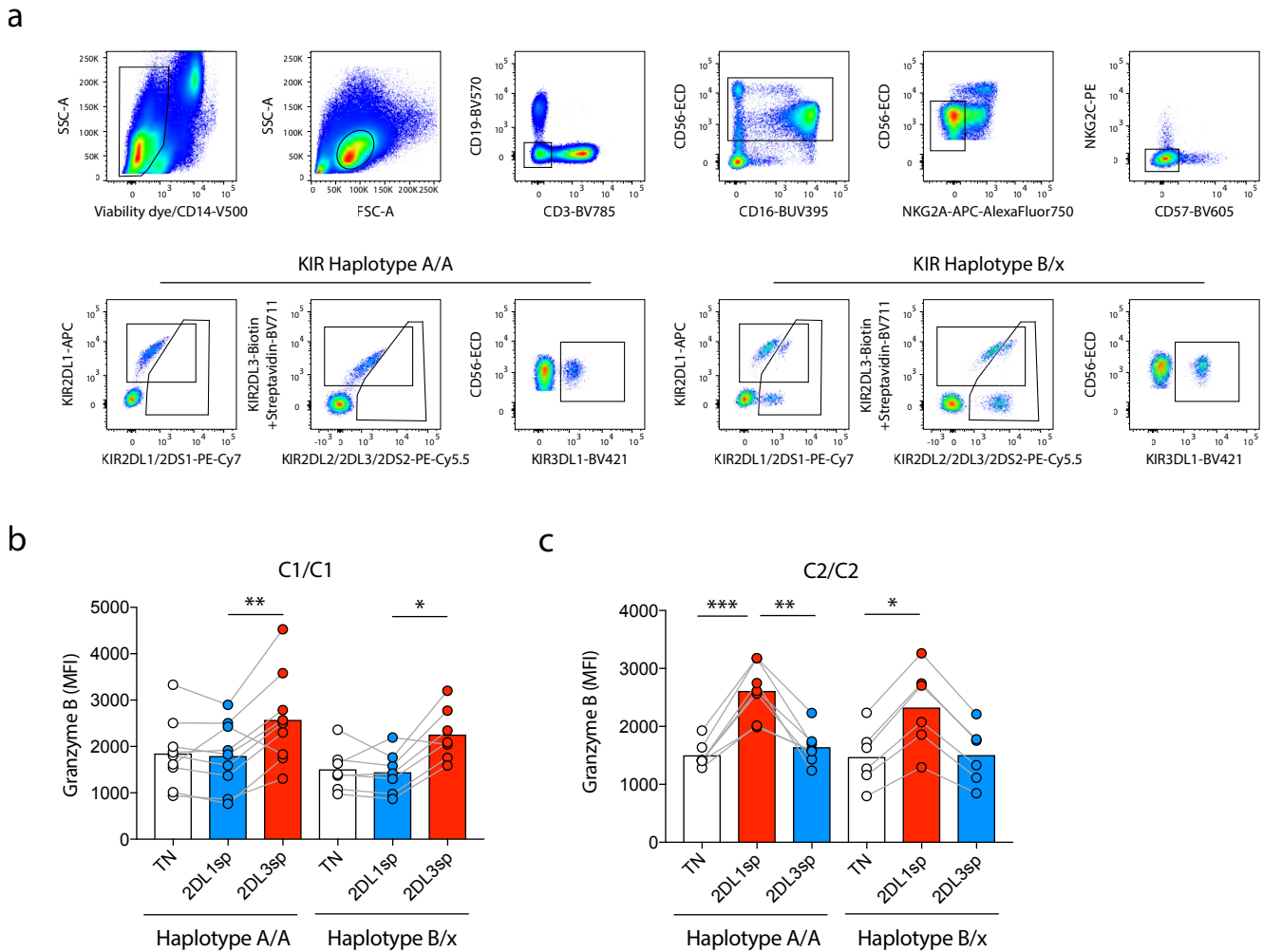


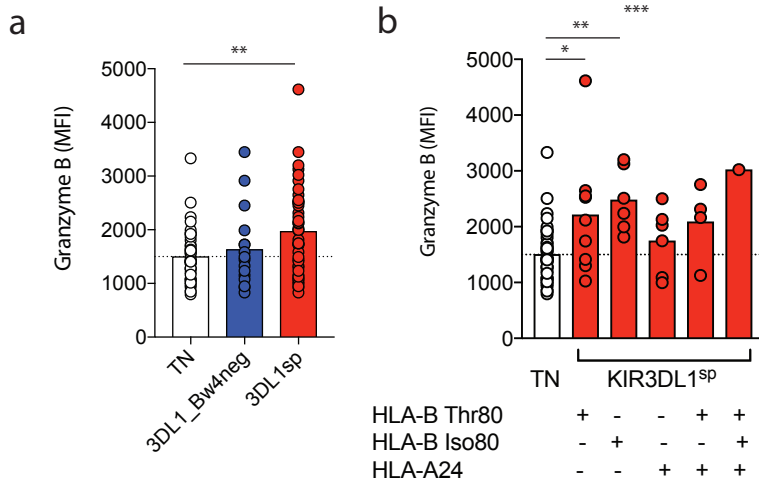
Supplementary Information

Remodeling of Secretory Lysosomes During Education Tunes Functional Potential in NK Cells

