

Supplementary Materials for

Novel reprogramming of neutrophils modulates inflammation resolution during atherosclerosis

Shuo Geng, Yao Zhang, Christina Lee, Liwu Li*

*Corresponding author. Email: lwli@vt.edu

Published 6 February 2019, *Sci. Adv.* **5**, eaav2309 (2019)

DOI: 10.1126/sciadv.aav2309

This PDF file includes:

- Fig. S1. Subclinical endotoxin up-regulates MPO level in HFD-fed mice.
- Fig. S2. Subclinical endotoxemia exacerbates atherosclerotic pathogenesis in RD-fed mice.
- Fig. S3. Subclinical endotoxin causes neutrophil expansion in atherosclerotic mice.
- Fig. S4. Subclinical endotoxin primes neutrophils into a proinflammatory state in atherosclerotic mice.
- Fig. S5. Subclinical endotoxin induces oxCAMKII elevation in vivo.
- Fig. S6. Neutrophils maintain viability after in vitro polarization.
- Fig. S7. Transfusion of superlow-dose LPS-polarized neutrophils elevates plasma lipid levels and modulates lesional macrophages.
- Fig. S8. Superlow-dose LPS and oxLDL treatment elevates ROS accumulation in neutrophils.
- Fig. S9. 4-PBA reverses superlow-dose LPS-induced differential regulation of miR-24 and miR-126 in neutrophils.
- Fig. S10. Transfusion of 4-PBA-polarized neutrophils down-regulates plasma lipid levels and reduces lesional macrophage activation.

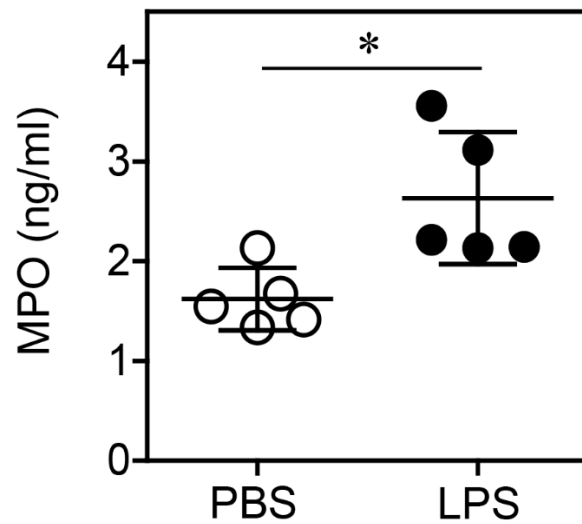


Fig. S1. Subclinical endotoxin up-regulates MPO level in HFD-fed mice. ApoE^{-/-} mice were administrated with PBS or super-low-dose LPS together with HFD for 4 weeks. Circulating MPO level was determined by ELISA. Data are representative of two independent experiments, and error bars represent means \pm s.e.m. * $P < 0.05$; Student's t -test ($n = 5$ for each group).

