

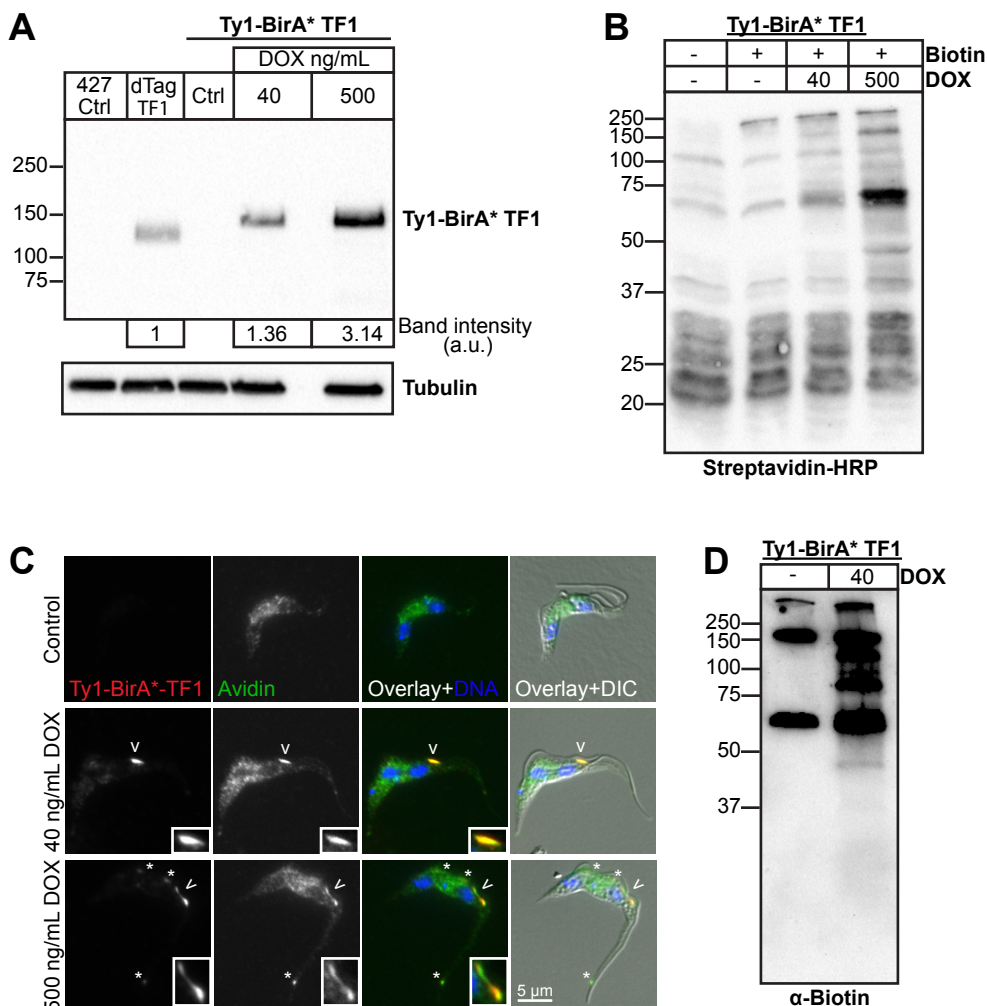
Supplemental Figures for:

Identification of TOEFAZ1-interacting proteins reveals key regulators of  
*Trypanosoma brucei* cytokinesis

