTATCACTTCAC ACAATCAGGAGTGGAACCTCAATGGAAAACAACACCACACGTATGATCTTACTAATAAAAAGAACA TGAACGTTCTCAGCGCGAACGTTTCGATTCTGCGCCTTCGAGCACAGGATAAGTTGCAGGAAG CCATCACATCTATGCAACGATTATCACGACACAACCTTGCCGCCGAGAAAACGTCCGTGGAGAA CCATTCGGTTCGATTGCTTCCCACGGAACGAGTGGACTGAAACTTAAGACTGCCCTCTCTTTCC GCGGAATCGCTCATGCCGCGGTGCCACAACATTTTCAGATTTACTAAGACTCCGGTGTAGTGTGG GCTCACCCGCGGGTCGTGATCACGATTCAGCACCCCTTTGGACTCGTGATTCCTGGAAAAGGTCA GATCTGTATTCTACTTATGGTAATGTGCTAAAAAAGAGAAATGTCTCCGCGGAACCTTCTAC GCCGTGGTACGATATCTGTTGAACGTAATCTGAGCAATACAACAAGGCCTCTCCTATACATAT ATAATTGTGATGTGCATAACCTTATGGCTGTTCGCTTTTATTATCATACCTTAGAATGAAATCCAA AAGAAAAAGTCACGCAAAGTTGCAAAACATAACAACCTGTGTTAGTTATCACTCGACTTTGTTA TTCTAATTATAAATAAATTGGCAACTAGGAACTTCGAAAAAGAAAATAAAGACCCCTTTAAGTT TTCGTATCCGCTCGTTCGAAAGACTTTAGACAAAA
P <sub>SNQ2</sub>	GATCTCACAGATGAAGAACTAAAAGACGCAATTATAGTTTCGAAGCCTGCCTTGGGTACTTGTTT GTACGAACAATAACACTAAACACCTTCTTGAACAACCTTTAAATATTTAAAGTTTACTCATAACC TCTTACACTCTATAAAGAAACAAGTTGGTGAGTCCAATAGAACCCATTCCAGCATCCGATGCAA CCATAGCCCGTTGATGCGAGTCAGAGTGGTCCGCGCACAGAGGTTATGTGACCATGGTAACC GTACACATTTTTCCACGGCAAGGAAGTGGCGCGAAAGAGCACCCGCCCTCCACGGATCACCCC ATTTGGAAGAAGACGGAGACATAAAAAAATTACCTTCACGCCAGACTATGTATGACTCGTAG AAATCTAGCATCCGCGGAGCTATTTAAGTTTCCGCGGATGCCTTTCGATCCTTTCCAGTCGTC TGGTTCGCGCACATATGACTAAATGTCCGCCATTTCCGTTTAAATCCGTTTCTATTTCTTTA GGTCTTTCCATTGTATTCCTGCAACCCCGTGGGTATTGGTATTACTAATAGTACTACGCAAC ATCAACATATAAAAACAATGATGAGTAGTGGCTCCCTCCCTCACCGCTATCGCCTCACCACTTTT ATGCTTGTATATGCTTGTAGATAAATAATACCTAACGAAAACAATATATACGAACCTAGTGTAT TTGTATCTTTTGTTTTTCAAGTTGAAAGTGTGCGAGGTCAAAAAAAAAAAGCTCACTGTAGGAT ATAGGAGTTACTATATCACCAAGTGCATTACATTCTCAGTGCATCCATCGTCTTCAACATTGATTA TTTTCTCCTTCCATTGATTAGAGTTCAAGCTCCCTGAGAAAACGAAGGTATAGCGGACGTACCCG CAGAGACATAAAAAAAGAAAACTATATCGAAGACCGAAAAGCAGTAAAAAAGTGGATAGAATA ACACAGCTACCAAAATACGTAAAGAGAATTCA
P <sub>YOR1</sub>	TTCTTTTCTTTTTTTTTTTTCTTTGACTGTTACCCGGTTGTTTATATTTGTAGGAAAACAACAAC GACAGAGAAAAATATCCTTGCAGTGGCGGCTAATTTGTTAGTTGACTGATTGATCACCTTCACTTA TTAAAGTAAAATCAGCATAACAAGAGATCAGAAGGGAGAAAGAGAGTGGGCAAGGCTATAGTACT TTGAAGAAAGCATCTTTGAACCGACCAGTTCTCTTCAAGCAAAATCTATATGACTAACCAGCA AGGGCAAAAGGGTGTGAGAGGGCCCGTCTTCTCCCGCTATAGCCGTCACTGGTATCCCTCCT GGCTGCACAAATCCGATAGAAAAGGGGAAGAAGGAAGTTTAGTGCCACCTTATAGCACGCAGTTA CTGTTTACGCTAAGGAGAGGCATACTCAATTTTTATTAGTCGCCTTCTTATGTTGCTGCGTTTTTA TCCACGGTTCTCTACTAAATGCTTGCATAAGCGCTTCTATTTTCTCCCCACCGCGAGGCGGAA ATGGCACATTTTTTTTCTTTTGTCTCTGTGCTTTTGTGTAATTTTTGGCATGTGCTATTGTATGAA GATAACCGTGGTTCCGTGGAATAGCCGGAATTTTGGCGGGAATATGACGGACATGATTTAA CACCCGTGGAATGAAAAAGCCAAGGTAAGAAAGTGGCAATATTTTTCTACAATAGATCTG CTGTCCCTTAAATGATTACCATACATATATATTTATTACACATCTGTCAGAGGTAGCTAGCGA AGGTGTCACTGAAATATTTTTGTTCCAGTTAGTATAAATACGGAGGTAGAACAGCTCTCCGCGT GTATATCTTTTTTGCCTATACAAGAACAGGAAGAACGCATTTCCATACCTTTTTCTCCTTACA GGTGCCCTCTGAGTAGTGTACGAACGAGGAAAAAGATTAATATTACTGTTTTTATATTTAAAAA GAGTAAAGCCGTTGCTATATACGAAT