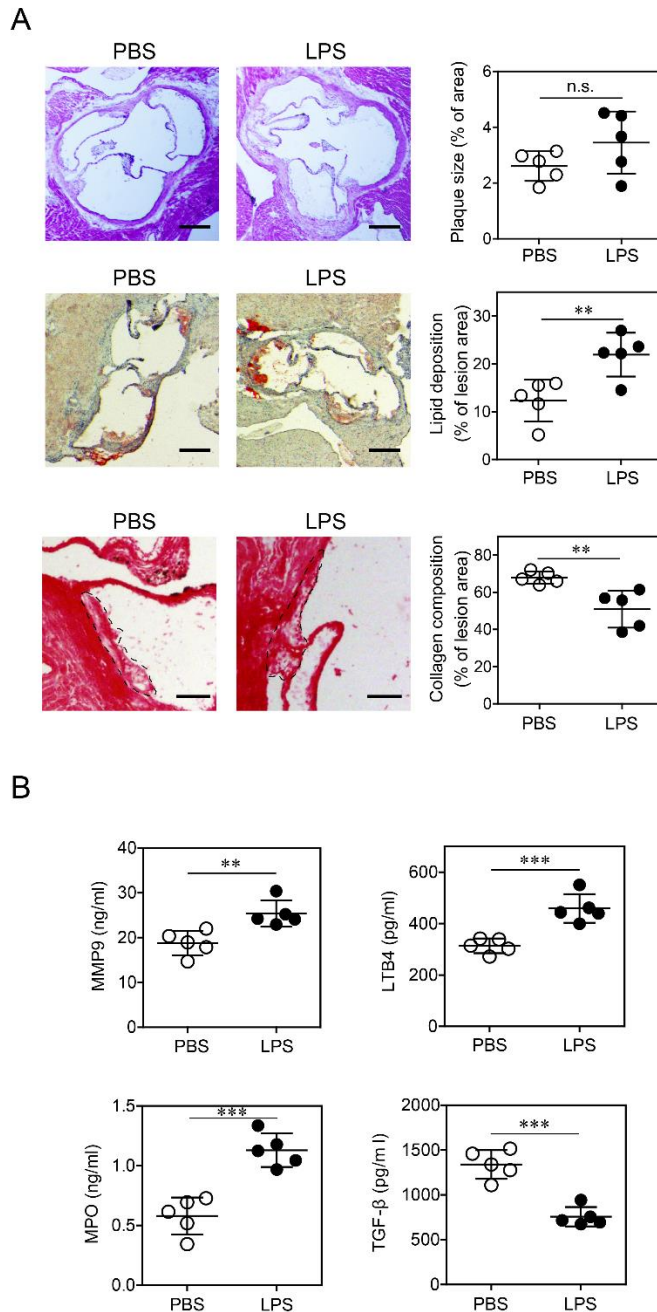


Fig. S2. Subclinical endotoxemia exacerbates atherosclerotic pathogenesis in RD-fed mice.

ApoE^{-/-} mice were administrated with PBS or super-low-dose LPS together with RD for 4 weeks. **(A)** Representative images of HE-stained atherosclerotic lesions and quantification of plaque size demonstrated as the percentage of lesion area within aortic root area (upper panels).

Representative images of Oil Red O-stained atherosclerotic plaques and quantification of lipid deposition within lesion area (middle panels). Representative images of Picrosirius Red-stained atherosclerotic plaques and quantification of collagen content within lesion area (lower panels).

Scale bar, 300 μ m (upper and middle panels) and 100 μ m (lower panels). **(B)** The levels of circulating MMP9, LTB4, MPO and TGF- β were determined by ELISA. Data are representative of two independent experiments, and error bars represent means \pm s.e.m. ***P* < 0.01 and ****P* < 0.001; Student's *t* test (*n* = 5 for each group).

