

Supporting Information

Pd-Catalyzed asymmetric allylic substitution cascade using α -(pyridin-1-yl)-acetamides formed *in situ* as nucleophiles

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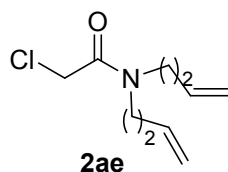
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1. General Information

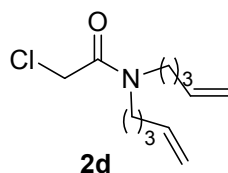
All the reactions were monitored by TLC using UV light to visualize the course of reaction. Anhydrous THF, DME, Et₂O, 1,4-dioxane and toluene were prepared by distillation over sodium-benzophenone prior to use. ¹H, ¹⁹F and ¹³C NMR spectra were obtained using a Varian MERCURY plus-400 or Bruker 500 spectrometer with TMS as an internal standard. HRMS was performed on a Bruker solariX FTICR Mass Spectrometer at the Instrumental Analysis Center of Shanghai Jiao Tong University. Melting points were measured with SGW X-4 micro melting point apparatus. Cinnamyl carbonates **1** were prepared according to literature procedures.^[1] All commercially available reagents were used as received.

2. Preparation of 2-Haloacetamides (2)

Preparation of 2-haloacetamides 2: To a solution of corresponding amine (10 mmol), triethylamine (2.02 g, 20 mmol) in DCM (50 mL) at 0 °C was added chloroacetyl chloride or bromoacetyl bromide (10 mmol) dropwise and stirred for 1 h. The solution was slowly warmed to room temperature and stirred overnight. The reaction mixture was diluted with DCM (100 mL), washed with brine, dried over Na₂SO₄ and then concentrated in vacuo. The residue was purified by column chromatography (PE/EtOAc = 10/1) to give the desired product. **2a**, **2b**, **2aa**, **2ab**, **2ac** and **2ad** were synthesized with this procedure and was consistent with the reported spectra^[2]. The data of **2ae** and **2d** were summarized below.



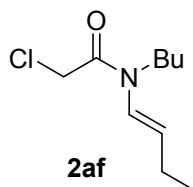
***N,N*-Di(but-3-en-1-yl)-2-chloroacetamide (2ae):** Yellow oil (1.68 g, 83%). ¹H NMR (400 MHz, CDCl₃): δ 5.86–5.65 (m, 2H), 5.19–4.96 (m, 4H), 3.82 (s, 2H), 3.46–3.21 (m, 4H), 2.44–2.19 (m, 4H); ¹³C NMR (100 MHz, CDCl₃): δ 166.8, 134.8, 133.7, 118.2, 117.0, 48.5, 45.7, 33.1, 31.5, 26.2; IR (KBr) cm⁻¹: 3426, 2978, 1643, 1461, 1256, 918; HRMS (APCI) [M+H]⁺ calcd 202.0993, found 202.0999.



2-Chloro-*N,N*-di(pent-4-en-1-yl)acetamide (2d): Yellow oil (1.72 g, 75%). ¹H NMR (400 MHz, CDCl₃): δ 5.86–5.69 (m, 2H), 5.12–4.88 (m, 4H), 4.03 (s, 2H), 3.39–3.18 (m, 4H), 2.15–1.95 (m, 4H), 1.75–1.59 (m, 4H); ¹³C NMR (100 MHz, CDCl₃): δ 166.3, 137.9, 137.1, 116.1, 115.3, 47.8, 45.9, 41.5, 31.2, 30.9, 28.2, 26.6; IR (KBr) cm⁻¹: 3453, 2976, 1651, 1462, 1432, 912; HRMS (APCI) [M+H]⁺ calcd 230.1306, found 230.1316.

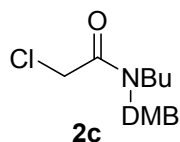
Preparation of 2af:^[3] A solution of *N*-butylidenebutan-1-amine (0.64 g, 5 mmol) in THF (10 mL) was stirred in a round bottom flask under a nitrogen atmosphere at room temperature. Then chloroacetyl chloride (0.57 g, 5 mmol) was added dropwise using a syringe. The mixture was stirred for 30 min at room temperature and then refluxed for a further 2 h. The reaction mixture

was concentrated in vacuo. The residue was purified by column chromatography (PE/EtOAc = 20/1) to give **2af** as a yellow oil.

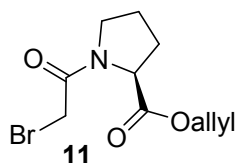


***N*-(But-1-en-1-yl)-*N*-butyl-2-chloroacetamide (**2af**):** Yellow oil (0.78 g, 76%). ¹H NMR (400 MHz, CDCl₃, major rotamer): δ 6.36 (dt, *J* = 14.0, 1.6 Hz, 1H), 5.33–5.22 (m, 1H), 4.14 (s, 2H), 3.57 (t, *J* = 7.6 Hz, 2H), 2.15–2.04 (m, 2H), 1.40–1.25 (m, 4H), 1.10–0.92 (m, 6H); ¹H NMR (400 MHz, CDCl₃, minor rotamer): δ 7.07 (d, *J* = 14.0 Hz, 1H), 5.19–5.10 (m, 1H), 4.13 (s, 2H), 3.49 (t, *J* = 8.0 Hz, 2H), 2.15–2.04 (m, 2H), 1.40–1.25 (m, 4H), 1.10–0.92 (m, 6H); ¹³C NMR (100 MHz, CDCl₃, major rotamer): δ 167.8, 126.1, 119.8, 44.5, 41.8, 29.1, 23.8, 20.3, 14.4, 14.0; ¹³C NMR (100 MHz, CDCl₃, minor rotamer): δ 167.3, 124.9, 115.5, 45.5, 41.4, 30.3, 22.9, 20.8, 14.7, 14.1; IR (KBr) cm⁻¹: 3427, 2934, 1666, 1441, 1260, 912; HRMS (APCI) [M+H]⁺ calcd 204.1150, found 204.1154.

Preparation of **2c and **11**:** To a solution of *N*-(2,4-dimethoxybenzyl)butan-1-amine or *L*-proline allyl ester hydrochloride (10 mmol) in water (5 mL) at 0 °C was added NaHCO₃ (2.52 g, 30 mmol) and the mixture was stirred for 10 min. Then, a solution of chloroacetyl chloride or bromoacetyl bromide (10 mmol) in toluene (5 mL) was added dropwise. The reaction was slowly warmed to room temperature and stirred for another 2 h. The reaction mixture was diluted with DCM (100 mL), washed with brine, dried over Na₂SO₄ and then concentrated in vacuo. The residue was purified by column chromatography to give the corresponding product.



***N*-Butyl-2-chloro-*N*-(2,4-dimethoxybenzyl)acetamide (**2c**):** Yellow oil (1.94 g, 65%). ¹H NMR (400 MHz, CDCl₃, mixture of two rotamers): δ 7.14 (d, *J* = 8.0 Hz, 0.3H), 6.94 (d, *J* = 8.0 Hz, 0.7H), 6.50–6.34 (m, 2H), 4.54 (s, 0.7H), 4.41 (s, 1.3H), 4.15 (s, 1.3H), 4.09 (s, 0.7H), 3.87–3.65 (m, 6H), 3.33–3.15 (m, 2H), 1.63–1.38 (m, 2H), 1.21–1.30 (m, 2H), 0.95–0.78 (m, 3H); ¹³C NMR (100 MHz, CDCl₃, mixture of two rotamers): δ 166.9, 166.6, 161.0, 160.5, 158.6, 158.5, 130.5, 128.8, 117.6, 116.6, 104.5, 104.2, 98.9, 98.5, 60.5, 59.5, 55.6, 55.4, 47.5, 47.2, 45.9, 42.8, 41.9, 41.5, 31.0, 29.4, 21.2, 20.3, 14.0, 13.9; IR (KBr) cm⁻¹: 3011, 1749, 1670, 1653, 922; HRMS (APCI) [M+H]⁺ calcd 300.1361, found 300.1378.



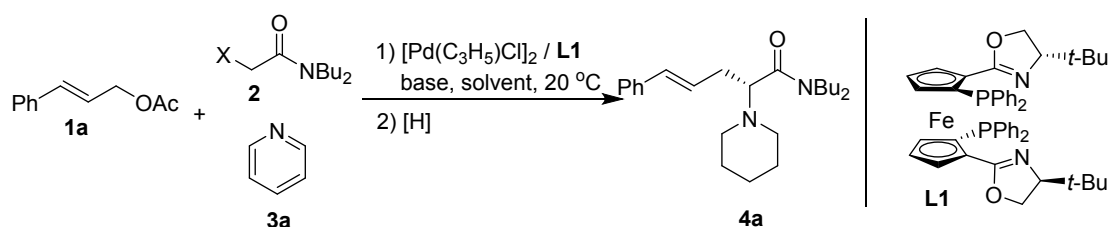
(*S*)-Allyl 1-(2-bromoacetyl)pyrrolidine-2-carboxylate (11**):** Yellow oil (2.41 g, 87%). ¹H NMR (500 MHz, CDCl₃, mixture of two rotamers): δ 6.06–5.79 (m, 1H), 5.47–5.15 (m, 2H), 4.74–4.49 (m, 3H), 4.05–3.51 (m, 4H), 2.39–1.86 (m, 4H); ¹³C NMR (126 MHz, CDCl₃, major rotamer): δ 171.4, 165.3, 131.8, 118.5, 65.8, 59.3, 47.5, 29.2, 26.9, 24.9; ¹³C NMR (126 MHz,

CDCl₃, minor rotamer): δ 171.2, 165.5, 131.2, 119.5, 66.4, 59.8, 47.0, 31.2, 27.0, 22.4; IR (KBr) cm⁻¹: 2958, 1743, 1648, 1418, 913; HRMS (APCI) [M+H]⁺ calcd 276.0230, found 276.0256.

3. Optimization of the Reaction Conditions

Our study began with the reaction of cinnamyl acetate (**1a**), 2-chloro-*N,N*-dibutylacetamide (**2a**) and pyridine (**3a**) using a catalytic system consisting of [Pd(η^3 -C₃H₅)Cl]₂ and a planar chiral phosphino-oxazoline ligand **L1** under a nitrogen atmosphere at room temperature for 12 h (Table S1). Unfortunately, no reaction occurred possible due to the low reaction activity for chloroacetamide. Then, 2-bromo-*N,N*-dibutylacetamide (**2b**) was employed instead of **2a** in the above reaction (entry 1). To our delight, the reaction processed smoothly with the desired product **4a** being obtained in 92% yield albeit with low enantioselectivity (entry 2). Subsequently, we examined the effect of solvent on the reaction. It was found that the reaction in protic alcohol solvents presented high yields but with low enantioselectivities (entries 1–4). Therefore, aprotic solvents such as toluene, THF, dioxane was examined and only trace amount of products were formed but with around 40% ee (entries 6–7). No product was observed at all when the reaction was carried out in toluene (entry 5). When MeCN was used, high yield with a little lower enantioselectivity were obtained (entry 8). We envisaged that the low yields might be attributed to low solubility of pyridine quaternary salt intermediates in ether or hydrocarbon solvents. So, some large-polar solvents were added to the above reaction as co-solvents. We were pleased to find that these co-solvents promoted the reaction efficiently (entries 9–11) and a mixed solvent of dioxane and DMSO (10/1, v/v) was found to be the best one (entry 10). Several bases were examined and 1,1,3,3-tetramethylguanidine (TMG) was undoubtedly the best choice (entries 12–16).