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Johan Malmberg^{1,2,3}



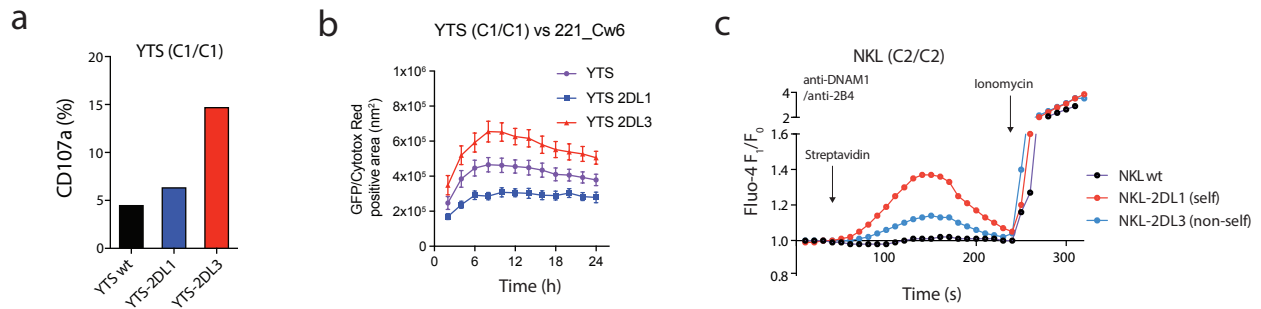
Supplementary Figure 1. Gating scheme and analysis of granzyme B expression in haplotype A/A and haplo-type B/x donors. (a) Example of gating scheme for analysis of expression of effector molecules in discrete NKG2A-CD57- NK cell subsets expressing single KIR. (b-c) Expression of granzyme B in NK cells triple negative (TN) for 2DL1, 2DL2/3, 3DL1 or expressing either of these KIR as their only KIR. Stratified analysis of donors with haplotype A/A and B/x in **(b)** HLA-C1/C1 and **(c)** HLA-C2/C2 donors. Red and blue colors indicate expression of self and non-self KIR, respectively. Friedman's test followed by Dunn's multiple comparison test were performed in panels b and c. **: $p < 0.001$; ** $p < 0.01$; and * $p < 0.05$.



Supplementary Figure 2. Granzyme B expression stratified based on KIR3DL1 and HLA-Bw4 ligands.

(a) Expression of granzyme B in the indicated NKG2A-CD57- NK cell subset from Bw4+ (n=26) and Bw4- donors (n=16). TN=triple negative for 2DL1, 2DL3 and 3DL1. 3DL1sp=3DL1 single-positive.

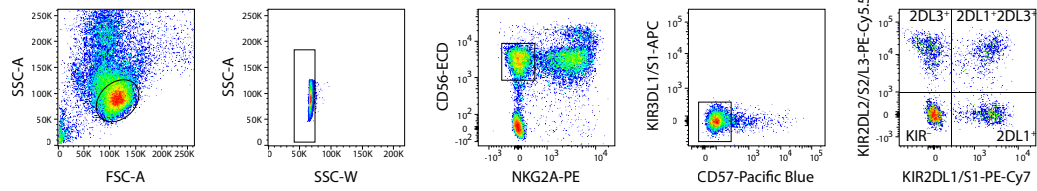
(b) Substratification showing expression of granzyme B in 3DL1sp NK cells based on the Bw4 ligands Bw4^{Thr80} (n=9), Bw4^{Iso80} (n=6), Bw4^{A24} (n=6), Bw4^{Thre80+A24} (n=4) and Bw4^{Iso80+A24} (n=1) were too rare to be analysed separately. One-way ANOVAs followed by Tukeys' multiple comparison test were performed in panels a and b. ***; p < 0.001; **p < 0.01; and *p < 0.05.



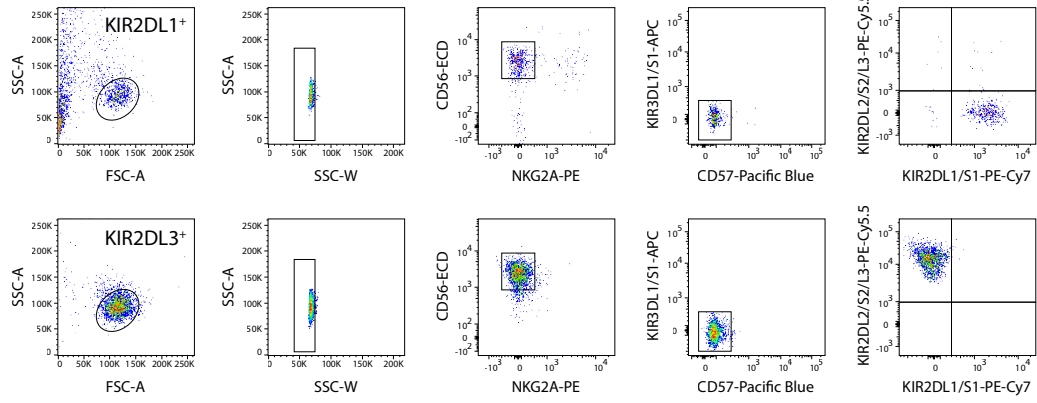
Supplementary Figure 3. Functional responses in KIR-transduced NK cell lines. (a) Degranulation in YTS cells transduced to express self (2DL3) and non-self (2DL1) KIR. (b) Long-term cyto-toxicity assay in the Incucyte showing killing (GFP+Cytotox Red+ area) of 221.Cw6.GFP+ cells by the KIR engineered YTS lines E:T ratio 1:1. Bars represent SD of triplicates. (c) Ca²⁺-flux in NKL cells transduced with self (2DL1) and non-self (2DL3) KIR in response to crosslinking of DNAM-1/2B4. Graphs in panels a-c show one representative of two experiments.

