

Supplemental Figure 1. Induction of wound inducible ER body genes *TSA1* and *BLGU18* is exclusive to the *ceh1* mutant.

Relative expression levels of wound inducible ER marker genes (*TSA1* and *BGLU18*) in *ceh1* and MEP pathway RNAi lines. Total RNA extracted from these genotypes was subjected to real-time quantitative-PCR analysis. The transcript levels were normalized to *At4g26410* (M3E9) measured in the same samples. Data are mean fold difference \pm SD of three biological replicates each with three technical repeats.



Supplemental Figure 2. Phenotypes of various genotypes (WT, *nai1*, *ceh1* and *ceh1/nai1*).



Supplemental Figure 3. MEcPP levels in various genotypes (WT, nai1, ceh1 and ceh1/nai1).



Supplemental Figure 4. Phenotypes of various genotypes (WT, *coi1*, *ceh1* and *ceh1/coi1*).



Supplemental Figure 5. MEcPP *does not* induce expression of genes associated with aliphatic glucosinolates (AGs) biosynthesis. (A) Schematic illustration of AGs biosynthesis pathway and their associated transcription factors (boxed) and metabolites (lassoed). (B) Heatmap of the expression levels of genes common to both the AGs and IGs biosynthesis, in *ceh1, ceh1/eds16*, *eds16* as compared to the WT levels using RNAseq data. (C) Heatmap of the expression of genes specifically associated with AGs biosynthesis in *ceh1, ceh1/eds16*, *eds16* relative to the WT levels using RNAseq data. The log₂-fold change for *ceh1* (left), *ceh1/eds16* (middle), and *eds16* (right) vs WT is represented by color, from -2 (blue) to 2 (yellow).



Supplemental Figure 6. Alteration of aliphatic glucosinolates *is not* specific to MEcPP. Measurement of AGs levels in WT transformed with empty vector (EV), RNAi lines silencing MEP-pathway genes individually and co-suppressed (*csHDS*). Data are mean \pm SEM; n=3. Asterisks show the significant differences relative to EV (P < 0.05). All the statistical analyses were performed using ANOVA.



Supplemental Figure 7. Production of NMOI3M in response to botrytis infection is most prominent in the *ceh1* mutant.

Gene ID	Name	Reference
AT3G09260	PYK10/BGLU23	(Nagano et al., 2008)
AT1G66270	BGLU21	(Nagano et al., 2008)
AT1G66280	BGLU22	(Nagano et al., 2008)
AT3G16420	PBP1/JAL30	(Nagano et al., 2008)
AT3G16430	JAL31	(Nagano et al., 2008)
AT3G16450	JAL33	(Nagano et al., 2008)
AT3G16460	JAL34	(Nagano et al., 2008)
AT3G16470	JAL35	(Nagano et al., 2008)
AT1G54000	GLL22	(Nagano et al., 2008)
AT2G22770	NAI1	(Matsushima et al., 2004)
AT4G27860	MEB1	(Yamada et al., 2013)
AT5G24290	MEB2	(Yamada et al., 2013)
AT1G52400	BGLU18	(Nakano et al., 2014)
AT5G23820	ML3	(Hakenjos et al., 2013)
AT1G54010	GLL23	(Nakano et al., 2016)
AT1G54030	MVP 1	(Agee et al., 2010; Nakano et al. 2012)
A T2 C1 50 50		
A13G15950	NAI2	(Yamada et al., 2008)

Supplemental Table 1. List of annotated ER body localized or associated proteins.

Supplemental table 2. List of annotated ER body localized or associated proteins altered in *ceh1*.

		Fold change	Fold change		
		(\log_2)		(\log_2)	
Gene ID	Name	Expression	P value	Protein	P value
AT1G52400	BGLU18	2.396541298	0.00299129	1.135421511	0.023237
AT3G09260	PYK10/BGLU23	1.175750063	0.00584943	2.419837605	0.000773
AT3G16420	PBP1/JAL30	n.d.	n.d.	1.222032104	0.003441
AT3G16450	JAL33	1.406667546	0.00579349	1.354872393	0.003285
AT3G16460	JAL34	0.732285878	0.2226334	1.929333173	0.031609
AT3G16470	JAL35	2.158159579	0.04639408	0.621660709	0.026101
AT1G54010	GLL23	n.d.	n.d.	2.344242453	0.026378
AT3G15950	NAI2	1.020693235	0.17069045	1.786527024	0.006863
AT4G27860	MEB1	3.250747779	0.03079126	1.781855467	0.025035
AT5G23820	ML3	1.368483384	0.01896147	0.281228153	0.144649

Supplemental Table 3. Transcriptomics analysis of GSLs synthesis genes in *ceh1* and *ceh1/eds16*.

