# Supplementary Information

### Early neurogenomic response associated with variation in guppy female mate preferences

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#### This PDF file includes:

Materials and Methods Figs. S1 to S8 Tables S1 to S11 Captions for databases S1 to S2 Supplementary references

#### Other Supplementary Materials for this manuscript includes the following:

Databases S1 to S2 as zipped archives:

- 1. Databases S1: Optic tectum normalized count data for differentially expressed genes
- 2. Databases S2: Telencephalon normalized count data for differentially expressed genes

Figure S1: Methodological overview of differentially expressed gene identification. The same procedure was carried out in Preference lines and Non-preference lines in parallel. Of all significant DE genes for each pairwise comparison, we retained only those with concordant changes in expression in all replicate samples (i.e. either increase in expression between treatment 1 and treatment 2 in ALL samples as illustrated in the graph inset, or decrease in expression between treatments in ALL samples). Once we determined the final set of DE genes for each pairwise treatment comparison, we defined Preference DE genes (or Non-preference DE genes in Non-preference lines), and Social DE genes as described in the lower Venn diagrams.



Figure S2: Heatmap of normalized expression correlations between samples. Upper half of heatmap corresponds to optic tectum (n= 20396 expressed transcripts) and lower half to telencephalon (n=19571 expressed transcripts). Dendrograms illustrate sample hierarchical clustering based on sample expression distance. Outliers removed from analyses are indicated with (\*).



Telencephalon

#### Figure S3: Hierarchical gene-expression clustering of Non-preference DE genes.

Clustering of samples for Non-preference DE genes, differentially expressed between attractive and dull male treatments in Non-preference females. We found 61 Non-preference DE genes in the optic tectum and 38 Non-preference DE genes in the telencephalon. Colors below dendrogram correspond to sample treatment and line as indicated in the legend. Values on top of nodes correspond to bootstrap Approximately Unbiased p-values<sup>1</sup>, computed by multiscale bootstrap resampling (all bootstrap values >68%, some not shown for clarity).



**Figure S4: Co-expression networks.** Overview of optic tectum (A) and telencephalon (B) co-expression networks. Genes highlighted in networks correspond to Preference DE genes in red, Non-preference DE genes in grey, known preference/social behavior genes in green, synaptic plasticity genes & immediate early genes in black/black edge.



• Non-preference DE genes

SPG and IEG

Figure S5: Co-expression gene module identification dendrograms. Gene dendrograms showing the co-expression modules identified by the WGCNA dynamic tree cut function. Top colors correspond to module color labels before merging and bottom colors after merging modules whose expression profiles are very similar. Co-expression similarity between modules was estimated by calculating module eigengene correlations.



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Figure S6: Social DE genes expression change between attractive and dull treatments in Preference lines relative to Non-preference lines. Scatterplots showing two different expression patterns of Social DE genes: Scatterplots of attractive:dull log<sub>2</sub> fold-change in Preference lines (x-axis) against Nonpreference lines (y-axis) for Social DE genes in the optic tectum (left) and telencephalon (right). Darker blue points correspond to higher log<sub>2</sub> fold-change values. In the optic tectum, the outlier gene GOPC was omitted from the graph to better visualize differences across genes. For this gene,  $log_2$  fold-change in Preference lines is -4.61 and log<sub>2</sub> fold-change in Non-preference lines is 3.45. Social DE genes could either play similar functions modulating social interactions in both Preference and Non-Preference lines, in which case we would expect them to change in the same direction between the attractive and dull male treatments in both line types. We found 174 optic tectum and 73 telencephalon genes fit this pattern (Fig. S6, quadrants I and III). In contrast, we would expect Social DE genes associated with the loss of female preference phenotype in Non-preference lines would show the opposite pattern. Consistent with this expectation, we identified 213 optic tectum and 88 telencephalon genes that changed in opposite directions between the attractive and dull male treatments in both Preference and Non-Preference lines (Fig. S6, quadrants II and IV).



