

SUPPLEMENTARY MATERIAL

Table S1. Oligonucleotides used in this work. The restriction sites are underlined.

Primer name	Primer sequence	Restriction enzyme
FV53_NdeI_F1	GGAA <u>ACATATGCCATATTTAGATA</u> AAAGTG	NdeI
FV53_BamHI_R1	TTGAG <u>GATCC</u> ATCTAGTCTTCCTTATTC	BamHI
FV53_XhoI_R1	CATCTAGTCTTC <u>CTCGAGTTCAGTC</u> ATTG	XhoI
FV53_Nde10F	TAAAGTGGT <u>ACATATGACAGT</u> TAAAC	NdeI
FV53_Bam10R	CAGCATTGGT <u>GGATCCTTAA</u> CCAAC	BamHI
FV53_Xho10R	CAGCATTGGTGG <u>ACTCGAGACCA</u> AC	XhoI
FV53_Nde20F	CCCAGCCTATT <u>CATATGGT</u> AGGTTC	NdeI
FV53_Bam20RN	GTCCAGA <u>AGGATCCTTTACCG</u> GATACG	BamHI
FV53_Xho20R	GAAGGACGTTGATCT <u>CGAGACGA</u> AC	XhoI
FV53_Nde25F	CTGTAGGTTTC <u>CATATGCCG</u> CTG)	NdeI
FV53_Bam25R	GTTGATCC <u>GGATCCGA</u> ACTTAGTGC	BamHI
FV53_Xho25R	GATCCGATACGCT <u>CGAGGTGC</u> AG	XhoI
FV53_Nde30F	GAAACTCCGCTG <u>CATATGG</u> CTATCC	NdeI
FV53_Bam30R	CGAAGTGCG <u>GATCCTTGTTT</u> TACAC	BamHI
FV53_Xho30R	GAAGTGCAGGGCGT <u>CTCGAGAC</u> ACCAG	XhoI
FV3_Bam50R	CTGATT <u>GGATCC</u> GATCTATGCACG	BamHI
FV3_Bam75R	CTGTCAGGATCCCTCAATTCTTAATG	BamHI
FV3_Bam100R	CGGAAGATCAGGATCCGACTACTTCATG	BamHI
FV3_Bam150R	CAACACCGTGGATCCTTCAGTAGAAG	BamHI
FV3_Bam200R	CACCTATGATAGGATCCTACGGAAAATC	BamHI
FV3_m_N_C_F	GATATACATATGGCTAGCACAGGTACCCCA TATTTAGATAAAAGTGG	NdeI, NheI, Acc65I
FV3_m_N_C_R	GTGGTGCTCGAGTTACGTACGTGTACTAGTT TCAGTCATTGCAACAGC	XhoI, Pfl23II, BcuI

Table S2. Constructs of gp053.

Mutant	Primers used for PCR fragment amplification	Expression vector	Amino acid residues, MW (kDa)	Description of self-assembling structures
Full-length gp053	FV53_NdeI_F1 FV53_BamHI_R1	pET-21a	458 (50.2)	Regular polysheaths, length up to ~1000 nm, width 27,8±2,7 nm.
gp053_N-his	FV53_NdeI_F1 FV53_BamHI_R1	pET-16b	479 (52.8)	Regular polysheaths, length up to ~300 nm, width ~28 nm.
gp053_C-his	FV53_NdeI_F1 FV53_XhoI_R1	pET-21a	466 (51.3)	Regular polysheaths, length up to ~1000 nm, width ~28 nm.
gp053_NΔ9	FV53_Nde10F FV53_BamHI_R1	pET-21a	449 (49.2)	Regular polysheaths, length up to ~600 nm, width ~28 nm.
gp053_CΔ11	FV53_NdeI_F1 FV53_Bam10R	pET-21a	447 (49.1)	Regular polysheaths, length up to ~600 nm, width ~28 nm.
gp053_NΔ9_CΔ11	FV53_Nde10F FV53_Bam10R	pET-21a	438 (48.1)	Regular polysheaths, length up to ~500 nm, width ~28 nm.
gp053_NΔ9-his_CΔ11	FV53_Nde10F FV53_Bam10R	pET-16b	459 (50.6)	Regular polysheaths, length up to ~800 nm, width ~28 nm.
gp053_NΔ9_CΔ11-his	FV53_Nde10F FV53_Xho10R	pET-21a	446 (49.2)	Regular polysheaths, length up to ~800 nm, width ~28 nm.
gp053_NΔ20	FV53_Nde20F FV53_BamHI_R1	pET-21a	438 (48.1)	Regular polysheaths, length up to ~500 nm, width ~28 nm.
gp053_NΔ20_CΔ20	FV53_Nde20F FV53_Bam20RN	pET-21a	418 (46.0)	Regular polysheaths, length up to ~600 nm, width ~28 nm.
gp053_NΔ20-his_CΔ20	FV53_Nde20F FV53_Bam20RN	pET-16b	439 (48.5)	Regular polysheaths, length up to ~1000 nm, width ~28 nm.
gp053_NΔ20_CΔ20-his	FV53_Nde20F FV53_Xho20R	pET-21a	424 (46.8)	Regular polysheaths, length up to ~1000 nm, width ~28 nm.
gp053_NΔ25_CΔ25	FV53_Nde25F FV53_Bam25R	pET-21a	408 (44.8)	Regular polysheaths, length up to ~800 nm, width ~28 nm.
gp053_NΔ25-his_CΔ25	FV53_Nde25F	pET-16b	429 (47.3)	Regular polysheaths, length up to ~800 nm, width ~28 nm.

