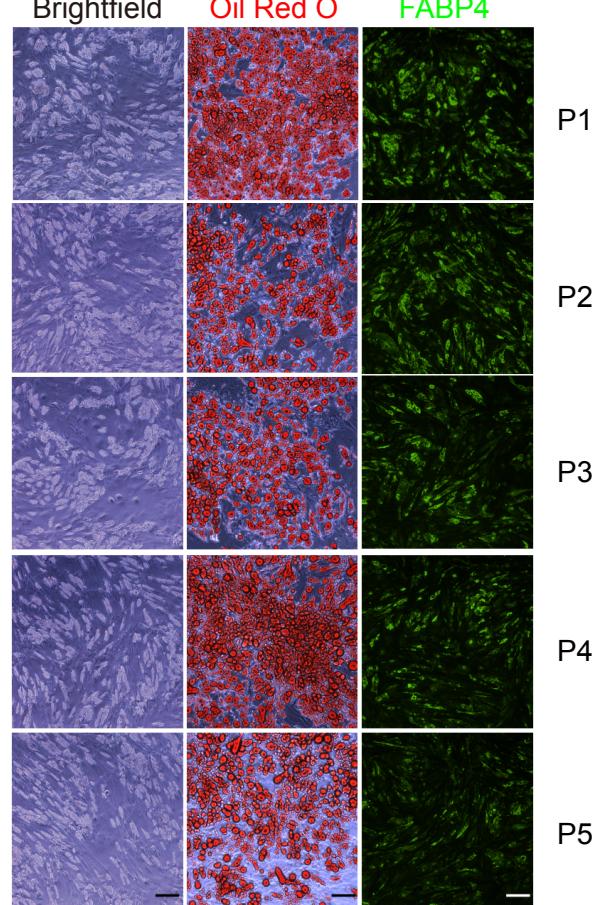
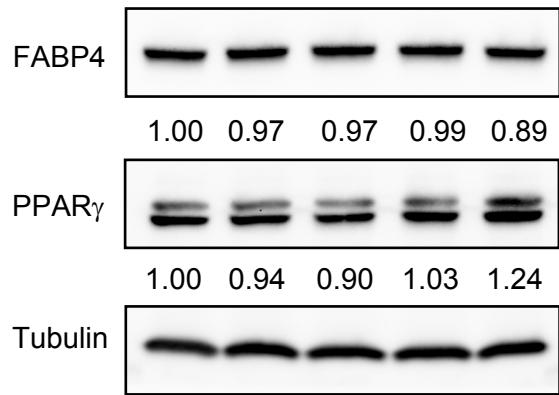