Table S1 The effect of solvent and base on the reaction^[a]



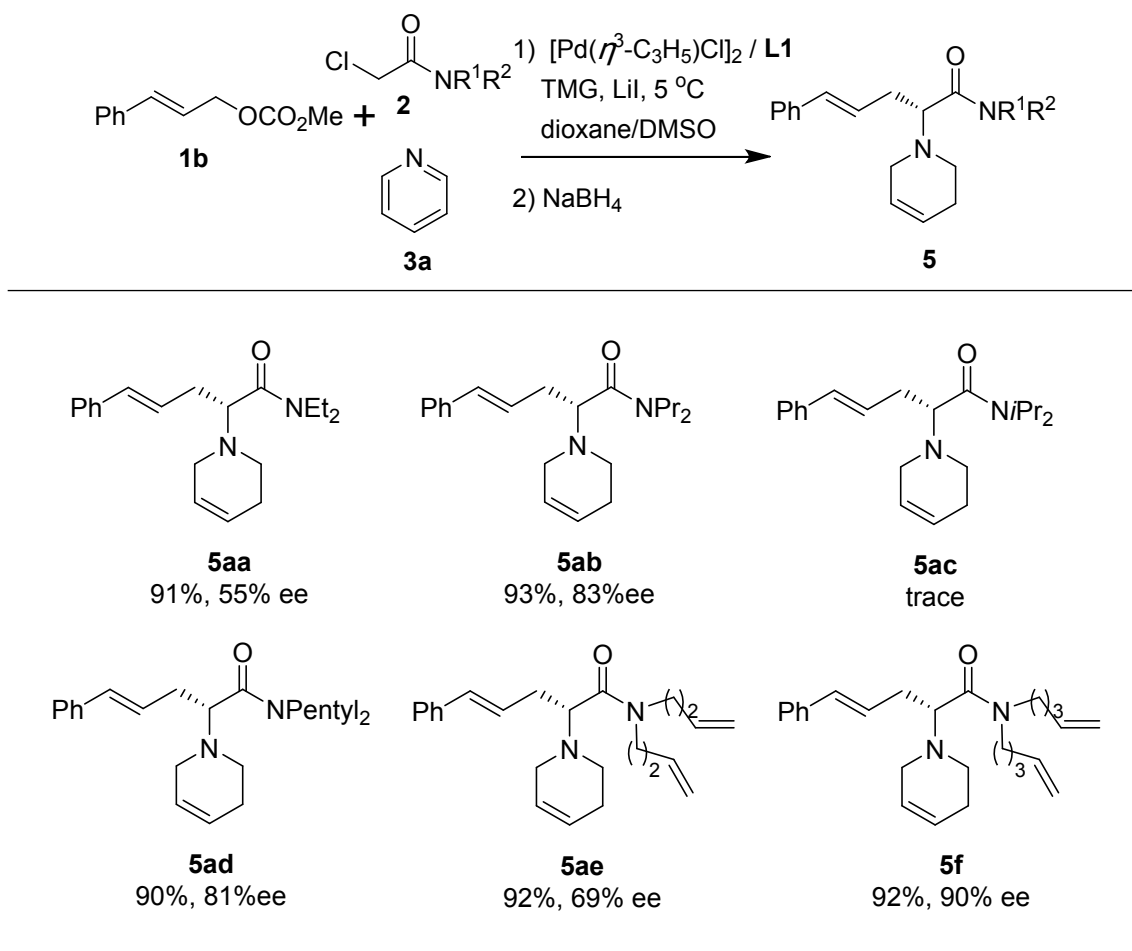
Entry	X	Solvent	Base	Yield (%) ^[b]	Ee (%) ^[c]
1	Cl	EtOH	TMG	NP	-
2	Br	EtOH	TMG	92	<10
3	Br	<i>n</i> -PrOH	TMG	95	<10
4	Br	<i>i</i> -PrOH	TMG	92	<10
5	Br	toluene	TMG	NP	-
6	Br	THF	TMG	trace	20
7	Br	dioxane	TMG	trace	43
8	Br	MeCN	TMG	91	41
9	Br	dioxane ^[d]	TMG	71	60
10	Br	dioxane ^[e]	TMG	92	77
11	Br	dioxane ^[f]	TMG	58	55
12	Br	dioxane ^[e]	Et ₃ N	NP	-

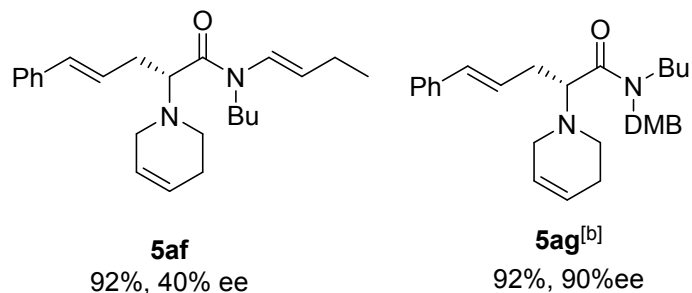
13	Br	dioxane ^[e]	Na ₂ CO ₃	NP	-
14	Br	dioxane ^[e]	Cs ₂ CO ₃	72	<10
15	Br	dioxane ^[e]	<i>t</i> -BuONa	NP	-
16	Br	dioxane ^[e]	DBU	88	24

[a] Reaction conditions: **1a** (0.1 mmol) with **2a** (0.2 mmol) and **3a** (0.5 mmol) under nitrogen atmosphere with a catalytic system of [Pd(η^3 -C₃H₅)Cl]₂ (2.5 mol%) and L* (6 mol%) in the presence of a base (0.16 mmol) in an indicated solvent (2 mL) at 20 °C for 12 h; The pyridine quaternary salt was reduced with Raney-Ni under H₂ atmosphere. [b] Isolated yield. [c] Determined by HPLC using a chiral Daicel column. [d] MeCN was added as a co-solvent. [e] DMSO was added as a co-solvent. [f] DMF was added as a co-solvent

Then, **2** with different *N*-substituents were taken into consideration (Table S2). Linear alkyl groups, such as Et, Pr, pentyl could give higher yields but with somewhat lower enantioselectivities (**5aa**, **5ab** and **5ad**) than that of Bu group (**4a**, 96% yield and 95% ee). When *i*-Pr group adopted, only trace amount of product was obtained (**5ac**). Linear unsaturated groups with different steric hindrance were subjected to the reaction, no better results were obtained (**5ae** and **5b**). Replacing one Bu group on the nitrogen atom of **2a** with butenyl group resulted in a dramatically decrease of enantioselectivity (**5af**). When DMB (2,4-dimethoxybenzyl) was used instead of Bu group on the nitrogen atom of **2a**, the desired product **5ag** was obtained in 92% yield and 90% ee.

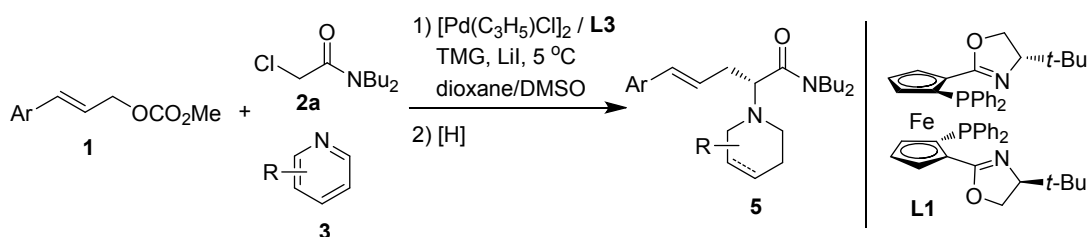
Table S2 Screen of **2** with different *N*-substituents^[a]





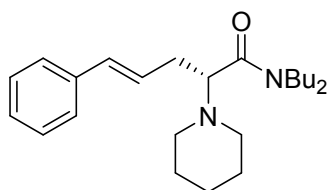
[a] Reaction conditions: **1a** (0.1 mmol) with **2a** (0.2 mmol) and **3a** (0.5 mmol) in a mixed solvent of dioxane/DMSO (10/1, v/v, 2mL) under nitrogen atmosphere with a catalytic system of $[\text{Pd}(\eta^3\text{-C}_3\text{H}_5)_2\text{Cl}]_2$ (2.5 mol%) and **L*** (6.0 mol%) with 1,1,3,3-Tetramethylguanidine (TMG) as a base (0.16 mmol) in the presence of LiI (0.1 mmol) at 5 °C for 12 h; The pyridine quaternary salt was reduced by NaBH_4 ; Isolated yields; ees were determined by HPLC using a chiral Daicel column. [b] Data and spectra were collected after removing the DMB group with TFA, see compound **10**.

4. General Procedure: Pd-Catalyzed Three-Component Allylic Substitution



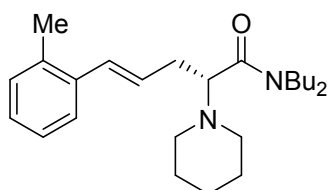
To an oven-dried glassware were added $[\text{Pd}(\text{C}_3\text{H}_5)_2\text{Cl}]_2$ (0.9 mg, 2.5 mol%), **L1** (4.8 mg, 5.5 mol%) and anhydrous dioxane (1.0 mL). To another dry glassware were added **2** (0.2 mmol), **3** (0.5 mmol), ultra-dry LiI (0.1 mmol, 13.3 mg) and anhydrous dioxane (1.0 mL). Then the first glassware was stirred at RT and second glassware was stirred at 100 °C under a nitrogen atmosphere for 1 h. Allylic carbonate **1** (0.1 mmol) was added to the first glassware. DMSO (0.2 mL) was added to the second glassware followed by 1,1,3,3-Tetramethylguanidine (TMG, 0.16 mmol). The solution in the first glassware was added dropwise to another one. The reaction was stirred at 5 °C for 12 h. The product was reduced *in situ* by one of the following methods. (1) Reduced by H_2 : Raney-Ni (water slurry, 50 μm , 0.2 mL) was added to the reaction mixture and the mixture was stirred at H_2 atmosphere (10 bar) for 1 h. The reaction mixture was filtrated, diluted with EtOAc, washed with brine, dried over anhydrous Na_2SO_4 and then concentrated in vacuo. The residue was then purified by flash column chromatography (PE/EtOAc = 10/1) to give the corresponding products. (2) Reduced with NaBH_4 : The solvent was removed in vacuo. MeOH (2 mL) was added and the resulting mixture was cooled to -20 °C quickly. Then, NaBH_4 (0.5 mmol) was added in one portion and the reaction was stirred for 2 h. The reaction mixture was diluted with EtOAc, washed with brine, dried over anhydrous Na_2SO_4 and then concentrated in vacuo. The residue was purified by flash column chromatography (PE/EtOAc = 10/1) to give the corresponding products. (3) Reduced with NaBH_4 for alkyl substituted substrates (**5a**, **5b**, **5c**, **5d** and **5e**): The solvent was removed in vacuo. MeOH (2 mL) was added and the resulting mixture was cooled to -40 °C quickly. Then, NaBH_4 (0.5 mmol) was added in one portion and the reaction was stirred for 2 h. The reaction mixture was diluted with EtOAc, washed with brine, dried over anhydrous Na_2SO_4 and then concentrated in vacuo. The residue was purified by flash column

chromatography (PE/EtOAc = 10/1) to give the corresponding products.



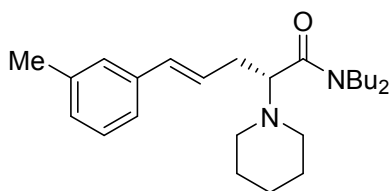
(E)-N,N-Dibutyl-5-phenyl-2-(piperidin-1-yl)pent-4-enamide (4a): Yellow oil (35.6 mg, 96%).

$[\alpha]_D^{25} = +59.9$ (*c* 0.18, acetone); $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 7.33–7.21 (m, 4H), 7.20–7.10 (m, 1H), 6.41 (d, $J = 16$ Hz, 1H), 6.14 (dt, $J = 15.6, 7.6$ Hz 1H), 3.51–3.33 (m, 3H), 3.22–3.00 (m, 2H), 2.83–2.72 (m, 1H), 2.71–2.58 (m, 2H), 2.53–2.32 (m, 3H), 1.64–1.30 (m, 10H), 1.33–1.19 (m, 4H), 0.86 (q, $J = 7.2$ Hz 6H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 170.5, 137.7, 131.6, 128.4, 128.2, 126.8, 125.9, 65.8, 50.3, 47.4, 45.6, 31.5, 29.8, 28.9, 26.6, 24.5, 20.3, 20.2, 13.9, 13.8; IR (KBr) cm^{-1} : 2976, 2905, 1671, 1456, 1382, 879; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd 371.3057, found 371.3088; Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 90/10, 0.8 mL/min, 254 nm: $t_{\text{major}} = 24.379$ min, $t_{\text{minor}} = 18.314$ min.



(E)-N,N-Dibutyl-2-(piperidin-1-yl)-5-(o-tolyl)pent-4-enamide (4b): Yellow oil (36.2 mg, 94%).

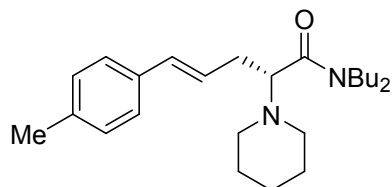
$[\alpha]_D^{25} = +45.9$ (*c* 0.22, acetone); $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 7.35 (d, $J = 5.6$ Hz, 1H), 7.13–7.06 (m, 3H), 6.66–6.54 (d, $J = 15.6$ Hz, 1H), 6.03 (dt, $J = 15.6, 7.6$ Hz 1H), 3.54–3.34 (m, 3H), 3.19–3.05 (m, 2H), 2.88–2.73 (m, 1H), 2.72–2.59 (m, 2H), 2.57–2.40 (m, 3H), 2.29 (s, 3H), 1.52–1.18 (m, 14H), 0.87 (t, $J = 7.2$ Hz, 6H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 170.5, 136.8, 134.9, 130.1, 129.6, 126.8, 125.9, 125.4, 109.9, 65.8, 50.4, 47.4, 45.6, 31.6, 29.8, 29.3, 26.6, 24.5, 20.3, 20.2, 19.8, 13.9, 13.8; IR (KBr) cm^{-1} : 2971, 2849, 1660, 1507, 938; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd 385.3213, found 385.3230; Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 95/5, 0.8 mL/min, 254 nm: $t_{\text{major}} = 25.141$ min, $t_{\text{minor}} = 14.969$ min.



(E)-N,N-Dibutyl-2-(piperidin-1-yl)-5-(m-tolyl)pent-4-enamide (4c): Yellow oil (35.3 mg, 92%).

