

Supporting Information

Linear and Differential Ion Mobility Separations of Middle-Down Proteoforms

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Table S1. Table of features distinguished in TIMS spectra for all variants (from MeOH solution, $t_{\text{ramp}} = 500$ ms): cross sections and measured resolving power values.

Figure S1. Relative (approximate) cross sections for K9ac (dominant peaks). Lines guide the eye through trends below and above the transition region.

Figure S2. TWIMS spectra for selected mixtures of me3, p, and me variants measured in N₂ using solvent (i) (solid black lines) with fits by scaled individual traces from Fig. 1 (colored lines) and their computed additions (dotted lines).

Figure S3. TWIMS spectra for selected variant mixtures measured in N₂ with solvent (i) using the traveling wave speeds of 650, 1000, and 1900 m/s.

Figure S4. Pairwise linear correlations between transit times for ac and me₃ variants at the traveling wave speeds of 650 and 1000 m/s. Variants with only three data points and p variants with partial data at $s = 1000$ m/s are not included.

Figure S5. TWIMS spectra measured with different ESI solvents: (a) four me₃ variants ($z = 9$) with solvents (i, ii, iii), (b) K23me₃ ($z = 8$ and 9) with solvents (i, iv).

Figure S6. TWIMS spectra for four me₃ variants in $z = 12$ measured in N₂ using solvent (ii) and $s = 650$ m/s. No signal at 12+ was found for the K4me₃ variant.

Figure S7. TIMS spectra (on the cross section scale) measured for exemplary me₃, ac, p, and me variant species using (a) $t_{\text{ramp}} = 100 - 500$ ms from MeOH/H₂O solutions, (b) from MeOH/H₂O and aqueous solutions.

Figure S8. TIMS spectra for selected mixtures of p variants and traces for the two components.

Figure S9. Linear correlations between transit times for me₃, ac, and p variants across charge state pairs (major peaks from Fig. 1). Variants for me (with three data points) are not included.

Figure S10. Linear correlations between separation parameters in TIMS for the acetylated and phosphorylated peptide variants in different charge states.

Figure S11. Linear correlations between separation parameters in Synapt and TIMS for the

acetylated and phosphorylated peptide variants.

Figure S12. Linear correlations between FAIMS and TIMS (left column) or TWIMS (right column) separations for (a) acetylated and (b) phosphorylated variants, r^2 marked. To facilitate comparison, the left column in (a) repeats the data from Fig. 6 in the text.

Figure S13. Linear correlations between FAIMS and TIMS (left) or TWIMS (right) separations for same acetylated (a) and phosphorylated (b) variants with different charge states selected in the two dimensions (r^2 marked).

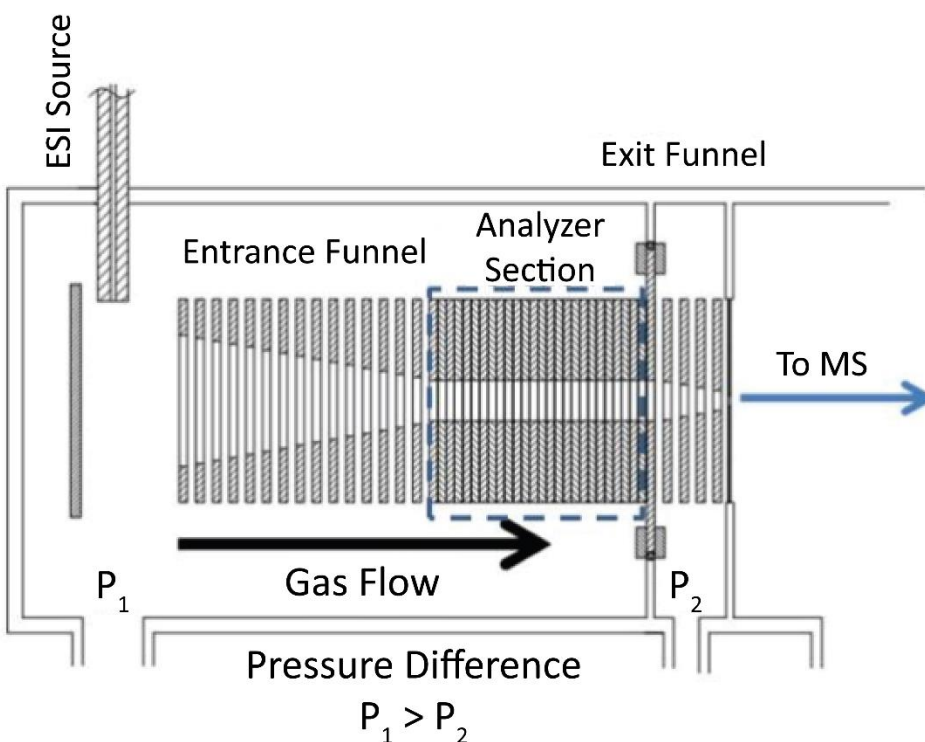
Details of nESI/TIMS hardware and mobility calibration

Nano-ESI source

Custom nESI emitters were made from quartz glass capillaries (I.D. 0.7 mm, O.D. 1.0 mm) utilizing a P-2000 micropipette laser puller (Sutter Instruments, Novato, CA). The inlet capillary was a stainless steel tube ($1/16 \times 0.020''$, IDEX Health Science, Oak Harbor, WA).

TIMS cell

The TIMS cell is an electrodynamic ion funnel with three regions (see scheme below): entrance, analyzer tunnel (46 mm length), and exit. The rf voltage creates a quadrupolar field in the tunnel and dipolar field in other regions.



Mobility calibration in TIMS

Multiple ion species are first trapped and thermalized at different E values set up by voltage profile along the tunnel, then pushed out by “ramping” (decreasing) E in steps with each species eluting at a characteristic voltage ($V_{\text{elution}} - V_{\text{out}}$). The total time an ion spends in the system (from gating into the TIMS cell to registration at the ToF detector) is:

$$t_{\text{total}} = t_{\text{trap}} + (V_{\text{elution}}/V_{\text{ramp}}) \times t_{\text{ramp}} + t_{\text{ToF}} = t_0 + (V_{\text{elution}}/V_{\text{ramp}}) \times t_{\text{ramp}},$$

where t_{trap} is the trapping time, t_{ToF} is the flight time from cell exit to detector (through MS vacuum), V_{ramp} and t_{ramp} are the voltage range and time of ramp, and t_0 is the constant time before and after the separation step. The V_{elution} and t_0 values for each m/z of interest are determined simultaneously by varying t_{ramp} from 100 to 500 ms at fixed V_{ramp} . More details on this procedure are found in Ref. [50].

Table S1.

Collision cross sections (CCS, Å²) and resolving power (*R*) values for protonated histone tails measured using TIMS.

Peptide	Ion	CCS (Å ²), std. error of mean: ± 0.04%	<i>R</i>
K9me3	[M+6H] ⁶⁺	1303	137
	[M+7H] ⁷⁺	1398/1406	203/180
	[M+8H] ⁸⁺	1574/1607	96/102
	[M+9H] ⁹⁺	1685/1710/1735/1749	153/119/141/211
	[M+10H] ¹⁰⁺	1835	170
	[M+11H] ¹¹⁺	1921	201
K23me3	[M+6H] ⁶⁺	1308	92
	[M+7H] ⁷⁺	1405/1437	82/72
	[M+8H] ⁸⁺	1508/1535/1574/1609	188/284/146/223
	[M+9H] ⁹⁺	1684/1707/1751	167/76/233
	[M+10H] ¹⁰⁺	1830/1839	218/236
	[M+11H] ¹¹⁺	1926/1943	244/183
K36me3	[M+6H] ⁶⁺	1291/1357	128/137
	[M+7H] ⁷⁺	1394/1422/1441	59/133/119
	[M+8H] ⁸⁺	1483/1510	215/265
	[M+9H] ⁹⁺	1658/1702	240/122
	[M+10H] ¹⁰⁺	1833	176
	[M+11H] ¹¹⁺	1948	229
K9ac	[M+6H] ⁶⁺	1209	105
	[M+7H] ⁷⁺	1290	145
	[M+8H] ⁸⁺	1395/1404/1420/1435/1447/1473	199/187/202/144/90/184
	[M+9H] ⁹⁺	1529/1563/1572/1603	191/223/197/255
	[M+10H] ¹⁰⁺	1673/1694/1715	222/121/159
	[M+11H] ¹¹⁺	1784	133
K14ac	[M+6H] ⁶⁺	1205	120
	[M+7H] ⁷⁺	1298/1322	137/85
	[M+8H] ⁸⁺	1404/1411/1427/1434/1442/1451	134/168/153/163/170/45
	[M+9H] ⁹⁺	1541/1562/1572/1587/1625/1640	220/112/210/122/216/262
	[M+10H] ¹⁰⁺	1701	150
	[M+11H] ¹¹⁺	1785/1814	223/151
K18ac	[M+6H] ⁶⁺	1207	72
	[M+7H] ⁷⁺	1287/1310	239/218
	[M+8H] ⁸⁺	1396/1405/1418/1428/1435/1454/1480	208/156/153/111/197/117/159
	[M+9H] ⁹⁺	1531/1563/1579/1611	247/223/198/201
	[M+10H] ¹⁰⁺	1694	179
	[M+11H] ¹¹⁺	1786	192
K27ac	[M+6H] ⁶⁺	1210	145
	[M+7H] ⁷⁺	1294/1304/1326	223/215/221