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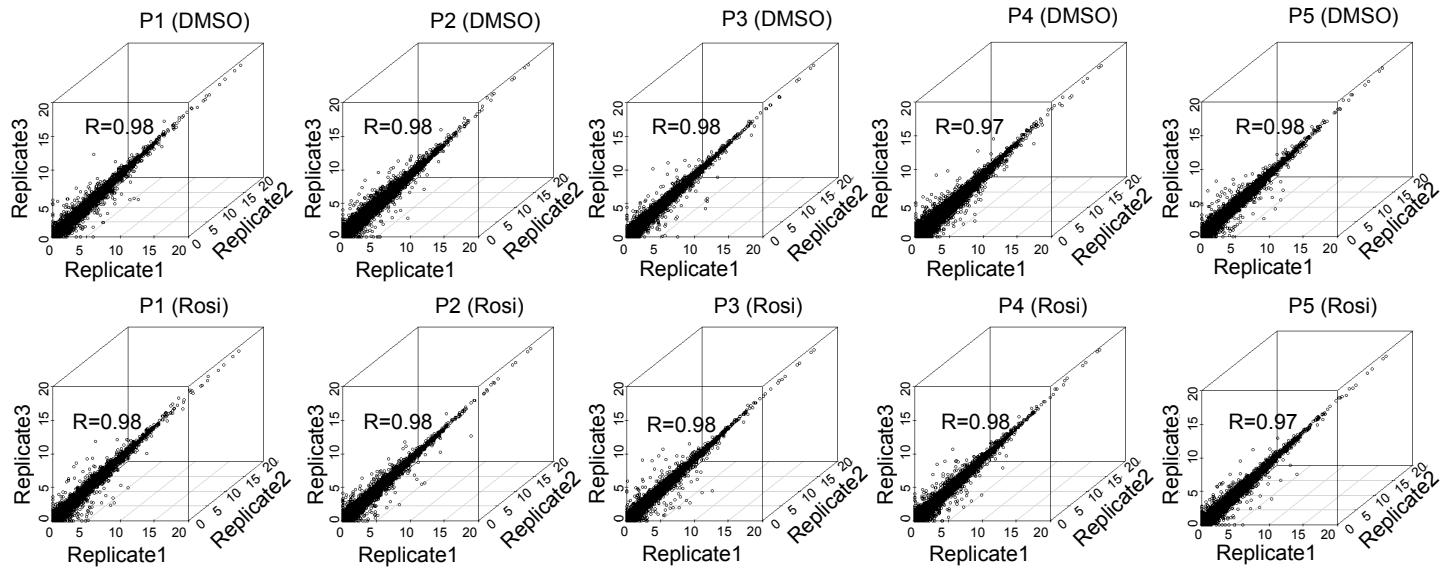
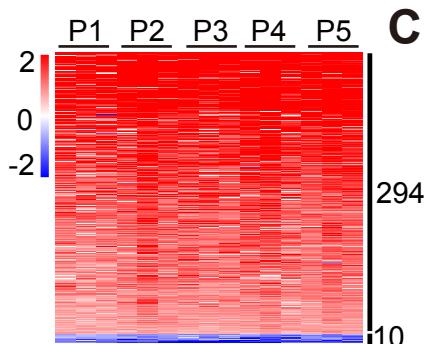
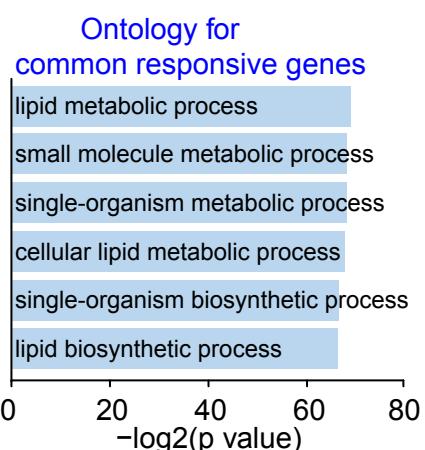
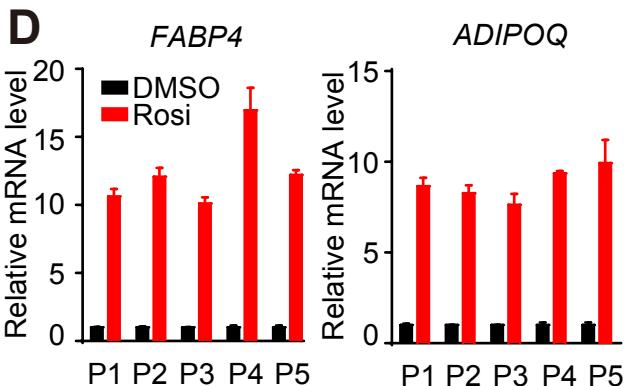
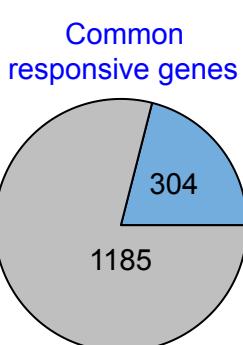
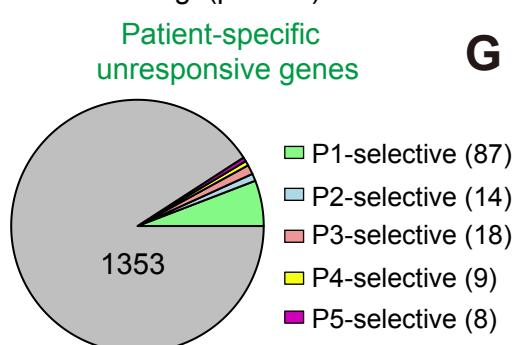
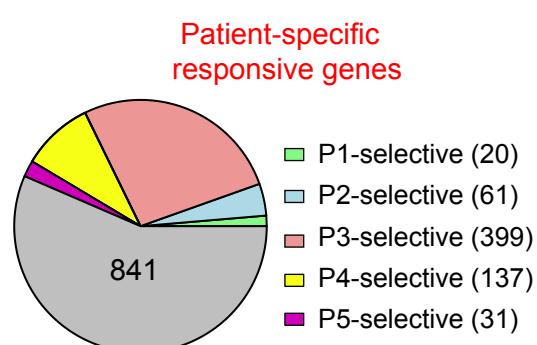