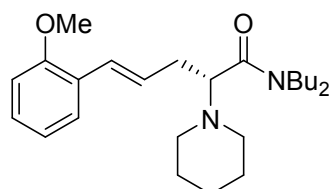
$[\alpha]_D^{25} = +35.5$ (*c* 0.28, acetone); $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 7.19–7.05 (m, 3H), 6.98 (d, $J = 6.8$ Hz, 1H), 6.37 (d, $J = 16.4$ Hz, 1H), 6.18–6.07 (m, 1H), 3.52–3.34 (m, 3H), 3.19–3.05 (m, 2H), 2.80–2.71 (m, 1H), 2.70–2.56 (m, 2H), 2.56–2.35 (m, 3H), 2.30 (s, 3H), 1.56–1.19 (m, 14H), 0.88 (q, $J = 6.8$ Hz, 6H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 170.5, 137.8, 137.6, 131.7, 128.2, 128.1, 127.5, 126.6, 123.1, 65.9, 50.3, 47.4, 45.6, 31.5, 29.7, 28.9, 26.5, 24.5, 21.3, 20.2, 20.1, 13.9, 13.8; IR (KBr) cm^{-1} : 2978, 2906, 1716, 1418, 668; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd 385.3213, found

385.3221 Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 95/5, 0.8 mL/min, 254 nm: $t_{\text{major}} = 19.604$ min, $t_{\text{minor}} = 16.423$ min.



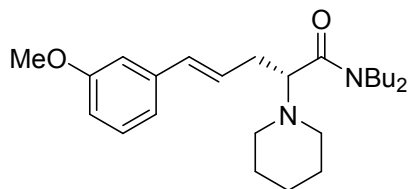
(E)-N,N-Dibutyl-2-(piperidin-1-yl)-5-(p-tolyl)pent-4-enamide (4d): Yellow oil (33.2 mg, 86%).

$[\alpha]_D^{25} = +48.4$ (*c* 0.33, acetone); $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 7.18 (d, $J = 8$ Hz, 2H), 7.03 (d, $J = 7.6$ Hz, 2H), 6.41–6.31 (d, $J = 16$ Hz, 1H), 6.16–6.01 (m, 1H), 3.52–3.34 (m, 3H), 3.22–3.04 (m, 2H), 2.84–2.71 (m, 1H), 2.70–2.58 (m, 2H), 2.59–2.35 (m, 3H), 2.29 (s, 3H), 1.55–1.11 (m, 14H), 0.85 (q, $J = 6.8$ Hz, 6H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 170.5, 136.4, 134.9, 131.4, 129.0, 127.2, 125.8, 65.8, 50.3, 47.3, 45.6, 31.5, 29.7, 28.9, 26.5, 24.5, 21.1, 20.2, 20.1, 13.9, 13.8; IR (KBr) cm^{-1} : 2925, 1733, 1261, 1045, 879; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd 385.3213, found 385.3223; Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 95/5, 0.8 mL/min, 254 nm: $t_{\text{major}} = 27.475$ min, $t_{\text{minor}} = 21.027$ min.



(E)-N,N-Dibutyl-5-(2-methoxyphenyl)-2-(piperidin-1-yl)pent-4-enamide (4e): Yellow oil (34.1 mg, 85%).

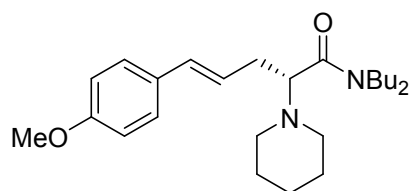
$[\alpha]_D^{25} = +68.2$ (*c* 0.18, acetone); $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 7.35 (d, $J = 7.6$ Hz, 1H), 7.13 (t, $J = 8$ Hz, 1H), 6.90–6.76 (m, 2H), 6.70 (d, $J = 16$ Hz, 1H), 6.20–6.07 (m, 1H), 3.81 (s, 3H), 3.53–3.32 (m, 3H), 3.21–3.05 (m, 2H), 2.82–2.71 (m, 1H), 2.72–2.58 (m, 2H), 2.57–2.37 (m, 3H), 1.57–1.17 (m, 14H), 0.96–0.79 (m, 6H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 170.7, 156.1, 128.9, 127.8, 126.7, 126.3, 125.9, 120.5, 110.6, 65.9, 55.3, 50.3, 47.4, 45.6, 31.5, 29.7, 29.3, 26.5, 24.5, 20.2, 20.1, 13.9, 13.8; IR (KBr) cm^{-1} : 2978, 2360, 1716, 1507, 1086, 1046, 879; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd 401.3163, found 401.3173; Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 85/15, 0.8 mL/min, 254 nm: $t_{\text{major}} = 30.257$ min, $t_{\text{minor}} = 14.089$ min.



(E)-N,N-Dibutyl-5-(3-methoxyphenyl)-2-(piperidin-1-yl)pent-4-enamide (4f): Yellow oil (38.2 mg, 95%).

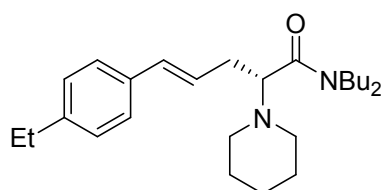
$[\alpha]_D^{25} = +44.9$ (*c* 0.25, acetone); $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 7.16 (t, $J = 8$ Hz, 1H), 6.88 (d, $J = 7.6$ Hz, 1H), 6.84–6.80 (m, 1H), 6.71 (dd, $J = 8.4, 2.4$ Hz, 1H), 6.37 (d, $J = 16$ Hz, 1H), 6.19–6.07 (m, 1H), 3.77 (s, 3H), 3.51–3.35 (m, 3H), 3.20–3.04 (m, 2H), 2.83–2.70 (m, 1H), 2.71–2.56 (m, 2H), 2.53–2.36 (m, 3H), 1.60–1.17 (m, 14H), 0.95–0.82 (m, 6H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 170.4, 159.7, 139.1, 131.5, 129.3, 128.7, 118.6, 112.5, 111.2, 65.8, 55.1,

50.3, 47.3, 45.6, 31.5, 29.7, 28.8, 26.5, 24.5, 20.2, 20.1, 13.9, 13.8; IR (KBr) cm^{-1} : 2978, 1716, 1558, 1261, 1085, 879; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd 401.3163, found 401.3168; Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 90/10, 0.8 mL/min, 254 nm: $t_{\text{major}} = 23.437$ min, $t_{\text{minor}} = 19.105$ min.



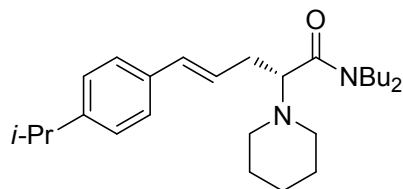
(*E*)-*N,N*-Dibutyl-5-(4-methoxyphenyl)-2-(piperidin-1-yl)pent-4-enamide (4g): Yellow oil

(33.5 mg, 84%). $[\alpha]_D^{25} = +46.6$ (*c* 0.32, acetone); ^1H NMR (400 MHz, CDCl_3): δ 7.22 (d, $J = 8.8$ Hz, 2H), 6.79 (d, $J = 8.4$ Hz, 2H), 6.34 (d, $J = 16.0$ Hz, 1H), 6.04–5.92 (m, 1H), 3.77 (s, 3H), 3.56–3.33 (m, 3H), 3.19–3.04 (m, 2H), 2.67 (dd, $J = 60.1, 42.9$ Hz, 6H), 1.60–1.16 (m, 14H), 0.96–0.78 (m, 6H); ^{13}C NMR (100 MHz, CDCl_3): δ 170.6, 158.6, 131.0, 130.5, 129.2, 127.0, 126.0, 113.8, 113.6, 65.8, 55.2, 50.3, 47.4, 45.6, 31.5, 29.7, 29.0, 26.5, 24.5, 20.2, 20.1, 13.9, 13.8; IR (KBr) cm^{-1} : 2979, 1716, 1558, 879; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd 401.3163, found 401.3173; Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 90/10, 0.8 mL/min, 254 nm: $t_{\text{major}} = 21.605$ min, $t_{\text{minor}} = 21.077$ min.



(*E*)-*N,N*-Dibutyl-5-(4-ethylphenyl)-2-(piperidin-1-yl)pent-4-enamide (4h): Yellow oil (35.9

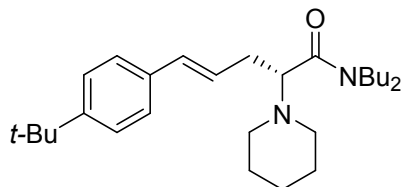
mg, 90%). $[\alpha]_D^{25} = +69.2$ (*c* 0.20, acetone); ^1H NMR (400 MHz, CDCl_3): δ 7.21 (d, $J = 8.0$ Hz, 2H), 7.08 (d, $J = 8.0$ Hz, 2H), 6.37 (d, $J = 16.0$ Hz, 1H), 6.14–6.03 (m, 1H), 3.52–3.34 (m, 3H), 3.20–3.04 (m, 2H), 2.79–2.71 (m, 1H), 2.70–2.52 (m, 4H), 2.53–2.34 (m, 3H), 1.32–1.14 (m, 17H), 0.89–0.82 (m, 6H); ^{13}C NMR (100 MHz, CDCl_3): δ 170.6, 142.9, 135.1, 131.5, 127.8, 127.3, 125.9, 65.8, 50.3, 47.3, 45.6, 31.5, 29.7, 28.9, 28.5, 26.5, 24.5, 20.3, 20.2, 15.5, 13.9, 13.8; IR (KBr) cm^{-1} : 2934, 1716, 1558, 1456, 880; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd 399.3370, found 399.3380; Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 95/5, 0.8 mL/min, 254 nm: $t_{\text{major}} = 21.694$ min, $t_{\text{minor}} = 19.572$ min.



(*E*)-*N,N*-Dibutyl-5-(4-isopropylphenyl)-2-(piperidin-1-yl)pent-4-enamide (4i): Yellow oil

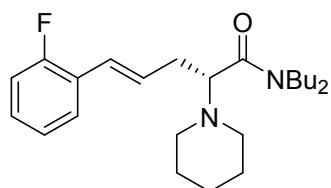
(38.6 mg, 94%). $[\alpha]_D^{25} = +71.9$ (*c* 0.22, acetone); ^1H NMR (400 MHz, CDCl_3): δ 7.23 (d, $J = 7.6$ Hz, 2H), 7.11 (d, $J = 7.6$ Hz, 2H), 6.37 (d, $J = 16.0$ Hz, 1H), 6.16–6.02 (m, 1H), 3.49–3.35 (m, 3H), 3.21–3.03 (m, 2H), 2.96–2.57 (m, 4H), 2.59–2.32 (m, 3H), 1.66–1.10 (m, 20H), 0.98–0.79

(m, 6H); ^{13}C NMR (100 MHz, CDCl_3): δ 170.6, 147.6, 135.3, 131.5, 127.4, 126.4, 125.9, 65.9, 50.4, 47.4, 45.6, 33.7, 31.5, 29.8, 28.9, 26.6, 24.5, 23.9, 20.3, 20.2, 13.8, 13.8.; IR (KBr) cm^{-1} : 2979, 1684, 1507, 1146, 879; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd 413.3526, found 413.3538; Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 95/5, 0.8 mL/min, 254 nm: t_{major} = 16.575 min, t_{minor} = 18.078 min.



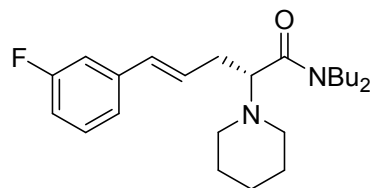
(E)-N,N-Dibutyl-5-(4-(tert-butyl)phenyl)-2-(piperidin-1-yl)pent-4-enamide (4j): Yellow oil 10

(39.7 mg, 93%). $[\alpha]_D^{25} = +37.2$ (*c* 0.20, acetone); ^1H NMR (400 MHz, CDCl_3): δ 7.32–7.18 (m, 4H), 6.38 (d, $J = 15.6$ Hz, 1H), 6.16–6.05 (m, 1H), 3.53–3.34 (m, 3H), 3.18–3.05 (m, 2H), 2.80–2.71 (m, 1H), 2.71–2.59 (m, 2H), 2.54–2.36 (m, 3H), 1.51–1.14 (m, 23H), 0.87 (q, $J = 7.2$ Hz, 6H); ^{13}C NMR (100 MHz, CDCl_3): δ 170.5, 149.8, 134.9, 131.3, 127.4, 125.6, 125.2, 65.9, 50.3, 47.3, 45.6, 34.4, 31.5, 31.2, 29.7, 28.9, 26.5, 24.50, 20.2, 20.1, 13.9, 13.8; IR (KBr) cm^{-1} : 2977, 1653, 1418, 1046, 879; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd 427.3683, found 427.3685; Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 95/5, 0.8 mL/min, 254 nm: t_{major} = 13.086 min, t_{minor} = 17.201 min.



