

Cell Reports, Volume 26

Supplemental Information

**TETs Regulate Proepicardial Cell Migration
through Extracellular Matrix Organization
during Zebrafish Cardiogenesis**

Yahui Lan, Heng Pan, Cheng Li, Kelly M. Banks, Jessica Sam, Bo Ding, Olivier Elemento, Mary G. Goll, and Todd Evans

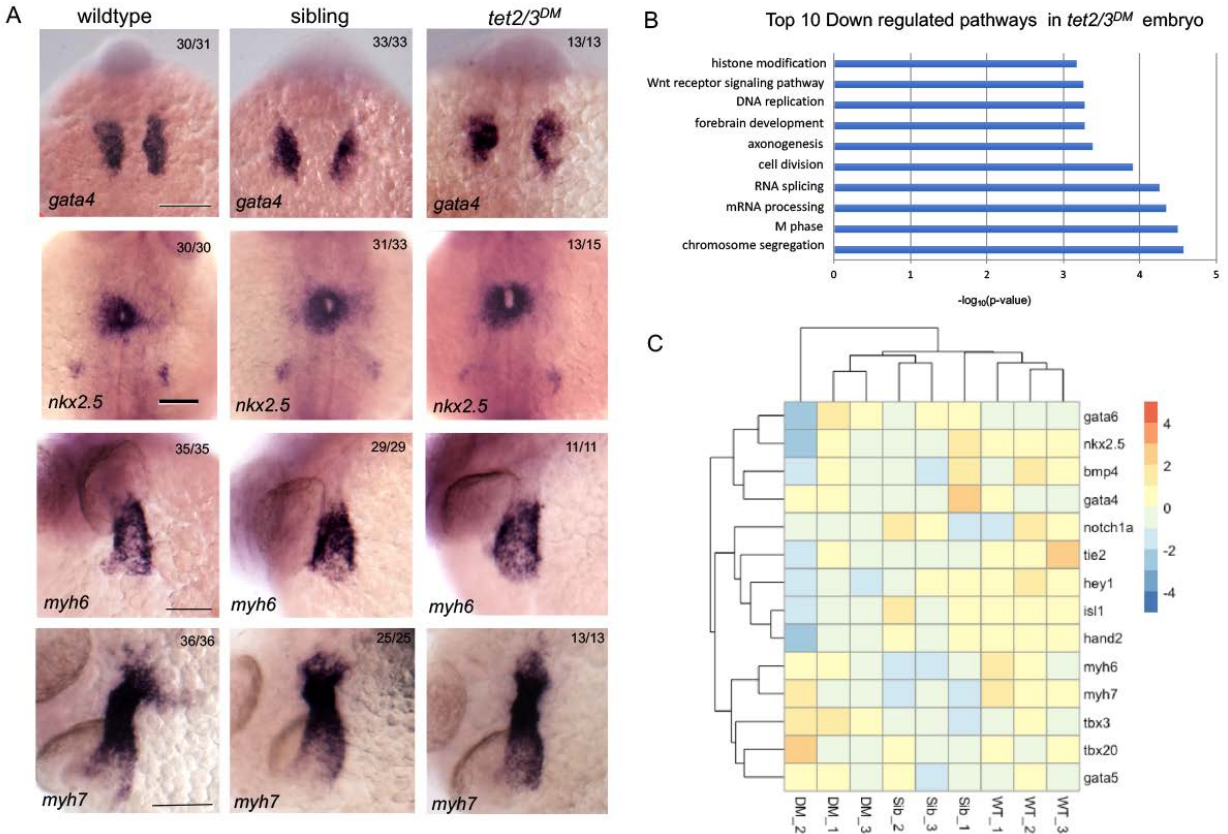


Figure S1: *tet2/3^{DM}* Larvae Show Neuronal but not Cardiac Defects at 28 hpf. Related to Figure 1.

(A) Markers of cardiac progenitors are similarly expressed in wildtype, sibling and *tet2/3^{DM}* larvae. WISH for *gata4* was performed at 18 hpf, *nkx2.5* was performed at 22-hpf, atrial myosin marker *myh6* and myosin marker *myh7* were performed at 28-hpf. Scale bar: 100 μm .