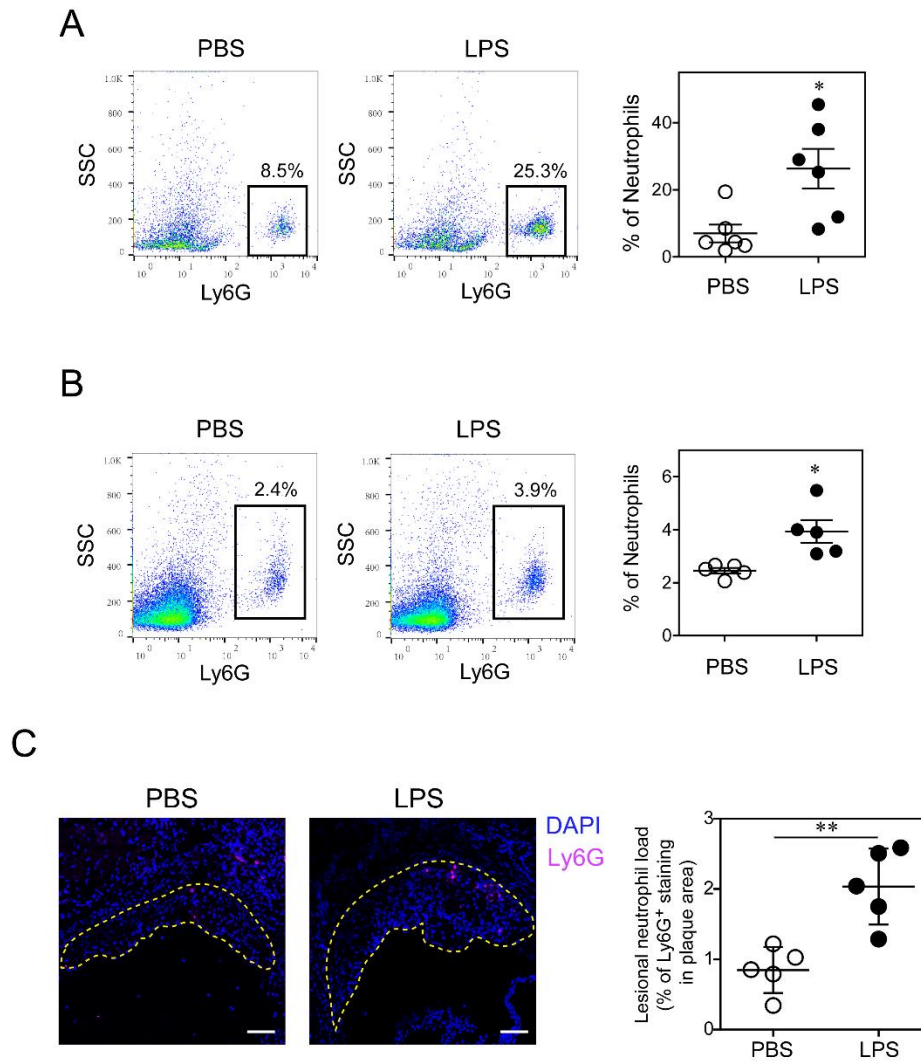


Fig. S3. Subclinical endotoxin causes neutrophil expansion in atherosclerotic mice. ApoE^{-/-} mice were administrated with PBS or super-low-dose LPS together with HFD for 4 weeks. The frequency of Ly6G⁺ neutrophils in the peripheral blood (**A**) and spleen (**B**) was analyzed with flow cytometry. (**C**) Representative confocal images and quantification of Ly6G⁺ neutrophils in atherosclerotic plaques. Scale bar, 100 μ m. Data are representative of two independent experiments, and error bars represent means \pm s.e.m. * $P < 0.05$ and ** $P < 0.01$; Student's t-test (n= 5 to 6 for each group).