(E)-N,N-Dibutyl-5-(2-fluorophenyl)-2-(piperidin-1-yl)pent-4-enamide (4k): Yellow oil (36.1 mg, 93%) 10

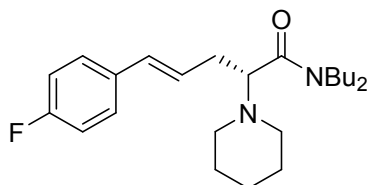
$[\alpha]_D^{25} = +50.6$ (*c* 0.25, acetone); ^1H NMR (400 MHz, CDCl_3): δ 7.37 (t, $J = 8.0$ Hz, 1H), 7.17–7.07 (m, 1H), 7.05–6.89 (m, 2H), 6.55 (d, $J = 16.0$ Hz, 1H), 6.30–6.16 (m, 1H), 3.56–3.29 (m, 3H), 3.20–3.05 (m, 2H), 2.85–2.74 (m, 1H), 2.74–2.57 (m, 2H), 2.57–2.32 (m, 3H), 1.63–1.16 (m, 14H), 0.87 (t, $J = 7.2$ Hz, 6H); ^{13}C NMR (100 MHz, CDCl_3): δ 170.5, 159.9 ($J_{\text{C-F}} = 250$ Hz), 131.2 ($J_{\text{C-F}} = 2$ Hz), 128.0 ($J_{\text{C-F}} = 10$ Hz), 127.0 ($J_{\text{C-F}} = 2$ Hz), 125.4 ($J_{\text{C-F}} = 10$ Hz), 123.9 ($J_{\text{C-F}} = 4$ Hz), 123.8 ($J_{\text{C-F}} = 4$ Hz), 115.5 ($J_{\text{C-F}} = 22$ Hz), 65.9, 50.3, 47.4, 45.6, 31.5, 29.7, 29.1, 26.5, 24.4, 20.3, 20.2, 13.9, 13.8; ^{19}F NMR (376 MHz, CDCl_3): δ -118.9; IR (KBr) cm^{-1} : 2979, 1773, 1559, 1419, 1045, 879; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd 389.2963, found 389.2966; Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 95/5, 0.8 mL/min, 254 nm: t_{major} = 13.455 min, t_{minor} = 11.373 min.



(E)-N,N-Dibutyl-5-(3-fluorophenyl)-2-(piperidin-1-yl)pent-4-enamide (4l): Yellow oil (33.5 mg, 86%) 10

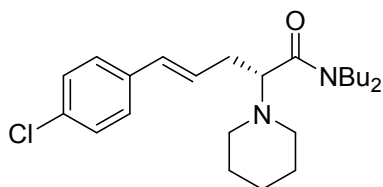
$[\alpha]_D^{25} = +69.2$ (*c* 0.26, acetone); ^1H NMR (400 MHz, CDCl_3): δ 7.19 (q, $J = 7.2$ Hz,

1H), 7.03 (d, $J = 7.6$ Hz, 1H), 6.98 (d, $J = 10.4$ Hz, 1H), 6.84 (t, $J = 9.2$ Hz, 1H), 6.36 (d, $J = 16.0$ Hz, 1H), 6.21–6.09 (m, 1H), 3.56–3.33 (m, 3H), 3.21–3.04 (m, 2H), 2.82–2.72 (m, 1H), 2.69–2.54 (m, 2H), 2.55–2.30 (m, 3H), 1.58–1.19 (m, 14H), 0.96–0.78 (m, 6H), 1.63–1.16 (m, 14H), 0.87 (t, $J = 7.2$ Hz, 6H); ^{13}C NMR (100 MHz, CDCl_3): δ 170.4, 163.0 ($J_{\text{C-F}} = 240$ Hz), 140.1 ($J_{\text{C-F}} = 10$ Hz), 130.6 ($J_{\text{C-F}} = 2$ Hz), 129.9, 129.7 ($J_{\text{C-F}} = 10$ Hz), 121.8 ($J_{\text{C-F}} = 2$ Hz), 113.5 ($J_{\text{C-F}} = 8$ Hz), 112.3 ($J_{\text{C-F}} = 8$ Hz), 65.7, 50.3, 47.3, 45.5, 31.5, 29.7, 28.6, 26.5, 24.4, 20.2, 20.1, 13.9, 13.8; ^{19}F NMR (376 MHz, CDCl_3): δ -113.9; IR (KBr) cm^{-1} : 2978, 1653, 1540, 1374, 879, 668; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd 389.2963, found 389.2967; Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 95/5, 0.8 mL/min, 254 nm: $t_{\text{major}} = 15.438$ min, $t_{\text{minor}} = 12.776$ min.



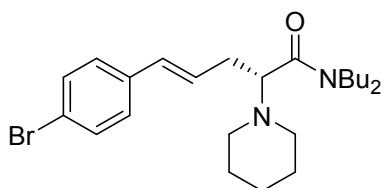
(E)-N,N-Dibutyl-5-(4-fluorophenyl)-2-(piperidin-1-yl)pent-4-enamide (4m): Yellow oil (36.1 mg, 93%).

$[\alpha]_D^{25} = +66.2$ (c 0.16, acetone); ^1H NMR (400 MHz, CDCl_3): δ 7.24 (dd, $J = 8.4, 5.2$ Hz, 2H), 6.93 (t, $J = 8.4$ Hz, 2H), 6.36 (d, $J = 16.0$ Hz, 1H), 6.10–5.97 (m, 1H), 3.54–3.30 (m, 3H), 3.20–3.04 (m, 2H), 2.83–2.72 (m, 1H), 2.72–2.57 (m, 2H), 2.55–2.31 (m, 3H), 1.70–1.13 (m, 14H), 0.98–0.77 (m, 6H); ^{13}C NMR (100 MHz, CDCl_3): δ 170.4, 161.8 ($J_{\text{C-F}} = 240$ Hz), 133.8 ($J_{\text{C-F}} = 4$ Hz), 130.4, 128.0 ($J_{\text{C-F}} = 2$ Hz), 127.3 ($J_{\text{C-F}} = 8$ Hz), 115.2 ($J_{\text{C-F}} = 22$ Hz), 65.7, 50.2, 47.3, 45.5, 31.5, 29.7, 28.7, 26.5, 24.4, 20.2, 20.1, 13.9, 13.8; ^{19}F NMR (376 MHz, CDCl_3): δ -115.7; IR (KBr) cm^{-1} : 2979, 2929, 1684, 1362, 879, 517; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd 389.2963, found 389.2960; Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 95/5, 0.8 mL/min, 254 nm: $t_{\text{major}} = 15.077$ min, $t_{\text{minor}} = 13.774$ min.



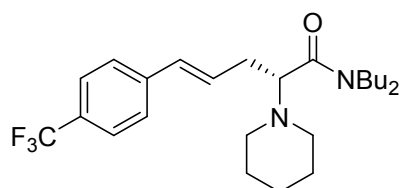
(E)-N,N-Dibutyl-5-(4-chlorophenyl)-2-(piperidin-1-yl)pent-4-enamide (4n): Yellow oil (36.5 mg, 90%).

$[\alpha]_D^{25} = +48.9$ (c 0.4, acetone); ^1H NMR (400 MHz, CDCl_3): δ 7.23–7.18 (m, 4H), 6.36 (d, $J = 14.8$ Hz, 1H), 6.20–6.01 (m, 1H), 3.60–3.25 (m, 3H), 3.21–3.07 (m, 2H), 2.92–2.25 (m, 6H), 1.53–1.19 (m, 14H), 0.91–0.84 (m, 6H); ^{13}C NMR (100 MHz, CDCl_3): δ 170.4, 136.2, 132.3, 130.4, 129.1, 128.5, 127.1, 65.7, 50.3, 47.3, 45.6, 31.5, 29.7, 26.5, 24.5, 22.7, 20.3, 20.2, 13.9, 13.8; IR (KBr) cm^{-1} : 2978, 2927, 1684, 1362, 879, 568; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd 405.2667, found 405.2668; Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 95/5, 0.4 mL/min, 254 nm: $t_{\text{major}} = 38.173$ min, $t_{\text{minor}} = 22.855$ min.



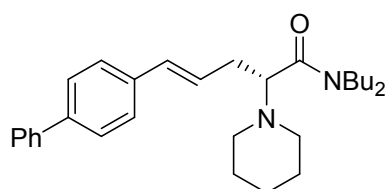
(E)-N,N-Dibutyl-5-(4-bromophenyl)-2-(piperidin-1-yl)pent-4-enamide (4o): Yellow oil (42.2 mg, 94%).

$[\alpha]_D^{25} = +29.9$ (c 0.36, acetone); $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 7.36 (d, $J = 8.4$ Hz, 2H), 7.15 (d, $J = 8.4$ Hz, 2H), 6.34 (d, $J = 16.0$ Hz, 1H), 6.13 (dt, $J = 15.2, 7.6$ Hz, 1H), 3.55–3.33 (m, 3H), 3.20–3.04 (m, 2H), 2.82–2.73 (m, 1H), 2.71–2.57 (m, 2H), 2.56–2.36 (m, 3H), 1.57–1.20 (m, 14H), 0.93–0.81 (m, 6H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 170.4, 136.6, 131.4, 130.5, 129.3, 127.4, 120.4, 65.7, 50.3, 47.3, 45.5, 31.5, 29.7, 28.7, 26.5, 24.4, 20.2, 20.1, 13.9, 13.8; IR (KBr) cm^{-1} : 2977, 2928, 1683, 1066, 879, 688; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd 449.2162, found 449.2164; Daicel Chiralpak IC-3 column, n -hexane/ i -PrOH = 95/5, 0.8 mL/min, 254 nm: $t_{\text{major}} = 25.302$ min, $t_{\text{minor}} = 17.077$ min.



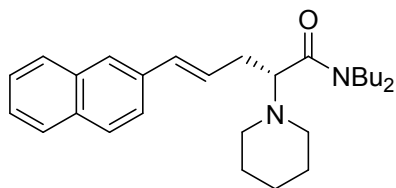
(E)-N,N-Dibutyl-2-(piperidin-1-yl)-5-(4-(trifluoromethyl)phenyl)pent-4-enamide (4p):

Yellow oil (33.0 mg, 75%). $[\alpha]_D^{25} = +73.3$ (c 0.24, acetone); $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 7.51 (d, $J = 8.4$ Hz, 2H), 7.39 (d, $J = 8.4$ Hz, 2H), 6.45 (d, $J = 16.0$ Hz, 1H), 6.33–6.23 (m, 1H), 3.57–3.36 (m, 3H), 3.19–3.06 (m, 2H), 2.88–2.79 (m, 1H), 2.76–2.62 (m, 2H), 2.61–2.39 (m, 3H), 1.54–1.18 (m, 14H), 0.92–0.85 (m, 6H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 170.3, 141.1, 131.4, 127.9 ($J_{\text{C-F}} = 190$ Hz), 130.4, 126.0, 125.3 (q, $J_{\text{C-F}} = 4$ Hz), 65.8, 50.3, 47.3, 45.5, 31.5, 29.7, 28.6, 26.5, 24.4, 20.2, 20.1, 13.9, 13.8; $^{19}\text{F NMR}$ (376 MHz, CDCl_3): δ -62.4; IR (KBr) cm^{-1} : 2978, 2923, 1670, 1653, 879; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd 439.2931, found 439.2928; Daicel Chiralpak AD-H column, n -hexane/ i -PrOH = 99.5/0.5, 0.8 mL/min, 254 nm: $t_{\text{major}} = 27.012$ min, $t_{\text{minor}} = 15.523$ min.



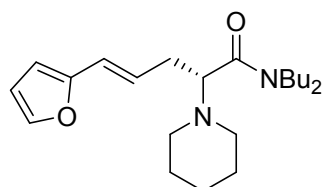
(E)-5-([1,1'-Biphenyl]-4-yl)-N,N-dibutyl-2-(piperidin-1-yl)pent-4-enamide (4q): Yellow oil

(40.7 mg, 91%). $[\alpha]_D^{25} = +49.9$ (c 0.25, acetone); $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 7.57 (d, $J = 8.4$ Hz, 2H), 7.50 (d, $J = 8.4$ Hz, 2H), 7.45–7.20 (m, 5H), 6.45 (d, $J = 16.0$ Hz, 1H), 6.31–6.08 (m, 1H), 3.62–3.30 (m, 3H), 3.24–3.01 (m, 2H), 2.90–2.75 (m, 1H), 2.75–2.59 (m, 2H), 2.57–2.26 (m, 3H), 1.38–1.18 (m, 14H), 0.85–0.94 (m, 6H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 170.5, 140.8, 139.5, 136.7, 131.2, 128.7, 128.5, 127.1, 127.0, 126.8, 126.3, 65.8, 50.3, 47.4, 45.6, 31.5, 29.6, 28.9, 26.5, 24.5, 20.3, 20.2, 13.9, 13.8; IR (KBr) cm^{-1} : 1748, 1540, 1472, 912; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd 447.3370, found 447.3380; Daicel Chiralpak IC-3 column, n -hexane/ i -PrOH = 95/5, 0.8 mL/min, 254 nm: $t_{\text{major}} = 27.566$ min, $t_{\text{minor}} = 24.641$ min.



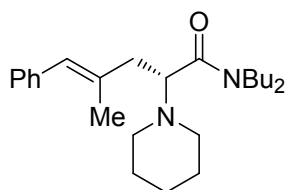
(E)-N,N-Dibutyl-5-(naphthalen-2-yl)-2-(piperidin-1-yl)pent-4-enamide (4r): Yellow oil (39.2 mg, 93%).