a

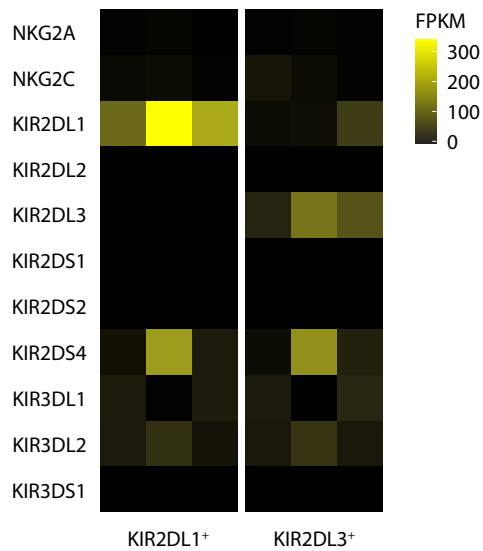
Before sort



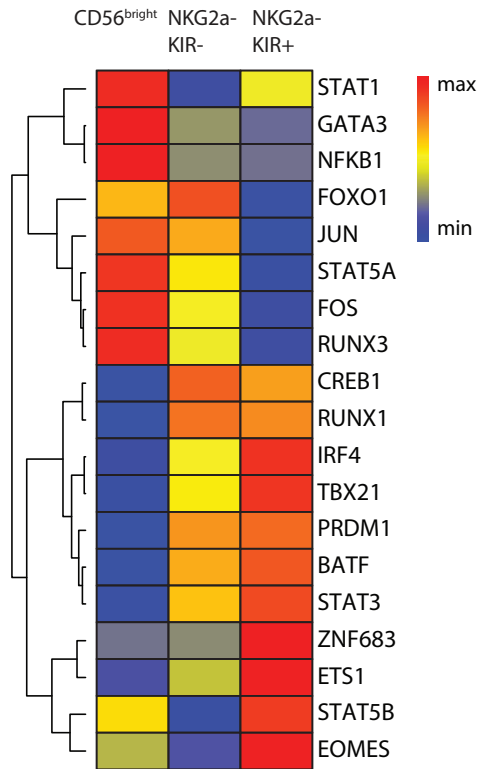
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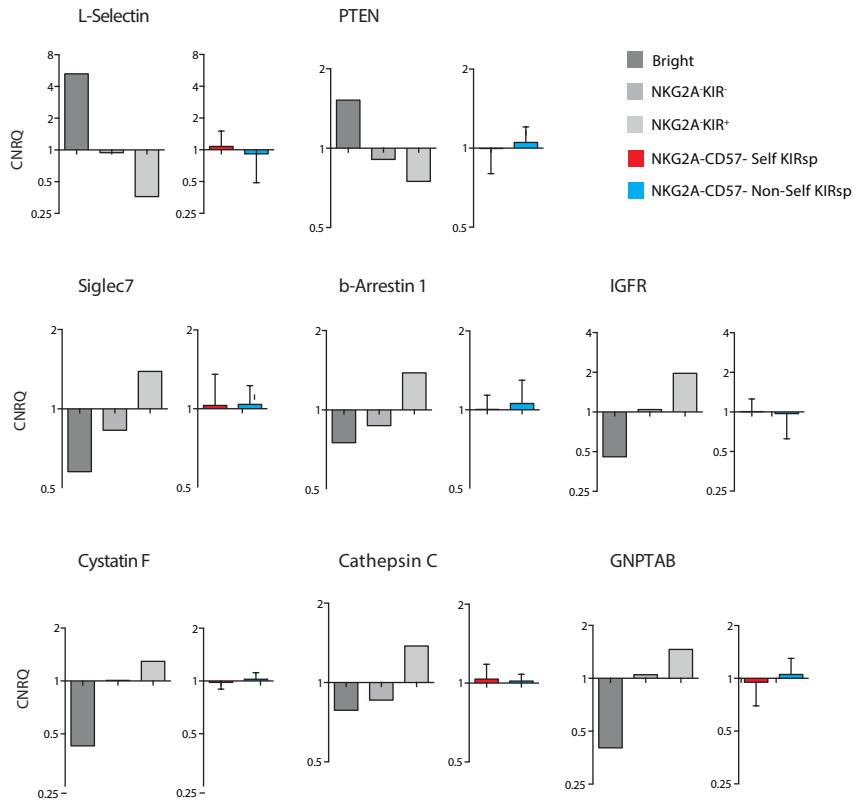
b



