

## *Supplementary Material*

### **Amplified host defense by Toll-like receptor-mediated downregulation of the glucocorticoid-induced leucine zipper (GILZ) in macrophages**

**Jessica Hoppstädter<sup>1\*</sup>, Britta Diesel<sup>1</sup>, Rebecca Linnenberger<sup>1</sup>, Nina Hachenthal<sup>1</sup>, Sara Flamini<sup>2</sup>, Marie Minet<sup>1</sup>, Petra Leidinger<sup>3</sup>, Christina Backes<sup>4</sup>, Friedrich Grässer<sup>5</sup>, Eckart Meese<sup>3</sup>, Stefano Bruscoli<sup>2</sup>, Carlo Riccardi<sup>2</sup>, Hanno Huwer<sup>6</sup>, Alexandra K. Kiemer<sup>1</sup>**

<sup>1</sup> Pharmaceutical Biology, Department of Pharmacy, Saarland University, Saarbrücken, Germany

<sup>2</sup> Pharmacology, Department of Medicine, Perugia University, Perugia, Italy

<sup>3</sup> Human Genetics, Department of Medicine, Saarland University, Homburg, Germany

<sup>4</sup> Chair for Clinical Bioinformatics, Saarland University, Saarbrücken, Germany

<sup>5</sup> Virology, Department of Medicine, Saarland University, Homburg, Germany

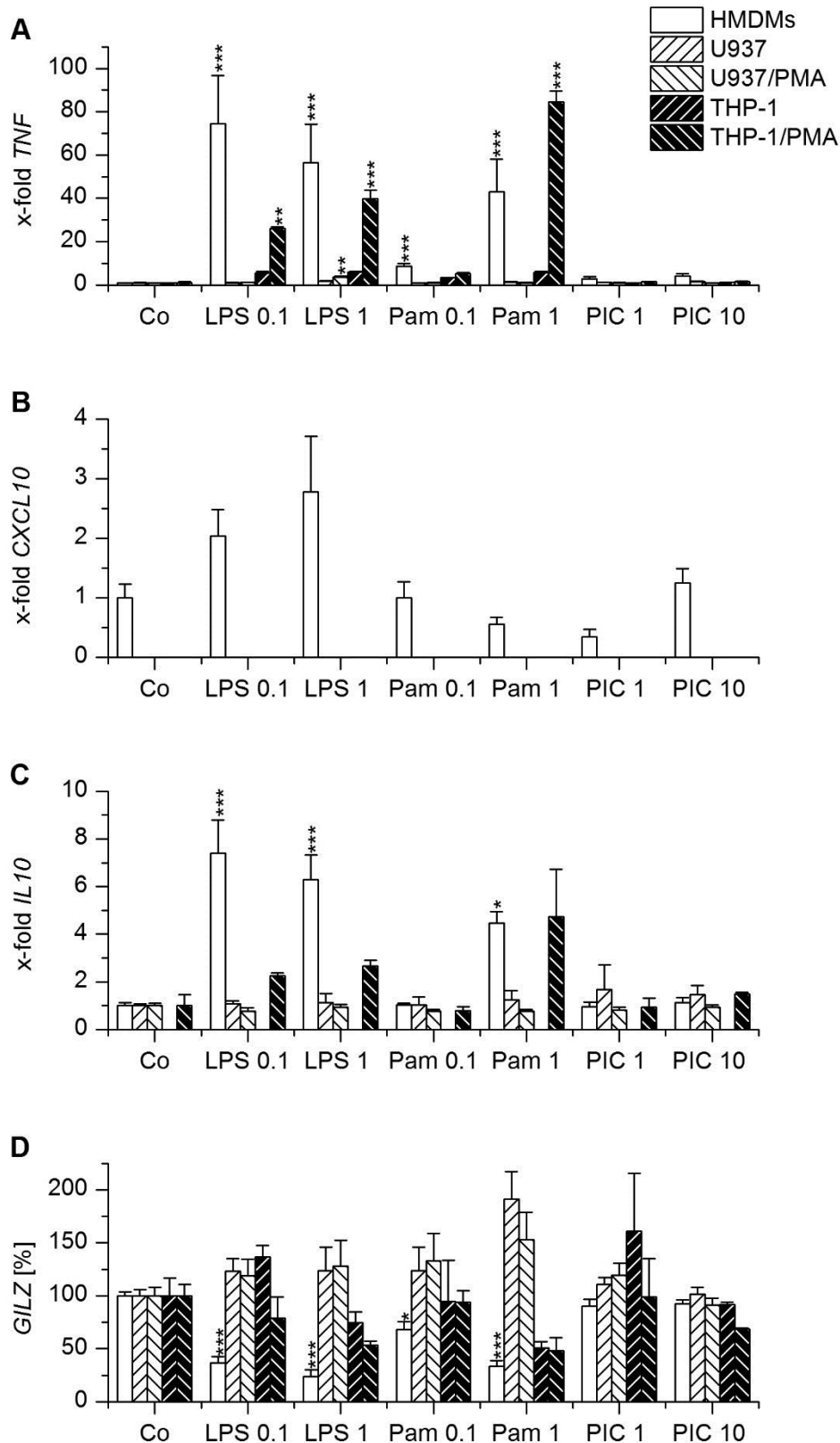
<sup>6</sup> Cardiothoracic Surgery, Völklingen Heart Centre, Völklingen, Germany