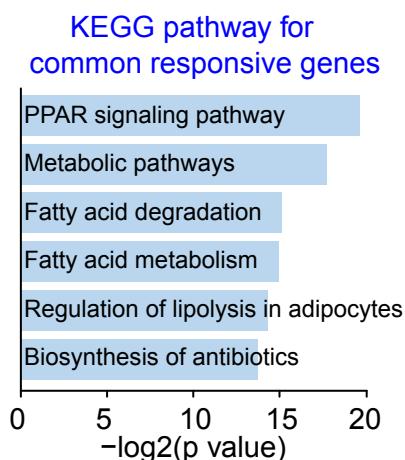
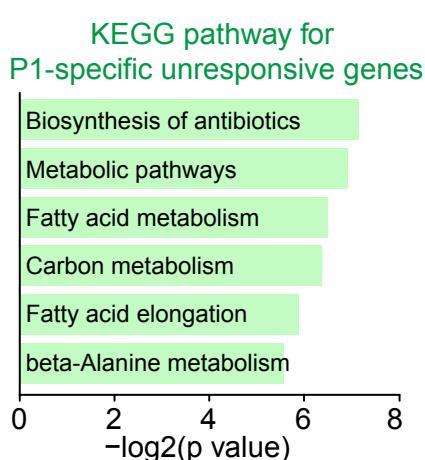
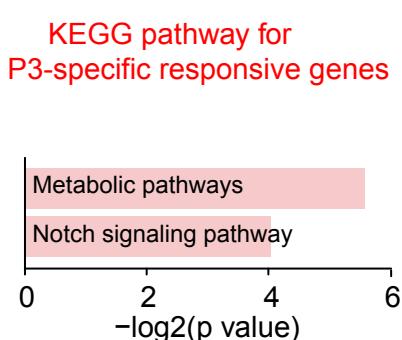
<u>Subject</u>	<u>Age</u>	<u>Gender</u>	<u>Race</u>	<u>BMI</u>
P1	46	Female	Black	37.2
P2	35	Female	Black	44.2
P3	22	Female	White	41.6
P4	49	Female	White	48.2
P5	49	Female	Black	54.9