**Table S2: GO Enrichment analysis of screen hits**

<b>Treatment Condition</b>	<b>GO Term</b>	<b>Description</b>	<b>P-value</b>
<b>3</b>	GO:0048583	regulation of response to stimulus	1.33E-04
	GO:0010646	regulation of cell communication	3.38E-04
	GO:0042147	retrograde transport, endosome to Golgi	3.45E-04
	GO:0010647	positive regulation of cell communication	4.67E-04
<b>2</b>	GO:0010468	regulation of gene expression	4.00E-05
	GO:0044774	mitotic DNA integrity checkpoint	6.25E-05
	GO:0031326	regulation of cellular biosynthetic process	1.28E-04
	GO:0009889	regulation of biosynthetic process	1.31E-04
	GO:0031570	DNA integrity checkpoint	1.35E-04
	GO:2000112	regulation of cellular macromolecule biosynthetic process	1.67E-04
	GO:0010556	regulation of macromolecule biosynthetic process	1.97E-04
	GO:0050789	regulation of biological process	2.09E-04
	GO:0019354	siroheme biosynthetic process	2.74E-04
	GO:0046156	siroheme metabolic process	2.74E-04
	GO:0050794	regulation of cellular process	2.81E-04
	GO:1901991	negative regulation of mitotic cell cycle phase transition	2.81E-04
	GO:0065007	biological regulation	3.00E-04
	GO:1901988	negative regulation of cell cycle phase transition	3.65E-04
	GO:0051171	regulation of nitrogen compound metabolic process	4.37E-04
	GO:0000075	cell cycle checkpoint	6.59E-04
GO:0000103	sulfate assimilation	6.65E-04	
GO:0007093	mitotic cell cycle checkpoint	7.31E-04	
GO:2000765	regulation of cytoplasmic translation	8.12E-04	
<b>1</b>	GO:0061188	negative regulation of chromatin silencing at rDNA	3.82E-06
	GO:0061187	regulation of chromatin silencing at rDNA	7.53E-06
	GO:0061186	negative regulation of chromatin silencing at silent mating-type cassette	2.19E-05
	GO:0090054	regulation of chromatin silencing at silent mating-type cassette	5.03E-05
	GO:0031939	negative regulation of chromatin silencing at telomere	7.16E-05
	GO:0031938	regulation of chromatin silencing at telomere	2.85E-04
	GO:2000217	regulation of invasive growth in response to glucose limitation	3.56E-04
	GO:0031936	negative regulation of chromatin silencing	3.56E-04
	GO:0045815	positive regulation of gene expression, epigenetic	4.39E-04
	GO:0060969	negative regulation of gene silencing	5.35E-04
	GO:0061408	positive regulation of transcription from RNA polymerase II promoter in response to heat stress	6.89E-04
	GO:0010646	regulation of cell communication	9.28E-04
	GO:2000219	positive regulation of invasive growth in response to glucose limitation	9.35E-04
<b>Overall</b>	GO:0065007	biological regulation	7.36E-07
	GO:0050794	regulation of cellular process	1.10E-06
	GO:0050789	regulation of biological process	2.09E-06
	GO:0031326	regulation of cellular biosynthetic process	2.42E-06
	GO:0009889	regulation of biosynthetic process	2.54E-06
	GO:0080135	regulation of cellular response to stress	2.72E-06
GO:0010556	regulation of macromolecule biosynthetic process	3.36E-06	