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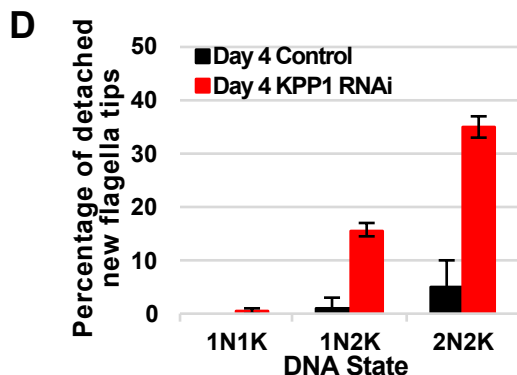
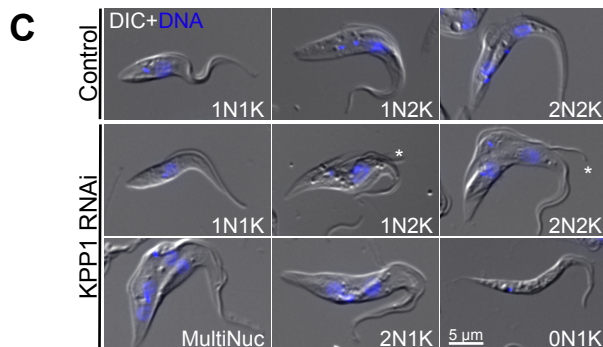
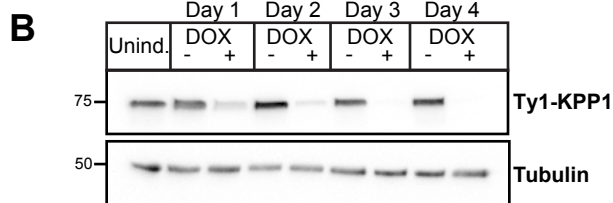
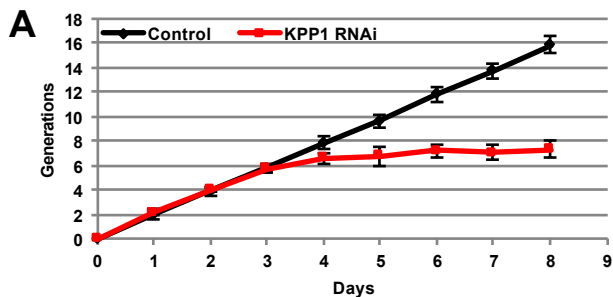
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# Figure S1



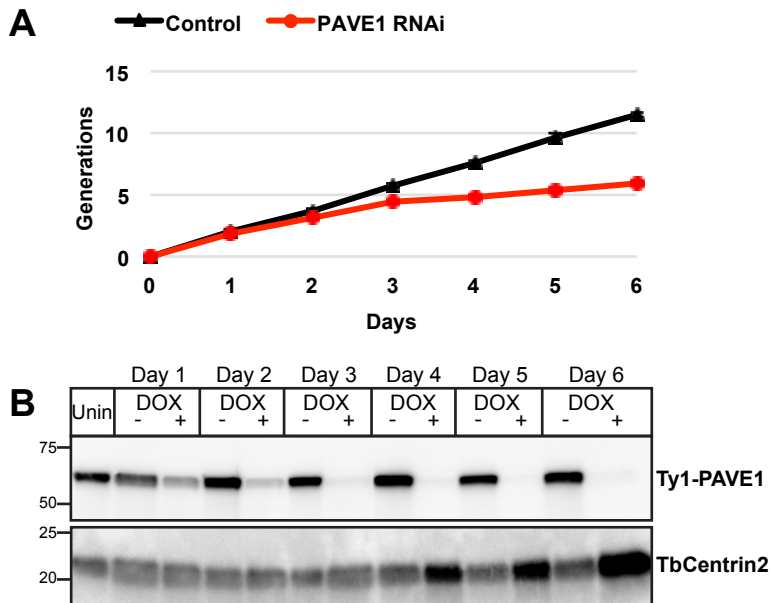
**Figure S1** Matching Ty1-BirA\* TOEFAZ1 expression to endogenous TOEFAZ1 levels. **A)** Western blot comparing the expression levels between a cell line in which both alleles of TOEFAZ1 are tagged with a Ty1 epitope tag (dTag TF1) and the Ty1-BirA\* cell line induced overnight with either 40 ng/mL or 500 ng/mL doxycycline. **B)** The Ty1-BirA\* cell line was induced as above, followed by incubation with 50  $\mu$ M biotin. Lysates from the induced Ty1-BirA\* TOEFAZ1 cells were collected and separated by SDS-PAGE followed by transfer to nitrocellulose for western blotting with streptavidin conjugated to HRP. **C)** The Ty1-BirA\* cell line was induced and incubated with biotin as above. Cells were methanol-fixed and stained with anti-Ty1 antibody and streptavidin. Arrowheads: Ty1-BirA\* TOEFAZ1 localization. Asterisks indicate spurious localization. DAPI was used to stain DNA. **D)** The Ty1-BirA\* TOEFAZ1 cell line was induced as in B) with 40 ng/mL doxycycline, followed by lysis and capture of biotinylated proteins, which were eluted and blotted using anti-biotin antibody.

# Figure S2



**Figure S2** KPP1 is required for cell division. **A)** KPP1 RNAi was induced for 8 d using 1  $\mu\text{g}/\text{mL}$  doxycycline, and cell concentration was counted every 24 h. **B)** Cell lysates were collected every 24 h from the RNAi timecourse in A), western blotted, and labeled with anti-Ty1 antibody. **C)** Cells carrying a KPP1 RNAi construct were treated with either vehicle control or 1  $\mu\text{g}/\text{mL}$  doxycycline to induce RNAi for four days. Cells were PFA-fixed and stained with DAPI for DNA. Asterisk indicates a detached flagellar tip. **D)** Quantification of detached new flagella tips in control and KPP1 RNAi cells in which KPP1 has been depleted for 4 d.