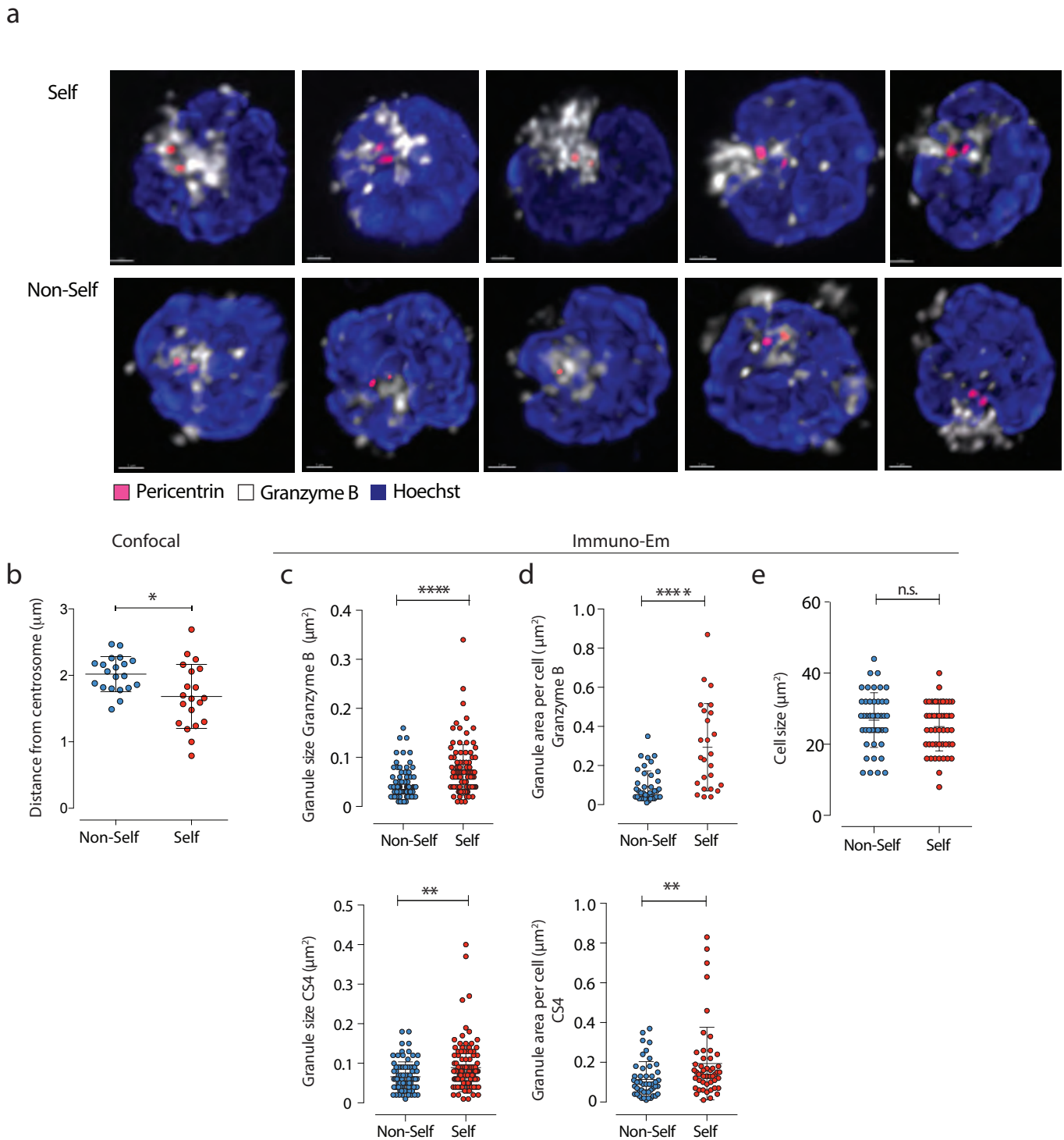
Supplementary Figure 4 .Purity of sorted subsets. (a) Representative example of NK cell phenotypes before and after flowcytometry-based sort from an haplotype A C1/C1 donor. **(b)** RNA-Seq of sorted 2DL1⁺ (left) or 2DL3⁺ (right) NKG2A⁻NKG2C⁻CD57⁻CD56^{dim} NK cell subsets. Data are from three representative donors. FPKM; fragments per kilobase of exon per million reads mapped.



Supplementary Figure 5. Transcriptional regulation of NK cell differentiation. Global RNA-Seq of sorted CD56^{bright}, NKG2A⁻panKIR⁻ and NKG2A⁻panKIR⁺ CD56^{dim} NK cell subsets. Shown are genes involved in the regulation of cytokine responsiveness and transcription of effector molecules, including gran-zyme B. Data are from one representative donor out of three independent donors.



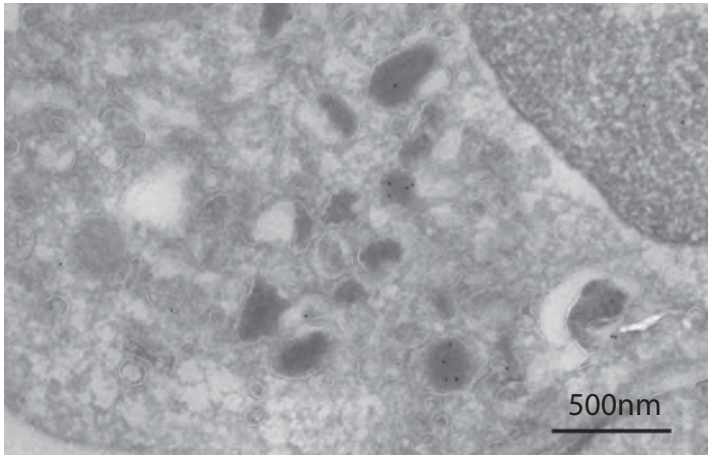
Supplementary Figure 6. Transcriptional dissociation of differentiation and education. Quantitative PCR of mRNA of the indicated markers in sorted CD56^{bright}, NKG2A^{pan}KIR⁻ and NKG2A^{pan}KIR⁺ CD56^{dim} NK cell subsets (left, 4 pooled samples) as well as in sorted NKG2A⁻CD57⁻ CD56^{dim} NK cells expressing self or non-self KIR (right, n=5 paired samples). Markers showing a positive relationship with differentiation were selected for comparison with education subsets. Whiskers show 5th to 95th percentile.