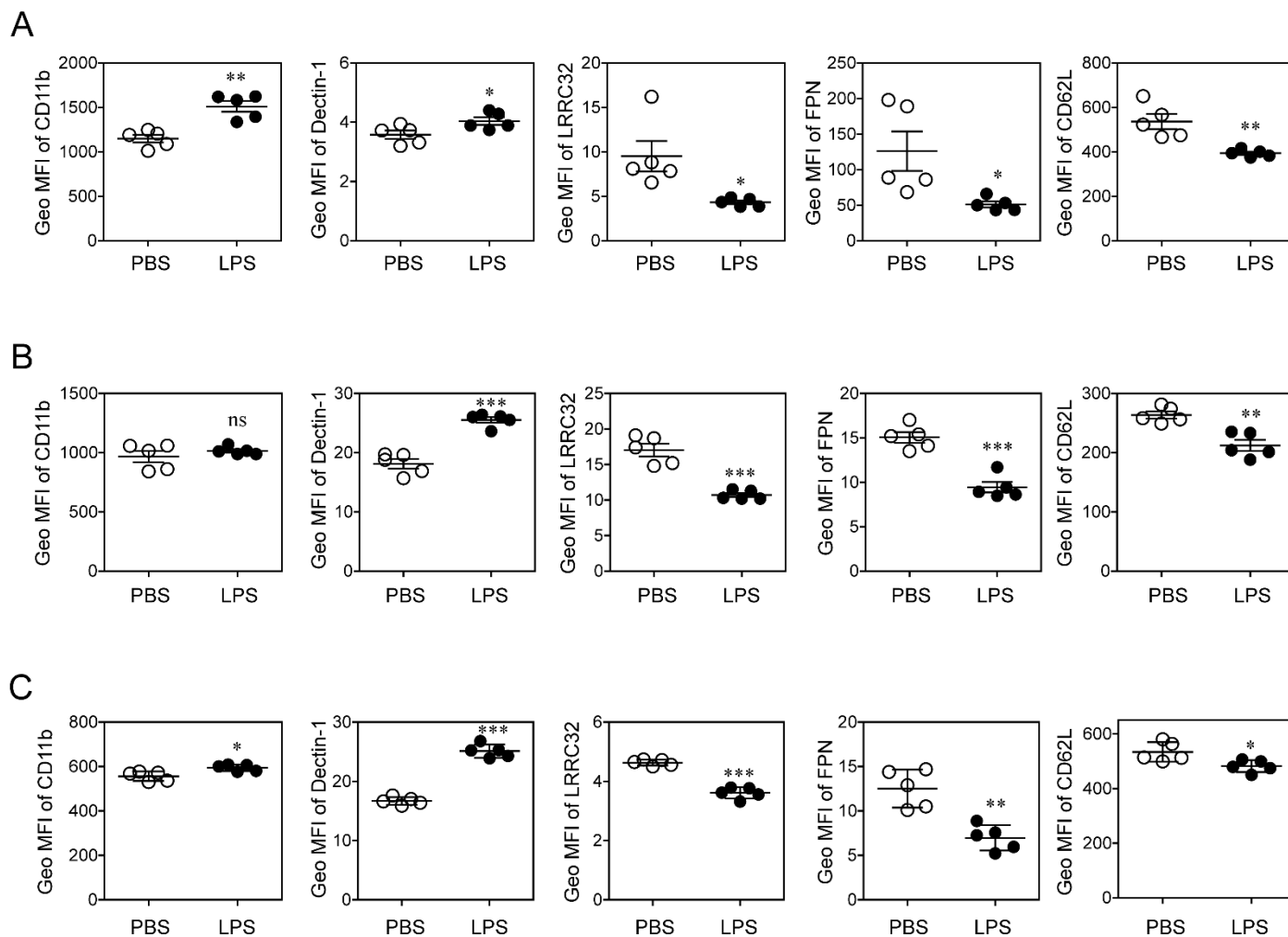


Fig. S4. Subclinical endotoxin primes neutrophils into a proinflammatory state in atherosclerotic mice. ApoE^{-/-} mice were administrated with PBS or super-low-dose LPS together with HFD for 4 weeks. The surface phenotypes of Ly6G⁺ neutrophils in the peripheral blood (A) spleen (B) and bone marrow (C) were analyzed by flow cytometry. The same data shown in Fig. 2 were re-analyzed with geometric mean fluorescence intensity (Geo MFI) as the parameter. Error bars represent means \pm s.e.m. * $P < 0.05$, ** $P < 0.01$ and *** $P < 0.001$; Student's t-test ($n = 5$ for each group).

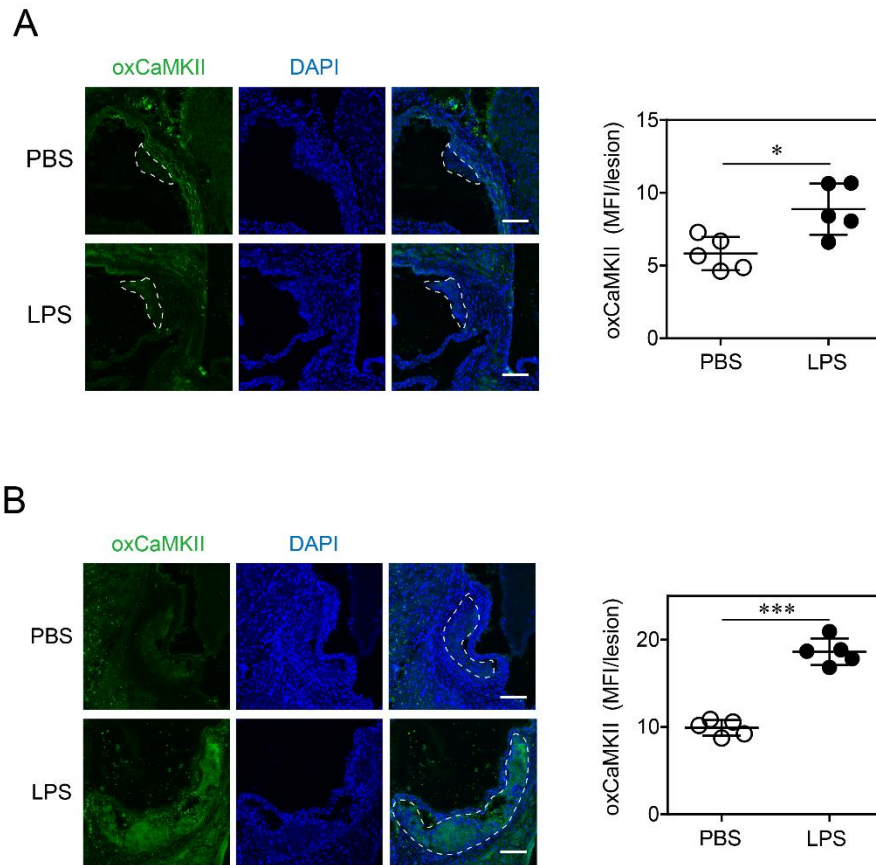


Fig. S5. Subclinical endotoxin induces oxCaMKII elevation in vivo. (A) ApoE^{-/-} mice were administrated with PBS or super-low-dose LPS together with RD for 4 weeks. Representative confocal images and quantification of lesional oxCaMKII levels are displayed. (B) ApoE^{-/-} mice were administrated with PBS or super-low-dose LPS together with HFD for 4 weeks. Representative confocal images and quantification of lesional oxCaMKII levels are displayed. Scale bar, 100 μ m. Data are representative of two independent experiments, and error bars represent means \pm s.e.m. * P < 0.05 and *** P < 0.001; Student's t-test (n = 5 for each group).