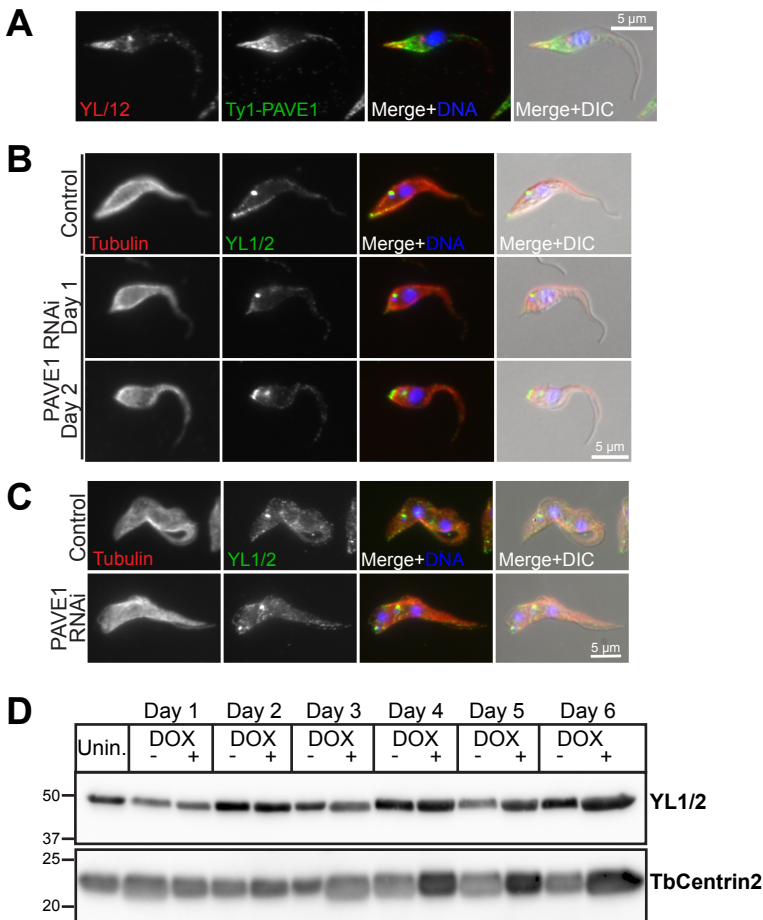
# Figure S3



**Figure S3** PAVE1 is necessary for cell division. **A)** PAVE1 RNAi cells were treated with either vehicle control or 1  $\mu\text{g}/\text{mL}$  doxycycline for 6 days and cell number was monitored every 24 h. **B)** Cell lysates were collected every 24 h from the time course in A). Lysates were western blotted and stained with anti-Ty1 antibody.



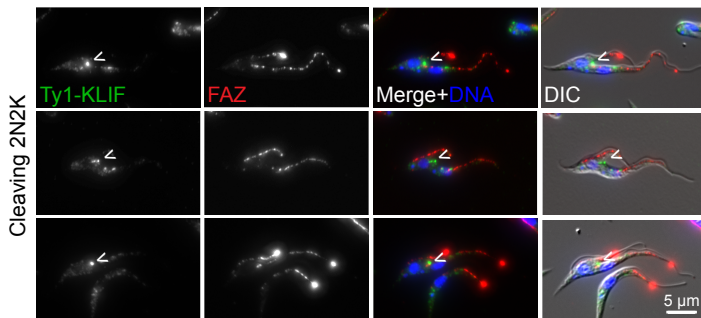
# Figure S4



**Figure S4** YL1/2 staining is not altered in PAVE1 RNAi cells. **A)** Immunofluorescence of methanol-fixed wherein PAVE1 was tagged at its endogenous locus with a Ty1 tag. Cells were stained for YL1/2 to detect tyrosinated tubulin, an indicator of growing microtubules, and Ty1 to label PAVE1. **B)** PAVE1 RNAi was induced with 1 µg/mL doxycycline for 1 and 2 d. Cells were methanol fixed and stained with anti-tubulin antibody and YL1/2. DAPI was used to stain DNA in all immunofluorescence panels. **C)** Dividing 2N2K cells from the experiment described in B). **D)** Lysates from the RNAi time course in Fig. S3A were western blotted and labeled with YL1/2 antibody.