Glucosinolate		Gene		ceh vs. wt			ceh/eds vs.	wt		eds vs. w	t	
Ali/Indol	Function	TAIR ID	NAME	log2 FC	q		log2 FC	q		log2 FC	q	
Indolic	Core structure	AT2G22330	CYP79B3	-0.2393	0.181		-0.0575	0.801		0.1242	0.708	
Indolic	Core structure	AT2G30860	GSTF9	1.00816	2E-08	*	1.34414	2E- 14	*	0.3424	0.242	
Indolic	Core structure	AT2G30870	GSTF10	1.51315	6E-16	*	2.26044	9E- 35	*	-0.008	0.99	
Indolic	Core structure	AT1G24100	UGT74B1	0.11227	0.653		0.47359	0.027		-0.027	0.961	
Indolic	Core structure	AT1G74100	SOT16	0.59503	8E-06		1.10667	5E- 18	*	-0.073	0.815	
Indolic	unknown	AT1G76790	AT1G76790	2.64603	3E-30	*	3.88737	1E- 65	*	1.4472	2E- 07	*
Indolic	Core structure	AT4G31500	CVP83B1	0 77821	3E-12		1.05165	1E- 21	*	-0.287	0.093	
Indolic		AT4637400	CVD81E3	-0 2013	0.661		-0.8608	0.045		0.1005	0.055	
Indolic	unknown	AT4037400	CVD81F4	-0.2015	0.001	*	1 95262	8E-		0.1003	0.913	
Indolic	unknown	A14G37410	CYP01F4	1.37819	0.014		1.65505	04		0.2978	NA 0.005	
Indolic	unknown	A14G37430	CYP81F1	0.80999	0.013		0.55/12	0.127 1E-		0.0976	0.905	
Indolic	Core structure	AT4G39950	CYP79B2	1.71285	7E-22	*	2.1347	33 2E-	*	-0.02	0.972	
Indolic	Core structure	AT5G05730	ASA1	1.00615	1E-16	*	1.43772	33 7E-	*	-0.212	0.343	
Indolic	Secondary Modification	AT1G21100	IGMT1	0.99226	7E-07		2.0832	28 3E-	*	-0.248	0.538	
Indolic	Secondary Modification	AT1G21120	IGMT2	1.21815	9E-06	*	2.0722	15 6E-	*	-1.049	0.016	*
Indolic	Secondary Modification	AT1G21110	IGMT3	1.20028	2E-06	*	1.9029	15 4E-	*	-0.28	0.584	
Indolic	Secondary Modification	AT1G21130	IGMT4	0.8836	1E-07		1.62861	24 6E-	*	-0.327	0.229	
Indolic	Core structure	AT5G54810	TSB1	0.48686	6E-04		1.20665	20 1E-	*	-0.053	0.877	
Indolic	Core structure	AT5G57220	CYP81F2	2.32941	1E-16	*	3.26757	32	*	-0.635	0.28	
Indolic	Core structure	AT5G05260	CYP79A2	0	NA		0	NA		0.5108	NA	
Aliphatic	Secondary modification	AT1G12140	OX5	-0.1466	0.369		-0.0554	0.78		-0.082	0.794	
Aliphatic	Core structure	AT1G16400	CYP79F2	0.317	0.25		0.09901	0.776		0.1188	0.839	
Aliphatic	Core structure	AT1G16410	CYP79F1	-0.9739	2E-06		-0.8671	3E- 05		0.143	0.741	
Aliphatic	Core structure	AT1G18590	SOT17	-0.3329	0.124		-0.2552	0.28		0.0627	0.896	
Aliphatic	Side-chain elongation	AT1G31180	IMDH1	-0.3635	0.006		-0.2598	0.068		0.2077	0.326	
Aliphatic	Secondary modification	AT1G62540	FMOGS- OX2	-0.2485	0.376		-0.4027	0.164		0.9407	9E- 04	
Aliphatic	Secondary modification	AT1G62560	FMOGS- OX3	-1.2003	7E-11	*	-1.2947	5E- 12	*	0.4138	0.146	
Aliphatic	Secondary modification	AT1G62570	FMOGS- OX4	-0.4049	0.208		0.19558	0.599		0.0302	0.971	
Aliphatic	Secondary modification	AT1G65860	FMOGS- OX1	-0.8613	8E-05		-0.9229	3E- 05		0.3327	0.366	
Aliphatic	Core structure	AT1G74090	SOT18	-0.7812	5E-06		-0.5582	0.002		0.0668	0.87	
Aliphatic	Core structure	AT1G78370	GSTU20	-0.9458	7E-06		-1.108	1E- 07	*	0.6143	0.033	
Aliphatic	Side-chain elongation	AT1G80560	IMDH2	0.0582	0.727		0.32618	0.018		0.2093	0.336	
Aliphatic	Secondary modificatio	AT2G25450	GSL-OH	1,29853	9E-25	*	1.7338	1E- 43	*	0.4221	0.017	
Aliphatic	Core structure	AT2G31790	UGT74C1	-0.8413	2E-05		-0 5995	0.004		0.062	0.896	
Aliphatic	Side-chain elongation	AT2G/3100		-0.768	15-04		-0.9607	1E- 06		0.002	0.000	
Aliphatic		AT2C02100	GSTE11	-0.700	0.002		-0.3007	0.100		0.3005	0.200	
Aliphotic		AT2C10710	DCATA	-0.7026	0.003		-0.4274	0.106		0.7727	0.007	
Апрпатіс	Silve-chain elongation	A13G19/10	BCA14	-0.4958	0.005		-0.4193	0.024 3E-		0.2858	0.314	
Aliphatic	Side-chain elongation	AT3G49680	BCAT3	-0.5933	5E-10		-0.6429	11 2E-		0.2755	0.041	
Aliphatic	Side-chain elongation	AT3G58990	IPMI1	-0.6562	0.004		-0.9515	05		0.2845	0.47	