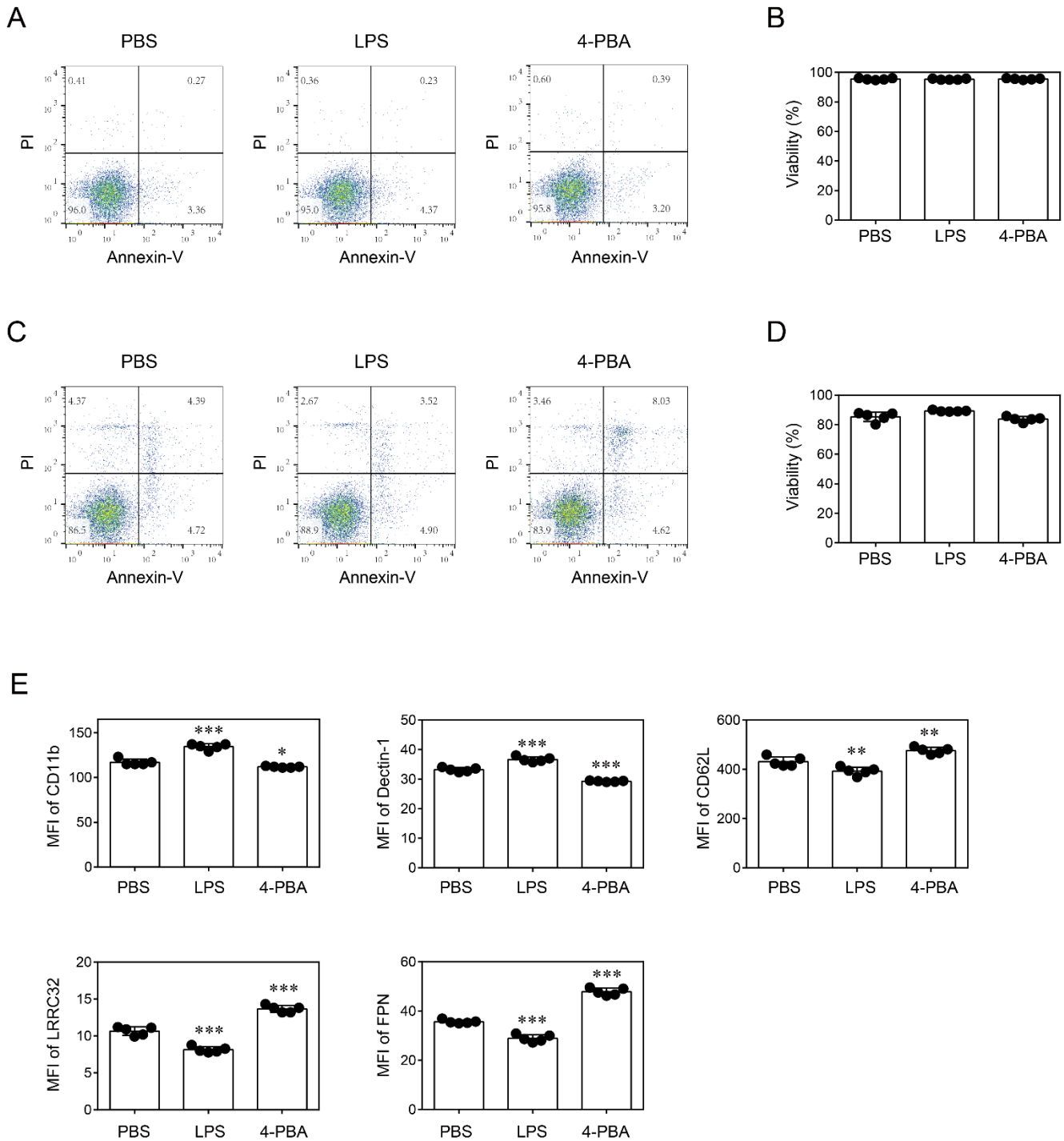


Fig. S6. Neutrophils maintain viability after in vitro polarization. Bone marrow neutrophils were purified and treated with PBS, super-low-dose LPS (100 pg/ml) or 4-PBA (1 mM) for 24 h (**A, B**) and 48 h (**C, D**). The cells were stained with Annexin-V and PI, and the viability (percentage of Annexin-V⁻/PI⁻ population) was determined by flow cytometry. (**E**) Surface phenotype of neutrophils was analyzed by flow cytometry after polarization for 24 h. Data are representative of three independent experiments, and error bars represent means \pm s.e.m. * $P < 0.05$, ** $P < 0.01$ and *** $P < 0.001$ as compared with PBS group; one-way ANOVA. (n = 5 for each group).