(B) Gene ontology (GO) analysis shows top ten down-regulated biological pathways in 28-hpf *tet2/3^{DM}* larvae by RNA sequencing.

(C) Heatmap of RNA sequencing data illustrating similar transcriptional expression of cardiac genes in *tet2/3^{DM}* (DM) compared with wildtype (WT) or sibling (Sib) larvae.

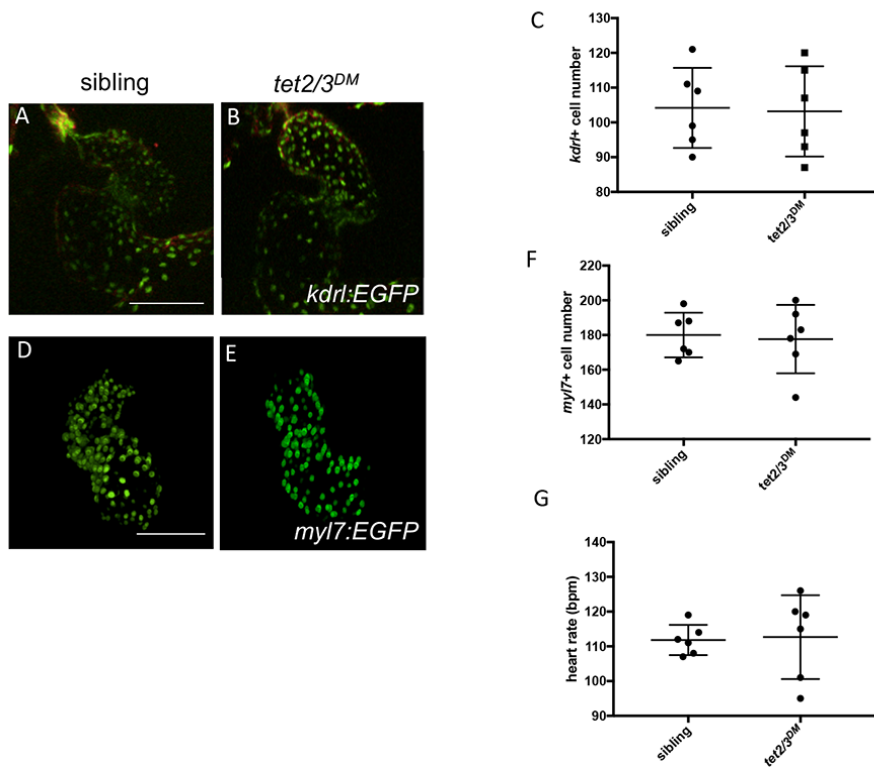


Figure S2: Normal Myocardium and Endocardium in 2-dpf *tet2/3^{DM}* Larvae. Related to Figure 1.

(A and B) GFP labeled endocardium in 2-dpf larvae carrying the Tg(*kdrl:EGFP-NLS*) transgene.

(C) Graph indicates the number of endocardial cells in 48-hpf sibling and *tet2/3^{DM}* larvae. Data are presented as the mean \pm SD.

(D and E) GFP labeled myocardium in 2-dpf larvae carrying the Tg(*myl7:EGFP*) transgene.

(F) Graph indicates the number of myocardial cells in 48-hpf sibling and *tet2/3^{DM}* larvae. Data are presented as the mean \pm SD.

(G) Graph indicates heart rate in 48-hpf sibling and *tet2/3^{DM}* larvae. Heart rate was measured in beats per minute. Data are presented as the mean \pm SD.

Scale bars: 100 μ m.