GO:0080134	regulation of response to stress	3.57E-06
GO:0010468	regulation of gene expression	6.25E-06
GO:2000112	regulation of cellular macromolecule biosynthetic process	1.17E-05
GO:0048583	regulation of response to stimulus	1.37E-05
GO:0051171	regulation of nitrogen compound metabolic process	1.75E-05
GO:0080090	regulation of primary metabolic process	3.56E-05
GO:0060255	regulation of macromolecule metabolic process	4.06E-05
GO:0019222	regulation of metabolic process	4.43E-05
GO:0031323	regulation of cellular metabolic process	4.87E-05
GO:0010646	regulation of cell communication	5.85E-05
GO:0016569	covalent chromatin modification	5.99E-05
GO:0006357	regulation of transcription from RNA polymerase II promoter	8.41E-05
GO:0010647	positive regulation of cell communication	1.24E-04
GO:0048522	positive regulation of cellular process	1.41E-04
GO:0061188	negative regulation of chromatin silencing at rDNA	1.63E-04
GO:0048523	negative regulation of cellular process	1.63E-04
GO:0006355	regulation of transcription, DNA-templated	1.83E-04
GO:2001141	regulation of RNA biosynthetic process	1.83E-04
GO:1903506	regulation of nucleic acid-templated transcription	1.83E-04
GO:0031936	negative regulation of chromatin silencing	1.86E-04
GO:0048518	positive regulation of biological process	2.08E-04
GO:0048519	negative regulation of biological process	2.19E-04
GO:0031939	negative regulation of chromatin silencing at telomere	2.31E-04
GO:0045815	positive regulation of gene expression, epigenetic	2.55E-04
GO:0061187	regulation of chromatin silencing at rDNA	3.14E-04
GO:0051252	regulation of RNA metabolic process	3.20E-04
GO:0060969	negative regulation of gene silencing	3.42E-04
GO:0009966	regulation of signal transduction	3.93E-04
GO:0023051	regulation of signaling	3.93E-04
GO:0006325	chromatin organization	7.69E-04
GO:0061186	negative regulation of chromatin silencing at silent mating-type cassette	8.72E-04
GO:0048584	positive regulation of response to stimulus	9.98E-04