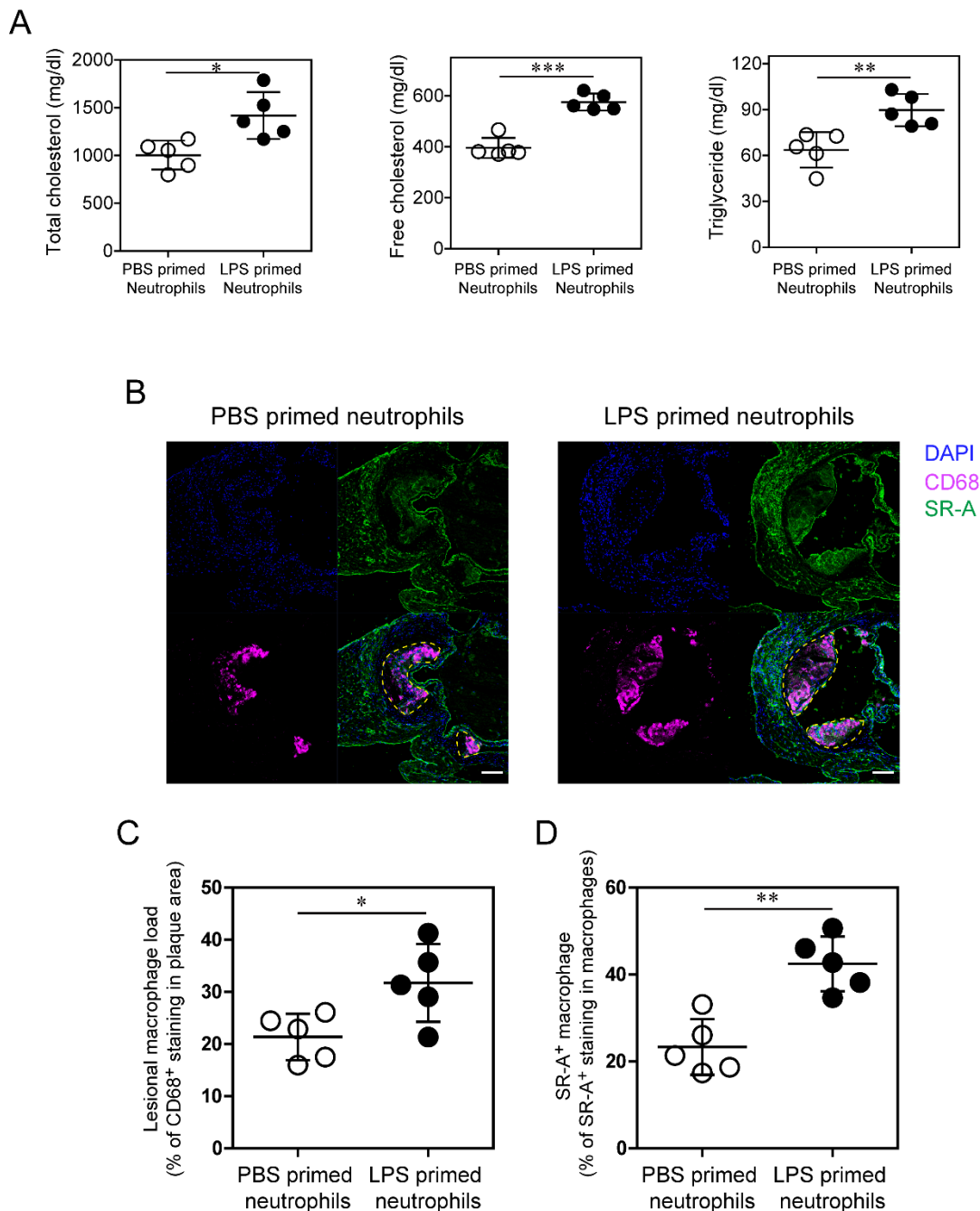


Fig. S7. Transfusion of superlow-dose LPS-polarized neutrophils elevates plasma lipid levels and modulates lesional macrophages. Neutrophils purified from ApoE^{-/-} mice were treated with PBS or super-low-dose LPS (100 pg/ml) for 24 h. PBS- or LPS-treated neutrophils (2×10^6 cells per mouse) were then adoptively transferred by intravenous injection to HFD-fed ApoE^{-/-} mice once a week for 4 weeks. **(A)** Plasma samples were collected 1 week after the last neutrophil transfer, and the levels of total cholesterol, free cholesterol and triglycerides were determined. **(B)** Representative confocal images of CD68 and SR-A staining in atherosclerotic plaques. Scale bar, 100 μ m. **(C)** Quantification of CD68⁺ macrophage load in plaques. **(D)** Quantification of the frequency of SR-A⁺ staining within macrophage population. Data are representative of two independent experiments, and error bars represent means \pm s.e.m. * $P < 0.05$, ** $P < 0.01$ and *** $P < 0.001$; Student's t -test ($n = 5$ for each group).