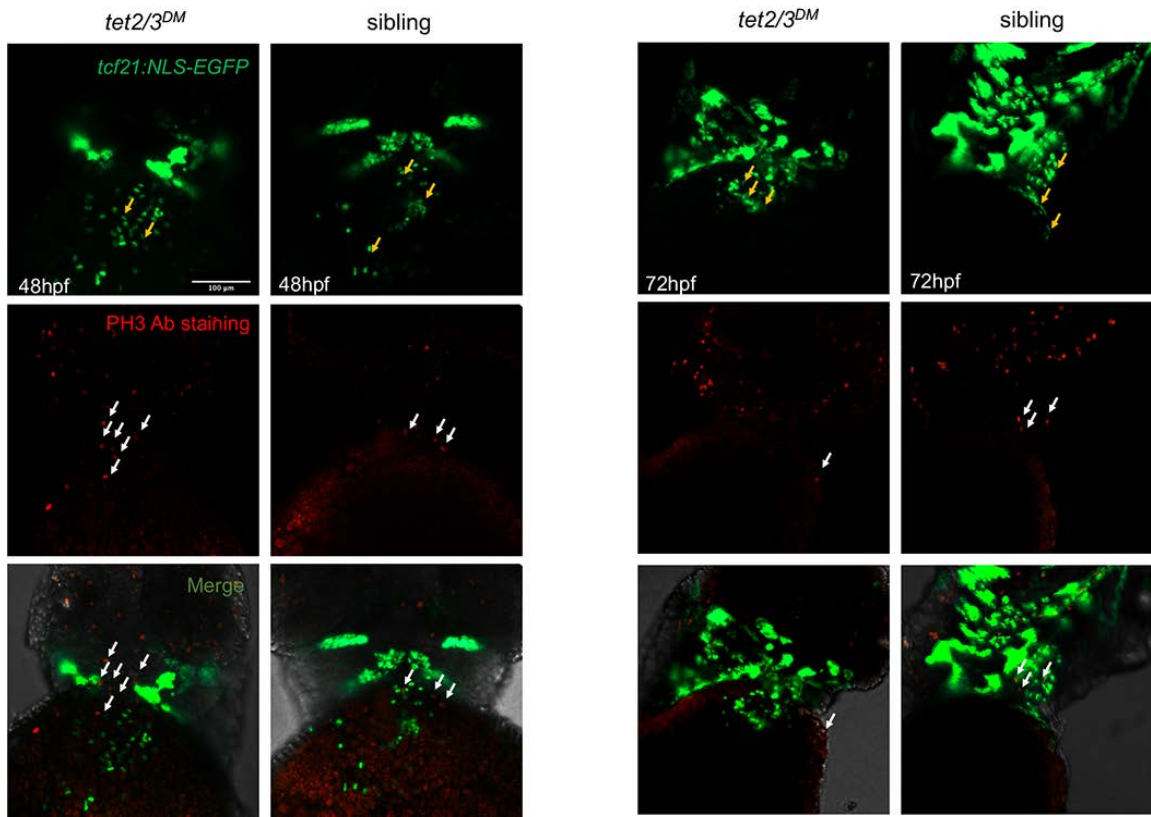


Figure S3: No PE Cell Proliferation at 48 and 72 hpf Sibling and *tet2/3^{DM}* Larvae. Related to Figure 1. Confocal images showing sibling and *tet2/3^{DM}* larvae carrying the Tg(*tcf21:NLS-EGFP*) transgene stained by anti-GFP antibody (Green) plus anti-pH3 antibody (Red). Yellow arrows indicate PE cells. White arrows indicate p3+ proliferating cells. Co-staining shows essentially no cell proliferation of *tcf21*+ PE cells at 48-hpf or 72-hpf. Scale bar: 100 µm.