**Table S3: qPCR validation of PDR transcriptional regulators**

Drug	Gene deleted	Systematic name	Brief Description	PDR5		SNQ2		YORI		Screen Result
				fc	p	fc	p	fc	p	
<b>1</b>	<i>hoΔ</i>	YDL227C	Site-specific endonuclease	15.61		6.90		5.68		
	<i>pdr1Δ</i>	YGL013C	Transcription factor that regulates the pleiotropic drug response	1.40	<b>0.000</b>	1.91	<b>0.001</b>	1.17	<b>0.000</b>	Upregulator
	<i>nbp2Δ</i>	YDR162C	Protein involved in the HOG (high osmolarity glycerol) pathway	8.91	<b>0.012</b>	6.03	0.152	4.38	0.095	Upregulator
	<i>cho2Δ</i>	YGR157W	Phosphatidylethanolamine methyltransferase (PEMT)	8.21	<b>0.005</b>	4.10	<b>0.007</b>	4.98	0.138	Upregulator
	<i>ypt6Δ</i>	YLR262C	Rab family GTPase	8.20	<b>0.037</b>	4.33	<b>0.024</b>	4.00	0.069	Upregulator
	<i>gyp1Δ</i>	YOR070C	Cis-golgi GTPase-activating protein (GAP) for yeast Rabs	9.56	<b>0.021</b>	4.03	0.072	3.99	0.064	Upregulator
	<i>gcn2Δ</i>	YDR283C	Protein kinase	22.90	<b>0.025</b>	11.94	<b>0.005</b>	8.26	<b>0.015</b>	Downregulator
	<i>mad1Δ</i>	YGL086W	Coiled-coil protein involved in spindle-assembly checkpoint	22.46	0.123	11.83	<b>0.022</b>	9.85	<b>0.014</b>	Downregulator
	<i>fyv10Δ</i>	YIL097W	Subunit of GID complex	17.98	0.148	11.64	<b>0.005</b>	6.05	0.324	Downregulator
	<i>sap30Δ</i>	YMR263W	Component of Rpd3L histone deacetylase complex	18.07	0.144	12.20	<b>0.005</b>	8.45	<b>0.013</b>	Downregulator
<i>tom7Δ</i>	YNL070W	Component of the TOM (translocase of outer membrane) complex	22.65	<b>0.023</b>	13.77	<b>0.002</b>	9.00	<b>0.009</b>	Downregulator	
<b>2</b>	<i>hoΔ</i>	YDL227C	Site-specific endonuclease	1.52		36.15		26.59		
	<i>pdr1Δ</i>	YGL013C	Transcription factor that regulates the pleiotropic drug response	1.37	0.381	20.14	<b>0.021</b>	23.76	0.303	Upregulator
	<i>yrr1Δ</i>	YOR162C	Zn2-Cys6 zinc-finger transcription factor	2.72	0.092	11.87	<b>0.002</b>	15.10	<b>0.041</b>	Upregulator
	<i>ssn2Δ</i>	YDR443C	Subunit of the RNA polymerase II mediator complex	2.91	0.073	20.28	<b>0.014</b>	15.79	<b>0.032</b>	Upregulator
	<i>met1Δ</i>	YKR069W	S-adenosyl-L-methionine uroporphyrinogen III transmethylase	1.46	0.454	21.07	<b>0.016</b>	10.58	<b>0.006</b>	Upregulator
	<i>met8Δ</i>	YBR213W	Bifunctional dehydrogenase and ferrochelatase	1.61	0.438	18.22	<b>0.010</b>	9.42	<b>0.006</b>	Upregulator
	<i>cho2Δ</i>	YGR157W	Phosphatidylethanolamine methyltransferase (PEMT)	1.66	0.412	20.42	<b>0.046</b>	14.92	<b>0.026</b>	Upregulator
	<i>spf1Δ</i>	YEL031W	P-type ATPase, ion transporter of the ER membrane	3.29	0.051	73.58	<b>0.016</b>	45.53	<b>0.033</b>	Downregulator
	<i>alg6Δ</i>	YOR002W	Alpha 1,3 glucosyltransferase	2.98	0.070	62.82	<b>0.048</b>	32.90	0.211	Downregulator
<i>mad2Δ</i>	YJL030W	Component of the spindle-assembly checkpoint complex	2.46	0.119	56.78	<b>0.026</b>	30.75	0.233	Downregulator	
<b>3</b>	<i>hoΔ</i>	YDL227C	Site-specific endonuclease	9.53		3.27		4.04		
	<i>pdr1Δ</i>	YGL013C	Transcription factor that regulates the pleiotropic drug response	1.83	<b>0.000</b>	1.42	<b>0.000</b>	1.46	<b>0.000</b>	Upregulator
	<i>uga2Δ</i>	YBR006W	Succinate semialdehyde dehydrogenase	8.31	0.079	2.69	<b>0.008</b>	3.29	<b>0.027</b>	Upregulator
	<i>cho2Δ</i>	YGR157W	Phosphatidylethanolamine methyltransferase (PEMT)	5.70	<b>0.001</b>	3.75	0.056	4.58	0.141	Upregulator
	<i>ykl053wΔ</i>	YLK053W	Dubious ORF	8.38	<b>0.021</b>	3.37	0.314	4.44	0.238	Downregulator
	<i>dph1Δ</i>	YIL103W	Protein required for synthesis of diphthamide	14.73	<b>0.005</b>	3.81	0.205	6.80	0.052	Downregulator
	<i>stf2Δ</i>	YBR103W	WD40 repeat-containing subunit of Set3C histone deacetylase complex	10.28	0.261	6.05	<b>0.000</b>	6.45	<b>0.001</b>	Downregulator