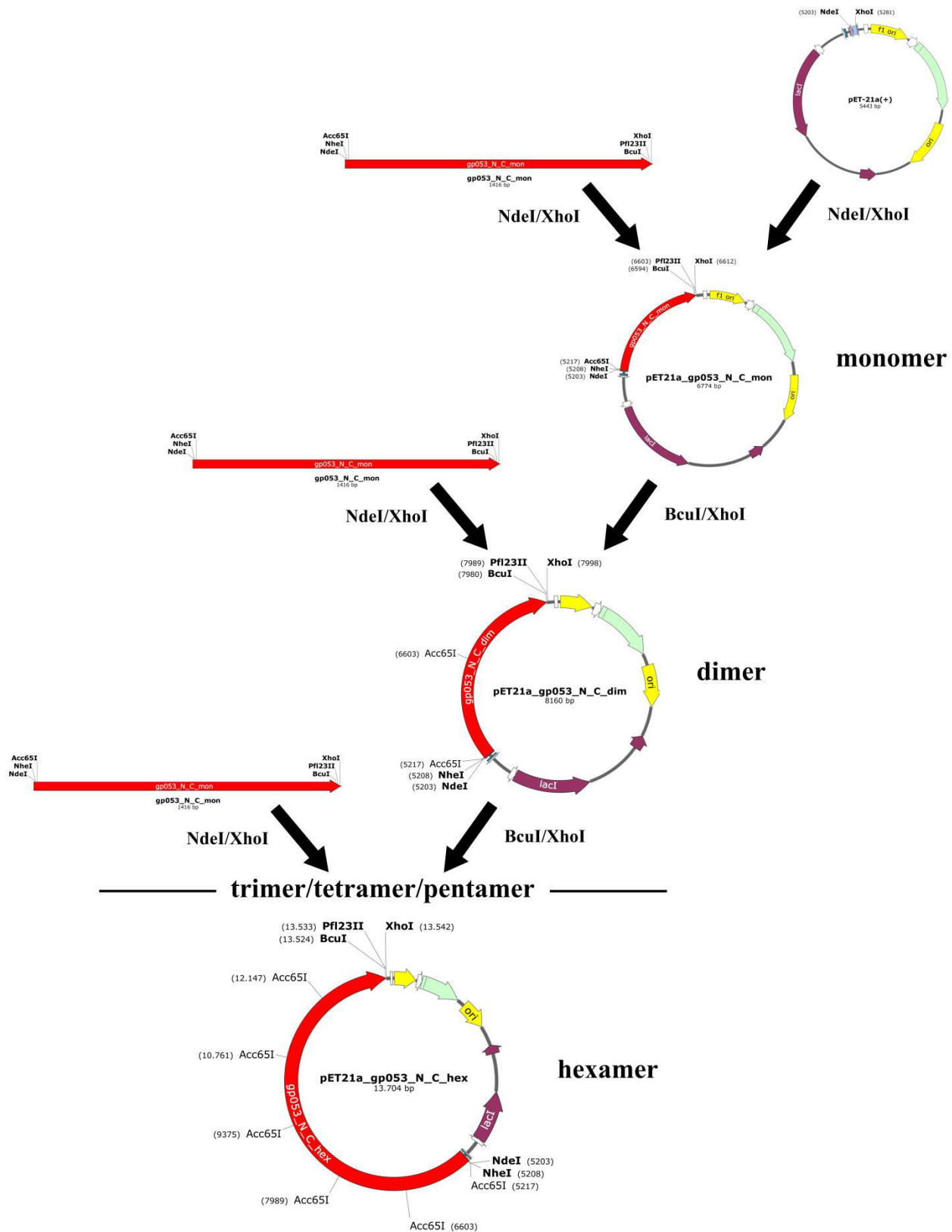
	FV53_Bam25RN			nm), width ~28 nm.
gp053_NΔ25_CΔ25-his	FV53_Nde25F FV53_Xho25R	pET-21a	416 (45.9)	Regular polysheaths, length up to ~800 nm, width ~28 nm.
gp053_NΔ29	FV53_Nde30F FV53_BamHI_R1	pET-21a	429 (47.1)	Soluble unordered protein ribbons
gp053_NΔ29_CΔ31	FV53_Nde30F FV53_Bam30R	pET-21a	398 (43.6)	Soluble unordered protein ribbons
gp053_NΔ29-his_CΔ31	FV53_Nde30F FV53_Bam30R	pET-16b	419 (46.1)	Insoluble protein
gp053_CΔ31	FV53_NdeI_F1 FV53_Bam30R	pET-21a	427 (46.8)	Regular polysheaths, length up to ~600 nm, width ~28 nm.
gp053_CΔ51	FV53_NdeI_F1 FV53_Bam50R	pET-21a	407 (44.6)	Regular polysheaths, length up to ~600 nm, width ~28 nm.
gp053_CΔ76	FV53_NdeI_F1 FV53_Bam75R	pET-21a	382 (41.9)	Regular polysheaths, length up to ~600 nm, width ~28 nm.
gp053_CΔ100	FV53_NdeI_F1 FV53_Bam100R	pET-21a	358 (39.3)	Regular polysheaths, length up to ~3000 nm, width ~32 nm.
gp053_CΔ152	FV53_NdeI_F1 FV53_Bam150R	pET-21a	306 (33.2)	Soluble unordered protein ribbons
gp053_CΔ200	FV53_NdeI_F1 FV53_Bam200R	pET-21a	258 (28.0)	Soluble unordered protein ribbons
gp053_C-strep	FV53_NdeI_F1 Pfl23Strep	pET21a	468 (51.54)	Regular polysheaths, length up to ~500 nm, width ~28 nm.
gp053_NΔ25_C-strep	FV53_Nde25F Pfl23Strep	pET21a	443 (48.84)	Insoluble protein
gp053_N_C_mon	FV3_m_N_C_F FV3_m_N_C_R	pET21a	468 (51.21)	Regular polysheaths, length up to ~500 nm, width ~28 nm.
gp053_N_C_dim	FV3_m_N_C_F FV3_m_N_C_R	pET21a	930 (101.75)	Irregular polysheaths, length up to ~200 nm after 48 hours of incubation <i>in vitro</i> at 22°C.
gp053_N_C_trim	FV3_m_N_C_F FV3_m_N_C_R	pET21a	1392 (152.29)	Irregular polysheaths, length up to ~200 nm after 48 hours of incubation <i>in vitro</i> at 22°C.