Aliphatic Secondary modification AT4G03060 AOP2 -0.8702 1E-05 -0.9745 06 8E 0.4443 0.134 Aliphatic Side-chain elongation AT4G12030 BASS5 -1.054 3E-06 • -1.1129 07 • 0.2665 0.523 Aliphatic Side-chain elongation AT4G13430 IIL1 -0.2764 0.063 -0.9715 0.11 07 • 0.2665 0.523 Aliphatic Core structure AT4G13430 IIL1 -0.2764 0.063 -0.971 0.13 -0.005 0.99 Aliphatic Core structure AT4G03050 AOP3 O NA 0.13791 NA 0.5537 0.005 Aliphatic Side-chain elongation AT5G14200 IMDH3 -0.9051 6E-10 -0.9231 10 0.3537 0.005 3E-5 Aliphatic Side-chain elongation AT5G23020 MAM1 -0.611 7E-05 -0.7496 06 0.3623 0.135 SE- Aliphatic and indolic Core structure AT4G30530 AT4G30530 0.61748 1E-04<	
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Aliphatic and indolic co-substrate AT4G23100 GSH1 -0.4229 0.004 0.10768 0.559 -0.097 0.753 Aliphatic and indolic co-substrate AT5G65940 CHY1 0.27467 0.08 0.21992 0.198 0.1716 0.531 Aliphatic and indolic co-substrate AT1G65880 BZ01 -0.8681 0.155 -0.2651 0.725 0.1042 NA Aliphatic and indolic co-substrate AT1G65880 BZ01 -0.8681 0.155 -0.2651 0.725 0.1042 NA Aliphatic and indolic co-substrate AT2G14750 APK1 -0.1153 0.445 -0.1953 0.187 0.2499 0.211 Aliphatic and indolic Transcription factor AT1G32640 MYC2 0.70496 35-04 1.45388 0.215 0.568	
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Aliphatic and indolic Transcription factor AT5G46760 MYC3 -0.158 0.409 0.24509 0.193 -0.292 0.276	
Aliphatic and indolic Transcription factor AT4G17880 MYC4 -1.4097 2E-15 * -1.548 17 * -0.177 0.616	
Aliphatic Transcription factor AT5G61420 MYB28 -0.9735 3E-09 -1.1561 12 * -0.028 0.952	
Aliphatic Transcription factor AT5G07690 MYB29 -1.6437 1E-17 * -1.7609 19 * 0.2147 0.564	
Aliphatic Transcription factor AT5G07700 MYB76 -0.908 0.009 -1.0325 0.005 * 0.3183 0.613	
Indolic Transcription factor AT5G60890 MYB34 -0.8344 8E-06 -0.9473 07 0.1147 0.777	
Indolic Transcription factor AT1G18570 MYB51 2.09471 1E-36 * 2.83585 67 * -0.529 0.055	
9E- Indolic Transcription factor AT1G74080 MYB122 2.0296 5E-04 * 2.54066 06 * -0.348 NA	
9E- Indolic Transcription factor AT4G17490 ERF6 2.26971 4E-10 * 3.6804 26 * 0.229 0.798	
3E- Unknown Transcription factor AT5G46830 MYC5 -1.9652 5E-05 * -2.1547 05 * 0.3971 NA	

