

Supplementary Information

Induction of human pluripotent stem cells into kidney tissues by synthetic mRNAs encoding transcription factors

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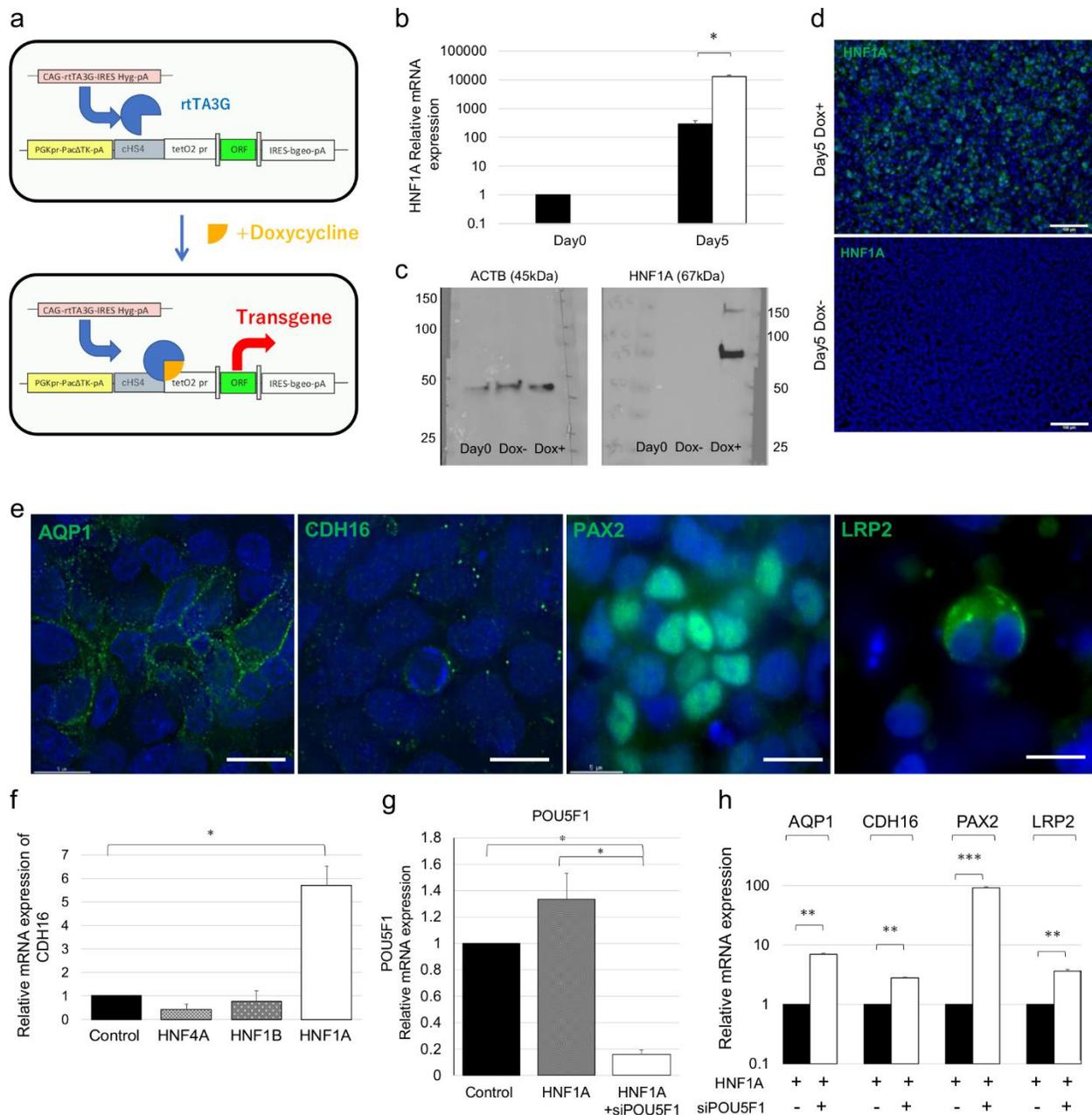