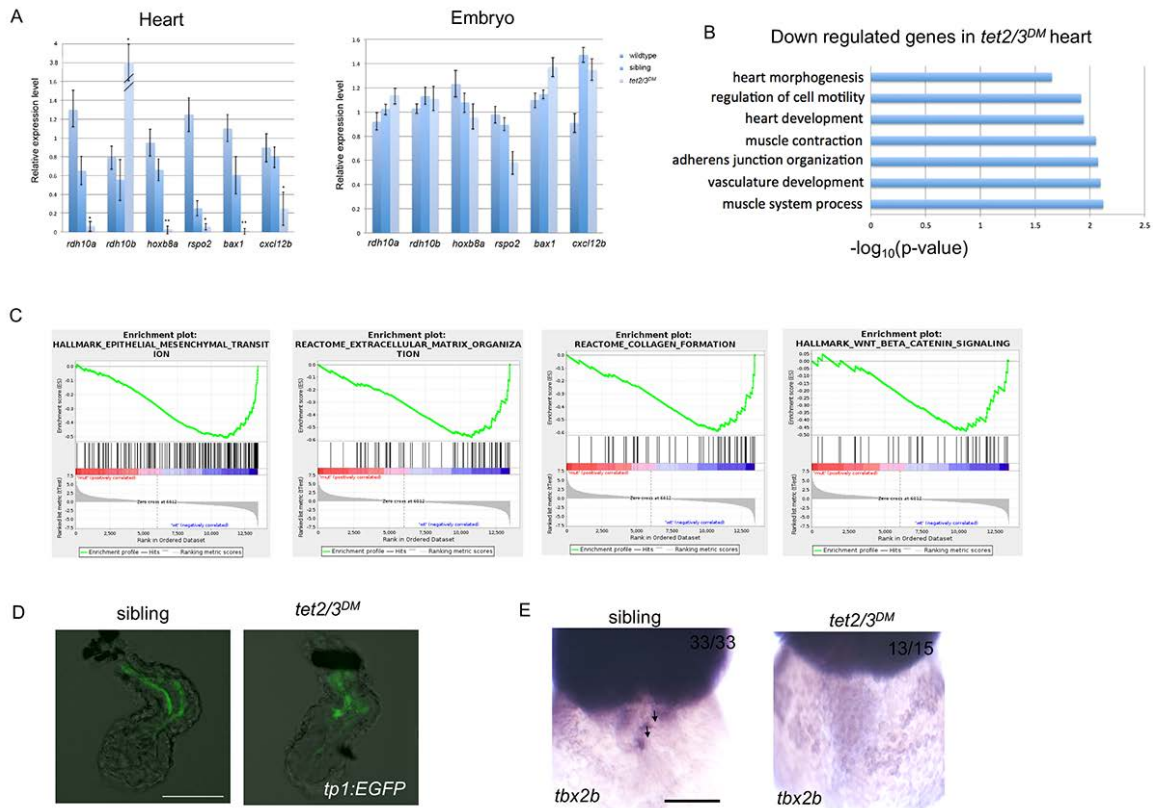


Figure S4: *tet2/3^{DM}* Larvae Show Cardiac AVC Defects at 48-hpf. Related to Figure 3 and Figure 4.

(A) Quantitative RT-PCR validation of top down-regulated genes from RNA sequencing. RT-PCR analysis of *rdh10a*, *hoxb8a*, *rspo2*, *bax1*, *cxcl12b* transcripts in 2-dpf embryonic heart or whole embryo. Notably, *rdh10b* apparently compensates for the *rdh10a* loss, with 4-fold increase in *tet2/3^{DM}* hearts, which is consistent with RNA sequencing results.

(B) GO analysis shows down-regulated biological pathways in 48 hpf *tet2/3^{DM}* hearts compared to wildtype hearts by RNA sequencing using isolated hearts.

(C) Gene set enrichment analysis shows down-regulated biological pathways in 48-hpf *tet2/3^{DM}* hearts compared to wildtype hearts by RNA sequencing using isolated hearts.

(D) Normal Notch Activity in *tet2/3^{DM}* endocardium. GFP-labeled atrial endocardium indicates Notch activity in sibling hearts as well as *tet2/3^{DM}* hearts. Hearts were dissected from 48 hpf larvae carrying the Tg(*tp1:EGFP*) Notch reporter transgene.

(E) WISH for AVC markers *tbx2b* at 48 hpf. Black arrows indicate AVC specific expression of *tbx2b* in sibling but not *tet2/3^{DM}* heart. Scale bars: 50 μm .