**Table S4: qPCR on *mad*Δ strains**

Drug	[Drug](μM)	Strain	PDR5		SNQ2		YOR1	
			FC	P	FC	P	FC	P
5	50	<i>ho</i> Δ	22.81		1.80		22.36	
		<i>mad1</i> Δ	37.83	<b>0.014</b>	2.94	<b>0.001</b>	46.05	<b>0.033</b>
		<i>mad2</i> Δ	34.27	0.058	2.41	<b>0.022</b>	43.01	<b>0.041</b>
		<i>mad3</i> Δ	26.60	0.083	2.57	<b>0.009</b>	36.52	0.068
6	10	<i>ho</i> Δ	2.51		1.24		3.26	
		<i>mad1</i> Δ	3.94	<b>0.019</b>	1.40	0.220	6.71	<b>0.001</b>
		<i>mad2</i> Δ	4.29	<b>0.007</b>	1.21	0.449	7.08	<b>0.001</b>
		<i>mad3</i> Δ	4.06	<b>0.010</b>	0.97	0.087	6.41	<b>0.007</b>
7	50	<i>ho</i> Δ	1.01		2.88		0.91	
		<i>mad1</i> Δ	1.28	0.079	4.90	<b>0.002</b>	1.26	<b>0.010</b>
		<i>mad2</i> Δ	0.93	0.127	5.70	<b>0.003</b>	0.84	0.268
		<i>mad3</i> Δ	1.17	<b>0.005</b>	3.74	<b>0.000</b>	1.17	<b>0.001</b>
8	50	<i>ho</i> Δ	3.11		1.70		2.03	
		<i>mad1</i> Δ	6.09	<b>0.001</b>	1.67	0.446	4.46	<b>0.004</b>
		<i>mad2</i> Δ	5.98	<b>0.003</b>	1.83	0.325	4.43	<b>0.003</b>
		<i>mad3</i> Δ	5.10	0.094	1.97	0.195	4.02	<b>0.014</b>
9	25	<i>ho</i> Δ	1.46		0.95		3.15	
		<i>mad1</i> Δ	2.22	<b>0.025</b>	0.98	0.357	5.75	<b>0.009</b>
		<i>mad2</i> Δ	1.79	0.153	1.05	0.297	5.01	<b>0.001</b>
		<i>mad3</i> Δ	1.47	0.469	0.97	0.414	4.07	<b>0.019</b>

**Table S5:** Description of all genes tested in qPCR experiments\*

Gene	Systematic Name	Gene name	Brief Description	Description
<i>ho1</i>	YDL227C	HOthollic switching endonuclease	Site-specific endonuclease	Site-specific endonuclease; required for gene conversion at the MAT locus (homothallic switching) through the generation of a ds DNA break; expression restricted to mother cells in late G1 as controlled by Swi4p-Swi6p, Swi5p, and Ash1p
<i>pdr1Δ</i>	YGL013C	Pleiotropic Drug Resistance	Transcription factor that regulates the pleiotropic drug response	Transcription factor that regulates the pleiotropic drug response; zinc cluster protein that is a master regulator involved in recruiting other zinc cluster proteins to pleiotropic drug response elements (PDREs) to fine tune the regulation of multidrug resistance genes; relocates to the cytosol in response to hypoxia; PDR1 has a paralog, PDR3, that arose from the whole genome duplication
<i>yrr1Δ</i>	YOR162C	Yeast Reveromycin-A Resistant	Zn2-Cys6 zinc-finger transcription factor	Zn2-Cys6 zinc-finger transcription factor; activates genes involved in multidrug resistance; paralog of Yrm1p, acting on an overlapping set of target genes; YRR1 has a paralog, PDR8, that arose from the whole genome duplication
<i>ssn2Δ</i>	YDR443C	Suppressor of SNf1	Subunit of the RNA polymerase II mediator complex	Subunit of the RNA polymerase II mediator complex; associates with core polymerase subunits to form the RNA polymerase II holoenzyme; required for stable association of Srb10p-Srb11p kinase; essential for transcriptional regulation