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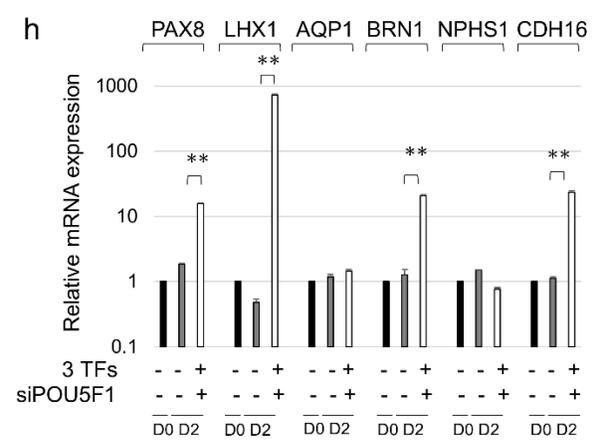
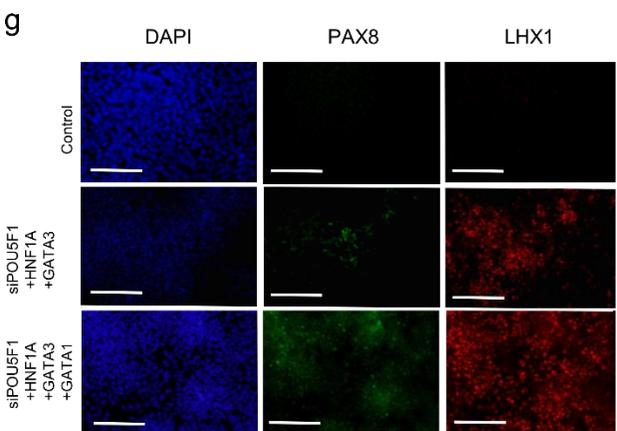
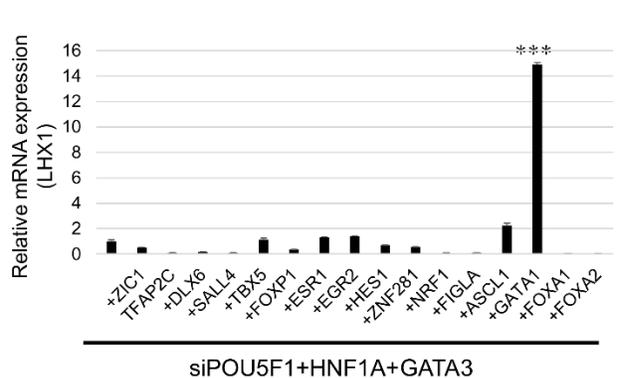
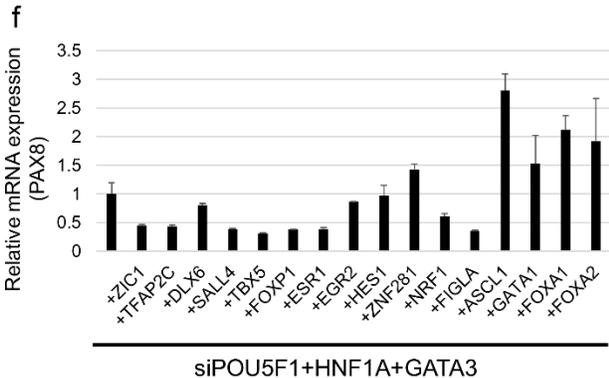