gp053_N_C_tetr	FV3_m_N_C_F FV3_m_N_C_R	pET21a	1854 (202.83)	Irregular polysheaths, length up to ~150 nm after 48 hours of incubation <i>in vitro</i> at 22°C.
gp053_N_C_pent	FV3_m_N_C_F FV3_m_N_C_R	pET21a	2316 (253.38)	Irregular polysheaths, length up to ~200 nm after 48 hours of incubation <i>in vitro</i> at 22°C.
gp053_N_C_hex	FV3_m_N_C_F FV3_m_N_C_R	pET21a	2778 (303.92)	Irregular polysheaths, length up to ~150 nm after 48 hours of incubation <i>in vitro</i> at 22°C.

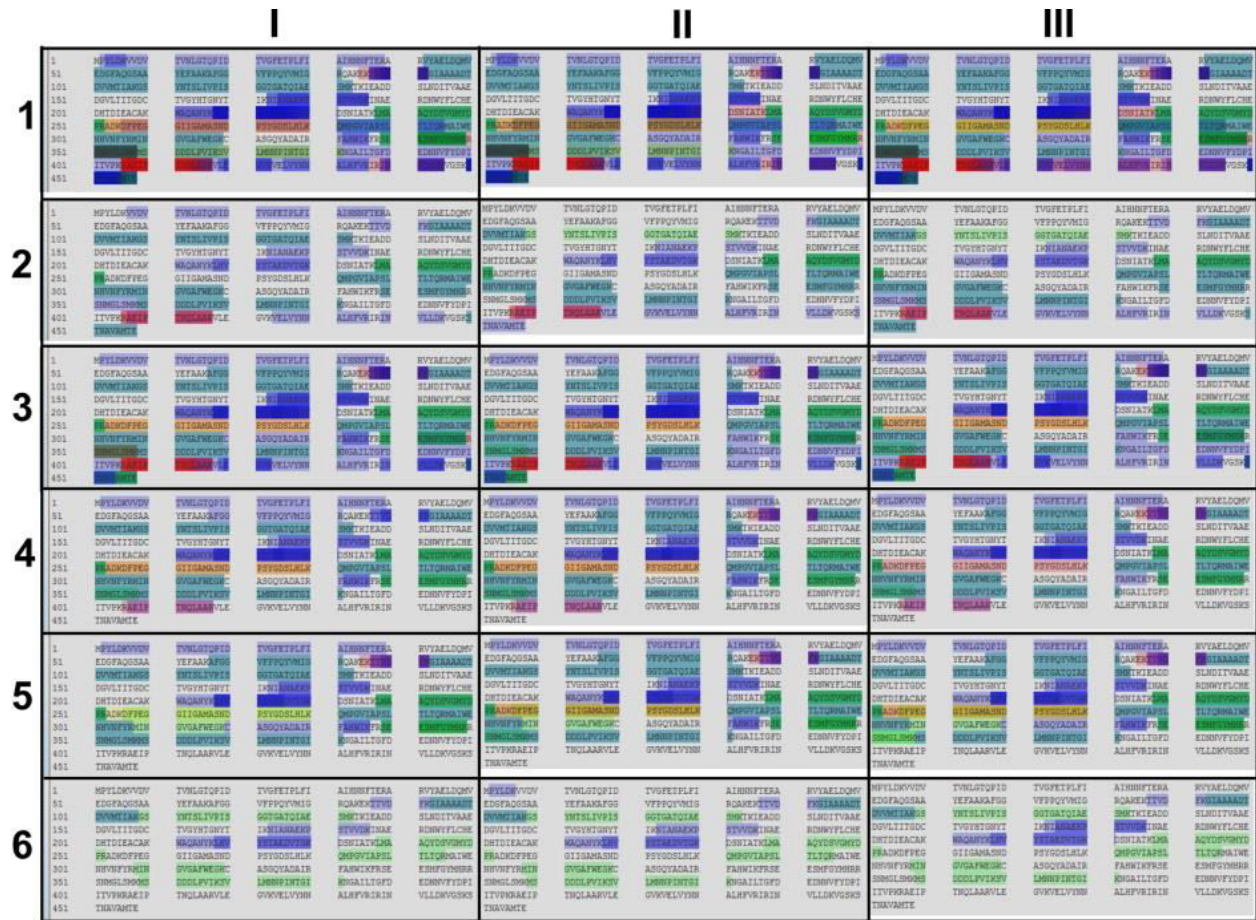
Supplementary Table S3. Homology of gp053 from FV3 with proteins from other bacteriophages.