$[\alpha]_D^{25} = +61.4$ (*c* 0.11, acetone); $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 7.88–7.59 (m, 4H), 7.52 (d, $J = 8.8$ Hz, 1H), 7.39 (p, $J = 7.2$ Hz, 2H), 6.57 (d, $J = 15.6$ Hz, 1H), 6.36–6.22 (m, 1H), 3.62–3.29 (m, 3H), 3.23–3.05 (m, 2H), 2.91–2.74 (m, 1H), 2.78–2.59 (m, 2H), 2.60–2.27 (m, 3H), 1.60–1.16 (m, 14H), 0.97–0.75 (m, 6H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 170.6, 135.2, 133.6, 132.6, 131.7, 128.8, 127.9, 127.7, 127.5, 126.0, 125.4, 125.3, 123.5, 65.8, 50.3, 47.3, 45.6, 31.5, 29.7, 29.0, 26.5, 24.5, 20.2, 20.1, 13.9, 13.8; IR (KBr) cm^{-1} : 1558, 962, 688; $[\text{M}+\text{H}]^+$ calcd 421.3213, found 421.3216; Daicel Chiralpak AD-H column, *n*-hexane/*i*-PrOH = 99/1, 1 mL/min, 254 nm: $t_{\text{major}} = 19.661$ min, $t_{\text{minor}} = 15.967$ min.



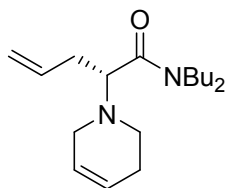
(E)-N,N-Dibutyl-5-(furan-2-yl)-2-(piperidin-1-yl)pent-4-enamide (4s): Yellow oil (32.9 mg, 91%).

$[\alpha]_D^{25} = +40.7$ (*c* 0.21, acetone); $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 7.25 (s, 1H), 6.32–6.25 (m, 1H), 6.22 (d, $J = 16.0$ Hz, 1H), 6.11–5.97 (m, 2H), 3.52–3.32 (m, 3H), 3.20–3.03 (m, 2H), 2.79–2.68 (m, 1H), 2.68–2.52 (m, 2H), 2.54–2.31 (m, 3H), 1.56–1.14 (m, 14H), 0.94–0.80 (m, 6H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 170.4, 153.1, 141.2, 127.2, 120.3, 110.9, 106.1, 65.5, 50.2, 47.4, 45.6, 31.5, 29.7, 28.7, 26.5, 24.4, 20.2, 20.1, 13.9, 13.8; IR (KBr) cm^{-1} : 2975, 2926, 1683, 1653, 1048, 880; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd 361.2850, found 361.2853; Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 95/5, 0.8 mL/min, 254 nm: $t_{\text{major}} = 43.549$ min, $t_{\text{minor}} = 30.185$ min.



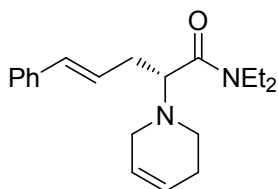
(E)-N,N-Dibutyl-4-methyl-5-phenyl-2-(piperidin-1-yl)pent-4-enamide (4t): Yellow oil (36.1 mg, 94%).

$[\alpha]_D^{25} = +45.9$ (*c* 0.60, acetone); $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 7.33–7.24 (m, 2H), 7.22–7.09 (m, 3H), 6.30 (s, 1H), 3.65–3.55 (m, 1H), 3.54–3.36 (m, 2H), 3.26–3.05 (m, 2H), 2.86–2.64 (m, 3H), 2.61–2.37 (m, 3H), 1.88 (s, 3H), 1.58–1.20 (m, 14H), 0.95–0.78 (m, 6H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 170.6, 138.5, 136.5, 128.7, 127.9, 127.1, 125.8, 63.9, 50.3, 47.5, 45.8, 36.1, 31.6, 29.8, 26.5, 24.5, 20.3, 20.2, 18.5, 13.9, 13.8; IR (KBr) cm^{-1} : 2973, 2925, 1677, 1250, 819; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd 385.3213, found 385.3213; Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 97/3, 0.8 mL/min, 254 nm: $t_{\text{major}} = 16.562$ min, $t_{\text{minor}} = 13.513$ min.

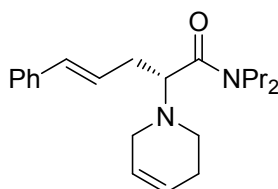


***N,N*-Dibutyl-2-(5,6-dihydropyridin-1(2*H*)-yl)pent-4-enamide (5a):** Yellow oil (22.2 mg, 76%).

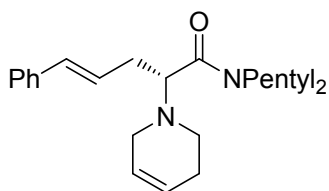
$[\alpha]_D^{20} = +25.0$ (c 0.26, acetone); $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 5.81–5.61 (m, 3H), 5.11–4.94 (m, 2H), 3.57–3.29 (m, 3H), 3.24–3.01 (m, 4H), 2.84–2.55 (m, 3H), 2.39–2.24 (m, 1H), 2.18–1.94 (m, 2H), 1.57–1.16 (m, 8H), 0.89 (q, $J = 6.8$ Hz, 6H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 170.4, 136.1, 125.6, 125.0, 116.5, 64.0, 48.7, 47.4, 45.7, 45.4, 31.5, 29.8, 29.7, 26.8, 20.2, 20.1, 13.9, 13.8; IR (KBr) cm^{-1} : 2978, 1698, 1558, 1046, 879; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd 293.2587, found 293.2605; Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 95/5, 0.8 mL/min, 210 nm: $t_{\text{major}} = 22.716$ min, $t_{\text{minor}} = 23.954$ min.



***(E)*-2-(5,6-Dihydropyridin-1(2*H*)-yl)-*N,N*-diethyl-5-phenylpent-4-enamide (5aa):** Yellow oil (28.4 mg, 91%). $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 7.35–7.13 (m, 5H), 6.44 (d, $J = 16.0$ Hz, 1H), 6.25–6.09 (m, 1H), 5.79–5.56 (m, 2H), 3.63–3.51 (m, 1H), 3.51–3.35 (m, 2H), 3.36–3.17 (m, 3H), 3.17–3.06 (m, 1H), 2.91–2.65 (m, 3H), 2.57–2.45 (m, 1H), 2.20–2.07 (m, 2H), 1.24–1.02 (m, 6H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 170.2, 137.5, 132.0, 128.4, 127.6, 126.9, 126.0, 125.5, 125.1, 64.4, 48.7, 45.8, 41.9, 40.4, 29.7, 26.8, 14.6, 13.0; IR (KBr) cm^{-1} : 2918, 2848, 1636, 694; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd 313.2274, found 313.2273.

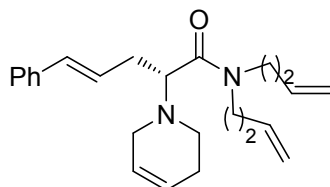


***(E)*-2-(5,6-Dihydropyridin-1(2*H*)-yl)-5-phenyl-*N,N*-dipropylpent-4-enamide (5ab):** Yellow oil (31.6 mg, 93%). $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 7.32–7.17 (m, 5H), 6.43 (d, $J = 15.6$ Hz, 1H), 6.26–6.08 (m, 1H), 5.78–5.56 (m, 2H), 3.64–3.53 (m, 1H), 3.46–3.34 (m, 2H), 3.31–3.22 (m, 1H), 3.21–3.06 (m, 3H), 2.91–2.66 (m, 3H), 2.56–2.44 (m, 1H), 2.16–2.07 (m, 2H), 1.61–1.45 (m, 4H), 0.89–0.81 (m, 6H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 170.6, 137.6, 131.9, 128.4, 127.8, 126.9, 126.0, 125.6, 125.1, 64.4, 49.5, 48.8, 47.7, 45.6, 30.2, 29.4, 26.8, 22.6, 20.9, 11.4; IR (KBr) cm^{-1} : 3236, 2853, 1636, 1384; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd 341.2587, found 341.2589.

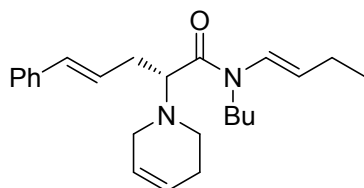


***(E)*-2-(5,6-Dihydropyridin-1(2*H*)-yl)-*N,N*-dipentyl-5-phenylpent-4-enamide (5ad):** Yellow oil

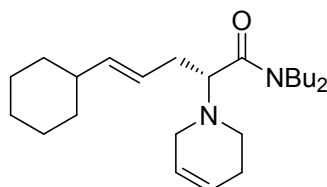
(35.6 mg, 90%). ¹H NMR (400 MHz, CDCl₃): δ 7.38–7.11 (m, 5H), 6.43 (d, *J* = 16.4 Hz, 1H), 6.23–6.09 (m, 1H), 5.77–5.55 (m, 2H), 3.62–3.51 (m, 1H), 3.49–3.34 (m, 2H), 3.35–3.07 (m, 4H), 2.91–2.62 (m, 3H), 2.54–2.43 (m, 1H), 2.19–2.03 (m, 2H), 1.45–1.16 (m, 12H), 0.92–0.76 (m, 6H); ¹³C NMR (100 MHz, CDCl₃): δ 170.4, 137.6, 132.0, 128.4, 127.8, 126.9, 126.0, 125.6, 125.1, 64.4, 48.8, 47.8, 46.0, 45.6, 29.4, 29.2, 29.18, 29.15, 27.4, 26.8, 22.4, 22.4, 14.0, 13.9; IR (KBr) cm⁻¹: 2929, 1631, 1507, 1456, 1265, 965, 879, 784; HRMS (APCI) [M+H]⁺ calcd 397.3213, found 397.3211.



(*E*)-*N,N*-Di(but-3-en-1-yl)-2-(5,6-dihydropyridin-1(2*H*)-yl)-5-phenylpent-4-enamide (5ad): Yellow oil (33.4 mg, 92%). ¹H NMR (400 MHz, CDCl₃): δ 7.34–7.14 (m, 5H), 6.44 (d, *J* = 15.6 Hz, 1H), 6.24–6.09 (m, 1H), 5.85–5.59 (m, 4H), 5.16–4.84 (m, 4H), 3.65–3.43 (m, 3H), 3.36–3.17 (m, 3H), 3.16–3.05 (m, 1H), 2.92–2.76 (m, 2H), 2.75–2.62 (m, 1H), 2.53–2.41 (m, 1H), 2.38–2.23 (m, 4H), 2.17–2.06 (m, 2H); ¹³C NMR (100 MHz, CDCl₃): δ 170.6, 137.6, 135.6, 134.7, 132.0, 128.4, 127.8, 126.9, 126.0, 125.5, 125.1, 117.1, 116.4, 64.8, 48.6, 47.4, 45.7, 45.6, 33.6, 32.2, 28.7, 26.8; IR (KBr) cm⁻¹: 2920, 2852, 1637, 1490, 1275, 748; HRMS (APCI) [M+H]⁺ calcd 365.2587, found 365.2586.

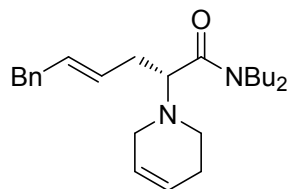


(*E*)-*N*-(But-1-en-1-yl)-*N*-butyl-2-(5,6-dihydropyridin-1(2*H*)-yl)-5-phenylpent-4-enamide (5af): Yellow oil (33.6 mg, 92%). ¹H NMR (400 MHz, CDCl₃, mixture of two rotamers): δ 7.42–7.12 (m, 5.34H), 6.71 (d, *J* = 14.4 Hz, 0.66H), 6.43 (d, *J* = 15.6 Hz, 1H), 6.22–6.08 (m, 1H), 5.78–5.60 (m, 2H), 5.23–5.12 (m, 0.66H), 5.12–4.99 (m, 0.34H), 3.78–3.53 (m, 3H), 3.24 (d, *J* = 16.0 Hz, 1H), 3.12 (d, *J* = 16.0 Hz, 1H), 2.97–2.55 (m, 4H), 2.16–2.04 (m, 4H), 1.35–1.22 (m, 4H), 1.06–0.96 (m, 3H), 0.91–0.85 (m, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 169.7, 137.7, 132.3, 128.6, 127.5, 127.3, 127.2, 126.2, 125.7, 125.3, 117.0, 65.3, 48.8, 46.1, 44.1, 30.1, 29.4, 27.0, 23.8, 20.4, 14.4, 14.1; IR (KBr) cm⁻¹: 3290, 1644, 1265, 740; HRMS (APCI) [M+H]⁺ calcd 367.2744, found 367.2740.