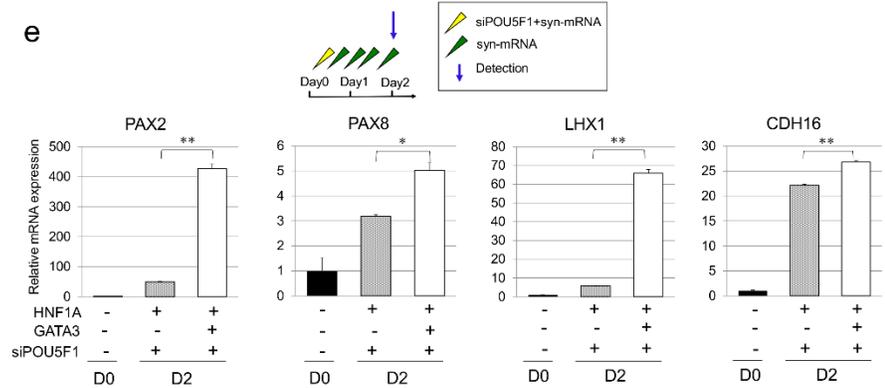
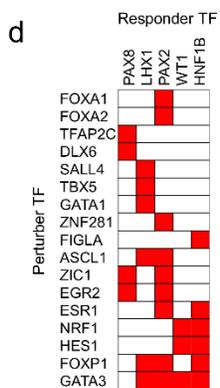
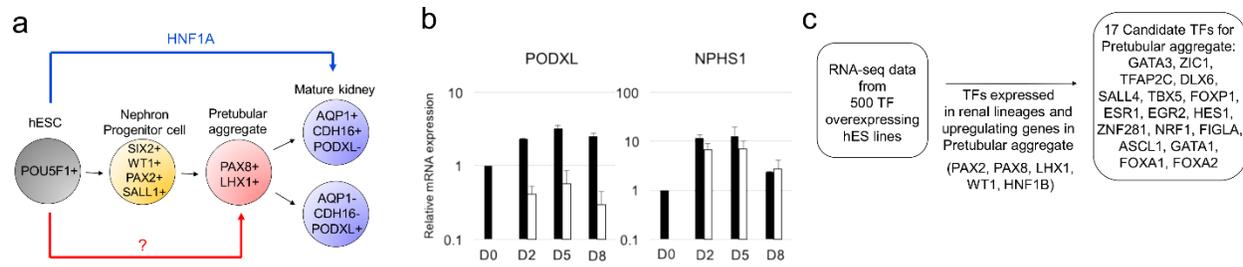
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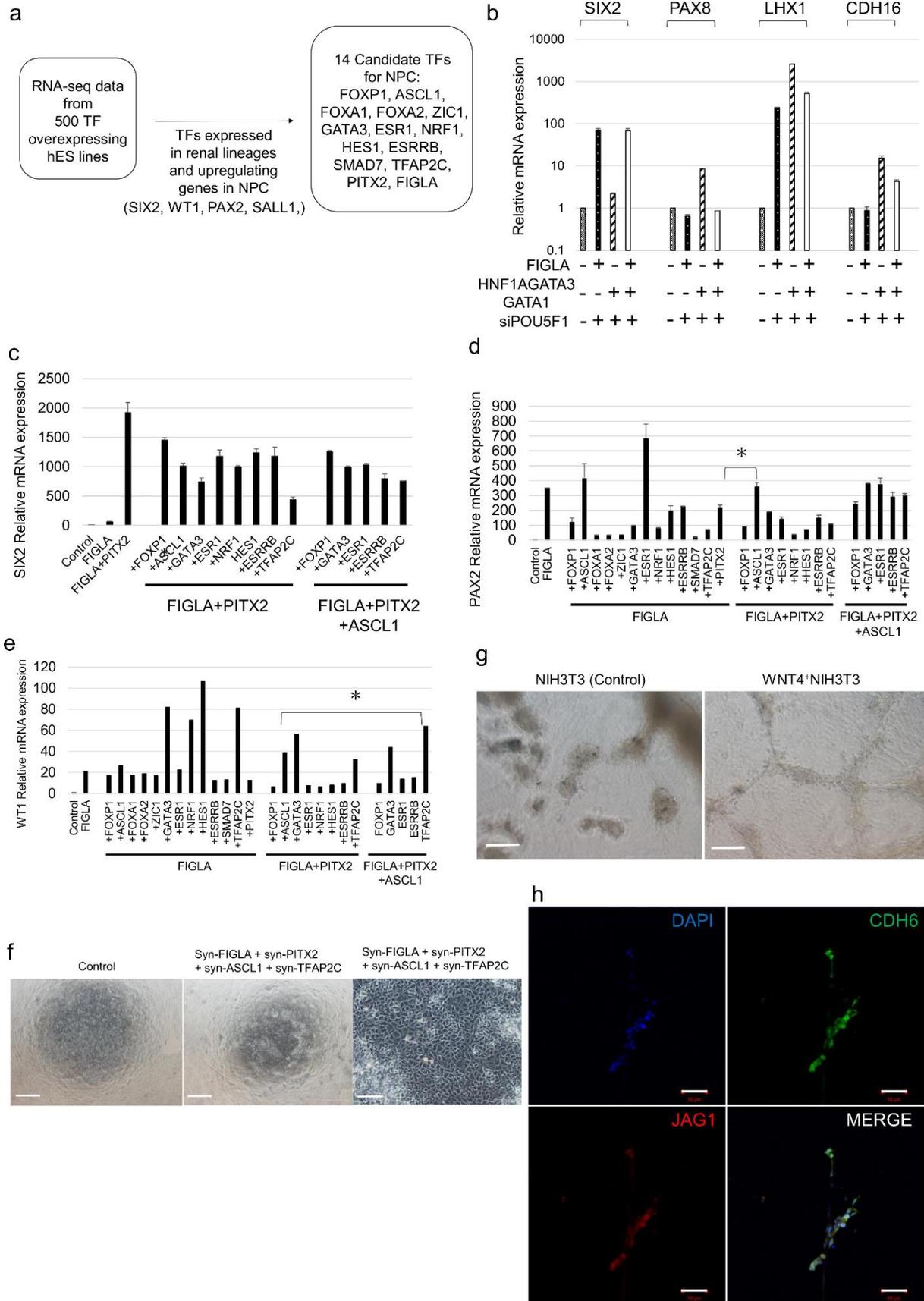
Supplementary Figure S1