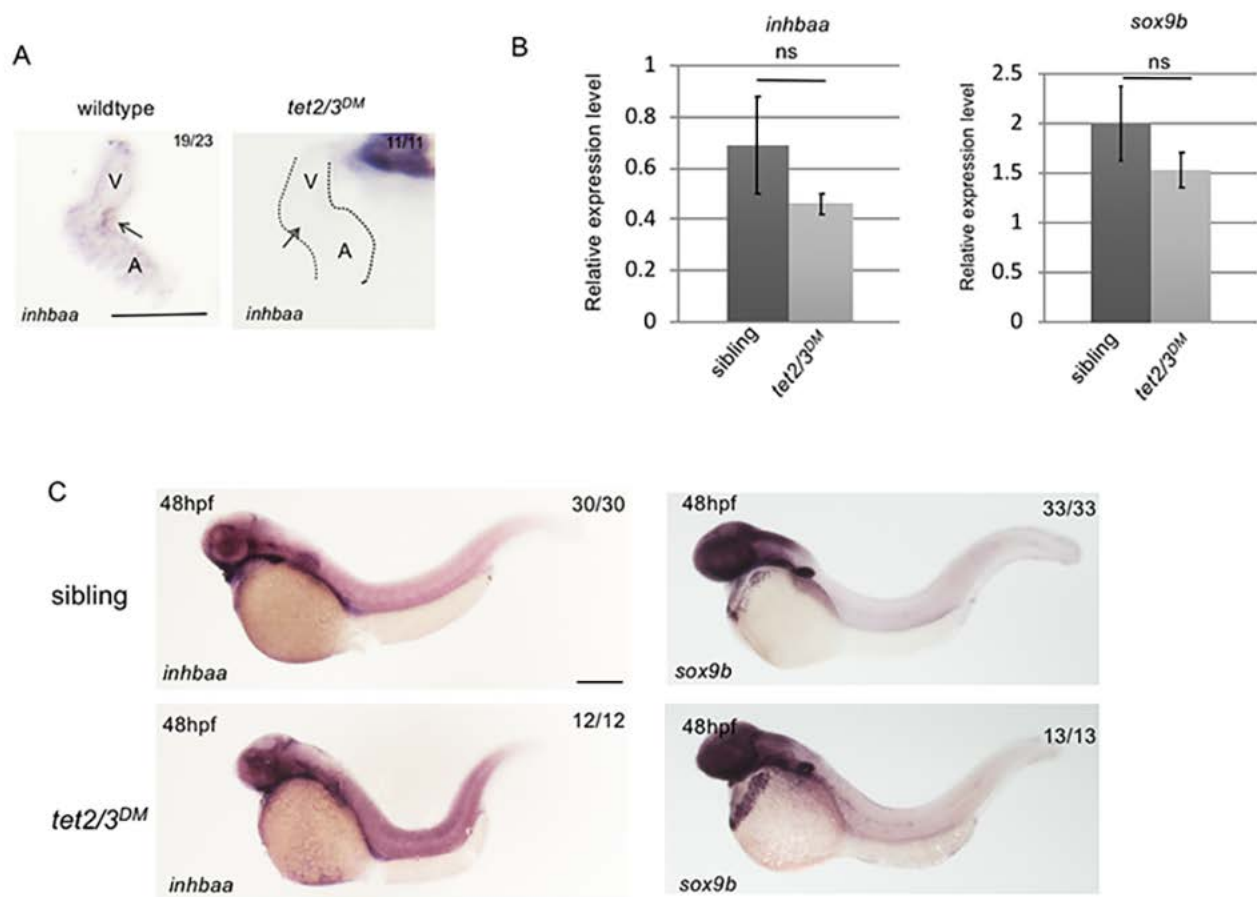


Figure S5: *inhbaa* and *sox9b* Expression Shows Cardiac Specific Defects in *tet2/3^{DM}* Larvae. Related to Figure 5.

(A) WISH for *inhbaa* at 48-hpf isolated hearts. Arrows indicate AVC endocardium with *inhbaa* transcripts. Scale bar: 100 μ m.

(B) RT-PCR analysis of *inhbaa* and *sox9b* transcripts in representative 48-hpf embryos.

(C) WISH for *inhbaa* and *sox9b* in representative 48-hpf embryos. Scale bar: 300 μ m.