B**C**

Supplemental Figure 1, related to Figure 1

Figure S1. Derive human adipocytes from obese patients, Related to Figure 1.

- (A) Information about the 5 patients studied, including age, gender, race, and BMI.
(B) Representative images of morphology, Oil Red O staining and FABP4 immunostaining of adipocytes from five patients at day 21. Scale bar, 100 μ m.
(C) Protein expression level of adipocyte marker genes *FABP4* and *PPAR γ* in adipocytes from five patients at day 21. The values below the western blot represent the relative expression level of each protein after normalization to tubulin.

A**B****C****D****E****F****G****H****I****J**

Supplemental Figure 2, related to Figure 1

Figure S2. Patient-derived adipocytes differentially respond to rosi treatment, Related to Figure 1.

(A) The correlation among three replicates in each patient and each condition. Each dot represents a gene. X-axis, Y-axis and Z-axis represent the normalized expression value of genes in three replicates.

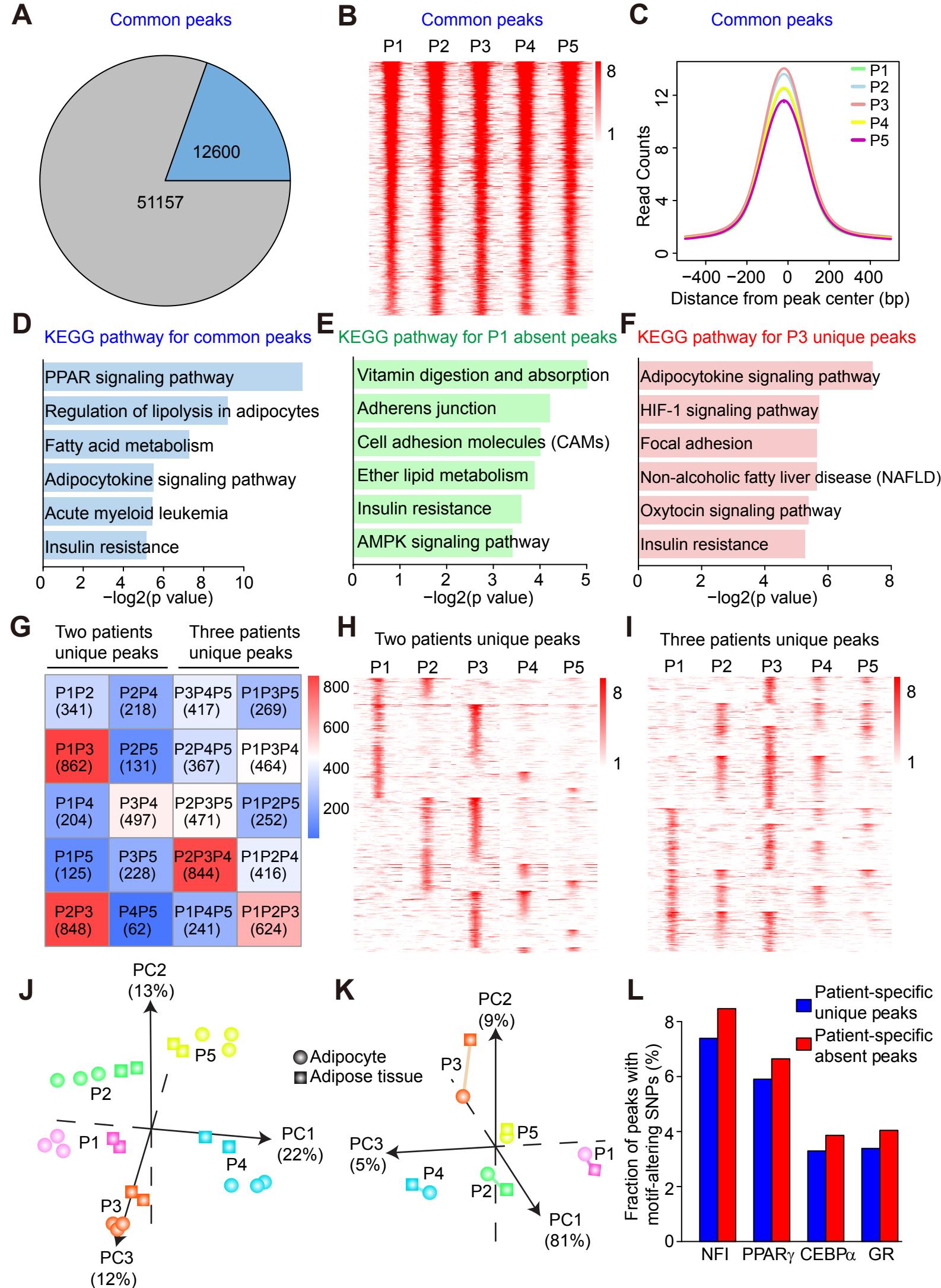
(B) Heat map of common rosi-responsive genes that are regulated by rosi in five patients. The color bar indicates the scale used to show log2FC(Rosi/DMSO) of genes across samples. Differentially expressed genes were identified using edgeR with adjusted *p* value < 0.01 and fold change \geq 1.5.

(C) Gene ontology for common rosi-responsive genes.

(D) mRNA expression of common rosi-responsive genes *FABP4* and *ADIPOQ* in adipocytes from five patients, normalized to *HPRT*, DMSO was set to 1, as measured by RT-qPCR.

(E-G) Proportion of common responsive genes (E), patient-specific unresponsive genes (F), and patient-specific responsive genes (G) in all five patients.

(H-J) KEGG pathways enriched in common responsive genes (H), P1-specific unresponsive genes (I), and P3-specific responsive genes (J).



Supplemental Figure 3, related to Figure 1

Figure S3. Common PPAR γ binding sites shared by these five patients, Related to Figure 1.

(A and B) Proportion (A) and Heat map (B) of common PPAR γ peaks that are detected similarly in all five patients. The color bar indicates the scale of relative normalized tag counts.