(a) Establishment of tetracycline-inducible gene expression system in hESCs. (b) Relative expression levels of HNF1A was determined by real-time PCR for Day0 hESCs (black bar), Day5 transgene un-induced hESCs (black bar), and Day5 transgene HNF1A-induced hESCs (white bar). (c) Western blot analysis of HNF1A expression in Day0 hESCs (left), Day5 transgene un-induced hESCs (middle), and Day5 transgene HNF1A-induced hESCs (right). (d) Expression of HNF1A at Day5 after the activation of single transgene, HNF1A. (e) Expression of marker genes at Day5 after the activation of single transgene, HNF1A. (f) The human kidney master regulatory TF is different from that of other species. $n = 2$. Data represents mean \pm s.e.m. (g) The expression of POU5F1 was downregulated with siPOU5F1 treatment. $n = 2$. Data represent mean \pm s.e.m. (h) siPOU5F1 accelerates the differentiation of hESCs towards the renal lineage. Values shown are the mean \pm SEM. P-values were determined by a Student's t-test. * $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$.



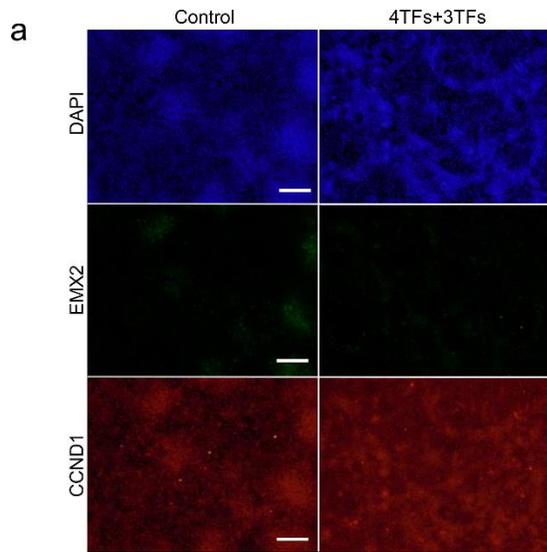
Supplementary Figure S2

(a) Diagram of differentiation into pretubular aggregate. (b) Relative mRNA expression levels of podocyte markers (PODXL, NPHS1) in transgene HNF1A-induced hESCs (white bars) and in transgene un-induced hESCs (black bars). Data represent mean +s.e.m. (c) Flowchart of an approach to identify pretubular aggregate-specific TFs. (d) 17 candidate TFs that upregulate pretubular aggregate markers. (e) Relative mRNA expression levels of pretubular aggregate markers (PAX2, PAX8, LHX1), and CDH16 after transfecting HNF1A and GATA3 mRNAs into hESCs. Data represent mean +s.e.m. (f) Relative mRNA expression levels of pretubular aggregate markers (PAX8, LHX1). The synergistic effect was observed after transfecting GATA1 in addition to HNF1A and GATA3. Expression of PAX8 and LHX1 are shown as fold induction relative to the expression levels after transfecting HNF1A and GATA3. Data represents mean +s.e.m. (g) Immunocytochemistry for PAX8 and LHX1 on day 2 of differentiation. Control indicates corresponding control cells without any TFs. Scale bar, 200 μ m. (h) Relative mRNA expression levels of pretubular aggregate markers (PAX8, LHX1), AQP1, BRN1, NPHS1, and CDH16 after transfecting HNF1A, GATA3 and GATA1 mRNAs into hESCs. Podocyte marker, NPHS1 is not upregulated after transfecting 3TFs for 2 days. Data represent mean +s.e.m.



Supplementary Figure S3

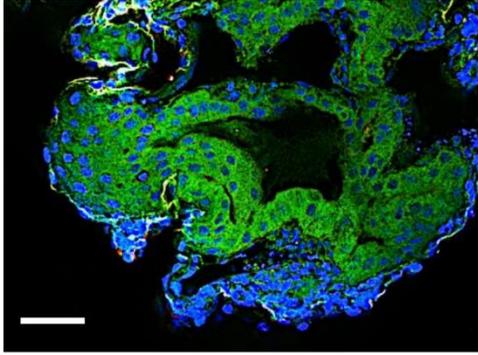
(a) Flowchart of an approach to identify NPC specific TFs. (b) Relative mRNA expression levels of SIX2, PAX8, LHX1 and CDH16 after transfecting FIGLA in addition to 3TFs. PAX8, LHX1, CDH16 were down regulated in combination with FIGLA. Data represents mean +s.e.m. (c) Relative mRNA expression levels of SIX2 after transfecting FIGLA, PITX2 and other candidate TF-encoding mRNAs into hESCs. Data represent mean +s.e.m. (d) Relative mRNA expression levels of PAX2 after transfecting FIGLA and other candidate TF-encoding mRNAs into hESCs. Data represents mean +s.e.m. (e) Relative mRNA expression levels of WT1 after transfecting FIGLA and other candidate TF-encoding mRNAs into hESCs. Data represents mean +s.e.m. (f) Representative bright-field images on day 2 of differentiation. Control indicates corresponding control cells without any TFs. Morphological changes of induced NPCs compared to control cells. Scale bar, 500 μ m (Left, Middle) 100 μ m (Right). (g) SIX2+SALL1+ cells formed branching structure in matrigel. Scale bar, 200 μ m. (h) Immunostaining showed branching structures partially expressed JAG1 and CDH6. Scale bar, 50 μ m. P-values were determined by a Student's t-test. *P < 0.05; **P < 0.01; ***P < 0.001.