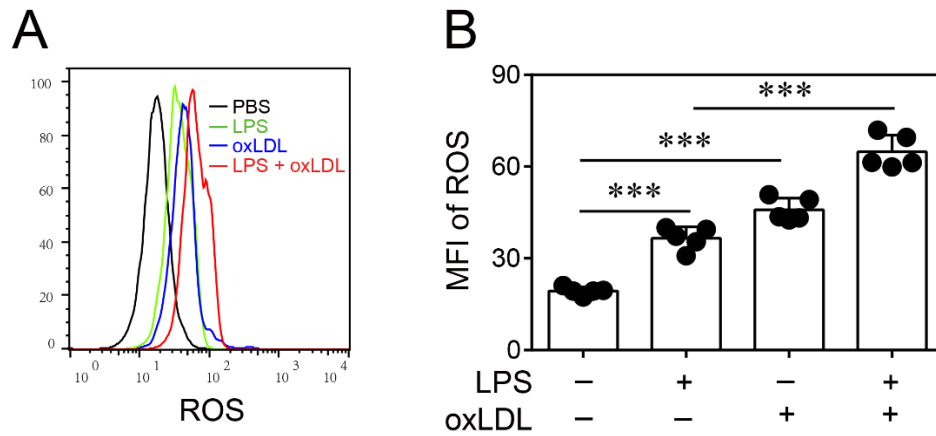


Fig. S8. Superlow-dose LPS and oxLDL treatment elevates ROS accumulation in neutrophils. Neutrophils were purified from the bone marrow of wild-type C57 BL/6 mice and treated with PBS, super-low dose LPS (100 pg/ml) and/or oxLDL (10 μ g/ml) for 2 d. **(A)** Representative histogram of ROS level determined by Cell ROX labeling. **(B)** Quantification of ROS level in neutrophils. Data are representative of three independent experiments, and error bars represent means \pm s.e.m. *** $P < 0.001$; ANOVA (n= 5 for each group).

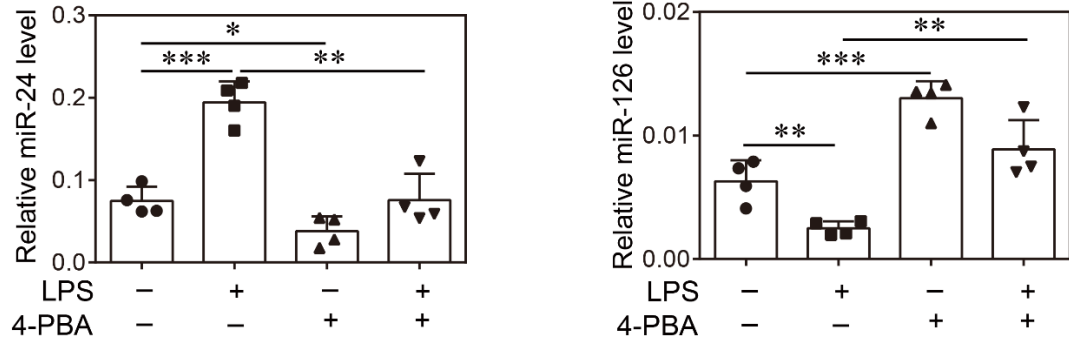


Fig. S9. 4-PBA reverses superlow-dose LPS-induced differential regulation of miR-24 and miR-126 in neutrophils. Neutrophils were purified from the bone marrow of wild-type C57 BL/6 mice and treated with PBS, super-low dose LPS (100 pg/ml) and/or 4-PBA (1 mM) for 2 d. The levels of miR-24 and miR-126 were determined by real-time RT-PCR. Data are representative of three independent experiments, and error bars represent means \pm s.e.m. * P < 0.05, ** P < 0.01 and *** P < 0.001; one-way ANOVA (n= 4 for each group).