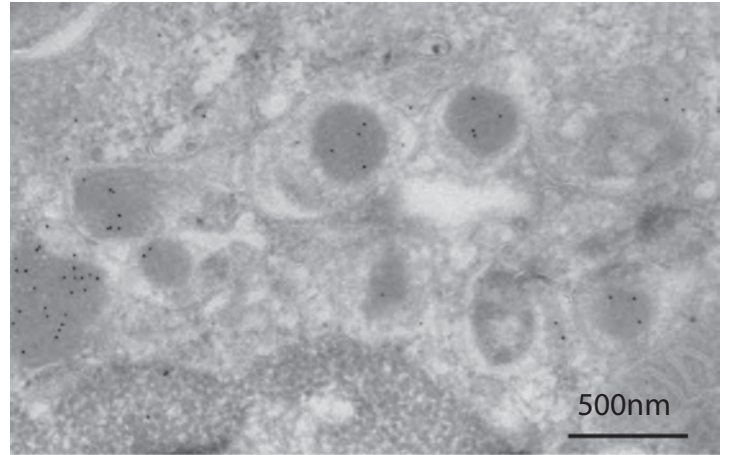
Supplementary Figure 7. Expression of self-KIR is associated with accumulation of large granzyme B dense secretory lysosomes. (a) Confocal Z-stacks showing Pericentrin (PCNT) in red and granzyme B staining in white in sorted CD56dim NKG2A-CD57- NK cells expressing Self (top) or Non-Self KIR (bottom). A panel of representative phenotypes acquired from one representative donor out of 5 is shown. (b) Average distance from the centrosome of the brightest lysosomal structures (top 25%) in confocal microscopy, n=20 cells per condition. (c) Granule size as determined by using grid overlay of electron dense structures staining positive for granzyme B (upper panel) and Chondroitin Sulphate-4 (CS4) (lower panel). (d) Granule area per cell as determined by taking total calculated granule area per cell for structures staining positive for granzyme B (upper panel) and CS-4 (lower panel). (e) Comparison of cell size between Self (n=50) and Non-Self (n=49) KIR+ NK cells. Whiskers show 5th to 95th percentile. Bars show the median. Non-parametric Mann-Whitney tests were performed in panels b-e. ****p < 0.0001; **p < 0.01; and *p < 0.05

a

Granzyme B



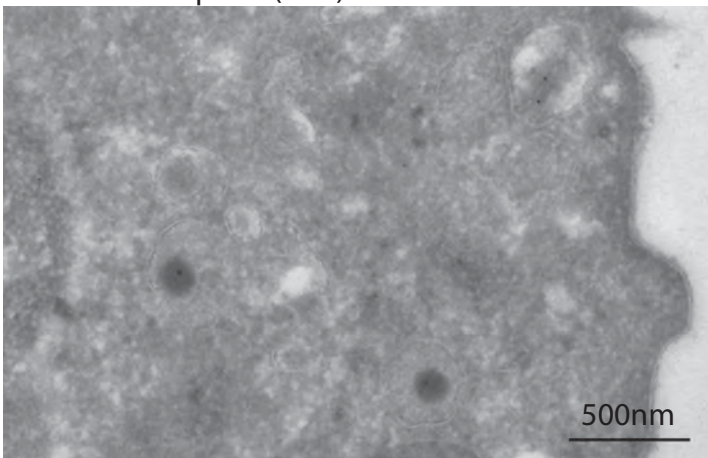
Non-Self



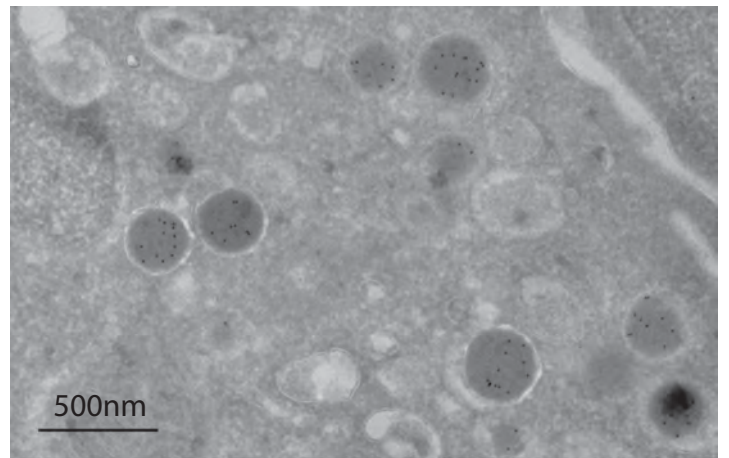
Self

b

Chondroitin Sulphate (CS-4)