Figure S7: Gene expression for Transcription factors among Social DE genes known to have TF motifs among Preference DE genes. Plots showing expression differences by treatment and line for transcription factors in Social DE genes found to have regulatory motifs among Preference DE genes for optic tectum (A) and telencephalon (B). Points indicate mean for treatment group and whiskers extend to minimum and maximum values. Significant pairwise treatment comparisons (based on GLM permutations within Preference lines and Non-preference lines) denoted with (\*).



Figure S8: Transcription factors motifs enriched among Preference DE genes. Transcription factor motifs associated with Preference DE genes in the optic tectum (A) and the telencephalon (B). TRANSFAC transcription factor binding sites collection as implemented in g:Profiler was used to determine TF motifs associated with each gene (FDR multiple testing correction and a p-value<0.05). Only transcription factors relevant to social behavior, synaptic plasticity or belonging to the same protein families as genes identified in social behavior studies were included. Color of grid is proportional to the number of genes within each module with the corresponding transcription factor motif.



Transcription factor motifs

### Table S1: Identity of in-network Preference DE genes.

**OPTIC TECTUM**







### Non-preference DE genes





#### **TELENCEPHALON**



FoxO signaling pathway



 $\delta$ Compiled from g:Profiler<sup>2</sup>



#### Table S2: Chromosome enrichment.

Numbers correspond to number of Preference DE genes in each chromosome. Remaining genes map to unplaced scaffolds. P-values correspond to one-tail fisher's exact test for gene enrichment in each chromosome considering the number of total genes mapped to each linkage group. Non-significant P-values are > 0.05.









**Table S5**: List of know Synaptic Plasticity genes (SPG) and Immediate Early genes (IEG) used in this study. Compiled from<sup>3</sup>.







\*Enrichment of modules in each GO category was determined using a p-value threshold of 0.1. Color squares indicate the gene enrichment for each module: modules enriched in Preference DE genes have a red square, non-preference DE genes a dark grey square, known preference/social behavior genes a green square and synaptic plasticity genes & immediate early genes a black square. Module size corresponds to number of genes within a module after filtering out genes without genes correlations >0.4.

## Table S7: Pathways for Preference DE genes.



#### Table S8: Pathways for Social DE genes.







 $\delta$ Whether transcription factors are down-regulated or up-regulated in females exposed to an attractive vs dull male. Details on expression patterns for each transcription factor can be found in Fig. S7

#### Table S10: Assembly statistics.



#### Table S11: Samples.



Samples were arranged in non-overlapping pools of 5 individuals for RNA-Seq. Number of female brains, and corresponding pools in parenthesis, used for each treatment in Preference and Non-preference lines. (\*) One pool excluded from this treatment as a statistical outlier.

#### CAPTIONS FOR DATABASES

#### Additional Data table S1 (separate file)

Optic tectum normalized count data for differentially expressed genes

### Additional Data table S2 (separate file)

Databases S2: Telencephalon normalized count data for differentially expressed genes