	[M+8H] ⁸⁺	1412/1420/1438/1452/1459/1484/1498	157/144/150/190/131/206/156
	[M+9H] ⁹⁺	1558/1572/1616	180/194/171
	[M+10H] ¹⁰⁺	1695	131
	[M+11H] ¹¹⁺	1789	145
	[M+6H] ⁶⁺	1217	124
	[M+7H] ⁷⁺	1294/1301/1315	193/239/198
K36ac	[M+8H] ⁸⁺	1373/1396/1406/1425/1444/1456/1475	177/47/201/88/79/209/88
	[M+9H] ⁹⁺	1526/1555/1575/1605/1621	209/216/108/189/80
	[M+10H] ¹⁰⁺	1683	155
	[M+11H] ¹¹⁺	1770/1792	197/201
	[M+6H] ⁶⁺	1305	161
	[M+7H] ⁷⁺	1386/1412	75/150
T3p	[M+8H] ⁸⁺	1556/1571/1596	134/118/114
	[M+9H] ⁹⁺	1653/1691/1734	221/72/73
	[M+10H] ¹⁰⁺	1786	194
	[M+11H] ¹¹⁺	1878	146
	[M+6H] ⁶⁺	1319	134
	[M+7H] ⁷⁺	1412	159
T6p	[M+8H] ⁸⁺	1500/1523/1532/1568/1581/1612	70/206/174/204/144/269
	[M+9H] ⁹⁺	1669/1714/1749	121/163/192
	[M+10H] ¹⁰⁺	1805	160
	[M+11H] ¹¹⁺	1921	209
	[M+6H] ⁶⁺	1300	122
	[M+7H] ⁷⁺	1401/1414/1425	246/267/285
S10p	[M+8H] ⁸⁺	1511/1520/1534/1553/1565/1582/1593/1611	232/179/197/219/142/115/306/298
	[M+9H] ⁹⁺	1665/1706/1736/1753/1767	86/74/235/428/89
	[M+10H] ¹⁰⁺	1783/1800/1852	244/243/197
	[M+11H] ¹¹⁺	1917	167
	[M+6H] ⁶⁺	1307/1327	82/140
	[M+7H] ⁷⁺	1380/1396/1423	88/170/169
S28p	[M+8H] ⁸⁺	1515/1554/1579	205/187/106
	[M+9H] ⁹⁺	1695/1726	138/149
	[M+10H] ¹⁰⁺	1829	116
	[M+11H] ¹¹⁺	1924	163
	[M+6H] ⁶⁺	1270	155
	[M+7H] ⁷⁺	1362/1379	91/136
Y41p	[M+8H] ⁸⁺	1464/1499/1529/1553	229/259/118/174
	[M+9H] ⁹⁺	1627/1691/1659/1744	258/148/210/208
	[M+10H] ¹⁰⁺	1824	182
	[M+11H] ¹¹⁺	1922/1869	180/228
	[M+6H] ⁶⁺	1218	129
K4me	[M+7H] ⁷⁺	1303/1315/1326	200/146/199
	[M+8H] ⁸⁺	1414/1428/1439/1452/1465/1493/1506/1517	89/291/175/278/144/138/232/124

	[M+9H] ⁹⁺	1627/1638/1651	148/252/229
	[M+10H] ¹⁰⁺	1695/1730/1750	213/122/221
	[M+11H] ¹¹⁺	1791/1799	167/189
	[M+6H] ⁶⁺	1212	135
	[M+7H] ⁷⁺	1301/1324	190/221
K9me	[M+8H] ⁸⁺	1426/1438/1473/1499/1511	247/173/142/180/140
	[M+9H] ⁹⁺	1574/1591/1628/1640	237/175/119/149
	[M+10H] ¹⁰⁺	1704	97
	[M+11H] ¹¹⁺	1787	167
	[M+6H] ⁶⁺	1218	151
	[M+7H] ⁷⁺	1302/1309/1326/1336	193/73/165/69
K23me	[M+8H] ⁸⁺	1416/1430/1439/1453/1468/1494/1507	153/164/212/153/120/199/260
	[M+9H] ⁹⁺	1561/1576/1625/1635	244/146/140/178
	[M+10H] ¹⁰⁺	1700	90
	[M+11H] ¹¹⁺	1800	143