Significant match protein accession number, organism (taxonomic lineage)	Identity aa %/ similarity aa% (length of the overlapping segment)	E value
AKO61920.1 , <i>Escherichia</i> phage APCEc02 (<i>Vequintavirinae</i> , V5virus)	99/99 (458)	0.0
ATW61401.1 , phage 203 (<i>Vequintavirinae</i> , V5virus)	99/99 (458)	0.0
YP_002003555.1 , <i>Escherichia</i> phage V5 (<i>Vequintavirinae</i> , V5virus)	99/99 (458)	0.0
YP_009177499.1 , <i>Escherichia</i> phage vB_EcoM_FFH2 (<i>Vequintavirinae</i> , V5virus)	99/99 (458)	0.0
YP_008530297.1 , <i>Escherichia</i> phage JES2013 (<i>Vequintavirinae</i> , V5virus)	99/99 (458)	0.0
ASJ80411.1 , <i>Escherichia</i> phage V18 (<i>Vequintavirinae</i> , V5virus)	98/99 (458)	0.0
AKU44144.1 , <i>Escherichia</i> phage Murica (<i>Vequintavirinae</i> , V5virus)	98/99 (458)	0.0
AKE47028.1 , <i>Escherichia coli</i> O157 typing phage 14 (<i>Vequintavirinae</i> , V5virus)	98/98 (458)	0.0
AVJ51863.1 , <i>Escherichia</i> phage PDX (<i>Vequintavirinae</i> , V5virus)	96/98 (458)	0.0
AXU22620.1 , <i>Escherichia</i> phage vB_EcoM-Pr121LW (<i>Vequintavirinae</i> , V5virus)	96/98 (458)	0.0
AXC42731.1 , <i>Escherichia</i> phage LL12 (<i>Vequintavirinae</i> , V5virus)	96/98 (458)	0.0
YP_006987047.1 , <i>Cronobacter</i> phage vB_CsaM_GAP31 (<i>Vequintavirinae</i> , Se1virus)	60/71 (468)	0.0

YP_009111160.1 , <i>Escherichia</i> phage Av-05 (unclassified myovirus)	59/72 (468)	0.0
YP_004893867.1 , <i>Salmonella</i> phage PVP-SE1 (<i>Vequintavirinae</i> , Se1virus)	59/73 (469)	0.0
YP_008857275.1 , Enterobacteria phage 4MG (<i>Vequintavirinae</i> , Se1virus)	59/71 (469)	0.0
ARB12543.1 , <i>Klebsiella</i> phage vB_KpnP_BIS47 (unclassified bacterial virus)	59/71 (469)	0.0
YP_009187676.1 , <i>Klebsiella</i> phage vB_KpnM_KB57 (unclassified myovirus)	57/68 (493)	0.0
AXN57735.1 , <i>Acinetobacter</i> phage ABPH49 (unclassified myovirus)	57/69 (468)	8e-174
ARB11491.1 , <i>Pectobacterium</i> phage vB_PatM_CB7 (<i>Vequintavirinae</i> , Cr3virus)	53/66 (464)	4e-159
YP_006383031.1 , <i>Cronobacter</i> phage CR3 (<i>Vequintavirinae</i> , Cr3virus)	53/66 (464)	7e-158
YP_009042253.1 , <i>Cronobacter</i> phage CR8 (<i>Vequintavirinae</i> , Cr3virus)	53/66 (464)	1e-157
YP_009014977.1 , <i>Cronobacter</i> phage CR9 (<i>Vequintavirinae</i> , Cr3virus)	53/66 (462)	1e-157
ATS93415.1 , <i>Pectobacterium</i> phage DU_PP_I (<i>Vequintavirinae</i> , Cr3virus)	53/65 (464)	2e-157
YP_007392684.1 , <i>Pectobacterium</i> phage phiTE (<i>Vequintavirinae</i> , Cr3virus)	52/66 (462)	3e-155



Supplementary Figure S1. Schematic representation of construction of gp053 oligomers. The PCR fragment of gene *053* mutant gp053_N_C_mon was obtained by amplification of phage DNA using FV3_m_N_C_F and FV3_m_N_C_R primers presented in Supplementary Table S1. Restriction endonucleases, which were used to obtain restricted fragments and cloning vectors, are shown beside the arrows.



Supplementary Figure S2. The protein sequence coverage map obtained from the PLGS-processed MS^E data. Rows (1-6) indicate the number of protein band, excised from the gel, columns (I-III) indicate three technical replicates of LC-MS/MS. Regions of the protein sequence that match peptides are highlighted in color: blue– matches to a peptide, red– matches to a partial peptide, green– matches to a modified peptide, yellow– matches to a partial modified peptide.