Asterisk (*) denote statistical significance.

Name	Primer	Note
NAI1-F	ACCATTCTCACTAAGATGGACAACAAATT	qRT Primer
NAI1-R	ACGTTTAACCACTGCGAAATTCACA	qRT Primer
NAI2-F	AACTTGAGTTCGAAGCACATATCAATGA	qRT Primer
NAI2-R	TTCACGTTCAATGGCTTCCAACA	qRT Primer
BGLU18-F	CGAGGTTCGGACTTTACTACATCGAT	qRT Primer
BGLU18-R	TGTCCCGATAGGATTCTTGATCATCT	qRT Primer
BGLU23-F	ACGTTACGAGAAAGAATCCGGCA	qRT Primer
BGLU23-R	TCTTGGTCACTCACAAAGTAAAACATAGGA	qRT Primer
TSA1-F	GGCTGACAAAGAAGAATCTGCAAAGA	qRT Primer
TSA1-R	GTTGGTATAACAATCGAGGAATCTTCGT	qRT Primer
JAL34-F	AAATCCAAGAGAGTTTGTGATTAACCATC	qRT Primer
JAL34-R	GCTGACCTCATAAGTGTTCAAGTTCGT	qRT Primer
GLL23-F	GAACGAATTGGCTAGAACAG	qRT Primer
GLL23-R	GCAACCATAAGCATCATGTG	qRT Primer
MEB1-F	AGGATGCCTTCTGCTTCTGT	qRT Primer
MEB1-R	GAGTGAAACGGGACCTCATT	qRT Primer
ERF6-F	ATTGTCTCCGTTGCCTACTA	qRT Primer
ERF6-R	GGTTTGGTTTCAAATTCAGA	qRT Primer
MYB51-F	AGCTCGTGGACTACCAGGAA	qRT Primer
MYB51-R	GGAGGTTATGCCCTTGTGTG	qRT Primer
MYB122-F	AACTTCATTGATCGGCGTCAC	qRT Primer
MYB122-R	ACCTCTTCGAATCTCCCCATC	qRT Primer
CYP81F2-F	GTCACAGGGAGACGCTACTAC	qRT Primer
CYP81F2-R	CACCACTGTTGTCATTGATGTC	qRT Primer
CYP81F4-F	TTGTTGAACCACCCAAAAGTTT	qRT Primer
CYP81F4-R	GGAGGTAAGGAAGGTTTGCT	qRT Primer
NSP1-F	ACTCCCTTGGGGCCTATGT	qRT Primer
NSP1-R	GCATCTTAGCCCTGGACCTT	qRT Primer
ESM1-F	AACGCCGTTCTTCGTAGGAT	qRT Primer
ESM1-R	TTCGAGTGCACGTTTCCCTT	qRT Primer

Supplemental table 4. List of primers used in this study.

References:

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