<i>met1Δ</i>	YKR069W	METHionine requiring	S-adenosyl-L-methionine uroporphyrinogen III transmethylase	S-adenosyl-L-methionine uroporphyrinogen III transmethylase; involved in the biosynthesis of siroheme, a prosthetic group used by sulfite reductase; required for sulfate assimilation and methionine biosynthesis
<i>met8Δ</i>	YBR213W	METHionine requiring	Bifunctional dehydrogenase and ferrochelatase	Bifunctional dehydrogenase and ferrochelatase; involved in the biosynthesis of siroheme, a prosthetic group used by sulfite reductase; required for sulfate assimilation and methionine biosynthesis
<i>cho2Δ</i>	YGR157W	CHoline requiring	Phosphatidylethanolamine methyltransferase (PEMT)	Phosphatidylethanolamine methyltransferase (PEMT); catalyzes the first step in the conversion of phosphatidylethanolamine to phosphatidylcholine during the methylation pathway of phosphatidylcholine biosynthesis
<i>spf1Δ</i>	YEL031W	Sensitivity to Pichia Farinosa killer toxin	P-type ATPase, ion transporter of the ER membrane	P-type ATPase, ion transporter of the ER membrane; required to maintain normal lipid composition of intracellular compartments and proper targeting of mitochondrial outer membrane tail-anchored proteins; involved in ER function and Ca <sup>2+</sup> homeostasis; required for regulating Hmg2p degradation; confers sensitivity to a killer toxin (SMKT) produced by Pichia farinosa KK1

<i>alg6Δ</i>	YOR002W	Asparagine-Linked Glycosylation	Alpha 1,3 glucosyltransferase	Alpha 1,3 glucosyltransferase; involved in transfer of oligosaccharides from dolichyl pyrophosphate to asparagine residues of proteins during N-linked protein glycosylation; C998T transition in human ortholog ALG6 causes carbohydrate-deficient glycoprotein syndrome type-Ic; wild-type human ortholog ALG6 can partially complement yeast <i>alg6</i> mutant
<i>mad2Δ</i>	YJL030W	Mitotic Arrest-Deficient	Component of the spindle-assembly checkpoint complex	Component of the spindle-assembly checkpoint complex; delays onset of anaphase in cells with defects in mitotic spindle assembly; forms a complex with Mad1p; regulates APC/C activity during prometaphase and metaphase of meiosis I; gene dosage imbalance between MAD1 and MAD2 leads to chromosome instability
<i>nbp2Δ</i>	YDR162C	Nap1 Binding Protein	Protein involved in the HOG (high osmolarity glycerol) pathway	Protein involved in the HOG (high osmolarity glycerol) pathway; negatively regulates Hog1p by recruitment of phosphatase Ptc1p the Pbs2p-Hog1p complex; interacts with Bck1p and down regulates the cell wall integrity pathway; found in the nucleus and cytoplasm, contains an SH3 domain and a Ptc1p binding domain (PBM)