Figure S1

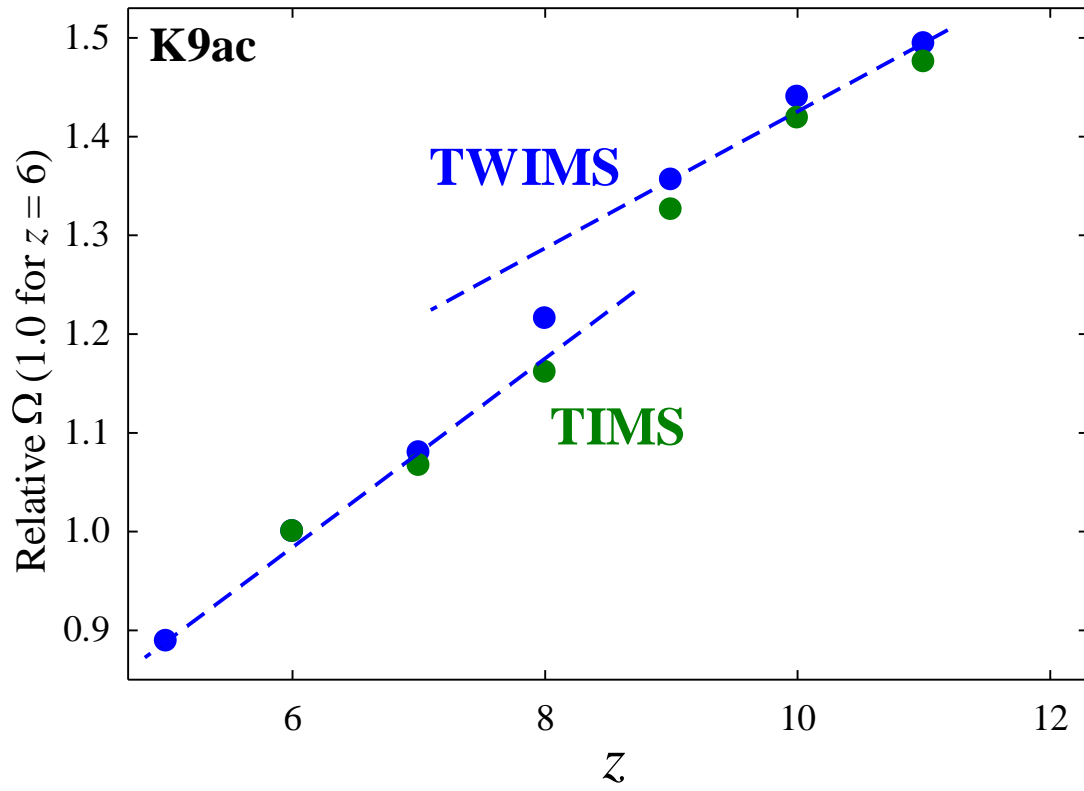


Figure S2

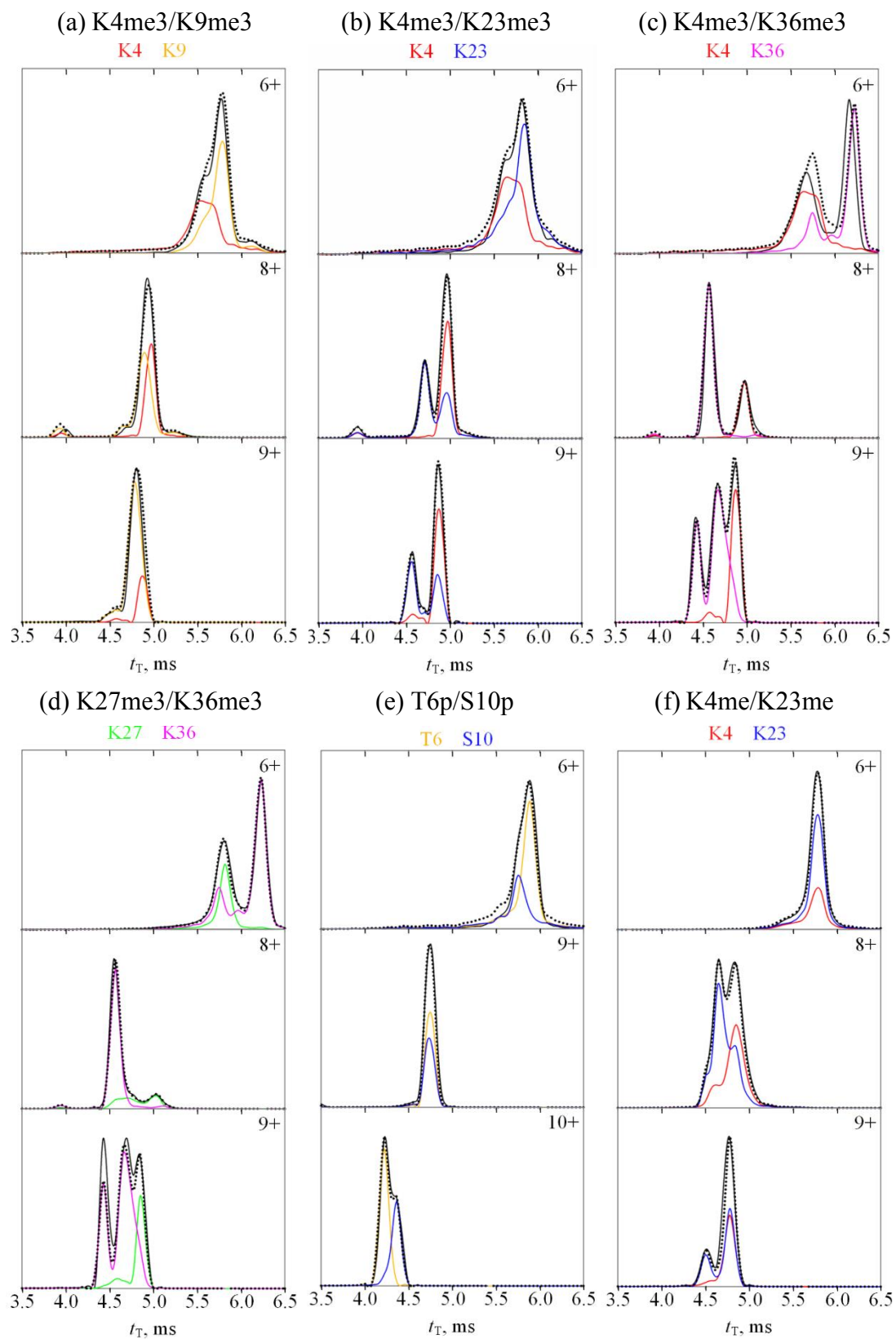


Figure S3

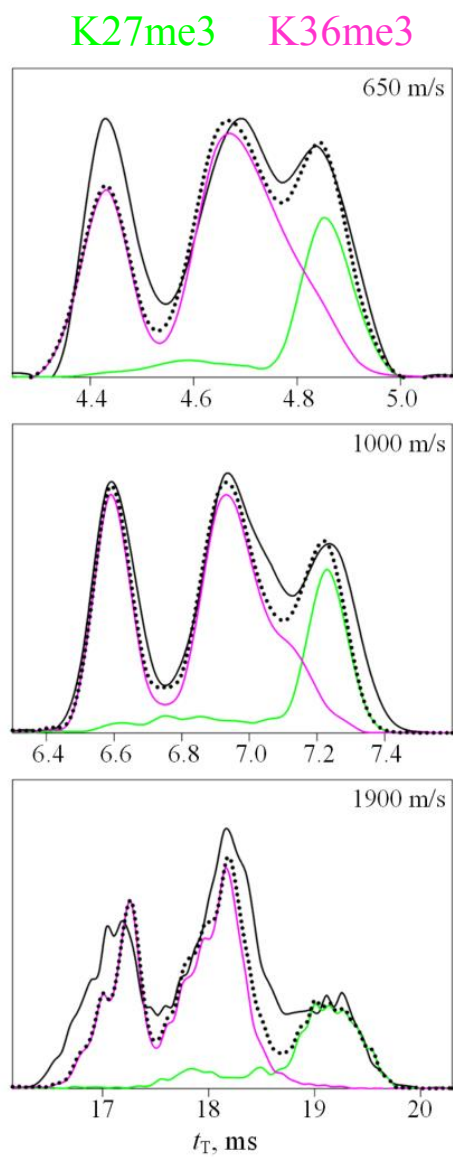


Figure S3 (continued)

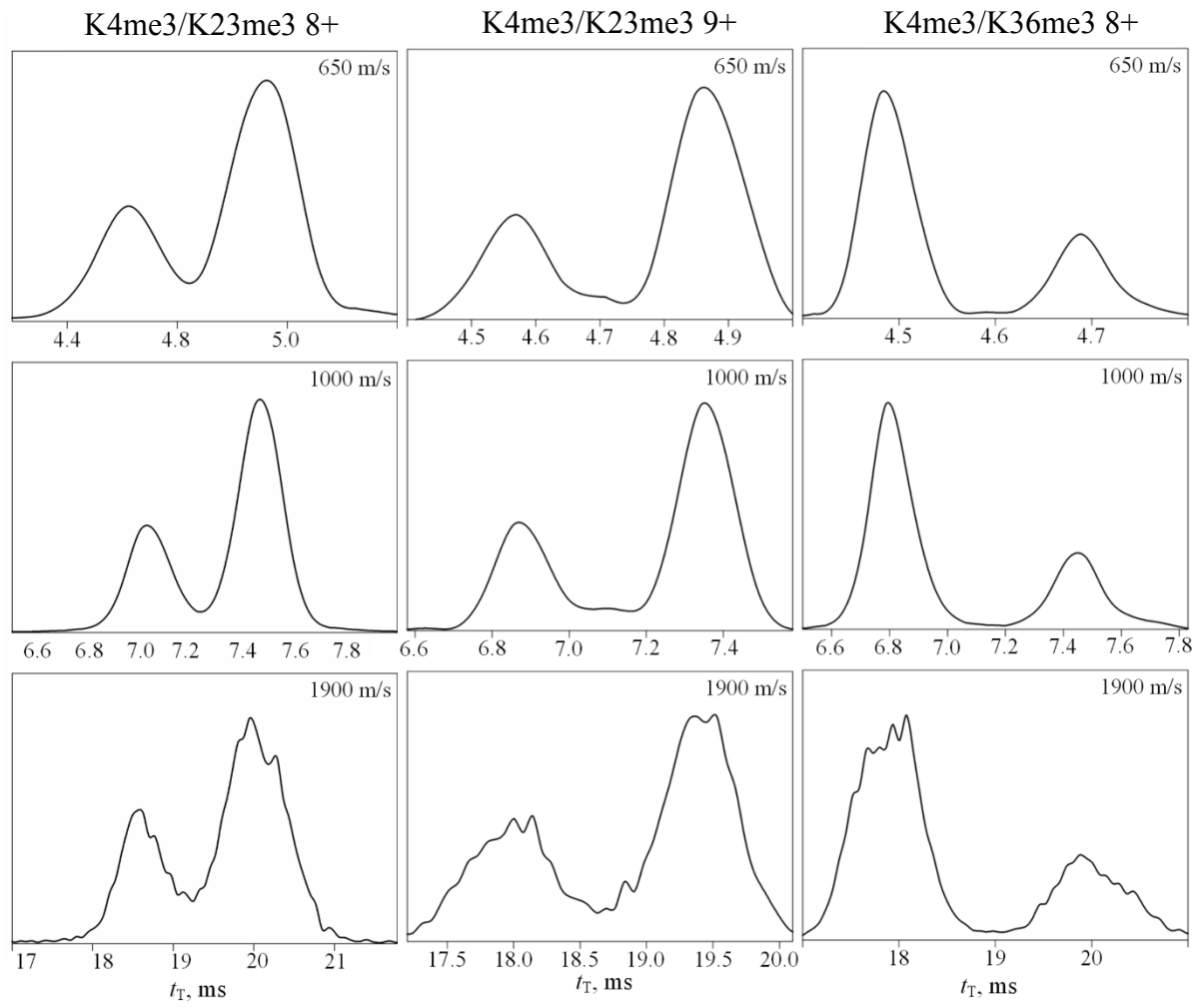


Figure S3 (continued)