#### **SUPPLEMENTARY REFERENCES**

- 1. Suzuki, R. & Shimodaira, H. Pvclust: an R package for assessing the uncertainty in hierarchical clustering. Bioinformatics 22, 1540-1542 (2006).
- 2. Reimand, J. et al. g:Profiler-a web server for functional interpretation of gene lists (2016 update). Nucleic Acids Res 44, W83-9 (2016).
- 3. Atluri, V. S. R., Kanthikeel, S. P., Reddy, P. V. B., Yndart, A. & Nair, M. P. N. Human Synaptic Plasticity Gene Expression Profile and Dendritic Spine Density Changes in HIV-Infected Human CNS Cells: Role in HIV-Associated Neurocognitive Disorders (HAND). PLoS ONE 8, e61399 (2013).
- 4. Cummings, M. E. The mate choice mind: studying mate preference, aversion and social cognition in the female poeciliid brain. Anim Behav 103, 249-258 (2015).
- 5. Ramsey, M. E., Maginnis, T. L., Wong, R. Y., Brock, C. & Cummings, M. E. Identifying Context-Specific Gene Profiles of Social, Reproductive, and Mate Preference Behavior in a Fish Species with Female Mate Choice. Frontiers in Neuroscience 6, 62 (2012).
- 6. Cummings, M. E. Looking for sexual selection in the female brain. Phil. Trans. R. Soc. Lond. B 367, 2348-2356 (2012).
- 7. Wong, R. Y. & Cummings, M. E. Expression patterns of neuroligin-3 and tyrosine hydroxylase across the brain in mate choice contexts in female swordtails. Brain Behav Evolut 83, 231-243 (2014).
- 8. Rittschof, C. C. et al. Neuromolecular responses to social challenge: Common mechanisms across mouse, stickleback fish, and honey bee. P Natl Acad Sci Usa 111, 17929-17934 (2014).
- 9. Larsen, K. B., Lutterodt, M. C., Møllgård, K. & Møller, M. Expression of the Homeobox Genes OTX2 and OTX1 in the Early Developing Human Brain. Journal of Histochemistry & Cytochemistry 58, 669-678 (2010).
- 10. Seong, E., Seasholtz, A. F. & Burmeister, M. Mouse models for psychiatric disorders. Trends in Genetics 18, 643-650 (2002).
- 11. Hamilton, S. M. et al. Multiple autism-like behaviors in a novel transgenic mouse model. Behav Brain Res 218, 29-41 (2011).
- 12. Ogura, H., Aruga, J. & Mikoshiba, K. Behavioral Abnormalities of Zic1 and Zic2 Mutant Mice: Implications as Models for Human Neurological Disorders. Behav Genet 31, 317-324 (2001).
- 13. Gerlai, R. et al. Forward Genetic Screening Using Behavioral Tests in Zebrafish: A Proof of Concept Analysis of Mutants. Behav Genet 47, 125-139 (2017).
- 14. Balamotis, M. A. et al. Satb1 ablation alters temporal expression of immediate early genes and reduces dendritic spine density during postnatal brain development. Mol. Cell. Biol. 32, 333-347 (2012).
- 15. Harrison, S. J., Nishinakamura, R., Jones, K. R. & Monaghan, A. P. Sall1 regulates cortical neurogenesis and laminar fate specification in mice: implications for neural abnormalities in Townes-Brocks syndrome. Disease Models & Mechanisms 5, 351-365 (2012).
- 16. Buechel, H. M. et al. Deep Sleep and Parietal Cortex Gene Expression Changes Are Related to Cognitive Deficits with Age. PLoS ONE 6, e18387 (2011).
- 17. Chung, S. J. et al. Genomic determinants of motor and cognitive outcomes in Parkinson's disease. Parkinsonism & Related Disorders 18, 881-886 (2012).
- 18. Lopes, J. S., Abril-de-Abreu, R. & Oliveira, R. F. Brain Transcriptomic Response to Social Eavesdropping in Zebrafish (Danio rerio). PLoS ONE 10, e0145801 (2015).
- 19. Swanberg, S. E., Nagarajan, R. P., Peddada, S., Yasui, D. H. & LaSalle, J. M. Reciprocal coregulation of EGR2 and MECP2 is disrupted in Rett syndrome and autism. Human Molecular Genetics 18, 525-534 (2009).
- 20. Poirier, R. et al. Paradoxical Role of an Egr Transcription Factor Family Member, Egr2/Krox20, in Learning and Memory. Front. Behav. Neurosci. 1, 6 (2007).
- 21. Roberts, J. L., Hovanes, K., Dasouki, M., Manzardo, A. M. & Butler, M. G. Chromosomal microarray analysis of consecutive individuals with autism spectrum disorders or learning disability presenting for genetic services. Gene 535, 70-78 (2014).
- 22. Laub, F. et al. Transcription factor KLF7 is important for neuronal morphogenesis in selected regions of the nervous system. Mol. Cell. Biol. 25, 5699-5711 (2005).