**\* Correspondence:**

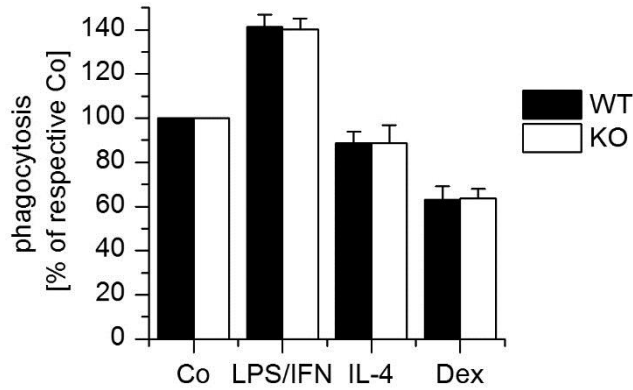
Jessica Hoppstädter, Ph.D.

[j.hoppstaedter@mx.uni-saarland.de](mailto:j.hoppstaedter@mx.uni-saarland.de)

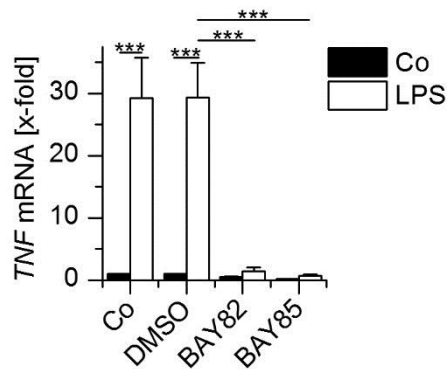
## Supplementary Figures



**Supplementary Figure 1.** Responsiveness of human macrophages and macrophage-like cells towards the TLR1/2 ligand Pam<sub>3</sub>CSK<sub>4</sub> and the TLR3 ligand Poly(I:C). Human monocyte-derived macrophages (HMDMs), as well as undifferentiated or PMA-differentiated U937 and THP-1 cells were left untreated (Co) or treated with LPS (0.1 or 1 μg/ml), Pam<sub>3</sub>CSK<sub>4</sub> (Pam, 0.1 and 1 μg/ml), or PIC (PIC, 1 and 10 μg/ml) for 2 h. and mRNA was quantified by qRT-PCR using *ACTB* as a housekeeping gene (n = 2-3, triplicates). \*p < 0.05, \*\*p < 0.01, \*\*\*p < 0.001 vs. Co.



**Supplementary Figure 2.** Different treatment schemes do not alter the effect of GILZ depletion on phagocytosis. WT and GILZ KO BMMs were left untreated (Co) or treated with LPS (1  $\mu\text{g/ml}$ ) and INF- $\gamma$  (IFN, 20 ng/ml), IL-4 (20 ng/ml) or dexamethasone (Dex, 1  $\mu\text{M}$ ) for 20 h, followed by incubation with fluorescent latex particles (diameter 1.75  $\mu\text{m}$ , 50 particles per cell) for 1 h. Particle uptake was quantified by flow cytometry. The total percentage of cells with particle-associated fluorescence in LPS/IFN, IL-4, or Dex-treated cells was normalized to the respective value for cells that were not treated otherwise prior to particle exposure.