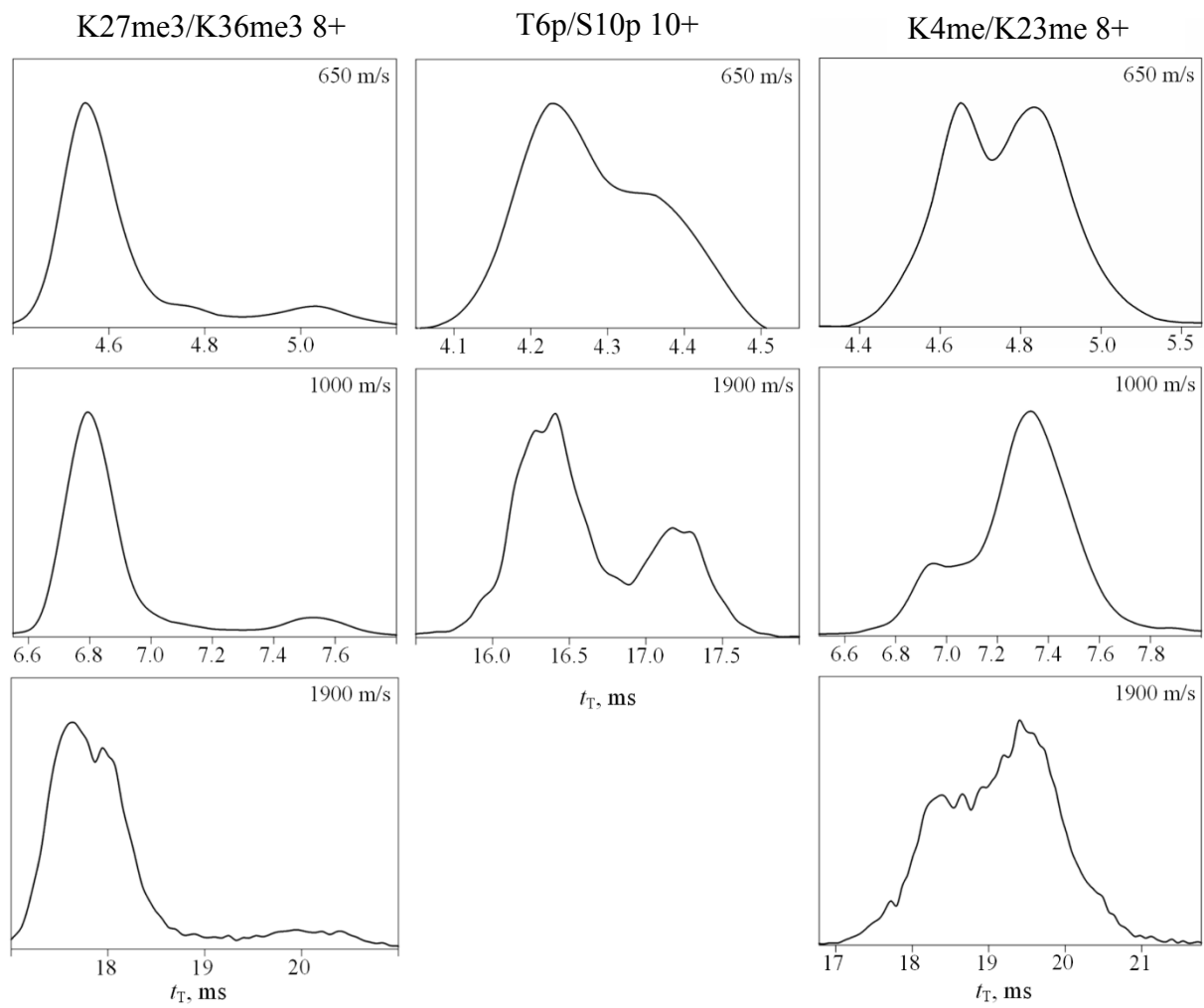


Figure S4

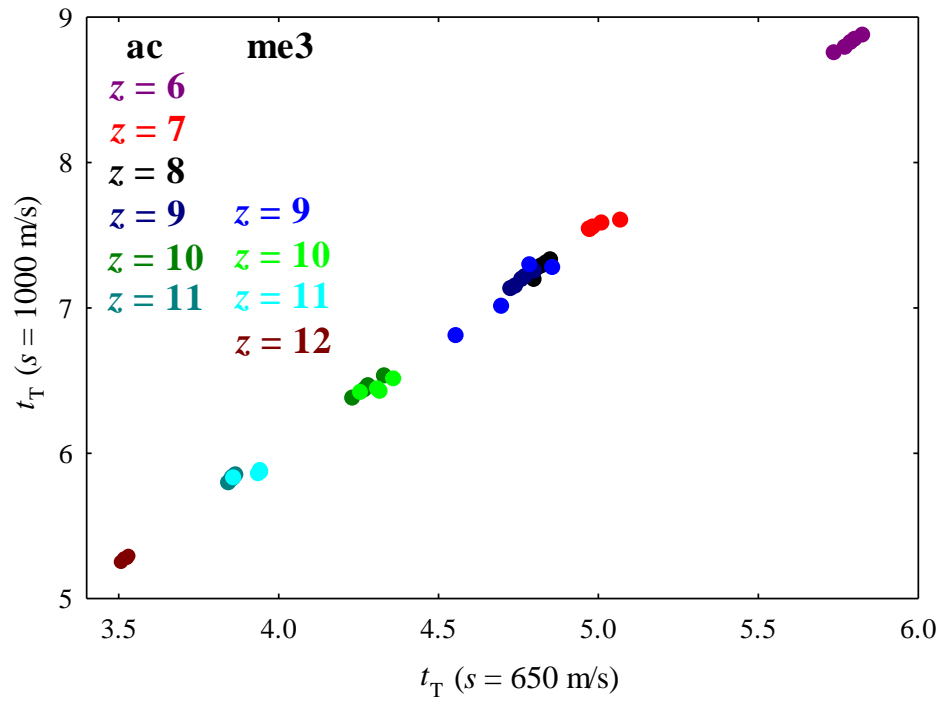
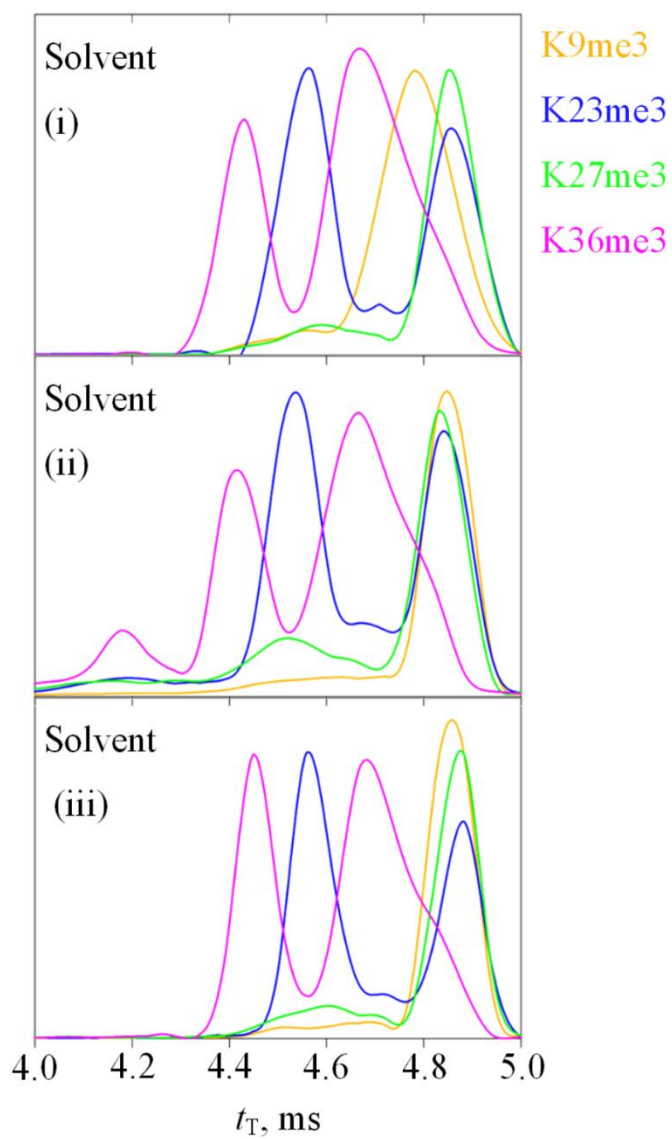


Figure S5

(a)



(b)