<i>ypt6Δ</i>	YLR262C	Yeast Protein Two	Rab family GTPase	Rab family GTPase; Ras-like GTP binding protein involved in the secretory pathway, required for fusion of endosome-derived vesicles with the late Golgi, maturation of the vacuolar carboxypeptidase Y; resides temporarily at the Golgi, dissociates into cytosol upon arrival of the Rab GTPase Ypt32p, which also functions in the late Golgi; Golgi-localized form is bound to GTP, while cytosolic form is GDP-bound; homolog of the mammalian Rab6
<i>gyp1Δ</i>	YOR070C	Gtpase-activating protein for YPt1p	Cis-golgi GTPase-activating protein (GAP) for yeast Rabs	Cis-golgi GTPase-activating protein (GAP) for yeast Rabs; the Rab family members are Ypt1p (in vivo) and for Ypt1p, Sec4p, Ypt7p, and Ypt51p (in vitro); involved in vesicle docking and fusion
<i>gcn2Δ</i>	YDR283C	General Control Nonderepressible	Protein kinase	Protein kinase; phosphorylates the alpha-subunit of translation initiation factor eIF2 (Sui2p) in response to starvation; activated by uncharged tRNAs and the Gcn1p-Gcn20p complex; contributes to DNA damage checkpoint control

<i>mad1Δ</i>	YGL086W	Mitotic Arrest-Deficient	Coiled-coil protein involved in spindle-assembly checkpoint	Coiled-coil protein involved in spindle-assembly checkpoint; required for inhibition of karyopherin/importin Pse1p (aka Kap121p) upon spindle assembly checkpoint arrest; phosphorylated by Mps1p upon checkpoint activation which leads to inhibition of anaphase promoting complex activity; forms a complex with Mad2p; gene dosage imbalance between MAD1 and MAD2 leads to chromosome instability
<i>fyv10Δ</i>	YIL097W	Function required for Yeast Viability	Subunit of GID complex	Subunit of GID complex; involved in proteasome-dependent catabolite inactivation of gluconeogenic enzymes FBPase, PEPCK, and c-MDH; forms dimer with Rmd5p that is then recruited to GID Complex by Gid8p; contains a degenerate RING finger motif needed for GID complex ubiquitin ligase activity in vivo, as well as CTLH and CRA domains; plays role in anti-apoptosis; required for survival upon exposure to K1 killer toxin
<i>sap30Δ</i>	YMR263W	Sin3-Associated Polypeptide	Component of Rpd3L histone deacetylase complex	Component of Rpd3L histone deacetylase complex; involved in silencing at telomeres, rDNA, and silent mating-type loci; involved in telomere maintenance

<i>tom7Δ</i>	YNL070W	Translocase of the Outer Mitochondrial membrane	Component of the TOM (translocase of outer membrane) complex	Component of the TOM (translocase of outer membrane) complex; responsible for recognition and initial import steps for all mitochondrially directed proteins; promotes assembly and stability of the TOM complex
<i>uga2Δ</i>	YBR006W	Utilization of GABA	Succinate semialdehyde dehydrogenase	Succinate semialdehyde dehydrogenase; involved in the utilization of gamma-aminobutyrate (GABA) as a nitrogen source; part of the 4-aminobutyrate and glutamate degradation pathways; localized to the cytoplasm
<i>yk1053wΔ</i>	YLK053W		Dubious ORF	
<i>dph1Δ</i>	YIL103W	DiPHthamide biosynthesis	Protein required for synthesis of diphthamide	Protein required for synthesis of diphthamide; required along with Dph2p, Kti11p, Jjj3p, and Dph5p; diphthamide is a modified histidine residue of translation elongation factor 2 (Eft1p or Eft2p); may act in a complex with Dph2p and Kti11p
<i>sif2Δ</i>	YBR103W	Sir4p-Interacting Factor	WD40 repeat-containing subunit of Set3C histone deacetylase complex	WD40 repeat-containing subunit of Set3C histone deacetylase complex; complex represses early/middle sporulation genes; antagonizes telomeric silencing; binds specifically to the Sir4p N-terminus

\*: All descriptions are directly adapted from Saccharomyces Genome Database (1).