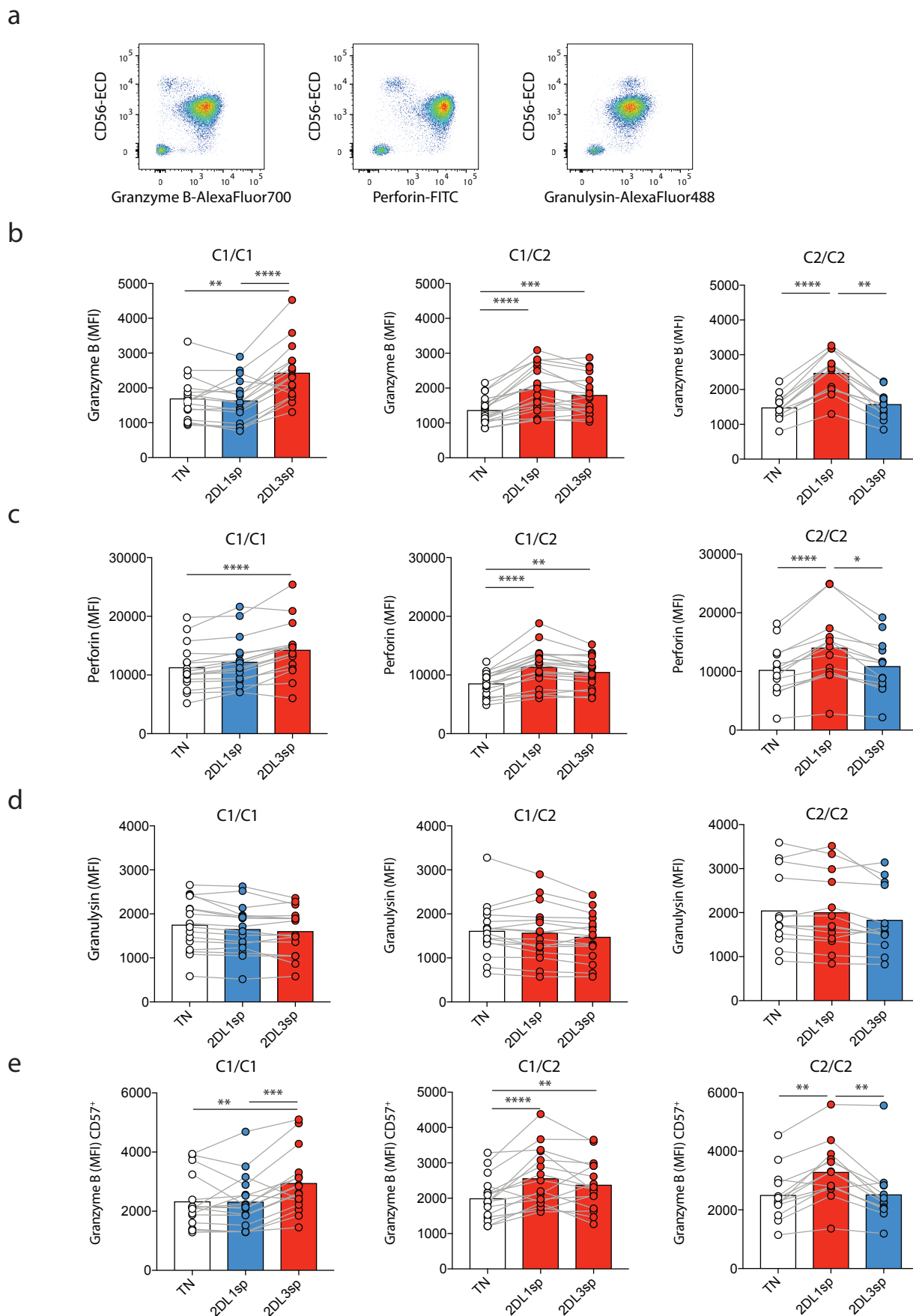


Non-self

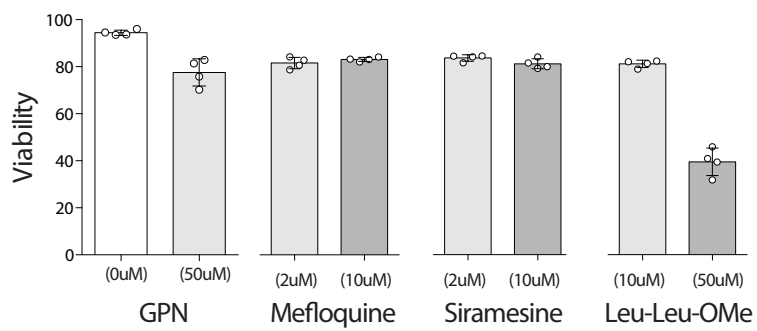


Self

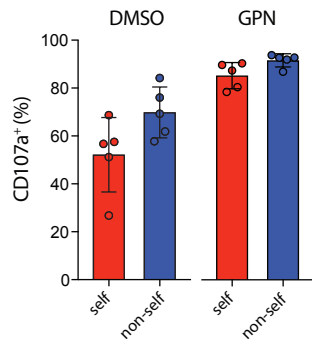
Supplementary Figure 8. Immuno-EM of sorted self/nonself KIR⁺NKG2A⁻CD57⁻ CD56^{dim}NK cells. Representative immuno-EM sections showing staining with gold-particle coated anti-granzyme B (a) and chondroitin Sulphate-4 (CS-4) (b) of sorted CD56^{dim} NKG2A⁻CD57⁻ NK cells expressing non-self (left) or self KIR (right). Immuno-EM images are from sections of representative cells from n=5 donors and n=5 experiments.



Supplementary Figure 9. NK cell education through self-KIR is associated with accumulation of granzyme B and perforin but not granulysin. (a) Representative examples showing expression of granzyme B, perforin and granulysin in CD56^{bright} and CD56^{dim} NK cells. Expression of (b) granzyme B (c) perforin and (d) granulysin in NKG2A⁻CD57⁻ NK cells expressing the indicated KIR in C1/C1 (n=16), C1/C2 (n=18) and C2/C2 (n=13) donors. KIR2DL1 single-positive (2DL1sp). (e) Expression of granzyme B in NKG2A⁻CD57⁺ NK cells expressing the indicated KIR in C1/C1 (n=16), C1/C2 (n=18) and C2/C2 (n=13) donors. Friedman's test followed by Dunn's multiple comparison test were performed in panels b-e. In panel d, there was no statistical accumulation of granulysin in any of the KIR⁺ subsets. ****p < 0.0001***; p < 0.001; **; p < 0.01 and *p < 0.05.



Supplementary Figure 10. Viability of NK cells exposed to lysosomotropic compounds. Viability was determined by titration of each of the indicated compounds during 4 hours by monitoring incorporation of a live/dead marker for cell viability using flow cytometry. Shown are the mean of 4 replicates. Whiskers show standard deviation.



Supplementary Figure 11 Compromising lysosomal activity increases the functional response PMA/Ionomycin stimulation. Resting NK cells were treated with PMA (1ng/mL) and Iono-mycin (0.5 μ M) for 4 hours in the presence of DMSO (control) or GPN at 50 μ M (n=5). Whiskers show standard deviation.