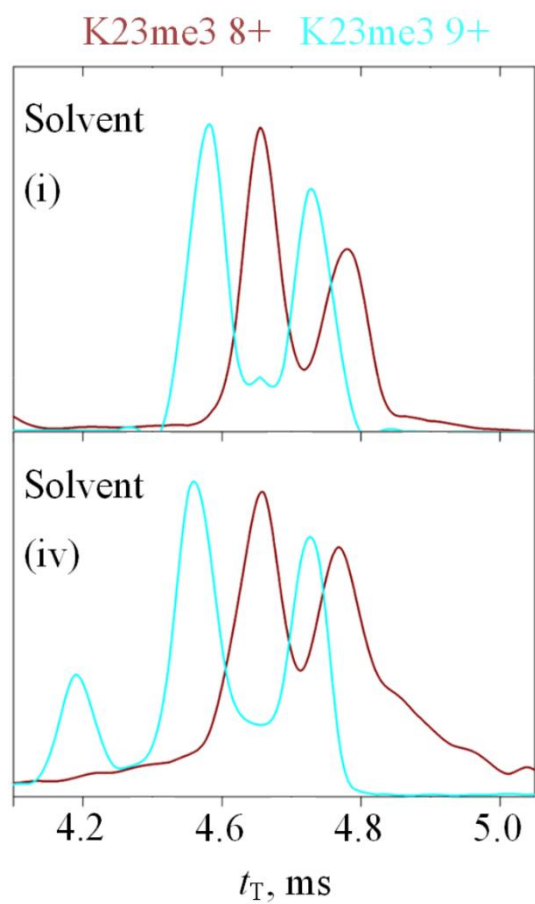


Figure S6

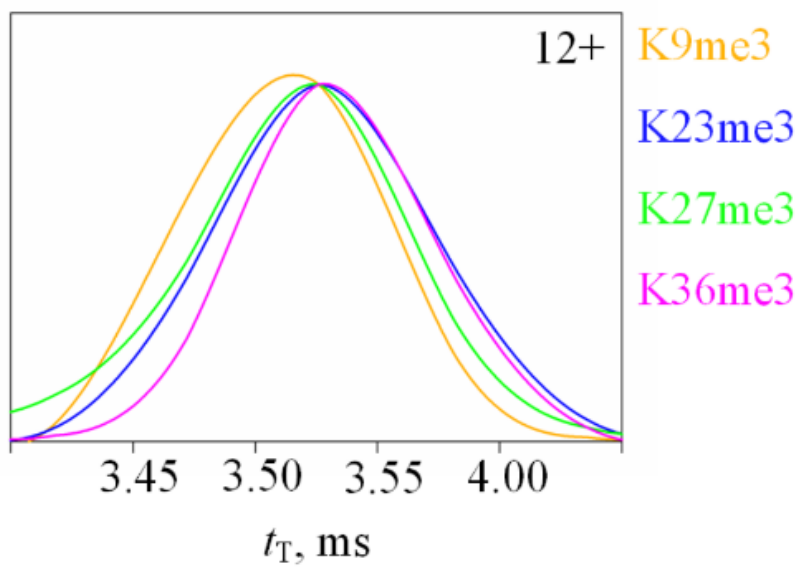


Figure S7

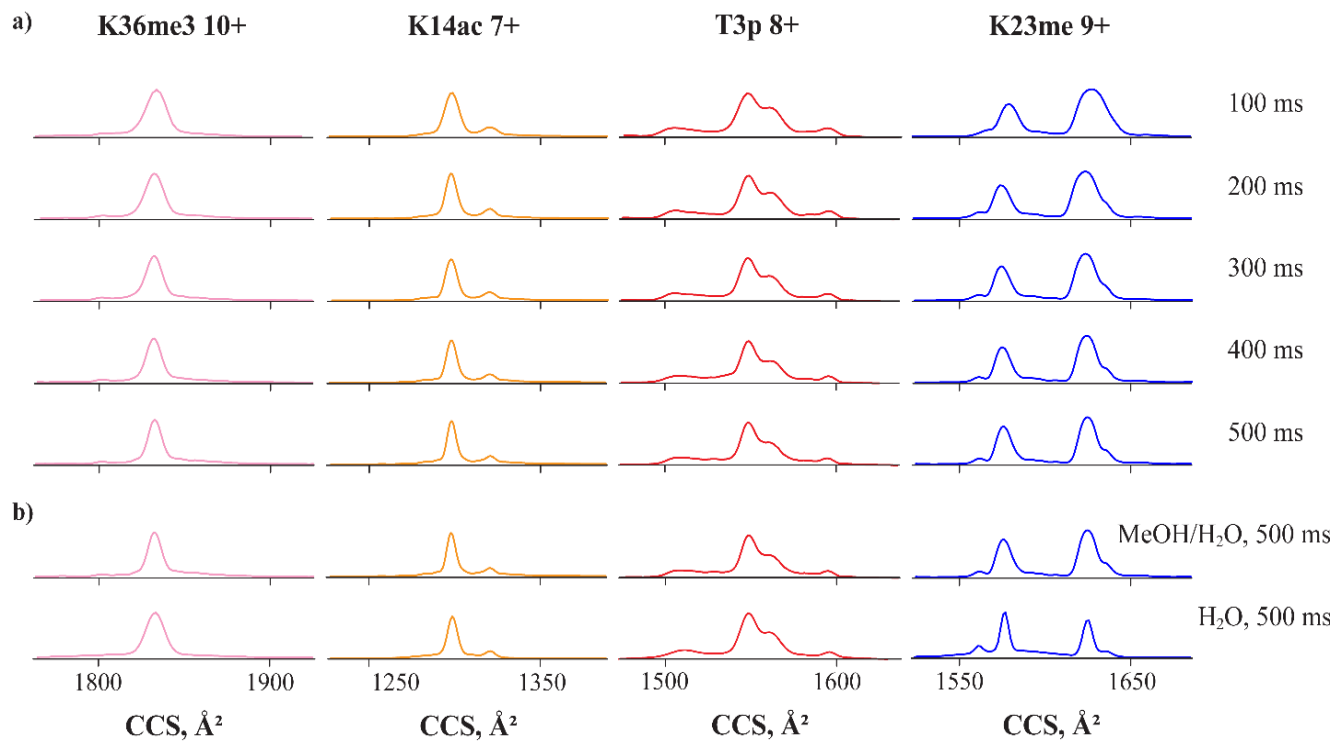
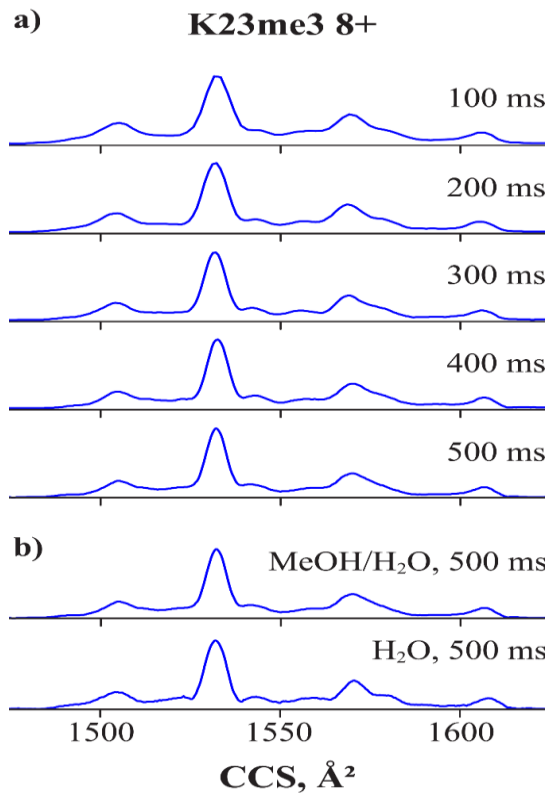