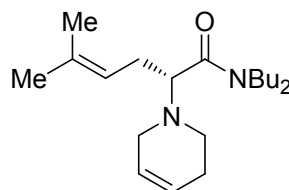
(*E*)-*N,N*-Dibutyl-5-cyclohexyl-2-(5,6-dihydropyridin-1(2*H*)-yl)pent-4-enamide (5b): Yellow oil (32.6 mg, 87%). [α]_D²⁰ = +31.3 (*c* 0.60, acetone); ¹H NMR (400 MHz, CDCl₃): δ 5.74–5.57 (m, 2H), 5.46–5.36 (m, 1H), 5.34–5.21 (m, 1H), 3.52–3.30 (m, 3H), 3.29–2.98 (m, 4H), 2.81–2.53 (m, 3H), 2.30–2.18 (m, 1H), 2.14–2.01 (m, 2H), 1.93–1.79 (m, 1H), 1.68–1.03 (m, 18H), 0.96–

0.85 (m, 6H); ^{13}C NMR (100 MHz, CDCl_3): δ 170.8, 138.9, 125.7, 125.1, 124.4, 64.4, 49.0, 47.6, 45.9, 45.5, 40.7, 33.02, 33.0, 31.7, 29.9, 29.4, 26.8, 26.2, 26.0, 20.2, 13.9, 13.8; IR (KBr) cm^{-1} : 2919, 2849, 1634, 1558, 1506, 914; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd 375.3370, found 375.3379; Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 98/2, 0.8 mL/min, 210 nm: $t_{\text{major}} = 29.191$ min, $t_{\text{minor}} = 33.756$ min.

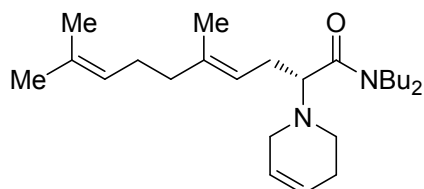


(E)-N,N-Dibutyl-2-(5,6-dihydropyridin-1(2H)-yl)-6-phenylhex-4-enamide (5c): Yellow oil

(28.4 mg, 74%). $[\alpha]_D^{20} = +10.8$ (*c* 0.80, acetone); ^1H NMR (400 MHz, CDCl_3): δ 7.25 (t, $J = 8.0$ Hz, 2H), 7.18 (d, $J = 7.2$ Hz, 1H), 7.16–7.11 (m, 2H), 5.77–5.58 (m, 3H), 5.51–5.39 (m, 1H), 3.52–3.34 (m, 3H), 3.30 (d, $J = 6.8$ Hz, 2H), 3.27–3.04 (m, 4H), 2.83–2.72 (m, 1H), 2.71–2.60 (m, 2H), 2.38–2.25 (m, 1H), 2.14–2.04 (m, 2H), 1.54–1.38 (m, 4H), 1.35–1.27 (m, 4H), 0.96–0.88 (m, 6H); ^{13}C NMR (100 MHz, CDCl_3): δ 170.8, 141.0, 131.6, 129.0, 128.7, 128.5, 126.1, 125.9, 125.3, 64.5, 49.1, 47.8, 46.1, 45.8, 39.3, 31.9, 30.1, 29.1, 27.1, 20.5, 14.2, 14.1; IR (KBr) cm^{-1} : 2956, 2920, 1646, 1558, 1265, 969; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd 383.3057, found 383.3059; Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 98/2, 0.8 mL/min, 230 nm: $t_{\text{major}} = 31.521$ min, $t_{\text{minor}} = 28.512$ min.



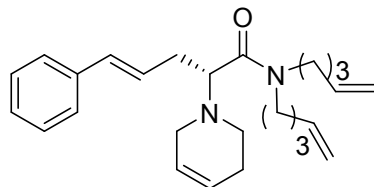
(E)-N,N-Dibutyl-2-(5,6-dihydropyridin-1(2H)-yl)-5-methylhex-4-enamide (5d): Yellow oil (16.8 mg, 52%). ^1H NMR (400 MHz, CDCl_3): δ 5.72–5.55 (m, 2H), 5.07–4.98 (m, 1H), 3.49–3.32 (m, 3H), 3.26–2.98 (m, 4H), 2.80–2.61 (m, 2H), 2.59–2.46 (m, 1H), 2.34–2.23 (m, 1H), 2.12–1.98 (m, 2H), 1.62 (s, 3H), 1.59 (s, 3H), 1.52–1.39 (m, 4H), 1.32–1.23 (m, 4H), 0.92–0.83 (m, 6H); ^{13}C NMR (100 MHz, CDCl_3): δ 170.9, 133.1, 125.7, 125.1, 121.3, 63.8, 48.9, 47.5, 45.8, 45.5, 31.5, 29.8, 26.8, 25.7, 24.8, 20.2, 20.1, 17.7, 13.9, 13.8; IR (KBr) cm^{-1} : 2919, 2849, 1634, 1507, 895, 739; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd 321.2900, found 321.2906.



(E)-N,N-Dibutyl-2-(5,6-dihydropyridin-1(2H)-yl)-5,9-dimethyldeca-4,8-dienamide (5e):

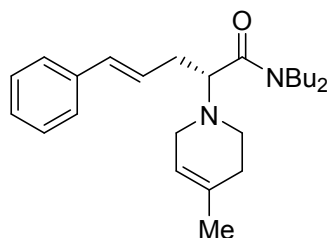
Yellow oil (21.2 mg, 54%). $[\alpha]_D^{20} = +10.7$ (*c* 0.50, acetone); ^1H NMR (400 MHz, CDCl_3): δ 5.74–5.59 (m, 2H), 5.13–5.01 (m, 2H), 3.52–3.33 (m, 3H), 3.31–3.01 (m, 4H), 2.84–2.74 (m, 1H), 2.73–2.63 (m, 1H), 2.62–2.51 (m, 1H), 2.39–2.29 (m, 1H), 2.15–2.06 (m, 2H), 2.06–1.98 (m, 2H), 1.98–1.90 (m, 2H), 1.65 (s, 3H), 1.62 (s, 3H), 1.57 (s, 3H), 1.55–1.39 (m, 4H), 1.34–1.25 (m, 4H),

0.98–0.78 (m, 6H); ^{13}C NMR (100 MHz, CDCl_3): δ 170.9, 136.9, 131.3, 125.7, 125.1, 124.3, 121.1, 63.8, 48.9, 47.6, 45.9, 45.6, 39.8, 31.6, 29.9, 26.8, 26.7, 25.7, 24.7, 20.3, 20.2, 17.6, 16.1, 13.9, 13.8; IR (KBr) cm^{-1} : 2958, 2927, 1646, 1558, 1265, 969; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd 389.3526, found 389.3531; Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 97/3, 0.8 mL/min, 210 nm: $t_{\text{major}} = 13.680$ min, $t_{\text{minor}} = 11.816$ min.



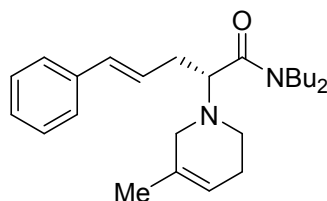
(E)-2-(5,6-Dihydropyridin-1(2H)-yl)-N,N-di(pent-4-en-1-yl)-5-phenylpent-4-enamide (5f):

Yellow oil (36.3 mg, 92%). $[\alpha]_D^{25} = +25.3$ (*c* 0.15, acetone); ^1H NMR (400 MHz, CDCl_3): δ 7.35–7.08 (m, 5H), 6.42 (d, $J = 15.6$ Hz, 1H), 6.25–6.07 (m, 1H), 5.96–5.55 (m, 4H), 5.16–4.84 (m, 4H), 3.62–3.50 (m, 1H), 3.50–3.32 (m, 2H), 3.30–2.99 (m, 4H), 2.91–2.74 (m, 2H), 2.75–2.63 (m, 1H), 2.54–2.38 (m, 1H), 2.22–1.79 (m, 6H), 1.74–1.44 (m, 4H); ^{13}C NMR (100 MHz, CDCl_3): δ 170.5, 138.0, 137.5, 137.4, 132.0, 128.4, 127.7, 126.9, 125.9, 125.5, 125.1, 115.4, 114.8, 64.5, 48.7, 47.2, 45.6, 45.5, 31.1, 31.0, 28.9, 28.3, 26.9, 26.8; IR (KBr) cm^{-1} : 2977, 2927, 1717, 1507, 880, 668; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd 393.2900, found 393.2924; Daicel Chiralpak AD-H column, *n*-hexane/*i*-PrOH = 98/2, 1 mL/min, 254 nm: $t_{\text{major}} = 15.029$ min, $t_{\text{minor}} = 11.897$ min.



(E)-N,N-Dibutyl-2-(4-methyl-5,6-dihydropyridin-1(2H)-yl)-5-phenylpent-4-enamide (5g):

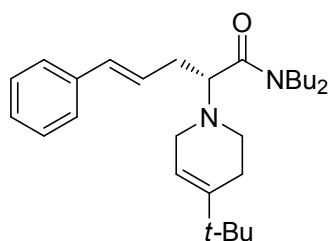
Yellow oil (34.5 mg, 90%). $[\alpha]_D^{25} = +29.3$ (*c* 0.22, acetone); ^1H NMR (400 MHz, CDCl_3): δ 7.31–7.21 (m, 4H), 7.20–7.12 (m, 1H), 6.42 (d, $J = 15.6$ Hz, 1H), 6.20–6.08 (m, 1H), 5.37–5.28 (m, 1H), 3.69–3.48 (m, 1H), 3.52–3.34 (m, 2H), 3.33–2.93 (m, 4H), 2.91–2.61 (m, 3H), 2.57–2.40 (m, 1H), 2.13–1.89 (m, 2H), 1.65 (s, 3H), 1.54–1.19 (m, 8H), 0.88 (t, $J = 7.2$ Hz, 3H), 0.83 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ 170.7, 137.8, 132.8, 132.1, 128.6, 128.0, 127.1, 126.2, 119.6, 67.3, 64.3, 49.1, 47.8, 46.1, 45.8, 31.8, 31.7, 30.1, 29.9, 23.2, 20.4, 14.1, 14.0; IR (KBr) cm^{-1} : 2977, 2927, 1684, 1653, 879, 668; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd 383.3057, found 383.3068; Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 95/5, 0.8 mL/min, 254 nm: $t_{\text{major}} = 26.938$ min, $t_{\text{minor}} = 30.912$ min.



(E)-N,N-Dibutyl-2-(3-methyl-5,6-dihydropyridin-1(2H)-yl)-5-phenylpent-4-enamide (5h):

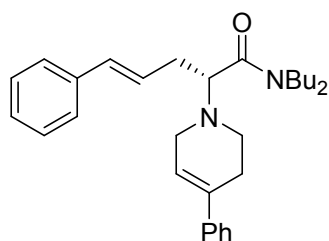
Yellow oil (32.5 mg, 85%). $[\alpha]_D^{25} = +64.9$ (*c* 0.30, acetone); ^1H NMR (400 MHz, CDCl_3): δ

7.36–7.28 (m, 4H), 7.24–7.16 (m, 1H), 6.47 (d, $J = 15.6$ Hz, 1H), 6.27–6.14 (m, 1H), 5.50–5.38 (m, 1H), 3.62 (dd, $J = 9.6, 4.0$ Hz, 1H), 3.51–3.39 (m, 2H), 3.25–3.12 (m, 3H), 3.05–2.96 (m, 1H), 2.93–2.84 (m, 1H), 2.83–2.75 (m, 1H), 2.73–2.64 (m, 1H), 2.58–2.48 (m, 1H), 2.14–2.05 (m, 2H), 1.66 (s, 3H), 1.49–1.31 (m, 8H), 0.93–0.87 (m, 6H); ^{13}C NMR (100 MHz, CDCl_3): δ 170.5, 137.6, 132.5, 132.0, 128.4, 127.7, 126.9, 126.0, 119.4, 64.3, 52.8, 47.6, 45.9, 45.7, 31.6, 29.9, 29.7, 26.5, 22.7, 21.0, 20.3, 13.9, 13.8; IR (KBr) cm^{-1} : 2978, 1684, 1653, 1338, 879, 668; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd 383.3057, found 383.3074; Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 95/5, 0.8 mL/min, 254 nm: $t_{\text{major}} = 35.416$ min, $t_{\text{minor}} = 30.117$ min.



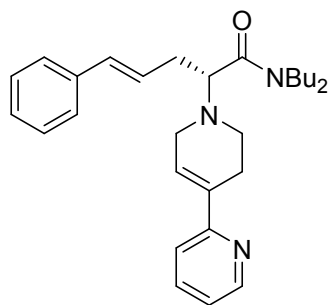
(*E*)-*N,N*-Dibutyl-2-(4-(tert-butyl)-5,6-dihydropyridin-1(2*H*)-yl)-5-phenylpent-4-enamide (5i):

Yellow oil (41.2 mg, 97%). $[\alpha]_D^{25} = +54.8$ (c 0.25, acetone); ^1H NMR (400 MHz, CDCl_3): δ 7.27–7.20 (m, 4H), 7.19–7.09 (m, 1H), 6.41 (d, $J = 15.6$ Hz, 1H), 6.21–6.08 (m, 1H), 5.42–5.28 (m, 1H), 3.62–3.52 (m, 1H), 3.38–3.08 (m, 6H), 2.91–2.65 (m, 3H), 2.57–2.39 (m, 1H), 2.19–2.00 (m, 2H), 1.50–1.25 (m, 8H), 0.99 (s, 9H), 0.89–0.82 (m, 6H); ^{13}C NMR (100 MHz, CDCl_3): δ 170.6, 144.1, 137.5, 131.9, 128.3, 127.7, 126.8, 125.9, 115.8, 64.0, 49.2, 47.6, 46.0, 45.8, 34.8, 31.5, 29.9, 29.8, 28.7, 26.0, 20.3, 20.2, 13.9, 13.8; IR (KBr) cm^{-1} : 2978, 2926, 1680, 1653, 879, 669; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd 425.3526; found 425.3529; Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 95/5, 0.8 mL/min, 254 nm: $t_{\text{major}} = 33.248$ min, $t_{\text{minor}} = 29.290$ min.