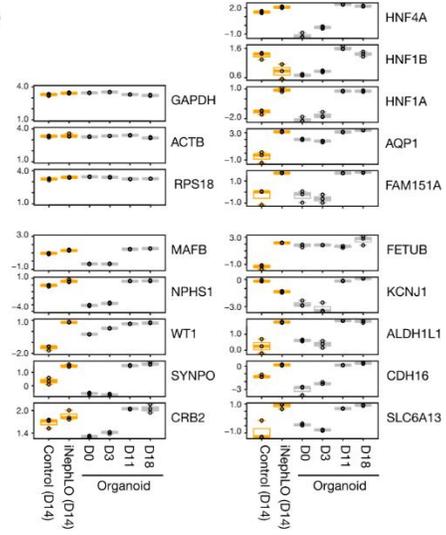
Supplementary Figure S4

Immunocytochemistry for EMX2 and CCND1, essential TFs for MET, on day 4 in cells differentiated from hPSCs using protocol depicted in; 4TFs, syn-FIGLA, syn-PITX2, syn-ASCL1, and syn-TFAP2C; 3TFs, syn-HNF1A, syn-GATA3, and syn-GATA1. Scale bar, 200 μ m.

a



b



c

Sample	Internal control										Podocytes										log10 (FPM)											
	Control (D14) rep1	Control (D14) rep2	Control (D14) rep3	NephroLO (D14) rep1	NephroLO (D14) rep2	NephroLO (D14) rep3	Organoid (D0) rep1	Organoid (D0) rep2	Organoid (D0) rep3	Organoid (D3) rep1	Organoid (D3) rep2	Organoid (D3) rep3	Organoid (D18) rep1	Organoid (D18) rep2	Organoid (D18) rep3	MAFB	NPHS1	WT1	SYNP	CRB2		ACTN	ITGB5	GALN	EHD3	CD2A	SH2D	TJP1	LAMB	ITGA8	DDN	LMO7
GAPDH	3.178	3.316	3.348	3.496	3.419	3.393	3.452	3.478	3.422	3.525	3.536	3.498	3.314	3.304	3.292	3.280	3.183	3.211	-3.00													
ACTB	0.293	0.397	0.384	0.678	0.293	0.293	0.297	0.316	0.290	0.344	0.308	0.348	0.418	0.403	0.376	0.295	0.136	0.160	-1.00													
RPP4	0.586	0.848	0.757	0.895	0.817	0.806	0.853	0.797	0.917	0.913	0.935	0.990	0.760	0.734	0.748	0.694	0.766	0.651	-1.00													
RPS18	3.134	3.310	3.250	3.425	3.410	3.357	3.446	3.454	3.424	3.415	3.449	3.379	3.256	3.227	3.252	3.274	3.283	3.277	0.00													
MAFB	0.835	0.951	0.965	1.282	1.266	1.401	-0.384	-0.325	-0.333	-0.343	-0.349	-0.302	1.445	1.463	1.447	1.553	1.523	1.529	1.00													
NPHS1	-0.031	-0.255	-0.007	0.827	0.441	0.550	-3.194	-3.013	-3.108	-2.801	-2.836	-2.666	0.587	0.638	0.584	0.707	0.573	0.625	2.00													
WT1	-1.373	-1.398	-1.736	0.819	0.868	0.816	-0.281	-0.283	-0.262	0.279	0.252	0.292	0.689	0.995	0.665	0.826	0.760	0.757	3.00													
SYNP	0.101	0.342	0.590	1.596	1.462	1.424	-0.587	-0.524	-0.443	-0.712	-0.601	-0.663	1.479	1.545	1.499	1.745	1.574	1.605	4.00													
CRB2	1.526	1.324	1.757	2.007	1.763	1.811	1.292	1.333	1.314	1.424	1.380	1.432	2.047	2.049	2.030	2.164	1.945	2.042	1.00													
ACTN4	2.299	2.403	2.458	2.707	2.517	2.514	2.386	2.409	2.379	2.207	2.204	2.222	2.560	2.587	2.553	2.899	2.732	2.778	0.00													
ITGB5	2.036	2.185	2.199	2.071	2.038	1.996	2.158	2.168	2.110	1.991	1.993	1.956	1.949	1.953	1.949	2.250	2.109	2.134	0.00													
GALNT10	1.520	1.795	1.701	2.128	2.147	2.169	2.001	2.059	2.031	1.848	1.856	1.833	1.963	1.993	1.946	2.108	1.992	2.034	0.00													
EHD3	1.238	1.403	1.394	1.500	1.354	1.394	1.507	1.530	1.446	1.644	1.644	1.642	1.336	1.325	1.283	1.267	1.283	1.169	0.00													
CD2A	1.522	1.897	1.788	1.372	1.630	1.683	1.483	1.455	1.499	1.344	1.368	1.396	1.825	1.773	1.829	1.894	1.863	2.329	0.00													
SH2D4	0.718	0.465	0.880	0.183	0.659	0.893	0.891	0.884	0.789	1.096	1.098	1.037	0.510	0.559	0.521	0.538	0.529	0.538	0.00													
TJP1	1.929	2.207	2.182	2.044	2.102	2.148	1.835	1.818	1.838	1.855	1.835	1.837	2.291	2.374	2.337	2.484	2.456	2.492	0.00													
LAMB2	1.927	1.972	1.973	2.439	2.256	2.352	1.515	1.564	1.519	1.748	1.752	1.772	2.431	2.520	2.435	2.545	2.333	2.438	0.00													
ITGA8	1.614	1.407	1.394	1.441	1.592	1.732	1.802	1.781	1.787	1.684	1.634	1.672	0.601	0.615	0.620	0.989	1.004	0.977	0.00													
DDN	-0.371	0.347	-0.008	-0.212	-1.378	-0.292	-2.845	-2.607	-2.539	-2.717	-2.353	-2.265	0.058	0.077	0.049	0.691	0.586	0.577	0.00													
LMO7	0.624	1.138	1.007	1.484	1.585	1.537	0.542	0.849	0.570	0.414	0.470	0.534	1.600	1.644	1.627	2.046	2.066	2.066	0.00													