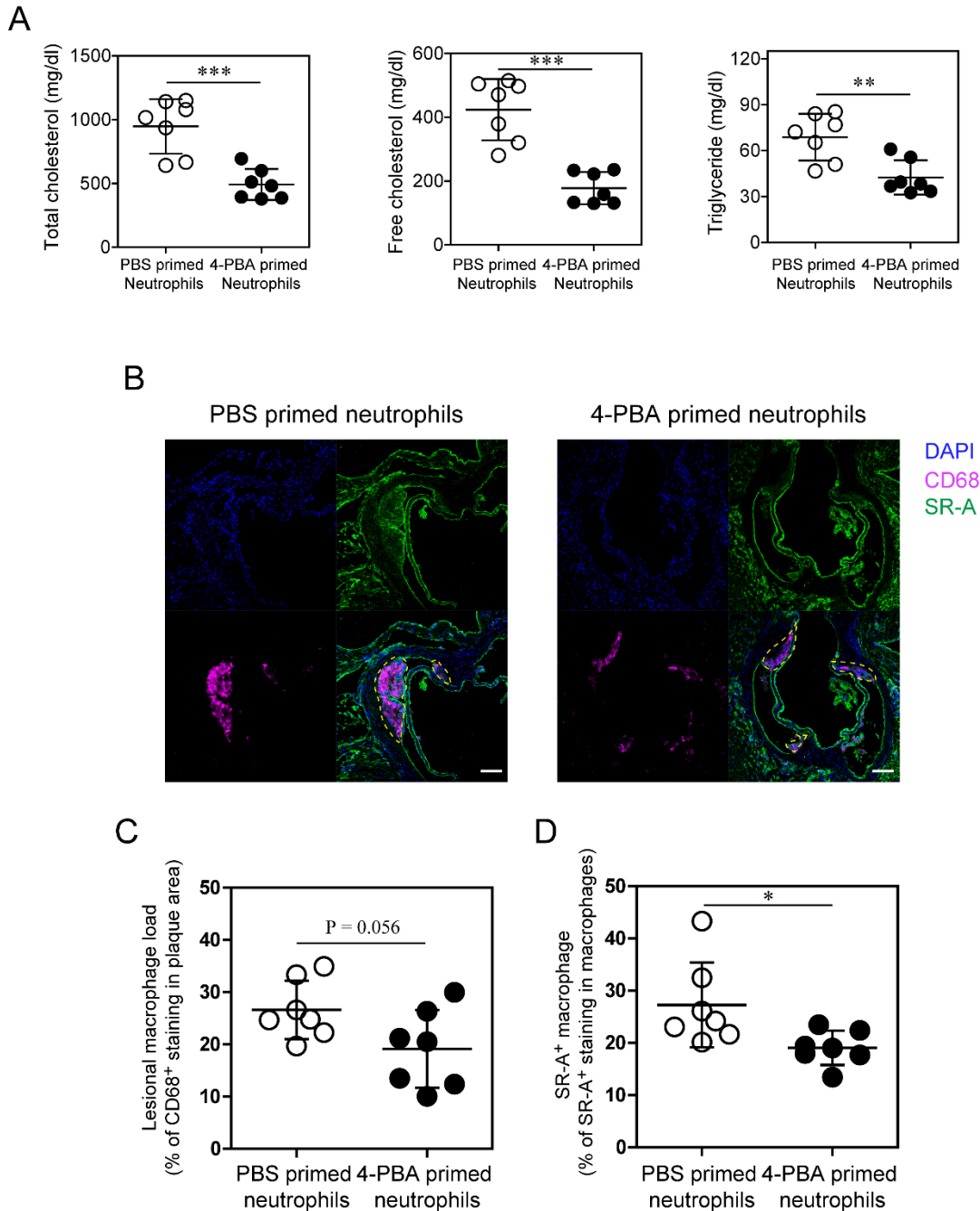


Fig. S10. Transfusion of 4-PBA-polarized neutrophils down-regulates plasma lipid levels and reduces lesional macrophage activation. Neutrophils purified from ApoE^{-/-} mice were treated with PBS or 4-PBA (1 mM) for 24 h. PBS- or 4-PBA-treated neutrophils (2×10^6 cells per mouse) were then adoptively transferred by intravenous injection to HFD-fed ApoE^{-/-} mice once a week for 4 weeks. **(A)** Plasma samples were collected 1 week after the last neutrophil transfer, and the levels of total cholesterol, free cholesterol and triglycerides were determined. **(B)** Representative confocal images of CD68 and SR-A staining in atherosclerotic plaques. Scale bar, 100 μ m. **(C)** Quantification of CD68⁺ macrophage load in plaques. **(D)** Quantification of the frequency of SR-A⁺ staining within macrophage population. Data are representative of two independent experiments, and error bars represent means \pm s.e.m. * $P < 0.05$, ** $P < 0.01$ and *** $P < 0.001$; Student's t -test ($n = 7$ for each group).