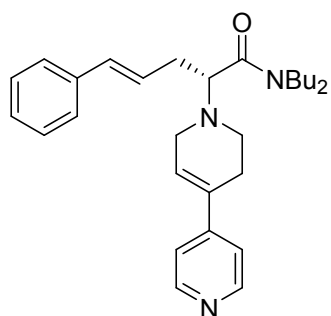
(*E*)-*N,N*-Dibutyl-5-phenyl-2-(4-phenyl-5,6-dihydropyridin-1(2*H*)-yl)pent-4-enamide (5j):

Yellow oil (42.3 mg, 95%). $[\alpha]_D^{25} = +46.6$ (c 0.28, acetone); ^1H NMR (400 MHz, CDCl_3): δ 7.52–7.06 (m, 10H), 6.45 (d, $J = 15.8$ Hz, 1H), 6.24–6.12 (m, 1H), 6.10–5.96 (m, 1H), 3.70–3.60 (m, 1H), 3.53–3.07 (m, 6H), 3.06–2.74 (m, 3H), 2.65–2.37 (m, 3H), 1.54–1.21 (m, 8H), 0.89 (t, $J = 7.6$ Hz, 3H), 0.84 (t, $J = 7.6$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ 170.4, 140.8, 137.5, 134.8, 132.0, 128.4, 128.2, 127.5, 126.9, 126.8, 125.9, 124.7, 122.0, 64.0, 49.2, 47.6, 45.8, 45.8, 31.6, 29.8, 29.7, 29.6, 28.7, 20.2, 13.9, 13.8; IR (KBr) cm^{-1} : 2978, 2928, 1640, 1684, 1670, 689; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd 445.3213, found 445.3208; Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 95/5, 0.8 mL/min, 254 nm: $t_{\text{major}} = 36.138$ min, $t_{\text{minor}} = 32.405$ min.



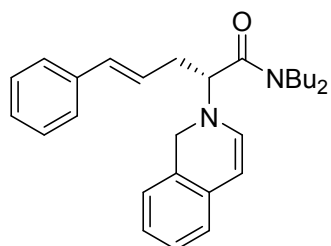
(E)-N,N-Dibutyl-2-(5',6'-dihydro-[2,4'-bipyridin]-1'(2'H)-yl)-5-phenylpent-4-enamide (5k):

Yellow oil (41.1 mg, 92%). $[\alpha]_D^{25} = +41.4$ (c 0.16, acetone); $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 8.58–8.49 (m, 1H), 7.60 (t, $J = 8.0$ Hz, 1H), 7.35–7.07 (m, 7H), 6.67–6.57 (m, 1H), 6.44 (d, $J = 15.6$ Hz, 1H), 6.24–6.10 (m, 1H), 3.70–3.59 (m, 1H), 3.58–3.31 (m, 4H), 3.22–3.06 (m, 2H), 3.04–2.79 (m, 3H), 2.68–2.43 (m, 3H), 1.54–1.26 (m, 8H), 0.88 (t, $J = 7.2$ Hz, 3H), 0.82 (t, $J = 7.2$ Hz, 3H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 170.2, 157.4, 148.8, 137.5, 136.2, 134.7, 132.0, 128.3, 127.6, 126.9, 125.9, 125.7, 121.6, 118.8, 64.0, 49.0, 47.5, 45.9, 45.8, 31.6, 30.1, 29.6, 29.2, 27.3, 20.2, 13.9, 13.8; IR (KBr) cm^{-1} : 2978, 2924, 1684, 1670, 1046, 879; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd 446.3166, found 446.3179; Daicel Chiralpak AD-H column, n -hexane/ i -PrOH = 98.5/1.5, 1 mL/min, 254 nm: $t_{\text{major}} = 42.850$ min, $t_{\text{minor}} = 50.916$ min.



(E)-N,N-Dibutyl-2-(5,6-dihydro-[4,4'-bipyridin]-1(2H)-yl)-5-phenylpent-4-enamide (5l):

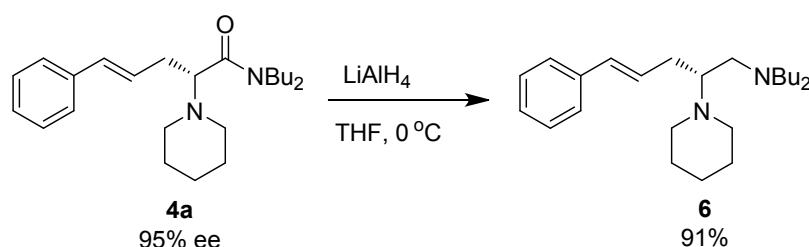
Yellow oil (42.8 mg, 96%). $[\alpha]_D^{25} = +31.1$ (c 0.18, acetone); $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 8.55 (d, $J = 5.6$ Hz, 2H), 7.35–7.15 (m, 7H), 6.49 (d, $J = 15.6$ Hz, 1H), 6.40–6.30 (m, 1H), 6.27–6.13 (m, 1H), 3.74–3.64 (m, 1H), 3.58–3.36 (m, 4H), 3.27–3.15 (m, 2H), 3.07–2.84 (m, 3H), 2.63–2.45 (m, 3H), 1.50–1.25 (m, 8H), 0.93 (t, $J = 7.2$ Hz, 3H), 0.88 (t, $J = 7.2$ Hz, 3H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 170.3, 149.9, 147.7, 137.4, 132.8, 132.2, 128.4, 127.2, 127.0, 126.0, 125.9, 119.3, 63.7, 49.1, 47.6, 45.9, 45.6, 31.6, 29.9, 29.7, 29.7, 27.8, 20.2, 13.9, 13.8; IR (KBr) cm^{-1} : 2978, 1684, 1653, 1084, 879, 668; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd 446.3166, found 446.3156; Daicel Chiralpak AD-H column, n -hexane/ i -PrOH = 95/5, 1 mL/min, 254 nm: $t_{\text{major}} = 30.526$ min, $t_{\text{minor}} = 38.624$ min.



(E)-N,N-Dibutyl-2-(isoquinolin-2(1H)-yl)-5-phenylpent-4-enamide (5m): Yellow oil (36.9 mg, 20

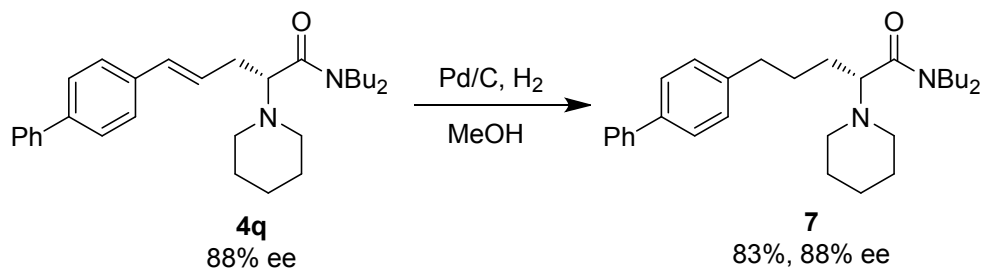
88%). $[\alpha]_D^{25} = +44.6$ (c 0.26, acetone); $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 8.39 (d, $J = 8.0$ Hz, 1H), 7.60 (t, $J = 7.2$ Hz, 1H), 7.52–7.33 (m, 3H), 7.22–6.95 (m, 4H), 6.52 (d, $J = 7.6$ Hz, 1H), 6.45 (d, $J = 15.6$ Hz, 1H), 6.20–6.08 (m, 2H), 3.56–3.28 (m, 3H), 3.29–3.05 (m, 3H), 3.07–2.97 (m, 1H), 2.96–2.81 (m, 1H), 2.76–2.63 (m, 1H), 1.53–1.24 (m, 8H), 0.86 (t, $J = 6.8$ Hz, 3H), 0.72 (t, $J = 6.8$ Hz, 3H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 168.7, 161.3, 137.0, 136.6, 133.7, 132.4, 128.3, 128.1, 128.0, 127.2, 126.8, 126.1, 125.9, 124.6, 106.7, 52.3, 47.5, 46.2, 36.1, 31.4, 31.2, 29.5, 20.1, 19.8, 13.7, 13.6; IR (KBr) cm^{-1} : 2978, 2928, 1698, 1684, 1085, 879; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd 417.2900, found 417.2897; Daicel Chiralpak IC-3 column, n -hexane/ i -PrOH = 95/5, 0.8 mL/min, 254 nm: $t_{\text{major}} = 17.766$ min, $t_{\text{minor}} = 26.716$ min.

5. Derivatization of 4a, 4q, 5f, 5j and 10



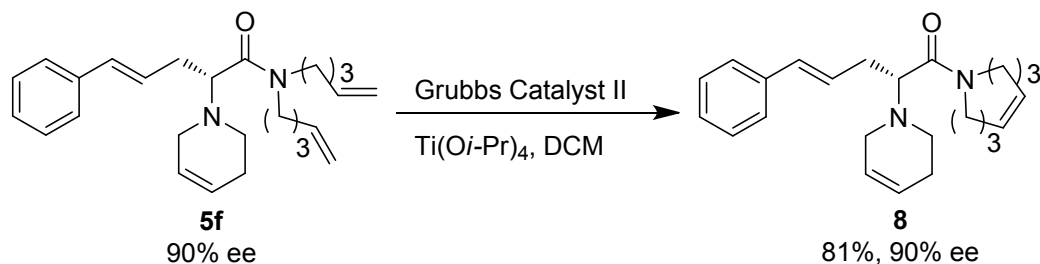
To a solution of **4a** (37.1 mg, 0.1 mmol) in anhydrous THF (1.0 mL) under N_2 was added LiAlH_4 (16.0 mg, 0.4 mmol) at 0 °C and the mixture was stirred at RT overnight. The reaction mixture was quenched with ethanol, diluted with DCM, washed with brine, dried over Na_2SO_4 and concentrated in vacuo. The residue was purified by flash column chromatography to give the yellow oil **6** in 91% yield. The two enantiomers are difficult to be separated on several kinds of 25

chiral column in HPLC analysis, the reported similar reactions showed no loss in ee.^[4] $[\alpha]_D^{25} = +45.9$ (c 0.3, acetone); $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 7.34–7.17 (m, 5H), 6.41–6.24 (m, 2H), 2.78–2.09 (m, 13H), 1.56–1.24 (m, 14H), 0.87 (t, $J = 7.2$ Hz, 6H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 138.0, 133.1, 130.3, 128.4, 126.6, 125.8, 63.6, 54.4, 50.1, 32.9, 29.6, 29.1, 26.6, 25.0, 20.7, 14.1; IR (KBr) cm^{-1} : 2978, 2928, 1748, 1489 1045, 879, 668; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd 357.3264, found 357.3275.



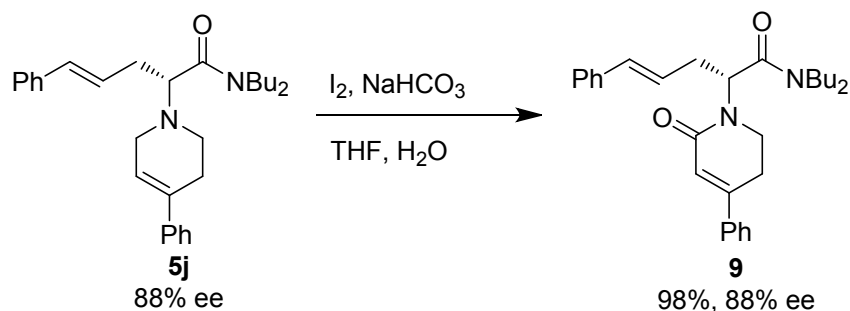
To a solution of **4q** (44.7 mg, 0.1 mmol) in MeOH (2.0 mL) was added Pd/C (10%, 10 mg) and the mixture was stirred under H_2 atmosphere (20 bar) at room temperature for 20 h. The reaction mixture was diluted with DCM, filtered, dried over anhydrous Na_2SO_4 and concentrated in vacuo. The residue was purified by flash column chromatography to give **7** as a yellow oil (83%, 88% ee).