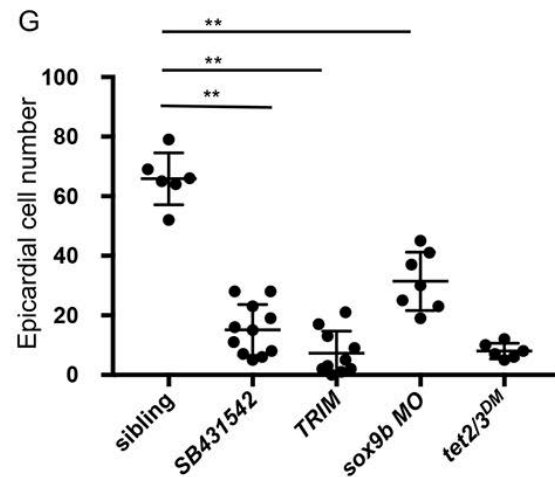
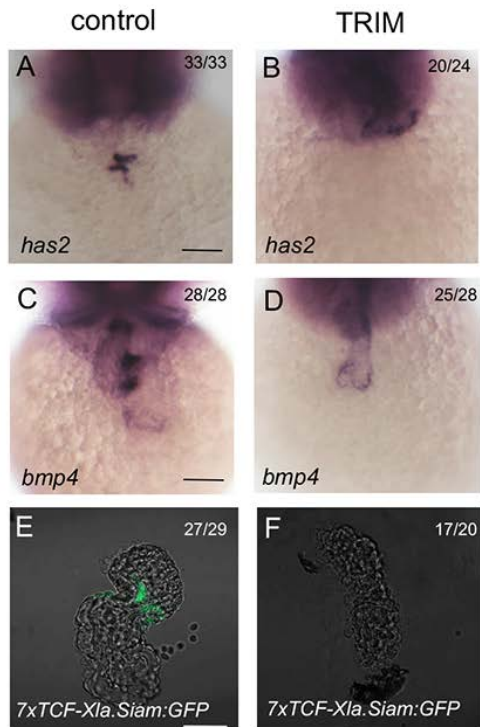


Figure S6: Inhibiting the Activin-A Pathway or *sox9b* Causes AVC Disruption and PE Migration Defect. Related to Figure 5.

(A-D) WISH for AVC markers *has2* and *bmp4* at 48-hpf in wildtype or TRIM treated larvae.

(E-F) GFP-labeled AVC endocardium represents Wnt activity in wildtype but not TRIM treated heart. Hearts were dissected from 48-hpf larvae carrying the Tg(*7xTCF-Xla.Siam:GFP*) transgene.

(G) Number of epicardial cells in the heart of 4-dpf sibling, sibling exposed to SB431542 or TRIM from 24 hpf, *sox9b* morphant and *tet2/3^{DM}* larvae carrying the Tg(*tcf21:NLS-EGFP*) transgene. Data are presented as the mean ± SD. The significance is indicated as **P < 0.01.

Scale bar: 50µm.

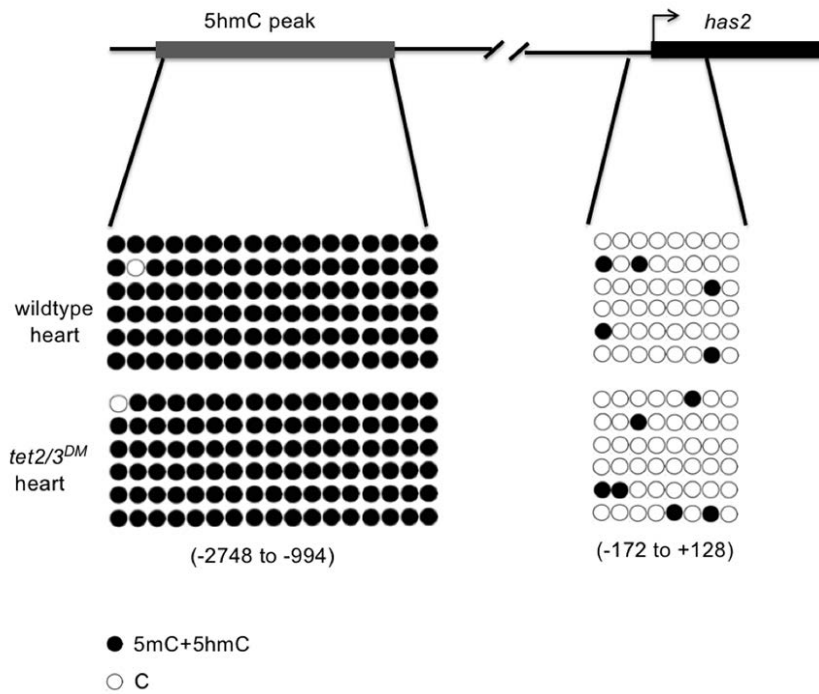


Figure S7. No Hypermethylation at the Promoter of the *has2* Gene in *tet2/3^{DM}* Hearts. Related to Figure 6.
 Diagram indicates *has2* locus and the associated regulatory regions. Gray box represents a 5hmC peak. Black box represents the coding sequence. Profiles of 5mC + 5hmC in the putative promoter regions revealed by bisulfite sequencing. Numbers indicate regions analyzed relative to the translation start site.