(C) For common PPAR γ peaks, the average binding profiles are shown in 1 kb windows across patients.

(D-F) KEGG pathways enriched in the genes surrounding common PPAR γ peaks (D), patient-specific absent peaks (E) and patient-specific unique peaks (F).

(G-I) The numbers of patient-specific sites shared by a subset of two or three individuals are shown in table (G). Heat map of PPAR γ peaks observed only in 2 patients (H) or 3 patients (I) with at least 2-fold differences in read counts compared to other patients.

(J) Principal component analysis (PCA) with computation of closest neighboring samples under ANOVA ($p=0.02$) showed more than 1000 genes with preserved patient-specific expression in adipocytes (circle) and adipose tissues (quadrilateral).

(K) PCA with computation of closest neighboring samples under ANOVA ($p=0.05$) showed more than 1000 PPAR γ binding sites with preserved patient-specific pattern in adipocytes (circle) and adipose tissues (quadrilateral).

(L) Fraction of PPAR γ binding sites that contain SNPs affecting PPAR γ motif and its cofactors NFI, CEBP α and GR motifs.

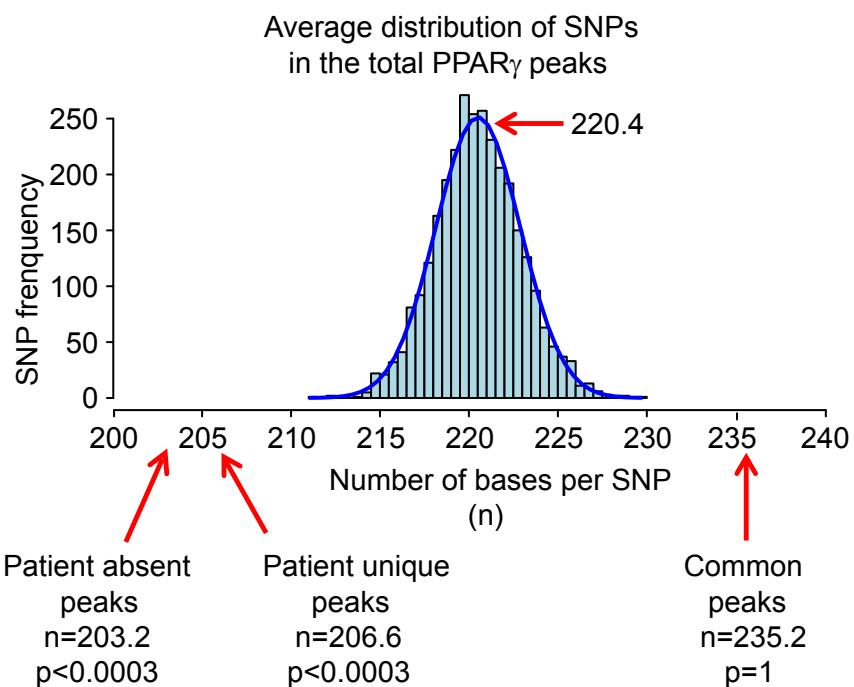
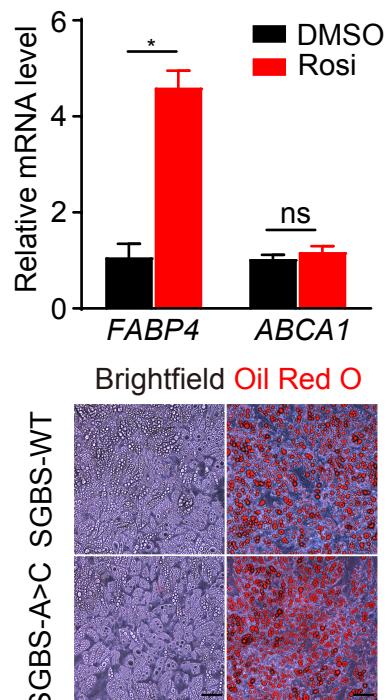
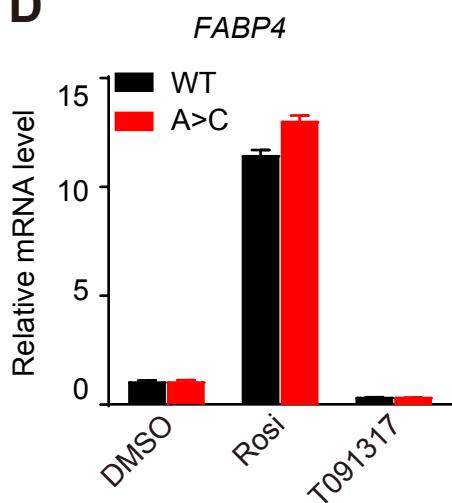
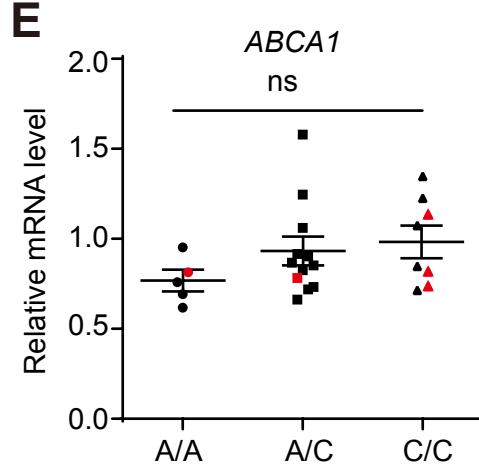
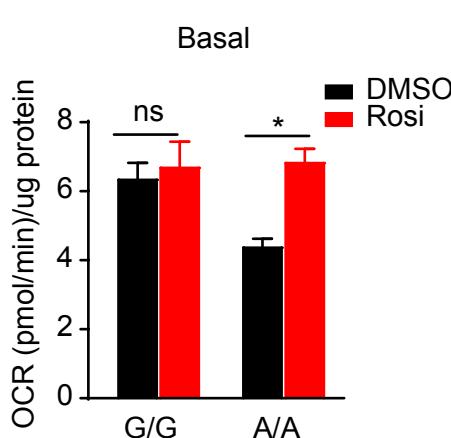
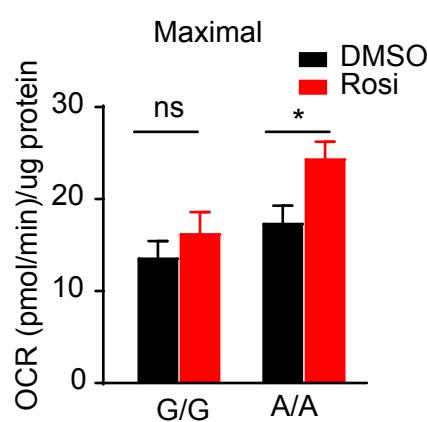
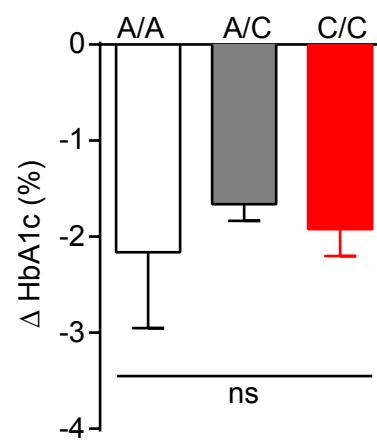
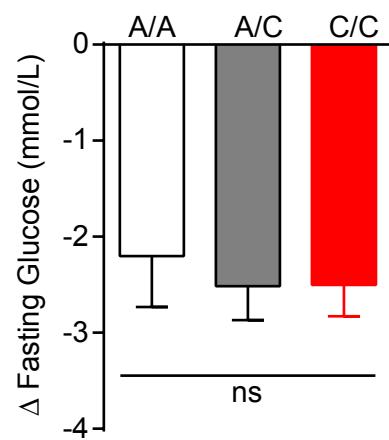
A**B****D****E****F****G****H****I****Supplemental Figure 4, related to Figure 3 and 4**

Figure S4. SNPs were enriched in patient-specific absent and unique sites and modulated rosi effect, Related to Figure 3 and 4.

(A) SNPs are enriched in patient-specific absent PPAR γ peaks, patient-specific unique PPAR γ peaks, but not enriched in common PPAR γ peaks based on random test. The length of bases in patient-specific sites or common peaks that contains one SNP was indicated by red arrows. *P*-values are determined by random test.

