Supplementary material

Ginsenosides from Korean red ginseng modulate T cell function via the regulation of NF-AT-mediated IL-2 production.

Le Ba Vinh^{a,b}, Jung Up Park^c, Le Xuan Duy^{a,e}, Nguyen Thi Minh Nguyet^a, Seo Young Yang^{a,**}, Young Ran Kim^{c,**}, and Young Ho Kim^{a,**}

^a College of Pharmacy, Chungnam National University, Daejeon 34134, Republic of Korea

^b Institute of Marine Biochemistry (IMBC), Vietnam Academy of Science and Technology (VAST), 18 Hoang Quoc Viet, Caugiay, Hanoi, Vietnam

^c College of Pharmacy and Research Institute of Drug Development, Chonnam National University, Gwangju, 500-757, Republic of Korea

^e Institute of Natural Products Chemistry, Vietnam Academy of Science and Technology (VAST), 18 Hoang Quoc Viet, Caugiay, Hanoi, Vietnam

** To whom correspondence should be addressed:

Email: whk@cnu.ac.kr (Kim, Y. H.);

CONTENTS

	Pages
General experimental procedures	3
Extraction and isolation	3

General experimental procedures

Optical rotations were determined on a JASCO P-2000 polarimeter. IR spectra were obtained on a Bruker TENSOR 37 FT-IR spectrometer (Bruker Optics, Ettlingen, Germany). The high-resolution electrospray ionization mass spectra (HR-ESI-MS) were gained using an AGILENT 6530 Accurate-Mass Q-TOF LC/MS system. The ¹H NMR (600 MHz) and ¹³C NMR (150 MHz) spectra were recorded on a Bruker Avance 600 MHz spectrometer (Billerica, MA). TMS was used as an internal standard. Column chromatography (CC) was performed on silica gel (Kieselgel 60, 70–230 mesh and 230–400 mesh, Merck, Darmstadt, Germany) and YMC RP-18 resins (30 - 50 μ m, Fuji Silysia Chemical Ltd., Kasugai, Aichi, Japan). Thin layer chromatography (TLC) was performed on silica gel 60 F₂₅₄ (1.05554.0001, Merck, Darmstadt, Germany) and RP-18 F_{254S} plates (1.15685.0001, Merck, Darmstadt, Germany) and compounds were visualized by spraying with aqueous 10% H₂SO₄ solution, followed by heating.

Extraction and isolation

Korean red ginseng (2.4 kg) was extracted three times in refluxing (H₂O). The water extract (600 g) was separated through a Diaion (HP-20) CC eluting with a gradient solvent mixture of MeOH in H₂O (0 %, 50 %, and 100 %) to yield three fractions (W1 to W3), based on TLC analysis. The fraction W2 (250 g) was crudely separated with silica gel CC eluting with gradient solvent systems of CH₂Cl₂-MeOH (25:1-1:0) to yield seven fractions (W2A-W2G), respectively. The fraction W2C (10 g) was then isolated by MPLC RP-18 CC using acetone-water (0.5:1, v/v) to give six subfractions (W2C1-W2C6). Purification of the subfraction W2C3 (3 g) using adsorbents Sephadex LH-20 resin, silica gel and the eluent CH₂Cl₂-MeOH (10:1) gave compounds **1** (8.7 mg), **2** (7.8 mg), and **3** (12.5 mg). Repeating the same steps above, subfraction W2C4 (2 g) was purified on a Sephadex LH-20 and silica gel columns with the eluent CH₂Cl₂-MeOH (10:1) to give compounds **4** (27.8 mg), and **5** (16.4 mg).

Similarly, the fraction W2E (12 g) was separated via MPLC RP-18 CC with a gradient solvent mixture of MeOH-H₂O (1:1-1:0, v/v) and purified with Sephadex LH-20 column by mixture of MeOH- acetone -H₂O (7.5:1:1, v/v/v) to obtain compounds **6** (15.7 mg), **9** (30.6 mg), and **11** (10.2 mg). Next, the fraction W2G was purified by silica gel CC using solvent mixture of CH₂Cl₂- EtOAc-MeOH (7:1:1, v/v/v) to yield four subfractions (W2G1-W2G4). The subfraction W2G2 was isolated by Sephadex LH-20 column and further purified with RP-18 column using MeOH-H₂O (4:1) to afford compounds **7** (7.5 mg), **8** (20.3 mg), and **10** (9.7 mg).