**Supplementary Figure 3.** *NF- $\kappa$ B* inhibition. After preincubation with BAY-11-7082 or BAY-11-7085 (5  $\mu\text{M}$ , 1 h), solvent (0.1% DMSO) or medium only (Co), AMs were treated with LPS (100 ng/ml) for 2 h. *TNF* mRNA expression was determined by qRT-PCR using ACTB as a housekeeping gene (n = 3, duplicates).

## Supplementary Tables

Supplementary Table 1: PCR conditions.

Gene	Sequence (5'→3') forward primer	Sequence (5'→3') reverse primer	Probe sequence (5'FAM→3'BHQ)	Probe [nM]	MgCl <sub>2</sub> [mM]	Annealing [°C]
<i>IL6</i>	AAT AAT AAT GGA AAG TGG CTA TGC	AAT GCC ATT TAT TGG TAT AAA AAC	TCC TTT GTT TCA GAG CCA GAT CAT TTC T	100	4	57
<b>IL8 (CXCL8)</b>	TGC CAG TGA AAC TTC AAG CA	ATT GCA TCT GGC AAC CCT AC	CAG ACC CAC ACA ATA CAT GAA GTG TTG A	100	4	59
<i>TNF</i>	CTC CAC CCA TGT GCT CCT CA	CTC TGG CAG GGG CTC TTG AT	CAC CAT CAG CCG CAT CGC CGT CTC	100	3	60
<b>GILZ (TSC22D3)</b>	TCC TGT CTG AGC CCT GAA GAG	AGC CAC TTA CAC CGC AGA AC	TCC CGA ATC CCC ACA AGT GCC CGA	100	4	60
<b>β-actin (ACTB)</b>	TGC GTG ACA TTA AGG AGA AG	GTC AGG CAG CTC GTA GCT CT	CAC GGC TGC TTC CAG CTC CTC	60	4	60
<i>CXCL10</i>	GAG CCT ACA GCA GAG GAA CC	AAG GCA GCA AAT CAG AAT CG	TCC AGT CTC AGC ACC ATG AAT CAA A	60	4	60
<i>IL10</i>	CAA CAG AAG CTT CCA TTC CA	AGC AGT TAG GAA GCC CCA AG	AGC CTG ACC ACG CTT TCT AGC TGT TGA G	100	4	60
<b>TTP (ZFP36)</b>	TCG CCA CCC CAA ATA CAA	TTC GCT AGG GTT GTG GAT	no probe	N/A	N/A	60

**Supplementary Table 2: Primers used to clone miRNA overexpression vectors.**

<b>miRNA</b>	<b>forward primer ( plus EcoR1 site)</b>	<b>reverse primer (plus BamH1 or BglIII site)</b>
hsa-miR-18a	GC GAA TTC ATG GGA AGC CAA GTT GGG CTT TAA	GA GGA TCC CAC CTA TAT ACT TGC TTG GCT
hsa-miR-19a	GC GAA TTC ATG ATC CAA TAA TTC AAG CCA AGC	GA GGA TCC GCA GAT TCT ACA TCG ACA CAA
hsa-miR-19b	GC GAA TTC ATG TAG CTG TAG AAC TCC AGC TTC	GA GGA TCC GGG TTT GAG TTT CCC TTA CTT
hsa-miR-34b	GC GAA TTC ATG TAC GCG TGT TGT GCG CTG CGA	GA GGA TCC AAC CGC GGG TTT CCT CGC ACT
hsa-miR-136	GC GAA TTC ATG AGC TCT TCC ATT TCC TGG AGT	GA GGA TCC TCT GCT CTG ATT AGT TGG GCA
hsa-miR-222	GC GAA TTC ATG GAA AAT ATG TGG CAC TTT ATT	GA GGA TCC CTT AAC ACC CTA GAA CTT GAC
hsa-miR-320d1	GC GAA TTC ATG CCT TCA AAT GAC AAA ACA CAC	GA GGA TCC TTC CTT TCC TAT TTC TTT CCT
hsa-miR-484	GC GAA TTC ATG AAA ACC GAC GCC CTT CTC TCC	GA GGA TCC GTC CAC GTC ACG AGC TCA TTC