A transcription factor PU.1 is critical for *Ccl22* gene expression in dendritic cells and macrophages

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Supplementary Table 1. Sequence of primers used in construction of reporter vectors.

5' position	Direction	Sequence
-597	Forward	CATAggtaccGCACACTCACTCTGGCTCGC
-245	Forward	CGTGggtaccATCCATGTGCCAGTGGCCAC
-101	Forward	CCACggtaccACAATCCTCCGGAGACCCAC
-65	Forward	CGTAggtaccAGTCCCCTCTGAGTAGTTC
-27	Forward	CGTAgattccGCTGGGGGGAAGTGGGACT
+108	Reverse	GTGCagatetGTCAGGTGTCTAATGACCAG

Mutation site	Direction	Sequence	
Site1	Forward	CTGGCCGGTACCGCTGGGACTAGTGGGACTTCTT	
		CTTTCAG	
	Reverse	CTGAAAGAAGAAGTCCCACTAGTCCCAGCGGTA	
		CCGGCCAG	
Site2	Forward	CCGCTGGGGGAAGTGGGACTAGATCTTTCAGGA	
		CATGAATGTC	
	Reverse	GACATTCATGTCCTGAAAGATCTAGTCCCACTTC	
		CCCCAGCGG	
Site1&2	Forward	CCGCTGGGACTAGTGGGACTAGATCTTTCAGGAC	
		ATGAATGTC	
	Reverse	GACATTCATGTCCTGAAAGATCTAGTCCCACTAG	
		TCCCAGCGG	

Supplementary Table 2. Sequence of primers used in ChIP assay.

<Mouse>

Amplified	Direction	Sequence
region		
-1780/-1714	Forward	CCAAAGGGTTTTGAGCTCTCA
	Reverse	GCTACTTCAGAGGGTAAAAAAAAGG
-1080/-1008	Forward	AAAGCCCACCGCATCATG
	Reverse	GCTGACTGTGTGCTCCAGGTT
-17/+52	Forward	GTGGGACTTCTTCTTCAGGACAT
	Reverse	CCCACTAGTTCAGGCACCTCTT

<Human>

Amplified	Direction	Sequence
region		
-2106/-2046	Forward	ACCAGCCCTGGCAACATAGT
	Reverse	CGCCCTGCTCATTTTTATTTCT
-153/-76	Forward	CCCCGCCAAAGAGAATTTC
	Reverse	GAGTCACTTGACATTCACATCTCAAA

*+1; Transcription start site

Supplemental figure

Materials and Methods

Materials

For detection of the mouse *Fizz-1/Retnla* and *Ym-1/Chil3* mRNAs, the following primers were used: mFizz1-F, 5'-ATCCCATGGCGTATAAAAGCAT-3',

mFizz1-R, 5'-GGGCAGAATCTCAGGAAAGGTT-3',

mYm1-F, 5'-TCTGGTGAAGGAAATGCGTAAA-3',

mYm1-R, 5'-GCAGCCTTGGAATGTCTTTCTC-3'.

Spi1/Pu.1 siRNA #1, 2 (Stealth Select RNAi, MSS247678 and MSS277025, respectively), and negative control siRNA (Stealth Negative Control Hi GC, 12935-400) were obtained from Invitrogen (Carlsbad, CA).

Methods

Preparation of Th2 cells and Migration assay

Naïve CD4⁺ T cells prepared from spleen using a MojoSort Ms CD4 Naïve T cell isolation kit (#480040, BioLegend) were polarized to Th2 cells by cultivation in the medium supplemented with 20 ng/ml recombinant mouse IL-4 (#094-03944, Wako) and 10 μ g/ml anti-IL-12p40 antibody (C17.8, BioLegend) in dishes coated with 1 μ g/ml anti-CD3 antibody (145-2C11, TONBO) and 10 μ g/ml anti-CD28 antibody (37.51, TONBO) for 5 days. The generated Th2 cells (5 x 10⁵/100 μ l) were subjected on upper site of a transwell (5 μ m pore size #3421, Corning), which were placed on a well filled

with 500 μ l culture supernatant of siRNA-introduced BMDCs. After 3 h incubation, the cell number on lower site of a transwell was counted.

Figure legends

Sup. Fig. 1. Western blotting profile of in vitro translated protein

Five µl of 20-fold diluted *in vitro* translated reaction mixture was loaded and detected by anti-PU.1 antibody.

Sup. Fig. 2. Expression of Fizz-1 (*Retnla*) and Ym-1 (*Chil3*) in *Spi1* siRNA introduced BMDMs

BMDMs were transfected with either negative control siRNA (siNega) or *Spi1* siRNA (siSpi1). At 48 h after transfection, relative mRNA levels were determined by quantitative RT-PCR after normalizing to mouse *Gapdh* mRNA levels. Data are expressed as the ratio of the expression level of the respective control siRNA-transfected cells. Results are means + S.D.s (n=3). Similar results were obtained in three independent experiments. *, p < 0.05.

Sup. Fig. 3. Effect of Spi1 siRNA on the migration activity

In vitro differentiated Th2 cells were seeded in the upper chambers of the transwell. The lower chambers were filled with culture supernatants from BMDCs transfected with either negative control siRNA (siNega) or *Spi1* siRNA (siSpi1) or culture medium as negative control (N.C.). After 3 h incubation, the cell number on lower site was

counted.

The numbers of migrated T cells are shown.

Sup. Fig. 4. Effect of Spi1 siRNA on the expression of Ccl22

BMDCs were transfected with either negative control siRNA (siNega) or *Spi1* siRNA (siSpi1). At 48 h after transfection, relative mRNA levels were determined by quantitative RT-PCR after normalizing to mouse *Gapdh* mRNA levels. Data are expressed as the ratio of the expression level of the respective control siRNA-transfected cells. Results are means + S.D.s (n=3). Similar results were obtained in three independent experiments. *, p < 0.05.

anti-PU.1





Sup. Fig. 2



Sup. Fig. 3



Sup. Fig. 4



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