CLDN5	-1.706	0.242	-1.146	1.697	1.482	1.315	-1.714	-1.174	-1.205	0.562	0.420	0.507	1.877	1.852	1.822	1.463	1.273	1.351	0.00													
NID2	1.195	1.709	1.114	2.644	2.512	2.598	2.271	2.283	2.244	1.784	1.801	1.788	2.259	2.310	2.278	2.417	2.323	2.398	0.00													
HNF4A	1.400	1.636	1.518	2.158	1.880	2.061	-0.846	-1.339	-1.318	-0.115	-0.229	-0.386	2.396	2.422	2.348	2.240	2.144	2.194	0.00													
HNF1B	1.216	1.443	1.413	1.058	0.557	0.828	0.682	0.669	0.730	0.786	0.819	0.846	1.585	1.678	1.578	1.847	1.320	1.408	0.00													
HNF1A	-1.192	-1.243	-1.583	1.089	0.710	0.872	-2.234	-1.816	-2.231	-1.323	-1.835	-1.770	0.785	0.820	0.739	0.862	0.711	0.773	0.00													
AQP1	-0.446	0.119	-0.296	3.620	3.628	3.506	1.930	2.099	2.192	1.811	1.579	1.791	3.198	3.311	3.082	3.998	3.842	3.979	0.00													
FAM151A	-1.167	0.068	0.032	1.782	1.715	1.723	-0.181	0.058	-0.955	-0.228	-0.974	-0.615	1.730	1.716	1.722	1.806	1.745	1.823	0.00													
SLC22A5	0.538	0.804	0.829	1.364	1.446	1.384	0.314	0.423	0.425	0.425	0.505	0.420	1.469	1.430	1.426	1.556	1.505	1.568	0.00													
SLC22A8	-1.432	-1.221	-1.304	-0.261	-0.213	0.248	-2.903	-2.785	-2.076	-2.857	-2.541	-2.714	-0.487	-0.562	-0.544	-0.950	-0.875	-0.836	0.00													
SLC22A2	-0.283	-0.189	0.003	-0.202	-0.244	0.196	-0.700	-0.482	-1.277	-1.311	-1.590	-1.399	0.194	0.184	-0.032	0.534	0.776	0.911	0.00													
SLC6A2	-1.557	-1.377	-0.306	0.915	0.933	0.675	-0.119	0.159	-0.118	0.007	-0.191	0.162	0.471	0.917	0.768	1.209	1.188	1.382	0.00													
FETUB	-1.261	-1.112	-1.726	2.153	2.163	2.197	1.987	1.519	1.953	1.815	1.850	1.855	1.426	1.391	1.393	1.814	1.816	2.296	0.00													
SLC12A1	-1.117	-0.280	-1.660	1.224	-1.125	-1.763	-3.886	-3.456	-3.523	-2.924	-2.996	-3.036	-0.874	-0.844	-0.917	0.820	-0.468	-0.498	0.00													
KCNJ1	-0.061	-0.226	-0.194	-1.352	-1.280	-1.470	-2.842	-2.903	-2.332	-3.523	-3.456	-2.559	-0.181	-0.193	-0.134	0.106	0.086	0.074	0.00													
ALDH1L1	0.249	0.704	-0.194	1.788	1.772	1.727	0.560	0.665	0.618	0.584	0.384	0.157	1.845	1.858	1.879	1.839	1.730	1.833	0.00													
CDH16	-1.396	-1.108	-1.382	0.139	0.048	0.197	-2.670	-3.745	-2.759	-2.062	-2.273	-2.288	0.113	0.103	0.074	0.459	0.280	0.336	0.00													
SLCA41	-1.226	-1.330	-1.506	-1.100	-0.215	-1.681	-1.381	-2.465	-2.116	-1.428	-0.764	-1.398	-1.186	-1.518	-1.189	-0.970	-0.247	-0.321	0.00													
SLC12A3	-1.489	-1.177	-1.557	-1.298	-1.158	-0.294	-1.559	-1.892	-1.941	-1.189	-1.224	-1.160	-0.772	-1.280	-0.775	-1.566	0.164	0.077	0.00													
CALB1	0.661	0.855	0.956	0.201	-1.134	0.892	0.500	0.739	-0.301	0.453	0.163	-0.414	0.671	-0.418	0.391	1.212	1.537	1.208	0.00													
ATP9B	0.117	0.456	0.338	0.102	0.837	0.199	-0.650	-0.318	-0.739	-0.303	0.140	0.028	0.784	0.929	0.910	0.376	0.577	1.026	0.00													
SLC6A13	-1.501	-1.288	-0.138	0.849	0.968	0.975	-0.482	-0.450	-0.351	-0.798	-0.777	-0.851	0.692	0.681	0.675	0.942	0.860	0.914	0.00													
PTH1R	-1.479	-1.496	-1.660	1.188	1.157	1.348	-0.128	-0.282	-0.333	0.471	0.354	0.457	1.710	1.696	1.694	2.272	2.125	2.146	0.00													
EMX1	-1.208	-1.272	-0.233	-1.206	0.159	-1.409	-2.759	-2.783	-2.796	-2.991	-2.955	-2.965	-0.851	-0.892	-0.823	-0.551	-0.816	-0.919	0.00													
CLCA4	-0.040	0.057	0.308	0.493	0.067	0.024	-1.288	-0.475	-0.504	-0.760	-0.789	-1.172	0.672	0.511	0.458	0.843	0.835	0.840	0.00													