bis-sequence primer		5'--3'
<i>inhbaa</i>	F	TTTGGGGGGGGGGTTTTTATTG
	R	AATAAAAAAACACATACTTAACATACTAC
<i>sox9b</i>	F	TGTGTTTATTTGTGTTGGTGATGTTGTGG
	R	ACAAAAAACTACAATAACCATACTAAAATCTC
<i>has2-promoter</i>	F1	GTATGTTTGTATTGTGGGGGAAATTTGAGGGTTTGGAG
	R1	TCATTATTTCAATTCATTTTCTTTTCAACTTAATCCC
	F2	AAAAGATTGTTGAGGTGTTGAAGG
	R2	CAAAATCAATTACACATAACCTCTCAATAATAC
<i>has2-exon1</i>	F	ATTTGAAGGGGTAGTATTTTTTTTTTTTAGATGTTG
	R	CATTCAAATTTTTTTTTTCCCAAAC

qRT-PCR primer		5'---3'
<i>inhbaa</i>	F	GCACATTCAGAAGCCGACTGCC
	R	TAAAGTCCGTCTGCCTGTGCGC
<i>sox9b</i>	F	AGGTGCTGAAGGGCTACGACTGGT
	R	GATTCCTCCGTCTGGGCTGGTATT
<i>cxcl12b</i>	F	CAAGTCATTGCCAAGCTGAAG
	R	CTCGTCTGTTTACTCTGAGCG
<i>rdh10a</i>	F	AGTTCACTGGGCCTTTTCAG
	R	ACACCAGCGTCATCTTGATAC
<i>hoxb8a</i>	F	ACAACCTGTTCCCGTGGATG
	R	TGTGACACTTCAATCCGACG
<i>rspo2</i>	F	TCCAACCATCGCTGAATCTAG
	R	TCGCTGTGTTGGTTCTGAG
<i>barx1</i>	F	CCGGATCAGAAGGTATCAAGTC
	R	GGAAACTTCAGGATACCCGTC
<i>b-actin</i>	F	CGAGCAGGAGATGGGAACC
	R	CAACGGAAACGCTCATTGC
<i>rdh10b</i>	F	GAGAGCCATACTAACAGACCAG
	R	AGGGTACATGCACTTATCAGC

mRNA generation primer	5'--3'		
EcoRI- <i>inhbaa</i> -F	CG GAATTC	AGTCCCAGACCTCTTACGAG	
XhoI- <i>inhbaa</i> -R	CTAACG CTCGAG	ttacgagcagccgattctt	
ECORI- <i>sox9b</i> -F	CG GAATTC	ttataacacacacgcgtgcg	
XhoI- <i>sox9b</i> -R	CTAACG CTCGAG	TCAGGGTCTGGACAGCTGTG	
BamHI- <i>has2</i> -F	CTGGGATCC	ATGAGATGTGATAAAGCGGTCAGC	
XhoI- <i>has2</i> -R	GTC ACTCGAG	CTATACGTCAAGAACCATGTC	

Table S1. PCR Primer Sequences Used in the Paper. Related to Figure 5 and Figure 6.

<i>alpi.1</i>
<i>amer2</i>
<i>ets2</i>
<i>foxa3</i>
<i>g6pc3</i>
<i>gmpr</i>
<i>inhbaa</i>
<i>lama4</i>
<i>nog2</i>
<i>pecam1</i>
<i>psid</i>
<i>pts</i>
<i>rtn4rl2a</i>
<i>slit1a</i>
<i>sox9b</i>
<i>spag1a</i>
<i>srl</i>
<i>trim69</i>
<i>tyr</i>
<i>zgc:153184</i>

Table S2. Genes have hyper-DMR in 2 kb upstream and downstream of transcription start sites as well as transcriptional downregulation in *tet2/3^{DM}* hearts (fold change >2). Related to Figure 5.