Supplementary Table 1

Effector Transcription	mTOR Signaling			
<i>NFATC1</i>	<i>AKT3</i>	<i>ATP6V1H</i>	<i>SKP2</i>	<i>STRADA</i>
<i>RUNX1</i>	<i>RRAGB</i>	<i>PDPK1</i>	<i>SLC3A2</i>	<i>ATP6V1F</i>
<i>IKAROS</i>	<i>LAMTOR5</i>	<i>CAB39</i>	<i>SOS1</i>	<i>EIF4E2</i>
<i>JUN</i>	<i>NPRL2</i>	<i>ATP6V1A</i>	<i>SOS2</i>	<i>ATP6V1G1</i>
<i>FOS</i>	<i>RRAGA</i>	<i>ATP6V1B1</i>	<i>BRAF</i>	<i>FNIP1</i>
<i>NFKB1</i>	<i>FZD10</i>	<i>ATP6V1B2</i>	<i>STK11</i>	<i>TTI1</i>
<i>MITF</i>	<i>CHUK</i>	<i>ATP6V1C1</i>	<i>TNF</i>	<i>DEPDC5</i>
<i>IRF4</i>	<i>ATP6V1G3</i>	<i>ATP6V1E1</i>	<i>TNFRSF1A</i>	<i>ULK2</i>
<i>BATF</i>	<i>SLC38A9</i>	<i>PIK3CA</i>	<i>TSC1</i>	<i>TELO2</i>
<i>EOMES</i>	<i>DVL1</i>	<i>PIK3CB</i>	<i>TSC2</i>	<i>TBC1D7</i>
<i>TBX21</i>	<i>DVL2</i>	<i>PIK3CD</i>	<i>WNT1</i>	<i>-LOC100130357</i>
<i>PRDM1</i>	<i>DVL3</i>	<i>PIK3CG</i>	<i>WNT2</i>	
<i>STAT1</i>	<i>EIF4B</i>	<i>PIK3R1</i>	<i>WNT3</i>	
<i>STAT3</i>	<i>EIF4E</i>	<i>PIK3R2</i>	<i>WNT5A</i>	
<i>STAT5A</i>	<i>EIF4EBP1</i>	<i>ATP6V1G2</i>	<i>WNT6</i>	
<i>STAT5B</i>	<i>FLCN</i>	<i>WNT4</i>	<i>WNT7A</i>	
<i>ETS1</i>	<i>AKT1</i>	<i>MIOS</i>	<i>WNT7B</i>	
<i>RUNX3</i>	<i>AKT2</i>	<i>DDIT4</i>	<i>WNT8A</i>	
<i>CREB1</i>	<i>RNF152</i>	<i>LAMTOR1</i>	<i>WNT8B</i>	
<i>GATA3</i>	<i>LPIN1</i>	<i>STRADB</i>	<i>WNT10B</i>	
<i>FOXO1</i>	<i>PIK3R5</i>	<i>PRR5</i>	<i>WNT11</i>	
	<i>ATP6V1C2</i>	<i>PRKAA1</i>	<i>WNT2B</i>	
	<i>MTOR</i>	<i>PRKAA2</i>	<i>WNT9A</i>	
	<i>RICTOR</i>	<i>PRKCA</i>	<i>WNT9B</i>	
	<i>EIF4E1B</i>	<i>PRKCB</i>	<i>FZD5</i>	
	<i>FZD2</i>	<i>PRKCG</i>	<i>MAPKAP1</i>	
	<i>ULK3</i>	<i>MAPK1</i>	<i>WDR59</i>	
	<i>RPS6KA6</i>	<i>MAPK3</i>	<i>FZD3</i>	
	<i>GRB2</i>	<i>MAP2K1</i>	<i>WNT10A</i>	
	<i>GRB10</i>	<i>MAP2K2</i>	<i>WNT5B</i>	
	<i>LAMTOR2</i>	<i>PTEN</i>	<i>NPRL3</i>	
	<i>GSK3B</i>	<i>RPTOR</i>	<i>SLC7A5</i>	
	<i>HRAS</i>	<i>FNIP2</i>	<i>CAB39L</i>	
	<i>IGF1</i>	<i>RRAGD</i>	<i>SEH1L</i>	
	<i>IGF1R</i>	<i>RAF1</i>	<i>FZD1</i>	
	<i>IKBKB</i>	<i>RHEB</i>	<i>FZD4</i>	
	<i>INS</i>	<i>RPS6</i>	<i>FZD6</i>	
	<i>INSR</i>	<i>RPS6KA1</i>	<i>FZD7</i>	
	<i>IRS1</i>	<i>RPS6KA2</i>	<i>FZD8</i>	
	<i>KRAS</i>	<i>RPS6KA3</i>	<i>FZD9</i>	
	<i>RHOA</i>	<i>RPS6KB1</i>	<i>SESN2</i>	
	<i>LAMTOR4</i>	<i>RPS6KB2</i>	<i>ULK1</i>	
	<i>LRP6</i>	<i>CLIP1</i>	<i>WDR24</i>	
	<i>LRP5</i>	<i>SEC13</i>	<i>AKT1S1</i>	
	<i>NRAS</i>	<i>RRAGC</i>	<i>PIK3R3</i>	
	<i>TBC1D7</i>	<i>MLST8</i>	<i>LAMTOR3</i>	
	<i>ATP6V1D</i>	<i>SGK1</i>	<i>WNT3A</i>	
	<i>WNT16</i>	<i>DEPTOR</i>	<i>ATP6V1E2</i>	