Sample	Internal control										Proximal tubules										Distal tubules										log10 (FPM)												
	GAPD	ACTB	RPP4	RPS1	O	ITGB5	NPHS1	WT1	SYNP	CRB2	ACTN	ITGB5	GALN	EHD3	CD2A	SH2D	TJP1	LAMB	ITGA8	DDN	LMO7	CLDN	NID2	HNF4A	HNF1B	HNF1A	AQP1	FAM151A	SLC2	SLC2		SLC3	FETU	SLC1	KCNJ	ALDH	CDH1	SLC4	SLC1	CALB	ATP9	SLC6	PTH1
Control (D14) rep1	3.18	3.20	3.59	3.13	0.84	-0.03	-1.37	0.10	1.53	2.30	2.04	1.52	1.24	1.52	0.72	1.93	1.93	1.16	-0.37	0.82	-1.70	1.20	1.26	-1.12	-0.06	0.25	-1.40	-1.23	-1.41	0.68	0.11	1.40	1.48	-1.27	-0.04	0.00							
Control (D14) rep2	3.32	3.34	3.85	3.31	0.95	-0.26	-1.40	0.34	1.72	2.40	2.18	1.80	1.40	1.70	0.46	2.21	1.97	1.41	0.35	1.14	0.24	1.71	-1.11	-0.26	-0.23	0.70	-1.11	-1.33	-1.19	0.85	0.46	-1.27	1.50	-1.27	0.06	0.00							
Control (D14) rep3	3.35	3.38	3.76	3.25	0.99	-0.01	-1.74	0.59	1.76	2.48	2.20	1.70	1.39	1.79	0.88	2.19	1.97	1.39	-0.01	1.01	-1.15	1.11	2.13	-1.22	-1.35	1.79	0.14	-1.10	-1.24	-0.20	0.10	0.65	1.19	-1.21	0.49	0.00							
NephroLO (D14) rep1	3.50	3.68	3.70	3.42	1.27	0.44	0.87	1.46	1.76	2.52	2.04	2.16	1.35	1.63	0.66	2.10	2.26	1.59	-1.38	1.59	1.48	2.51	2.16	-1.12	-1.28	1.77	0.05	-0.22	-1.16	-1.13	0.84	0.97	1.16	0.16	0.07	0.00							
NephroLO (D14) rep2	3.42	3.29	3.82	3.41	1.40	0.55	0.82	1.42	1.81	2.51	2.00	2.17	1.39	1.68	0.89	2.15	2.35	1.73	-0.29	1.54	1.32	2.60	2.20	-1.76	-1.47	1.73	0.20	-1.68	-0.29	0.08	0.20	0.97	1.35	-1.41	0.32	0.00							
NephroLO (D14) rep3	3.50	3.68	3.70	3.42	-0.38	-3.19	-0.26	-0.59	1.29	2.39	2.16	2.06	1.51	1.46	0.89	1.84	1.51	1.80	-2.84	0.54	-1.71	2.27	1.44	0.59	0.18	1.48	2.05	2.56	1.96	1.24													