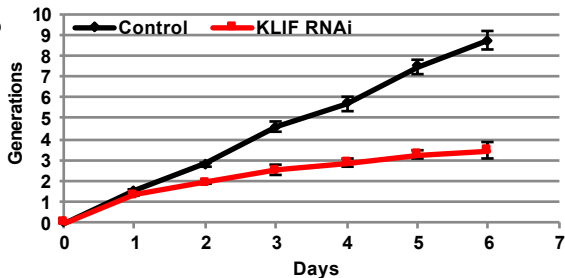
# Figure S5

**A**

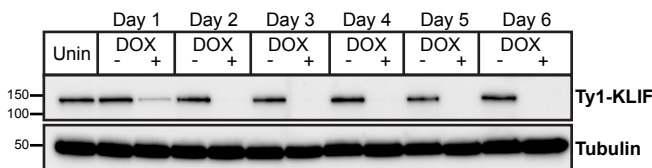


**Figure S5** KLIF is required for cell division. **A)** Cells in which KLIF was tagged at its endogenous locus with a Ty1 epitope tag were methanol-fixed and stained with anti-Ty1 antibody and 1B41 to label the FAZ. DAPI was used to label DNA. Arrowheads indicate Ty1-KLIF localization at the cleavage furrow. **B)** Cells containing the KLIF RNAi construct were treated with either vehicle control or 1  $\mu\text{g}/\text{mL}$  doxycycline. Cell concentration was counted every 24 h. **C)** Lysates were collected every 24 h from the RNAi timecourse in B), western blotted, and stained with anti-Ty1 antibody. **D)** Cells containing a KLIF RNAi construct were treated with either vehicle control or 1  $\mu\text{g}/\text{mL}$  doxycycline for 3 d. Cells were collected and PFA-fixed for morphological analysis. DAPI was used to stain DNA. **E)** TOEFAZ1 RNAi was induced in a cell line containing KLIF tagged at its endogenous locus with a Ty1 epitope tag. Lysates were collected every 12 h, western blotted, and stained with anti-Ty1 antibody.

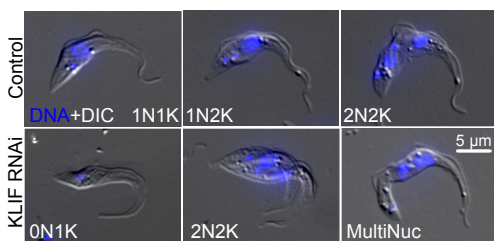
**B**



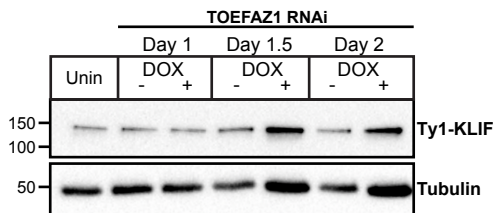
**C**



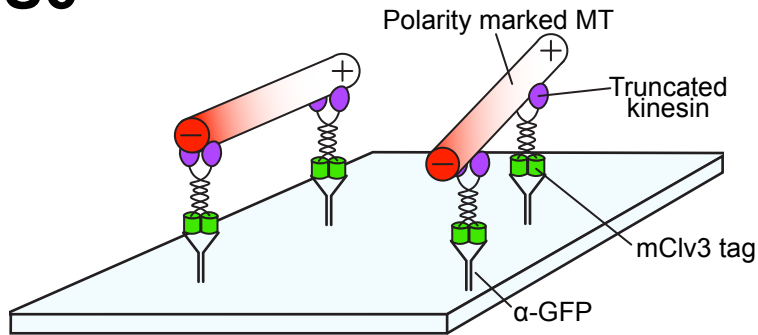
**D**



**E**



# Figure S6



**Figure S6** Schematic of the *in vitro* motility assay. KLIF (purple) containing a mClover3 tag (mClv3, green) is bound to an  $\alpha$ -GFP coated flow chamber (light blue) and blocked. Then, rhodamine-labeled polarity marked microtubules (red) containing enhanced fluorescent signal at their minus-ends are introduced to the flow chamber to interact with KLIF in the presence of MgATP.

# Figure S7

**Figure S7** Characterization of curated TOEFAZ1 BioID hits. A Ty1 epitope tag was integrated at either the N- or C-terminus of the endogenous locus of each hit from the TOEFAZ1 BioID screen. The size of each protein was determined by anti-Ty1 western blotting, and the correct integration of the Ty1 tag at the endogenous locus was established by loci PCR. Immunofluorescence analysis was conducted on each protein using anti-Ty1 antibody and, where applicable, using antibodies against known organelles for co-localization.