Figure S8

Phosphorylated variants, 10+

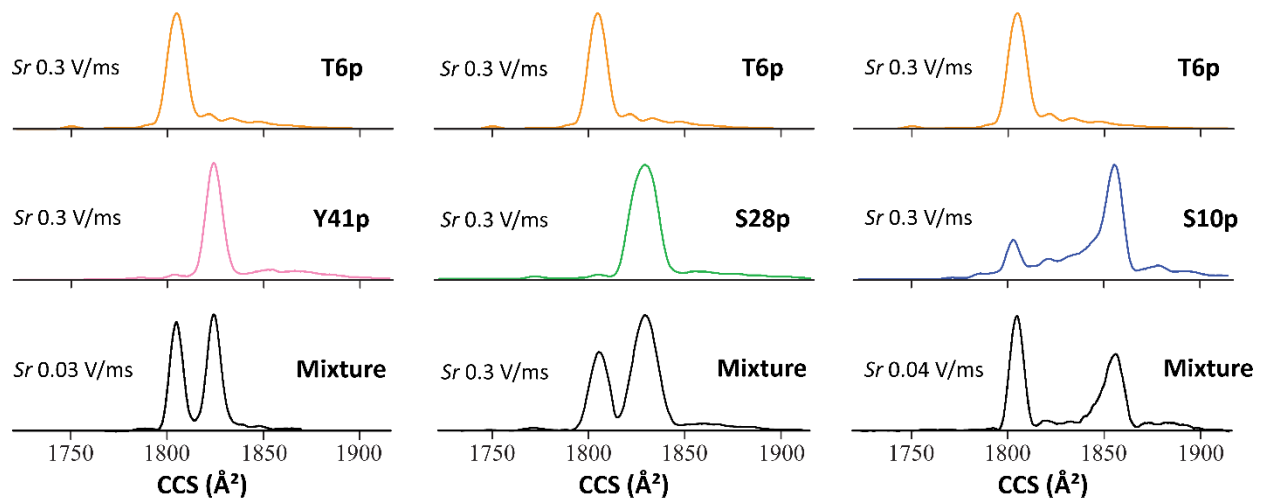


Figure S9

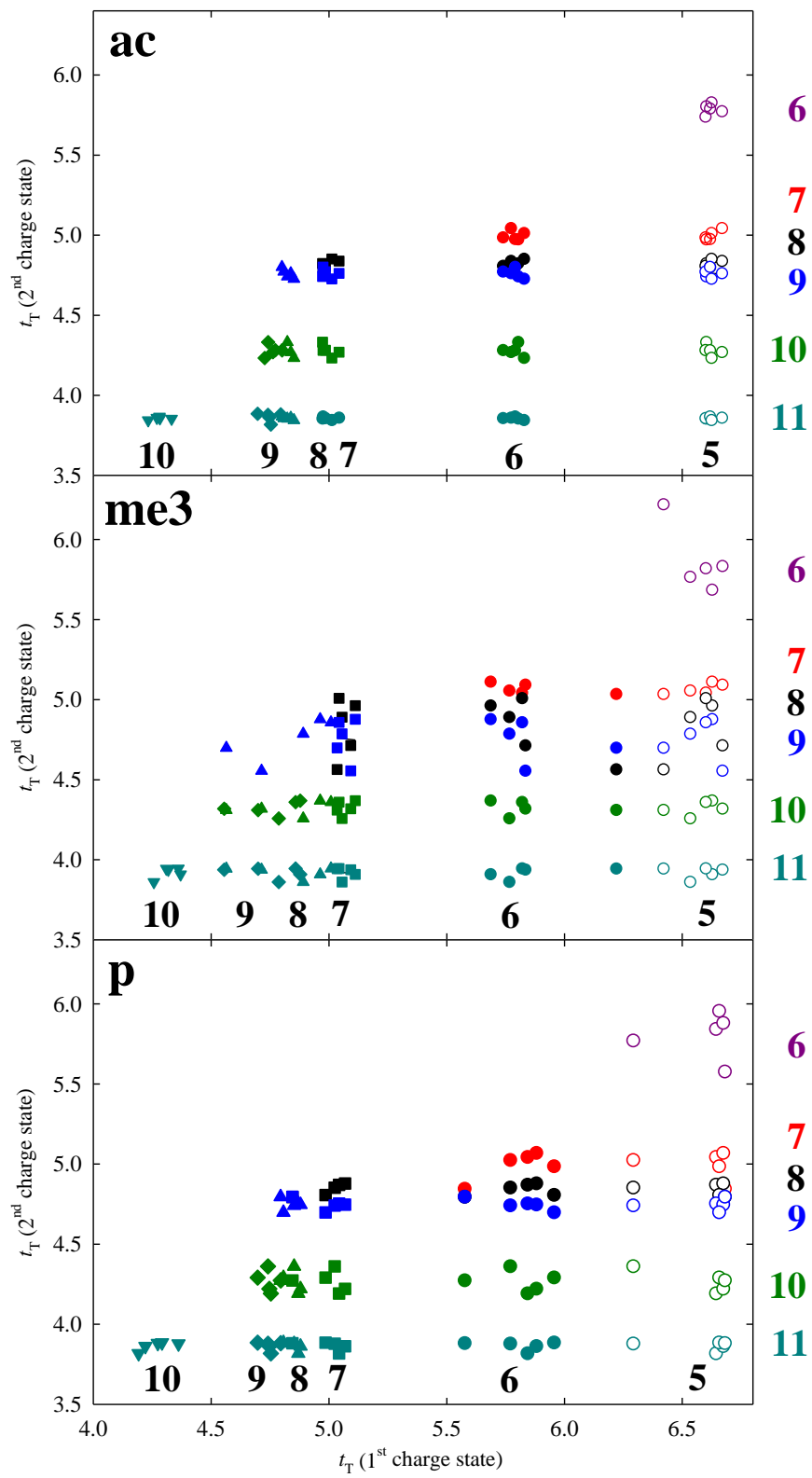
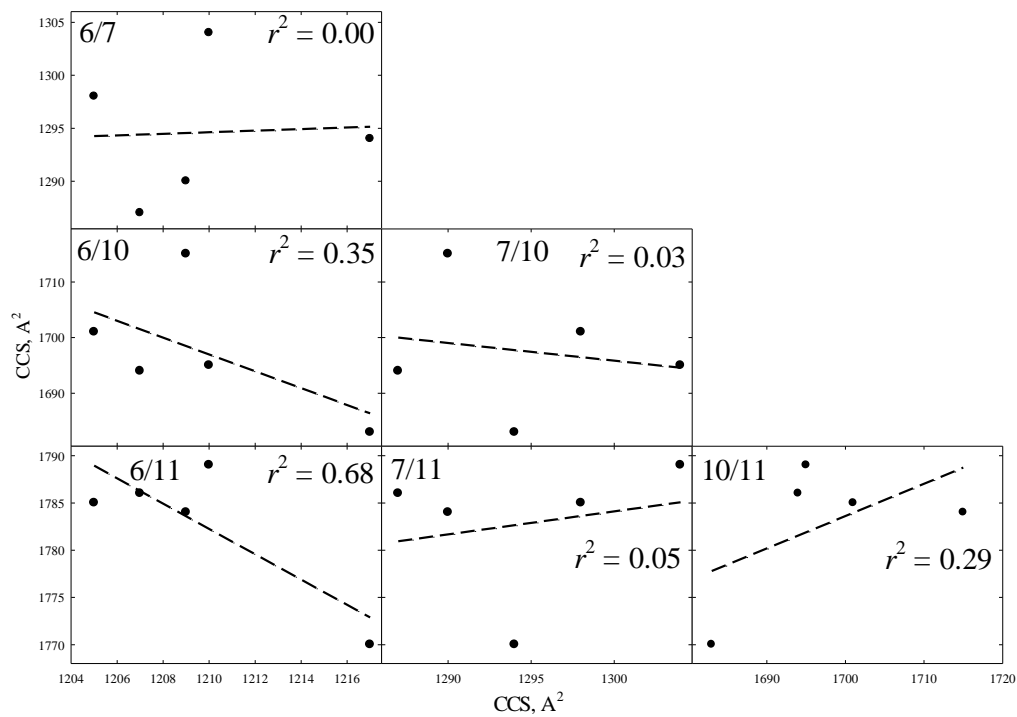