Supplementary Figure S5

(a) Immunocytochemistry for proximal tubule (LTL) that shows tubule-like structures on day 14. Scale bar, 50 μ m. (b) Principal component analysis for other human organs, corresponding control cell organoids (shown as Control (D14)) and iNephLOs (shown as iNephLO (D14)), kidney organoids reported previously (GSE70101, shown as organoid (D0, D3, D11, and D18) viewed from different directions. (c) Direct comparison between corresponding control cell organoids (shown as Control (D14)) and iNephLOs (shown as iNephLO (D14)), kidney organoids reported previously (GSE70101, shown as organoid (D0, D3, D11, and D18). Segment-specific makers are normalized by internal controls (GAPDH, ACTB, PRP40, RPS18). ; Podocyte markers; MAFB, NPHS1, WT1, SYNPO, CRB2, ACTN4, ITGB5, GALNT10, EHD3, CD2AP, SH2D4A, TJP1, LAMB2, ITGA8, DDN, LMO7, CLDN5, and NID2; Proximal tubular markers; HNF4A, HNF1B, HNF1A, AQP1, FAM151A, SLC2A5, SLC22A8, SLC22A2, and SLC36A2; Distal tubular markers; FETUB, SLC12A1, KCNJ1, ALDH1L1, CDH16, SLC4A1, SLC12A3, CALB1, ATP6V0D2, SLC6A13, PTH1R, EMX1, and CLCNKA.

Supplementary video 1

The three-dimensional (3D) image stacks were further processed using Imaris 3D software. It shows multi-segmented kidney structures containing podocytes (PODXL), proximal tubules (LTL), and distal tubules (CDH1) by day 14.

Supplementary data 1

Principal component analysis of transcriptome data obtained from various human organs, corresponding control cell organoids (shown as Control (D14)) and iNephLOs (shown as iNephLO (D14)), kidney organoids reported previously (GSE70101, shown as organoid (D0, D3, D11, and D18).

Supplementary Table S1. List of gene accession numbers

Transcription factor	Accession Number
FIGLA	BC148798
PITX2	BC013998
ASCL1	NM_004316.3
TFAP2C	BC051829
HNF1A	EU446660.1
GATA3	BC006793.1
GATA1	AB463674
EMX2	AB463605

Supplementary Table S2. List of primary antibodies

Antibody	Source	Catalog or clone number	Dilution
AQP1	Santa cruz	H-55	1:100
CDH16	home made		1:50
PAX2	BioLegend	PRB-276P	1:500
LRP2	kindly provided by Dr. Michiko Sekine, Tokyo Metropolitan Institute of Medical Science		1:100
PAX8	Proteintech	10336-1-AP	1:500
LHX1	Developmental Studies Hybridoma Bank	4F2-c	1:50
SIX2	Proteintech	11562-1-AP	1:500
SALL1	R&D systems	PP-K9814-00	1:100
EMX2	Abnova	H00002018-M02	1:100
CCND1	Abcam	ab134175	1:50
CDH1	Abcam	ab11512	1:100
CDH6	Atlas antibodies	HPA007456	1:500
JAG1	Santa cruz	C-20	1:50
CDH2	Abcam	ab98952	1:100
PNA	Vector laboratories	RL-1071	1:200
PODXL	R&D systems	AF1658	1:500
BRN1	Santa cruz	C-17	1:100
LTL	Vector laboratories	B-1325	1:200
THP	Alfa aesar	J65429	1:150
KIM1	R&D systems	AF-1750	1:200
HNF1A	Abcam	ab96777	1:100

Supplementary Table S3. Primer sequences

Gene	Forward	Reverse
ACTB	CAATGTGGCCGAGGACTTTG	CATTCTCCTTAGAGAGAAGTGG
CDH16	GGTACAGGTCAAGGACATGGG	TGCGGGTATAGGACTTTGAGAT
AQP1	TCTCTGTAGCCCTTGGACACC	GCCAGGATGAAGTCGTAGATGAG
PAX2	GCAACCCCGCCTTACTAAT	AACTAGTGGCGGTCATAGGC
LRP2	TGTGATGCAGCCATCGAACT	TGCATTTGGGGAGGTCAGTC
POU5F1	TGGAGTTTGTGCCAGGGTTT	CTTCACCTCCCTCCAACCA
PODXL	GCAGAGAGGGCAAGAGTAAAACG	GAGTCATCTGTGTCCTCCATGCT
NPHS1	CGCAGGAGGAGGTGTCTTATTC	CGGGTTCCAGAGTGTCCAAG
PAX8	CCCCCTACTCCTCCTACAGC	GGCCTTGATGTGGAAGTGTAA
LHX1	ATGCAACCTGACCGAGAAGT	CAGGTCGCTAGGGGAGATG
BRN1	TGCCCAAAGGAAAGATGTC	CATTCTCCTCGCAACAGTCA
SIX2	AGGAAAGGGAGAACAACGAGAA	GGGCTGGATGATGAGTGGT
WT1	GGCAGCACAGTGTGTGAACT	CCAGGCACACCTGGTAGTTT
SNAI2	CTGCGGCAAGGCGTTTTCCAGA	CAGATGAGCCCTCAGATTTGAC
VIMENTIN	ACACCCTGCAATCTTTCAGACA	GATTCCACTTTGCGTTCAAGGT
HNF1B	AGAAGCGTGCCGCTCTGT	TGGTTGAATTGTCGGAGGATCT
HNF1A	CCCACCAAGCAGGTCTTCAC	AAGGTCTCGATGACGCTGTG