(B) mRNA expression of *FABP4* and *ABCA1* in SGBS-differentiated adipocytes, normalized to *HPRT*, as measured by RT-qPCR. Data are expressed as mean \pm SEM. (*) $p < 0.05$, (ns) $p > 0.05$ in Student's t-test. $n = 3$ per group.

(C) Representative images of morphology and Oil Red O staining of adipocytes from SGBS WT and SGBS A>C cell lines. Scale bar, 100 μ m.

(D) mRNA expression of *FABP4* in SGBS WT and SGBS A>C adipocytes treated with DMSO, 1 μ M rosi or 10 μ M T091317 ($n=3$), normalized to *HPRT*, as measured by RT-qPCR. Data are expressed as mean \pm SEM.

(E) mRNA expression of *ABCA1* in hASC-derived adipocytes from 25 patients without rosi treatment. Red dots represent patients P1-P5. Normalized to *HPRT*. (ns) $p > 0.05$ in one-way ANOVA test.

(F and G) Basal (F) and maximal (G) respiration calculated from OCRs in 3 rs2106146 G/G and 3 A/A patient adipocytes treated with DMSO or 1 μ M rosi. Data were normalized to total protein content per well. Data are expressed as mean \pm SEM from $n = 7$ technical replicates. (*) $p < 0.05$, (ns) $p > 0.05$ in Student's t-test.

(H and I) The change of HbA1c (H) and fasting glucose (I) levels after rosi treatment of patients carrying different rs4743771 genotypes. (ns) $p > 0.05$ in one-way ANOVA test.

Table S1. Primers used in this study, Related to STAR Methods.

Primer for RT-qPCR		
Primers	Forward (5'-3')	Reverse (5'-3')
FABP4	TCATGAAAGGCGTCACTTCC	GCTTGCTAAATCAGGGAAAACA
ADIPOQ	GATGAAGTCCTGTCTTGGAGG	CAGCACTTAGAGATGGAGTTGG
HPRT	TGACACTGGCAAACAAATGCA	GGTCCTTTCACCAGCAAGCT
ANGPTL4	CCTTCTCCACTGGGACCAG	AAACCACCAGCCTCAGAGA
AGPAT2	GAGGGTACTCGCAACGACAA	CCTGCACTGTGACTTTCCT
DGKQ	TGAGGCCAGACCGGGTTCTA	CAGGCAACGTCCAACACTAC
GUK1	GTATGGCACGAGCAAGGTG	GCGGCTGCACAGAGATGTAG
SLC7A10	CCGAGGAGATGGTTGACGC	ATGGCCGTGAAGTAGGCAATG
FAM160B2	CACATGCCTGCTGAGACCG	CCACCCGACTCTTCTTGCTC
ABCA1	ACCCACCCATGAACAAACATGA	GAGTCGGGTAACGGAAACAGG
SLC25A1	ACGGGGTTAGGGAGATTGTG	GCCTGCAATAGCTCCGAAGA
MSX1	ACACAAGACGAACCGTAAGCC	CACATGGGCCGTGTAGAGTC
NFIA	GCAGGCCCGAAAACGAAAATA	TTTGCAGAAGTCGAGATGCC
Primer for ChIP-qPCR		
Primers	Forward (5'-3')	Reverse (5'-3')
ABCA1	CCTGTAAACAGCTTTATGCTAG	CACCTCAGTGCATGATAGTTA
ALAS2	GGGAGGGGGTACACTAGAGG	GGGTTAGGACCTGGCATTAA
INS2	GCCCACCCCTCTGATGTATCT	AAAGTGACCAAGCTCCCTGTG
PDK4	GCAGAGTCAACAAGGGGAAG	ACTAGATGCCTGGAGCTGA
Primer for SNaPshot		
Primers	Forward (5'-3')	Reverse (5'-3')
ABCA1 PCR	GCAACATACGGTTCTGTGATTG	AGAACACCTCAGTGCATGATAG
ABCA1 SNaPshot	GACTGACTtactgaattgtccctgc	
Primer for Genotyping		
Primers	Forward (5'-3')	Reverse (5'-3')
rs4743771-F	GCAACATACGGTTCTGTGATTG	AGAACACCTCAGTGCATGATAG
rs2106146-F	AGCCCTAAATCATAAATCACCAGAG	GAACCATGCCCTCTCAGAT
rs76932545-F	GGATTCACTGAGCCCCACA	TCACCGATCGTGTCCCTGTC
3C probe and primers		
Primers/Probe	Forward (5'-3')	Reverse (5'-3')
ABCA1 probe	ctggaagtgcgtggagg	
ABCA1_constant_F	catagaagaggcccaagcag	
ABCA1_1R		ggacatctggagtatagatgcaa
ABCA1_2R		aagcaatgcgtacttattgac
ABCA1_3R		gtcccccttaactgtcca
ABCA1_4R		tcccttcctctgatacactgac
ABCA1_5R		cttcaaggcagtgtcagcag
ABCA1_6R		gagaagagcaatcctaagattaga
TBP probe	tggAACCCCAACTAGGTCGG	
TBP_constant_F	tctttcctgcgtctgctgag	
TBP_1R		tgaccagccacagttacttatac
Oligos for CRISPR		
ABCA1 sgRNA	5' CACCGagttcgagagaatatgtc 3'	5' AACGcacatattctctgcaactc 3'
ssODN	5'atacggttctgtgttggatttagaacaacacagagacattatactgaattgtcctcccccatttctgca actctcagctctgccttgtaaacagctttatgctagagca 3'	
gBlocks Gene for Luciferase reporter cloning		