**Table S6:** Plasmids used in this study

Name	Description
pCH81	pRS25_ <i>P<sub>PDR5</sub></i> -HIS3- <i>T<sub>CYC1</sub></i>
pCH82	pRS25_ <i>P<sub>SNQ2</sub></i> -HIS3- <i>T<sub>CYC1</sub></i>
pCH83	pRS25_ <i>P<sub>YOR1</sub></i> -HIS3- <i>T<sub>CYC1</sub></i>

**Table S7:** Strains used in this study

Strain	Description	Source	Notes
<b>JLY30</b>	<i>MAT<math>\alpha</math>/his3<math>\Delta</math>1/his3<math>\Delta</math>1 leu2<math>\Delta</math>0/leu2<math>\Delta</math>0 LYS2/lys2<math>\Delta</math>0 met15<math>\Delta</math>0/MET15 ura3<math>\Delta</math>0/ura3<math>\Delta</math>0 <math>\Delta</math>ho::KanMX4/<math>\Delta</math>ho::KanMX4</i>	Yeast deletion collection (2)	Aka <i>ho<math>\Delta</math></i> ; <i>ho<math>\Delta</math></i> BY4743
<b>JLY31</b>	JLY30 transformed with pCH81	This study	
<b>JLY32</b>	JLY30 transformed with pCH82	This study	
<b>JLY33</b>	JLY30 transformed with pCH83	This study	
<b>JLY34</b>	<i>MAT<math>\alpha</math>, pdr1-3, ura3, his1, <math>\Delta</math>yor1::hisG, <math>\Delta</math>snq2::hisG, pdr5-<math>\Delta</math>2::hisG, <math>\Delta</math>pdr10::hisG, <math>\Delta</math>pdr11::hisG, <math>\Delta</math>ycf1::hisG, pdr3-<math>\Delta</math>2::hisG, <math>\Delta</math>pdr15::hisG, pdr1-<math>\Delta</math>3::hisG, <math>\Delta</math>ho::KanMX4</i>	This study	Aka <i>ho<math>\Delta</math></i> AD1-9
<b>JLY35</b>	<i>MAT<math>\alpha</math>, pdr1-3, ura3, his1, <math>\Delta</math>yor1::hisG, <math>\Delta</math>snq2::hisG, pdr5-<math>\Delta</math>2::hisG, <math>\Delta</math>pdr10::hisG, <math>\Delta</math>pdr11::hisG, <math>\Delta</math>ycf1::hisG, pdr3-<math>\Delta</math>2::hisG, <math>\Delta</math>pdr15::hisG, pdr1-<math>\Delta</math>3::hisG, <math>\Delta</math>mad1::KanMX4</i>	This study	Aka <i>mad1<math>\Delta</math></i> AD1-9
<b>JLY36</b>	<i>MAT<math>\alpha</math>, pdr1-3, ura3, his1, <math>\Delta</math>yor1::hisG, <math>\Delta</math>snq2::hisG, pdr5-<math>\Delta</math>2::hisG, <math>\Delta</math>pdr10::hisG, <math>\Delta</math>pdr11::hisG, <math>\Delta</math>ycf1::hisG, pdr3-<math>\Delta</math>2::hisG, <math>\Delta</math>pdr15::hisG, pdr1-<math>\Delta</math>3::hisG, <math>\Delta</math>mad2::KanMX4</i>	This study	Aka <i>mad2<math>\Delta</math></i> AD1-9
<b>JLY37</b>	<i>MAT<math>\alpha</math>, pdr1-3, ura3, his1, <math>\Delta</math>yor1::hisG, <math>\Delta</math>snq2::hisG, pdr5-<math>\Delta</math>2::hisG, <math>\Delta</math>pdr10::hisG, <math>\Delta</math>pdr11::hisG, <math>\Delta</math>ycf1::hisG, pdr3-<math>\Delta</math>2::hisG, <math>\Delta</math>pdr15::hisG, pdr1-<math>\Delta</math>3::hisG, <math>\Delta</math>mad3::KanMX4</i>	This study	Aka <i>mad3<math>\Delta</math></i> AD1-9

All other deletions strains used in this study is retrieved from the yeast deletion collection.



**Supplementary Datasets:**

**Dataset 1. Z-scores for each drug under with each biosensor construct in MSNP collection.**

**Dataset 2. Identity, fold change and significance of all samples that pass significance threshold in screenings**

## Reference

1. Cherry JM, *et al.* (2012) Saccharomyces Genome Database: the genomics resource of budding yeast. *Nucleic acids research* 40(Database issue):D700-705.
2. Giaever G, *et al.* (2002) Functional profiling of the Saccharomyces cerevisiae genome. *Nature* 418(6896):387-391.