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## Supplementary Materials for

## Restriction of hepatitis B virus replication by c-Abl–induced proteasomal degradation of the viral polymerase

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Fig. S1. Capsid-associated viral DNA level in the treatment of TKIs.

(A-C) Same experiment as in Fig. 1A but treated with crizotinib (A) (n=4 per group), erlotinib (B)(n=3-4 per group) and ruxolitinib (C) (n=3 per group), respectively.



Fig. S2. c-Abl kinase reduces the level of HBV polymerase.

(A) Same experiment as in **Fig. 1G** but with HEK293T cells. (**B-C**) Immunoblot for polymerase protein level in WT or c-Abl KO Huh7 cells (**B**) or HepG2 cells (**C**). Total cell lysates were lysed 48 hrs after transfection and subjected to WB. (**D**) HEK293T cells were transfected with constructs expressing HA-polymerase, Flag-Arg, Flag-Arg-KR or empty vector control. After 24 hrs, cells were treated, or not treated, with 2  $\mu$ M imatinib and harvested 24 hrs later. Total cell lysates were then analyzed for the indicated proteins.



Fig. S3. c-Abl–CRL4<sup>Cdt2</sup> reduces HBV polymerase by promoting its ubiqutination but not transcription level.

(A) Quantitation of polymerase mRNA by real-time PCR. mRNA purified from transfected HEK293T cells were diluted and the relative polymerase mRNA level without co-transfected Flag-Abl was set to 100% and compared to others. Statistical significance is indicated by asterisks above the brackets (n=8-10 per group). (B) Ubiquitination of polymerase by IP HA in Huh7 cells expressing HA-polymerase, Flag-ub and siDDB1, siCUL4A, siCUL4B or a combination of siCUL4A and

siCUL4B and treated with MG132 for 8 hrs. HA immunoprecipitates (top) or total cell lysates (bottom) were then analyzed by western blot. (**C**) Quantitation of polymerase mRNA by real-time PCR. mRNA purified from transfected HEK293T cells were diluted and analyzed by real-time PCR (top). The relative polymerase mRNA level without co-transfected Flag-Abl was set to 100% and compared to others (n=3-4 per group). Whole-cell lysates were prepared to subject to WB (bottom). (**D**) Co-immunoprecipitation of Myc-DDB1 with HA-polymerase and Cdt2 in WT or c-Abl KO HepG2 cells over-expressing HA-polymerase and Myc-DDB1.



Fig. S4. Inhibition of CRL4 E3 ubiquitin ligase enhances HBV replication by stabilizing viral polymerase.

(A) HepG2 cells and (B) Huh7 cells were co-transfected with plasmids expressing pHBV, pHBV(Pol-HA) $\Delta X$  and HBx, and capsid-associated viral DNA were quantitated. Mean copy number from cells transfected with pHBV was set to 100% and compared to others (n=3-4 per group). (C) HepG2 cells were and (D) Huh7 cells transfected with indicated siRNAs and compHBV. Whole-cell lysates were prepared to subject to WB (bottom) and capsid-associated viral DNA were quantitated (top). Mean copy number from cells transfected with control siRNA was set to 100% and compared to others (n=3-4 per group). (E) HepG2 cells and (F) Huh7 were transfected with siCdt2 and compHBV. Whole-cell lysates were prepared to subject to WB (bottom) and capsid-associated viral DNA were quantitated (top). Mean copy number from cells transfected with control siRNA was set to 100% and compared to others (n=3-4 per group). (G, H) Real-time PCR assay to determine the efficiency of Cdt2 knockdown in HepG2 cells (G) and Huh7 cells (H) (n=3-4 per group). (I) HepG2 cells and (J) Huh7 cells were transfected with compHBV and treated with DMSO, MG132 or MLN4924. Whole-cell lysates were prepared to subject to WB (bottom) and capsid-associated viral DNA were quantitated (top). Mean copy number from cells treated DMSO was set to 100% and compared to others (n=3-4 per group). (K) Detection of GFP in fresh liver tissue sections from the indicated mice hydrodynamic tail vein injected into GFP or not.



Fig. S5. Western blot-scanned films. Red boxes highlight lanes used in Fig. 1-4



Fig. S6. Western blot-scanned films. Red boxes highlight lanes used in Fig. 4-6 and fig. S1-4