rs4743771-A (5'-3')	CCGCTCGAGAGATGCACTGAATGAGTTAACAAATCAAGCACTTGAGAGA GCAACATACGGTTCTGTGATTGGATTAGAACACAGAGACATTATACT GAATTGTCCTCCTGCACATATTCTCTGCAACTCTCAGCTCTGCCTGTTTC CTGTAAACAGCTTTATGCTAGAGCAGGCATCTCCAACAGTGAATCAAG AGAAATCCTTCTATCAGAGATCTCC
rs4743771-C (5'-3')	CCGCTCGAGAGATGCACTGAATGAGTTAACAAATCAAGCACTTGAGAGA GCAACATACGGTTCTGTGATTGGATTAGAACACAGAGACATTATACT GAATTGTCCTCCTGCCCATATTCTCTGCAACTCTCAGCTCTGCCTGTTTC CTGTAAACAGCTTTATGCTAGAGCAGGCATCTCCAACAGTGAATCAAG AGAAATCCTTCTATCAGAGATCTCC
rs2106146-A (5'-3')	CCGCTCGAGGCGTGATCACGGCTCAGTGCAGCTTGACCTCCTGGCT CATGCGATCCTCTCATTCAACCTTCCAATAGCTGGGACTACAGACAT GTGCCACTTACCCAGTTAAATTAAATTGGTACAGACAGATTCTC CTTATGTTGCCCATGATGGTGTGAACGGGCTCAAGTGATGCACCTCG GCCTCAGATCTCC
rs2106146-G (5'-3')	CCGCTCGAGGCGTGATCACGGCTCAGTGCAGCTTGACCTCCTGGCT CATGCGATCCTCTCATTCAACCTTCCAATAGCTGGGACTACAGACAT GTGCCACTTGGCCAGTTAAATTAAATTGGTACAGACAGATTCTC CCTTATGTTGCCCATGATGGTGTGAACGGGCTCAAGTGATGCACCTCG GGCCTCAGATCTCC
rs76932545-A (5'-3')	CCGCTCGAGGGTCCTGACACGCAGTGGGAACTCAGCGCTGTTAAATGT GGATTCACTGAGCCCCACAGGCAGCCCTTAGGACCTCGACCCAAGG TCATTCTGGCCAAAGGGCGCTAGAGGGCAGCGGTGCTTTCCCTCTGC ATGAAGTTGGACAACGTCCGCAGTCAGCCTCGTCTGGTCCCAGGAG ATCTTCC
rs76932545-G (5'-3')	CCGCTCGAGGGTCCTGACACGCAGTGGGAACTCAGCGCTGTTAAATGT GGATTCACTGAGCCCCACAGGCAGCCCTTAGGACCTCGACCCAAGG TCATTCTGGCCAAAGGGCGCTAGAGGGCAGCGGTGCTTTCCCTCTGC ATGAAGTTGGACAACGTCCGCAGTCAGCCTCGTCTGGTCCCAGGAG ATCTTCC