Figure S10

Acetylated



Phosphorylated

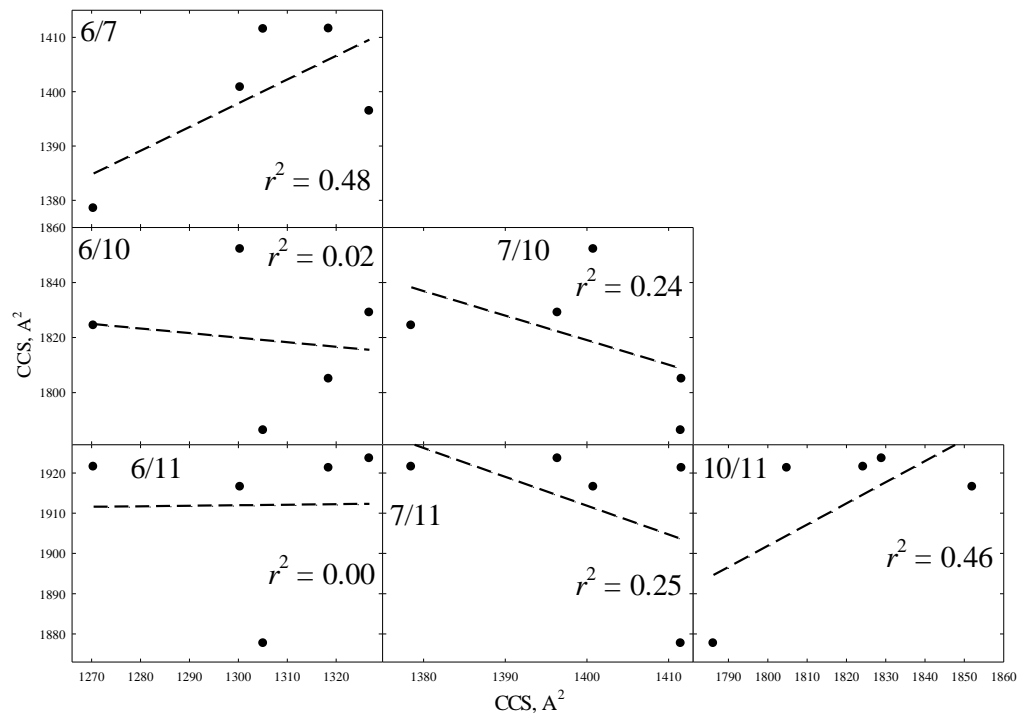


Figure S11

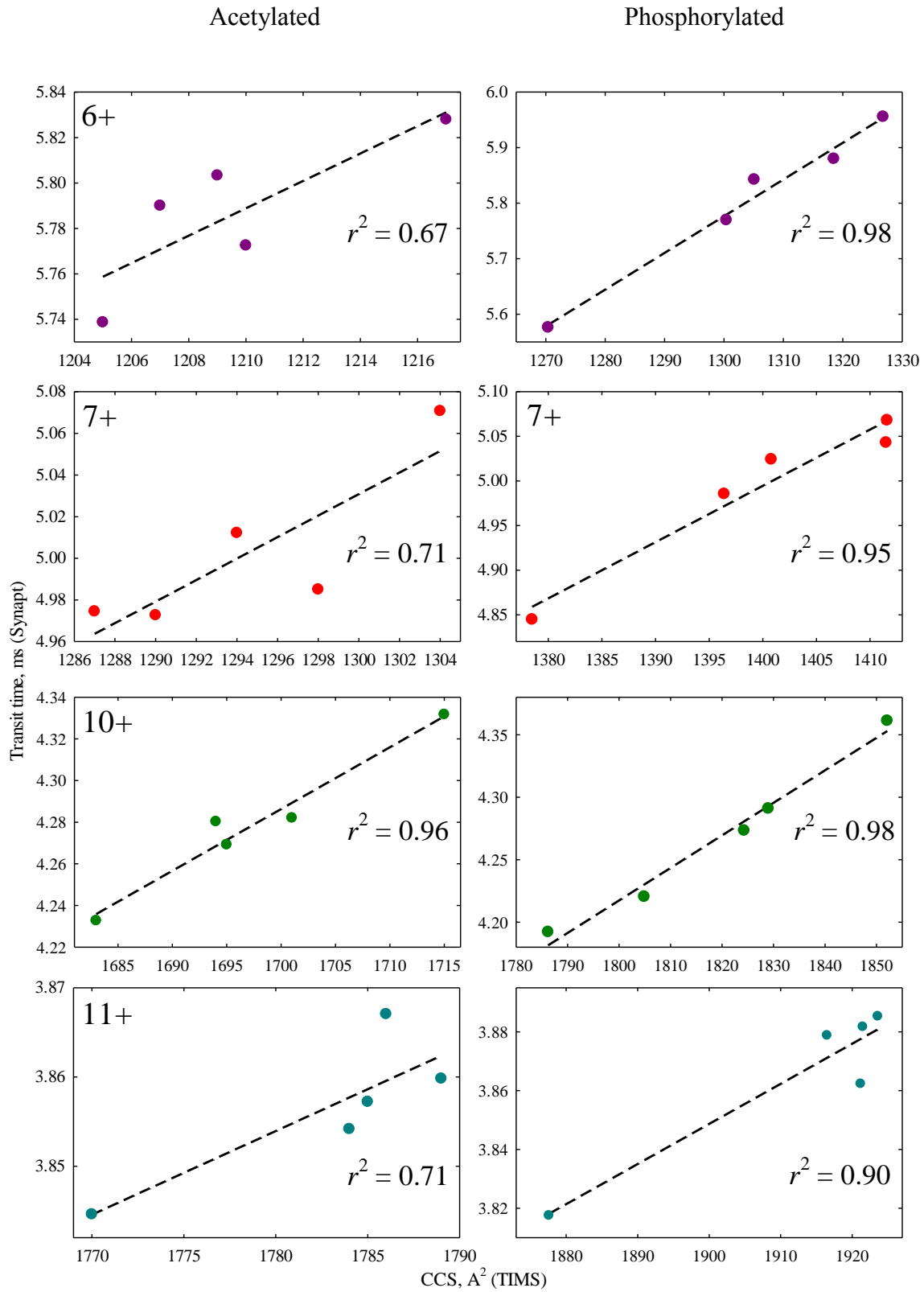


Figure S12a

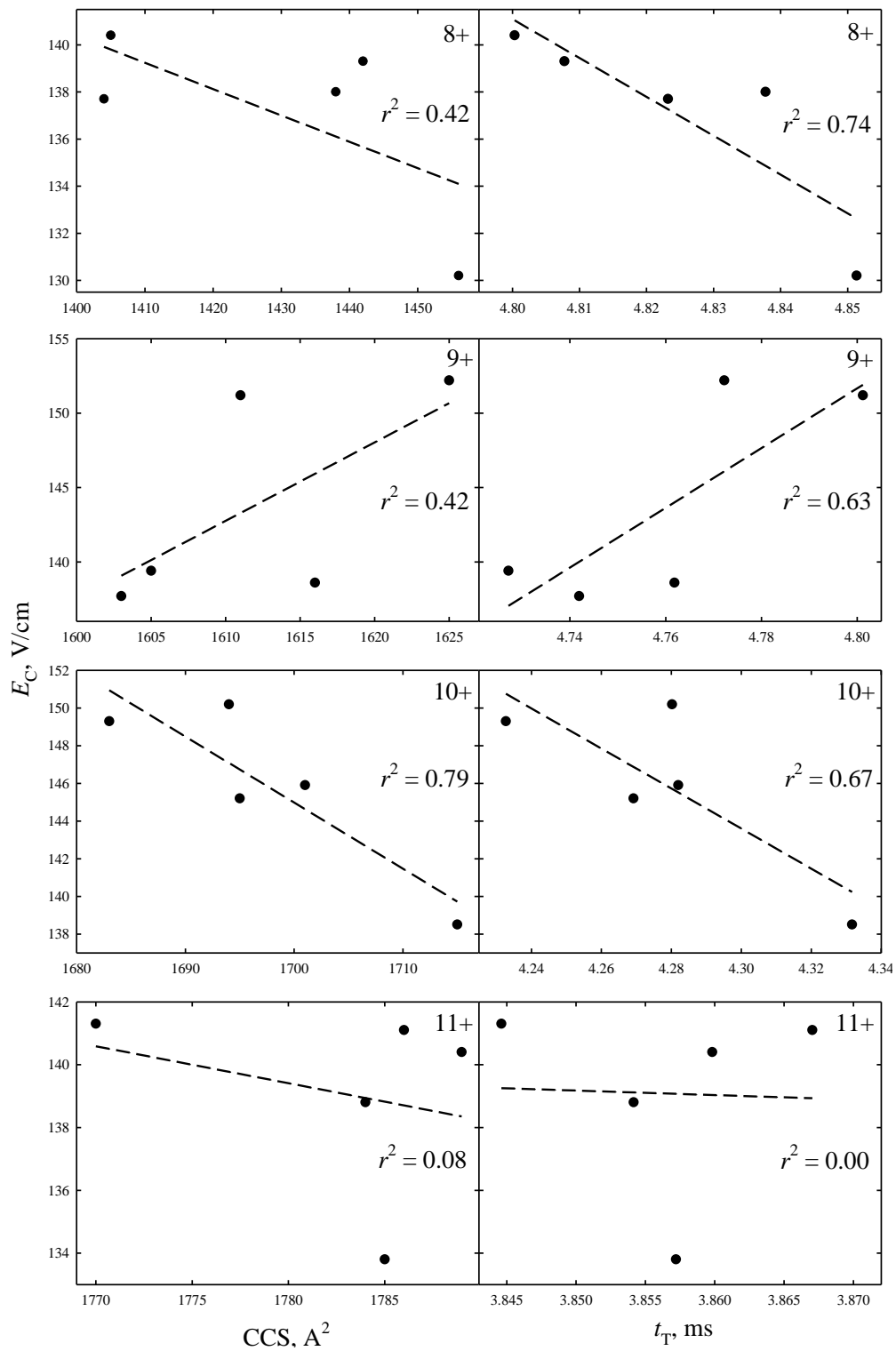


Figure S12b

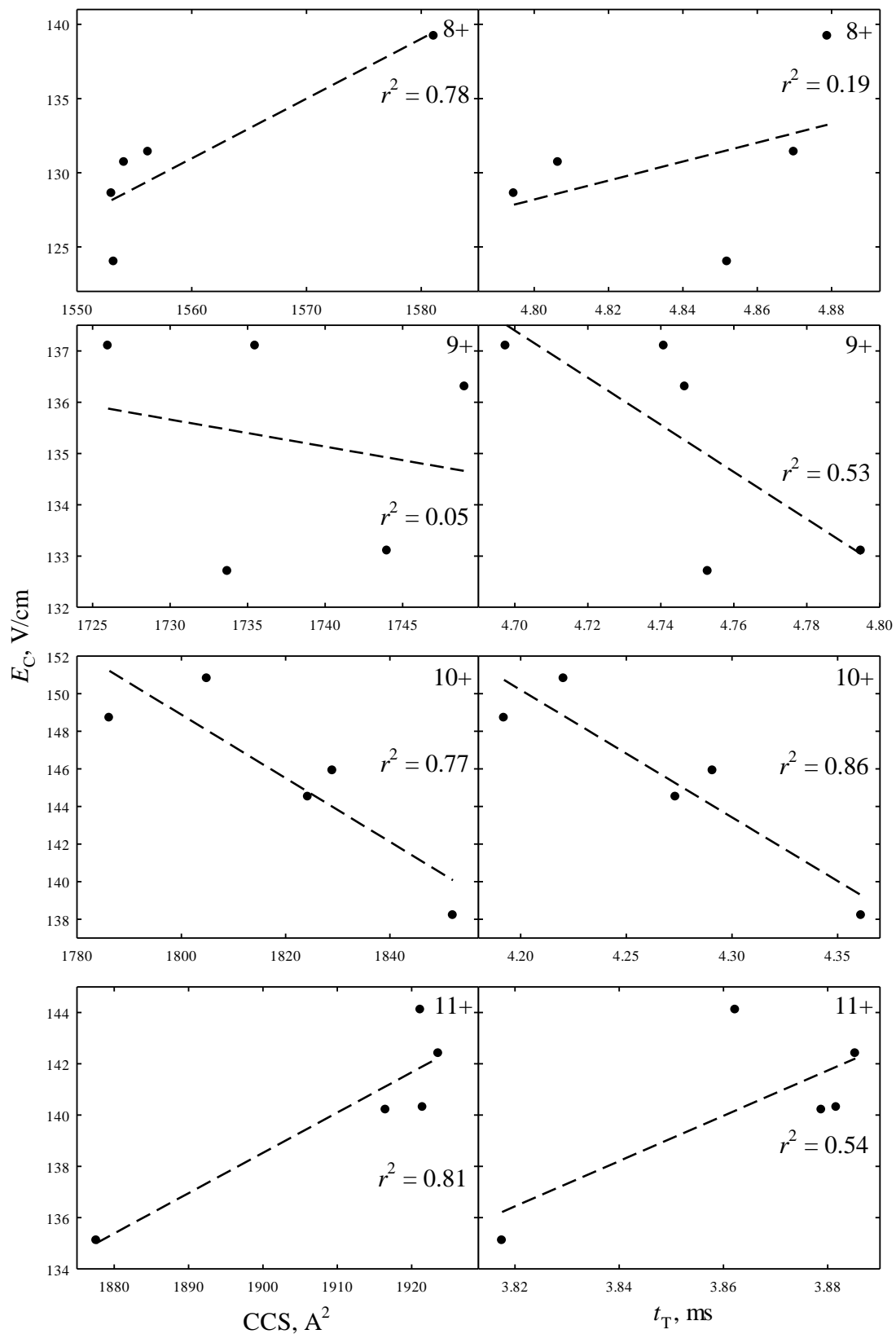