Lysosomal Biogenesis

<i>A2M</i>	<i>F2R</i>	<i>RAB26</i>	<i>TMED10</i>
<i>ABCA1</i>	<i>F2RL3</i>	<i>RAB27A</i>	<i>TMED2</i>
<i>ABCA12</i>	<i>F5</i>	<i>RAB4A</i>	<i>TOR1A</i>
<i>ABCC4</i>	<i>F8</i>	<i>RAB4B</i>	<i>VAMP1</i>
<i>ACTB</i>	<i>FGA</i>	<i>REST</i>	<i>VAMP2</i>
<i>ACTN1</i>	<i>FGB</i>	<i>SELP</i>	<i>VAMP7</i>
<i>ACTN2</i>	<i>FGG</i>	<i>SERPINA1</i>	<i>VAMP8</i>
<i>ACTN4</i>	<i>FN1</i>	<i>SERPINA5</i>	<i>VEGFA</i>
<i>ADAM8</i>	<i>GAL</i>	<i>SERPINE1</i>	<i>VPS33B</i>
<i>ALDOA</i>	<i>GAS6</i>	<i>SERPINF2</i>	<i>VWF</i>
<i>AP1G1</i>	<i>GHRL</i>	<i>SFTPD</i>	<i>ZP3</i>
<i>APOA1</i>	<i>HGF</i>	<i>SLC11A1</i>	
<i>APP</i>	<i>HPS4</i>	<i>SLC2A4</i>	
<i>AZU1</i>	<i>HRG</i>	<i>SNAP23</i>	
<i>BLOC1S1</i>	<i>HSPD1</i>	<i>SNAPIN</i>	
<i>BLOC1S2</i>	<i>IGF1</i>	<i>SNCA</i>	
<i>BLOC1S3</i>	<i>IGF2</i>	<i>SNX19</i>	
<i>BRCA2</i>	<i>INS</i>	<i>SNX19</i>	
<i>CAPZB</i>	<i>ITGA2B</i>	<i>SPACA1</i>	
<i>CASC5</i>	<i>ITGB3</i>	<i>SPACA3</i>	
<i>CCDC64</i>	<i>KIT</i>	<i>SPARC</i>	
<i>CD36</i>	<i>KLK7</i>	<i>SPESP1</i>	
<i>CD63</i>	<i>LRGUK</i>	<i>SPINK5</i>	
<i>CD9</i>	<i>LYST</i>	<i>SRGN</i>	
<i>CHGA</i>	<i>MPO</i>	<i>SRI</i>	
<i>CLCN3</i>	<i>MYO5A</i>	<i>STX3</i>	
<i>CLU</i>	<i>OLFM4</i>	<i>STX4</i>	
<i>CPE</i>	<i>PAFAH1B1</i>	<i>STX7</i>	
<i>CTSG</i>	<i>PDGFA</i>	<i>STXBP1</i>	
<i>CTSH</i>	<i>PDGFB</i>	<i>STXBP3</i>	
<i>CXADR</i>	<i>PECAM1</i>	<i>TEKT3</i>	
<i>CYBA</i>	<i>PLG</i>	<i>TF</i>	
<i>DENND4C</i>	<i>PRKCA</i>	<i>TGFB1</i>	
<i>DTNBP1</i>	<i>PTPRN</i>	<i>TGFB2</i>	
<i>EGF</i>	<i>RAB10</i>	<i>TGFB3</i>	
<i>ELANE</i>	<i>RAB11B</i>	<i>THBS1</i>	
<i>EXOC3</i>	<i>RAB13</i>	<i>TIMP1</i>	

Genes involved in effector transcription, mTOR signaling and lysosomal biogenesis.

Supplementary Table 2

Gene Symbol	Alias	Refseq #	Official Full Name	RT2 Catalog Number
PPC	PPC	SA_00103	Positive PCR Control	PPX63339
HGDC	HIGX1A	SA_00105	Human Genomic DNA Contamination	PPH65835
RTC	RTC	SA_00104	Reverse Transcription Control	PPX63340
18S rRNA		X03205	Human 18S ribosomal RNA	PPH05666
B2M	-	NM_004048	Beta-2-microglobulin	PPH01094
ARRB1	ARB1/ARR1	NM_004041	Arrestin, beta 1	PPH02341
GZMB	CCPI/CGL-1/CGL1/CSP-B/CSPB/CTLA1/CTSGL1/HLP/SECT	NM_004131	Granzyme B (granzyme 2, cytotoxic T-lymphocyte-associated serine esterase 1)	PPH02594
CST7	CMAF	NM_003650	Cystatin F (leukocystatin)	PPH05560
SELL	CD62L/LAM1/LECAM1/LEU8/LNHR/LSEL/LYAM1/PLNHR/TQ1	NM_000655	Selectin L	PPH00677
CTSC	CPP1/DPP-I/DPP1/DPPI/HMS/JP/JPD/PALS/PDON1/PLS	NM_001814	Cathepsin C	PPH06129
GNPTAB	GNPTA/ICD	NM_024312	N-acetylglucosamine-1-phosphate transferase, alpha and beta subunits	PPH19573
IGF2R	CD222/CIMPR/M6P-R/MPRI/MPRI	NM_000876	Insulin-like growth factor 2 receptor	PPH00351
PTEN	10q23del/BZS/CWS1/DEC/GLM2/MHAM/MMAC1/PTEN1/TEP1	NM_000314	Phosphatase and tensin homolog	PPH00327
TBX21	T-PET/T-bet/TBET/TBLYM	NM_013351	T-box 21	PPH00396
SIGLEC7	AIRM1/CD328/CDw328/D-siglec/QA79/SIGLEC-7/SIGLEC19P/SIGLECP2/p75/p75/AIRM1	NM_014385	Sialic acid binding Ig-like lectin 7	PPH15594
MCOLN1	TRP-ML1	NM_020533	Mucolipin 1	PPH19799A