Figure S13a

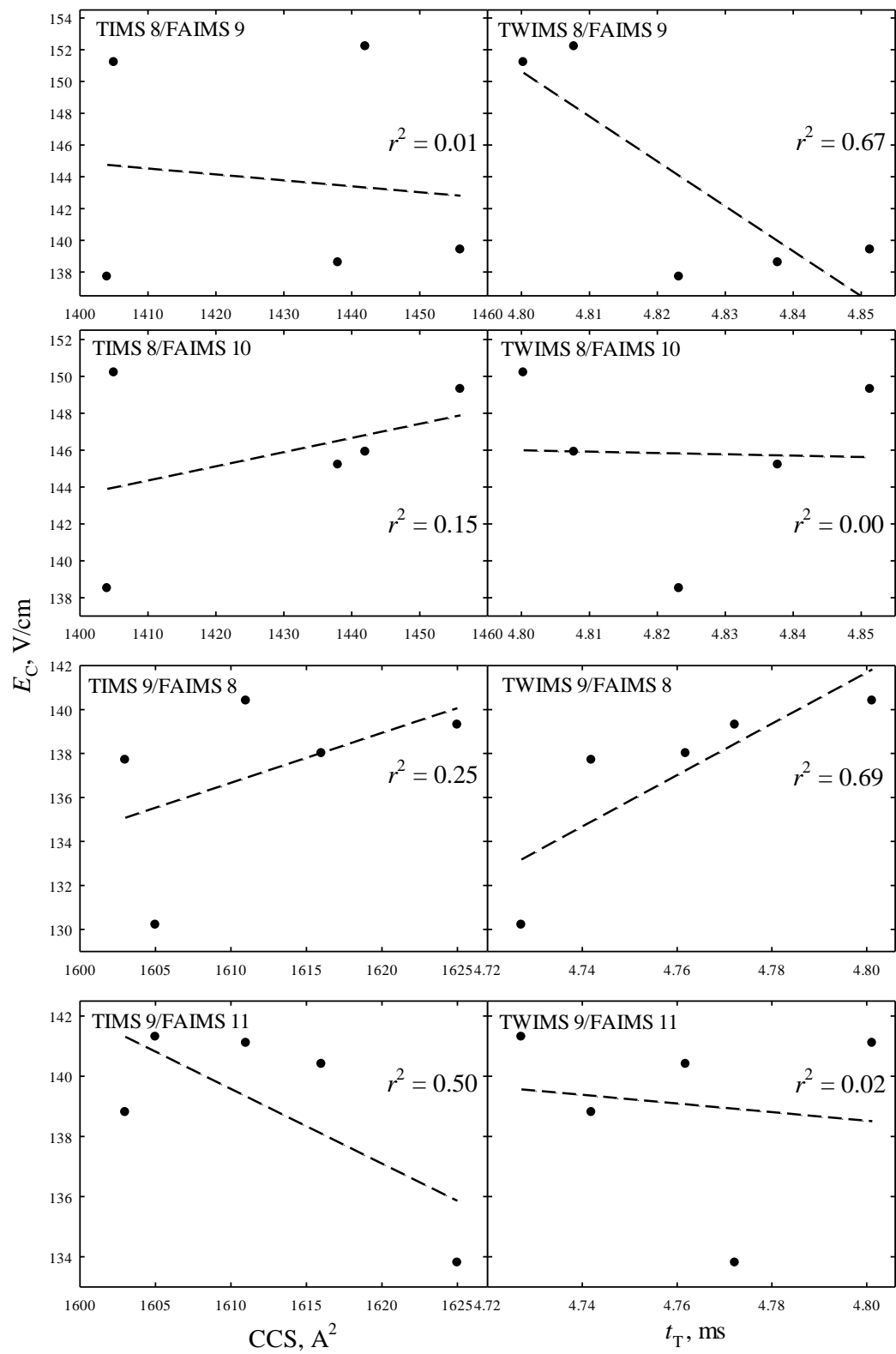


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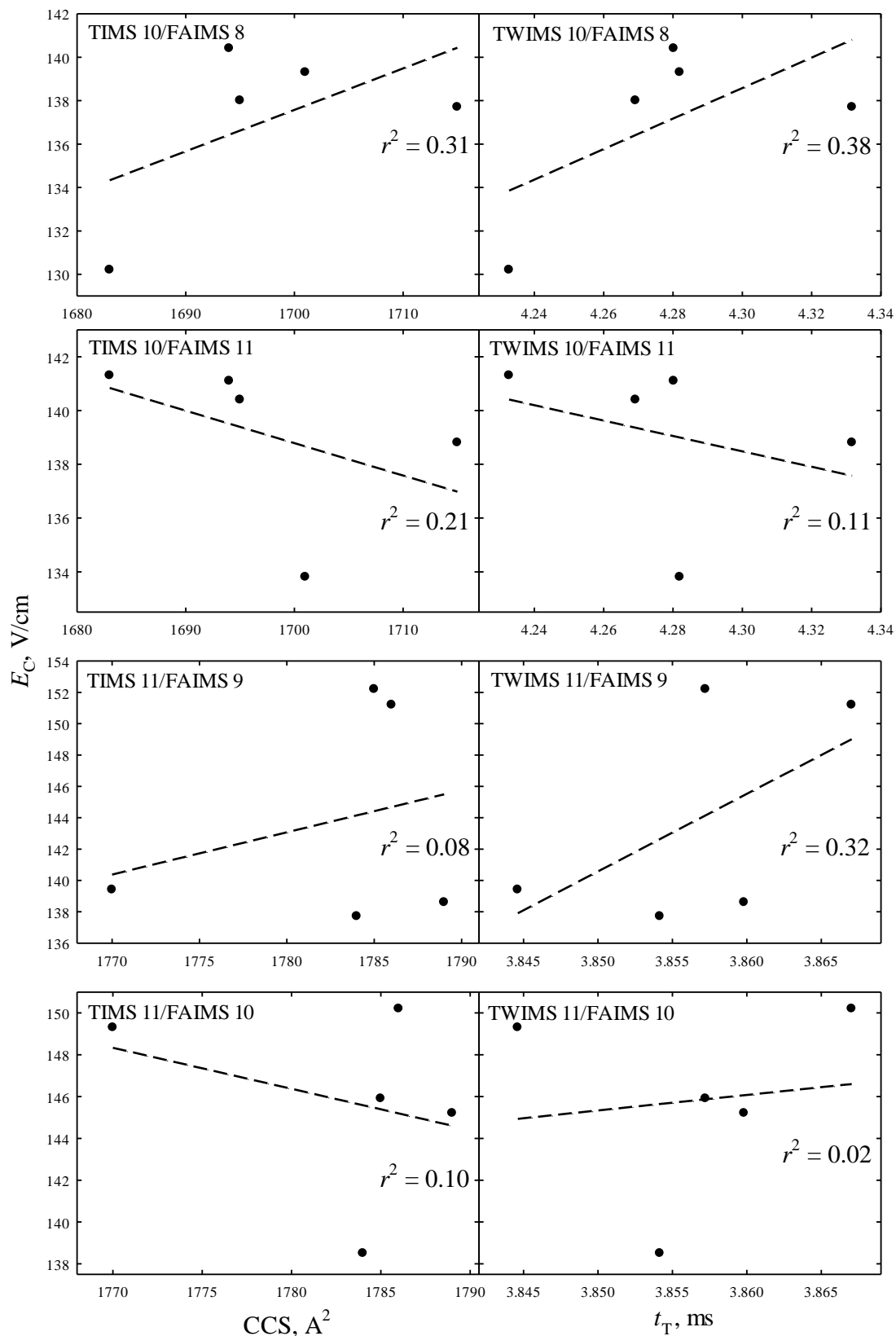


Figure S13b

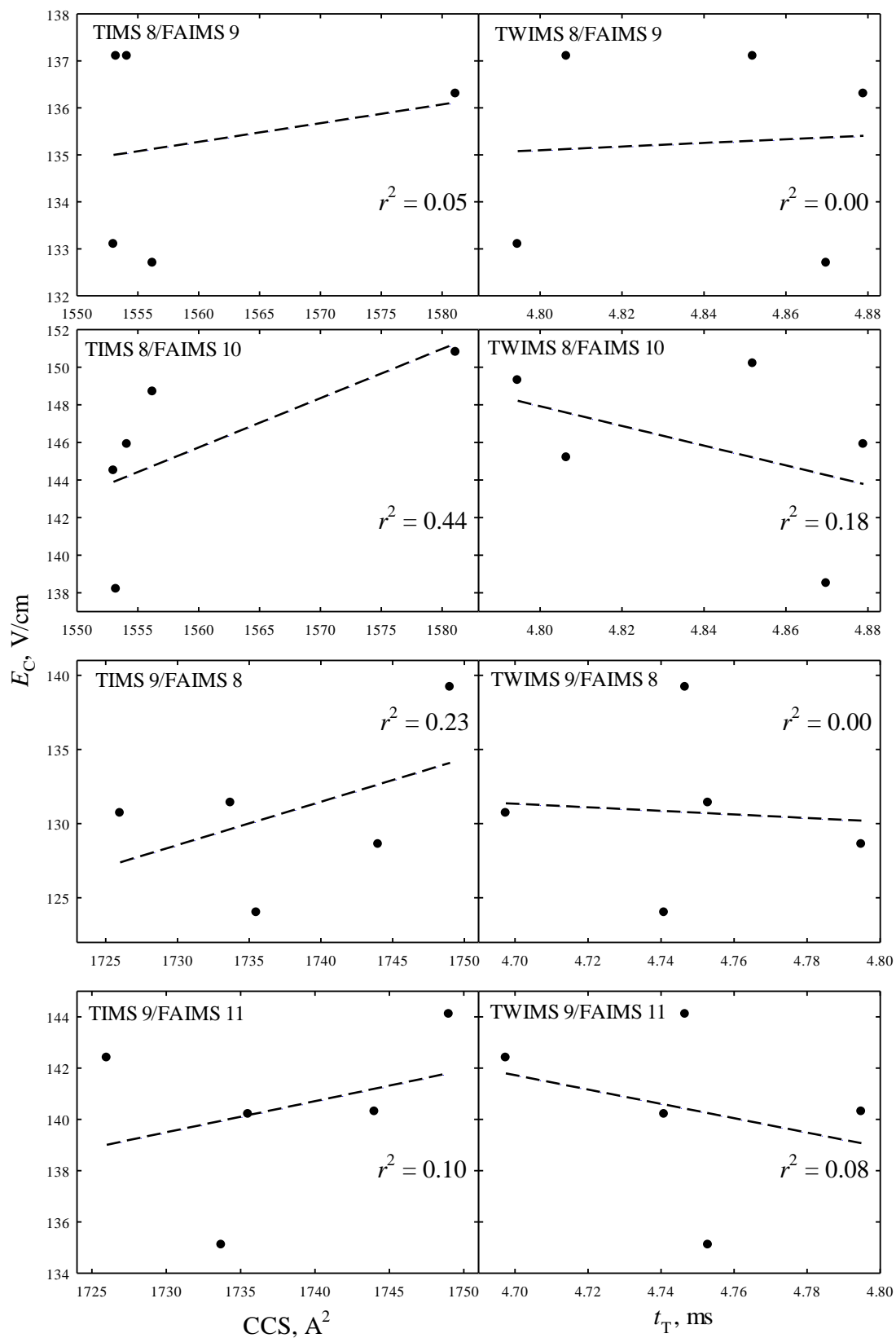


Figure S13b (continued)