$[\alpha]_D^{25} = +8.3$ (*c* 0.50, acetone); $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 7.57 (d, $J = 8.0$ Hz, 2H), 7.50 (d, $J = 8.0$ Hz, 2H), 7.42 (t, $J = 7.6$ Hz, 2H), 7.34–7.29 (m, 1H), 7.25–7.20 (m, 2H), 3.56–3.39 (m, 2H), 3.38–3.25 (m, 1H), 3.21–3.04 (m, 2H), 2.80–2.52 (m, 4H), 2.55–2.36 (m, 2H), 1.64–1.25 (m, 18H), 0.98–0.83 (m, 6H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 171.0, 141.6, 141.1, 138.6, 128.8, 128.7, 127.0, 126.9, 126.8, 65.0, 50.3, 47.3, 45.6, 35.8, 31.6, 29.8, 28.9, 26.5, 25.0, 24.5, 20.3, 20.2, 13.9; IR (KBr) cm^{-1} : 2920, 2850, 1635, 1558, 1471, 1373, 1265, 667; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd 449.3526, found 449.3527; Daicel Chiralpak IC-3 column, *n*-hexane /*i*-PrOH = 95/5, 0.8 mL/min, 254 nm: $t_{\text{major}} = 14.175$ min, $t_{\text{minor}} = 12.130$ min.



To a solution of **5f** (39.5 mg, 0.1 mmol) in anhydrous DCM (20 mL) under N_2 was added fresh distilled $\text{Ti}(\text{O}i\text{-Pr})_4$ (28.4 mg, 0.1 mmol) and stirred for 10 min, then Grubbs Catalyst II (8.5 mg, 0.01 mmol) was added. The resulting mixture was stirred under reflux for 10 h. The reaction mixture was concentrated in vacuo. The residue was purified by flash column chromatography to **8**

give the compound **8** (81%, 90% *ee* and *E/Z* = 2:1).^[5] $[\alpha]_D^{25} = +10.0$ (*c* 0.15, acetone); $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 7.34–7.11 (m, 5H), 6.46 (d, $J = 16.0$ Hz, 1H), 6.26–6.07 (m, 1H), 5.81–5.62 (m, 4H), 4.10–4.02 (m, 2H), 3.55–3.30 (m, 7H), 2.28–1.71 (m, 12H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3 , major isomer): δ 170.8, 137.6, 132.0, 131.6, 128.4, 128.2, 128.1, 126.9, 125.9, 125.6, 125.0, 65.4, 53.4, 50.6, 48.3, 47.6, 45.9, 28.3, 27.8, 26.8, 25.9, 23.6; $^{13}\text{C NMR}$ (101 MHz, CDCl_3 , minor isomer) δ 171.6, 137.5, 132.3, 130.0, 129.7, 127.2, 127.1, 127.0, 125.9, 125.6, 125.2, 63.7, 49.3, 48.7, 47.9, 47.3, 41.6, 30.2, 29.5, 26.9, 26.6, 23.2; IR (KBr) cm^{-1} : 2979, 1684, 1670, 1045, 879, 668; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd 365.2587, found 365.2591; Daicel Chiralpak AD-H column, *n*-hexane /*i*-PrOH = 98/2, 0.8 mL/min, 254 nm: $t_{\text{major}} = 24.955$ min, $t_{\text{minor}} = 18.207$ min.

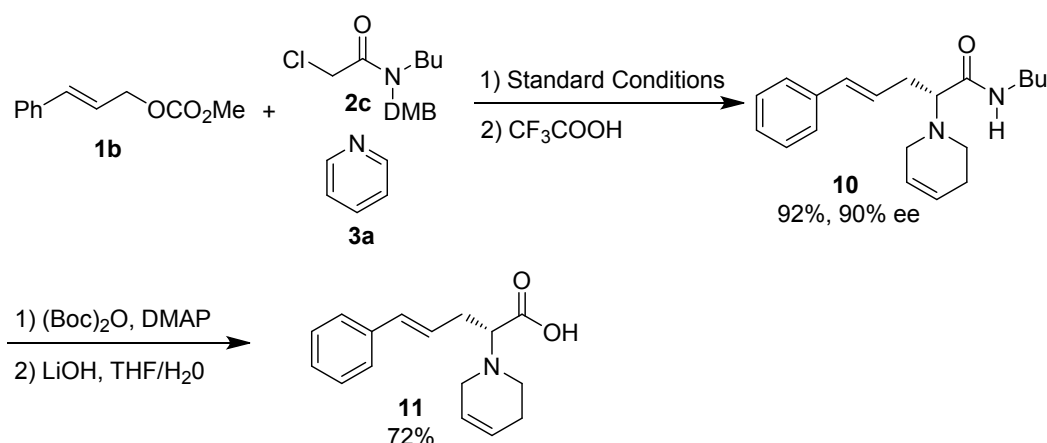


To a solution of **5j** (44.5 mg, 0.1 mmol) in THF (10.0 mL) and H_2O (4 mL) was added NaHCO_3 (84 mg, 1 mmol) and I_2 (190 mg, 0.75 mmol). The mixture was stirred at RT for 12 h. The reaction mixture was quenched with saturated Na_2SO_3 and saturated NaHCO_3 , extracted with DCM, dried over Na_2SO_4 and concentrated in vacuo. The residue was purified by flash column chromatography to **9**

chromatography to give the compound **9** (98%, 88% *ee*).^[6] $[\alpha]_D^{25} = +101.3$ (*c* 0.3, acetone); ^1H

NMR (400 MHz, CDCl₃) δ 7.57–7.09 (m, 10H), 6.51 (d, J = 15.6 Hz, 1H), 6.30 (s, 1H), 6.22–6.06 (m, 1H), 5.66 (t, J = 7.2 Hz, 1H), 3.75–3.65 (m, 1H), 3.61–3.41 (m, 3H), 3.25–3.08 (m, 2H), 2.87–2.58 (m, 4H), 1.61–1.43 (m, 4H), 1.39–1.21 (m, 4H), 1.02–0.75 (m, 6H); ¹³C NMR (101 MHz, CDCl₃) δ 169.8, 164.7, 150.3, 137.5, 137.4, 133.3, 129.8, 128.9, 128.7, 127.4, 126.4, 125.9, 119.7, 51.3, 47.5, 46.0, 40.6, 33.5, 31.7, 29.9, 27.0, 20.4, 20.2, 14.1, 14.1; IR (KBr) cm⁻¹: 2978, 2924, 1654, 1647 1374, 1045, 879, 668; HRMS (APCI) [M+H]⁺ calcd 459.3006, found 459.3023; Daicel Chiralpak AD-H column, *n*-hexane /*i*-PrOH = 95/5, 1 mL/min, 254 nm: t_{major} = 22.878 min, t_{minor} = 40.089 min.

6. Synthesis of Dipeptide and Unnatural Amino Acid

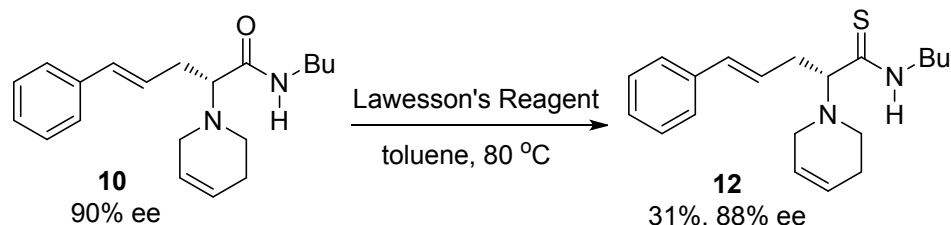


The reaction of **1b** (0.1 mmol), **2e** (0.2 mmol) and **3a** (0.5 mmol) was conducted using general procedure. The reaction mixture was filtrated through a short column of silica gel to remove DMSO and concentrated under vacuum. To the crude product was added TFA (2 mL) and the mixture was then stirred at 60 °C for 6 h. TFA was evaporate under vacuo and TEA (2 mL) was added. The mixture was stirred for further 0.5 h. The reaction mixture was diluted with DCM, the organic phase was washed with saturate NaHCO₃, dried over anhydrous Na₂SO₄ and concentrated in vacuo. The residue was purified by flash column chromatography (PE/EA = 5/1) to give **10**

(92%, 90% ee). Mp: 39–40 °C. [α] = -14.9 (c 0.20, MeOH); ¹H NMR (400 MHz, CDCl₃) δ 7.40–7.08 (m, 6H), 6.44 (d, J = 15.6 Hz, 1H), 6.29 (dd, J = 15.6, 8.4 Hz, 1H), 5.82–5.61 (m, 2H), 3.40–2.96 (m, 5H), 2.87–2.73 (m, 1H), 2.73–2.49 (m, 3H), 2.24–2.04 (m, 2H), 1.51–1.37 (m, 2H), 1.31–1.22 (m, 2H), 0.92–0.79 (m, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 172.5, 137.6, 132.1, 128.6, 127.6, 127.2, 126.2, 125.5, 125.4, 69.1, 49.8, 47.7, 38.9, 32.0, 31.8, 26.7, 20.3, 13.9; IR (KBr) cm⁻¹: 2974, 1843, 1715, 1646, 1615, 1435, 766, 720; HRMS (APCI) [M+H]⁺ calcd 313.2274, found 313.2291; Daicel Chiralpak OD-H column, *n*-hexane /*i*-PrOH = 95/5, 0.8 mL/min, 254 nm: t_{major} = 24.966 min, t_{minor} = 17.239 min.

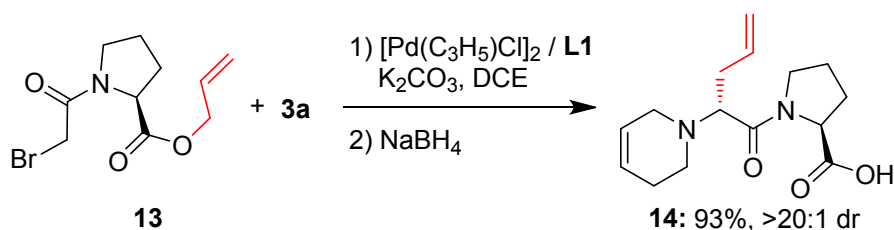
10 (0.13 mmol) and DMAP (0.26 mmol) were dissolved in (Boc)₂O (2 mL). The mixture was stirred at RT overnight before another portion of DMAP (0.13 mmol) and (Boc)₂O (2 mL) were added and the mixture was stirred at 80 °C for another 2 h. The reaction mixture was directly purified by flash column chromatography (PE/EA = 20/1) to give Boc-protected **10**, which was dissolved in THF (3 mL) followed by the addition of water (1 mL) and LiOH (0.78 mmol). The mixture was stirred at RT for 2 days. The reaction was quenched with acetic acid, concentrated in vacuo. The residue was purified by flash column chromatography (DCM/MeOH = 15/1) to give

the compound **11** (72%). mp: 46–48 °C. $[\alpha]_D^{25} = -4.0$ (*c* 0.2, MeOH); $^1\text{H NMR}$ (400 MHz, D_2O) δ 7.49–7.04 (m, 5H), 6.50 (d, $J = 15.6$ Hz, 1H), 6.15–6.00 (m, 1H), 5.91–5.76 (m, 1H), 5.65–5.48 (m, 1H), 3.79–3.49 (m, 3H), 3.42–3.10 (m, 2H), 2.88–2.62 (m, 2H), 2.46–2.15 (m, 2H); $^{13}\text{C NMR}$ (101 MHz, D_2O) δ 172.4, 136.7, 133.8, 128.8, 127.9, 126.3, 125.6, 123.0, 119.4, 68.7, 48.9, 48.7, 31.1, 22.3; IR (KBr) cm^{-1} : 3502, 1868, 1791, 1683, 1635, 1456, 457, 418; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd 258.1489; found 258.1501.



To a solution of **10** (31.2 mg, 0.1 mmol) in toluene (2.0 mL) was added Lawesson's reagent (80.8 mg, 0.2 mmol) under N_2 atmosphere and the mixture was stirred at 80 °C for 20 h. The reaction mixture was diluted with DCM, washed with saturated NaHCO_3 , dried over anhydrous Na_2SO_4 and concentrated in vacuo. The residue was purified by flash column chromatography to give **12**

as a yellow oil (31%, 88% ee). $[\alpha]_D^{25} = +22.9$ (*c* 0.20, acetone); $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 9.15 (s, 1H), 7.35–7.16 (m, 5H), 6.44 (d, $J = 15.6$ Hz, 1H), 6.21–6.04 (m, 1H), 5.87–5.72 (m, 1H), 5.73–5.61 (m, 1H), 3.73–3.48 (m, 3H), 3.43–3.19 (m, 1H), 3.15–2.93 (m, 2H), 2.89–2.53 (m, 3H), 2.27–2.02 (m, 2H), 1.63–1.46 (m, 2H), 1.40–1.26 (m, 2H), 0.85 (t, $J = 7.2$ Hz, 3H); $^{13}\text{C NMR}$ (100 MHz, CDCl_3): δ 202.0, 137.2, 132.8, 132.7, 128.5, 127.3, 126.1, 125.3, 125.1, 49.7, 49.6, 48.2, 44.8, 34.9, 30.1, 26.3, 20.3, 13.7; IR (KBr) cm^{-1} : 2920, 2850, 1683, 1635 1521, 1265, 895; HRMS (APCI) $[\text{M}+\text{H}]^+$ calcd 329.2046, found 329.2045; Daicel Chiralpak IC-3 column, *n*-hexane / *i*-PrOH = 95/5, 1 mL/min, 254 nm: $t_{\text{major}} = 11.591$ min, $t_{\text{minor}} = 13.361$ min.



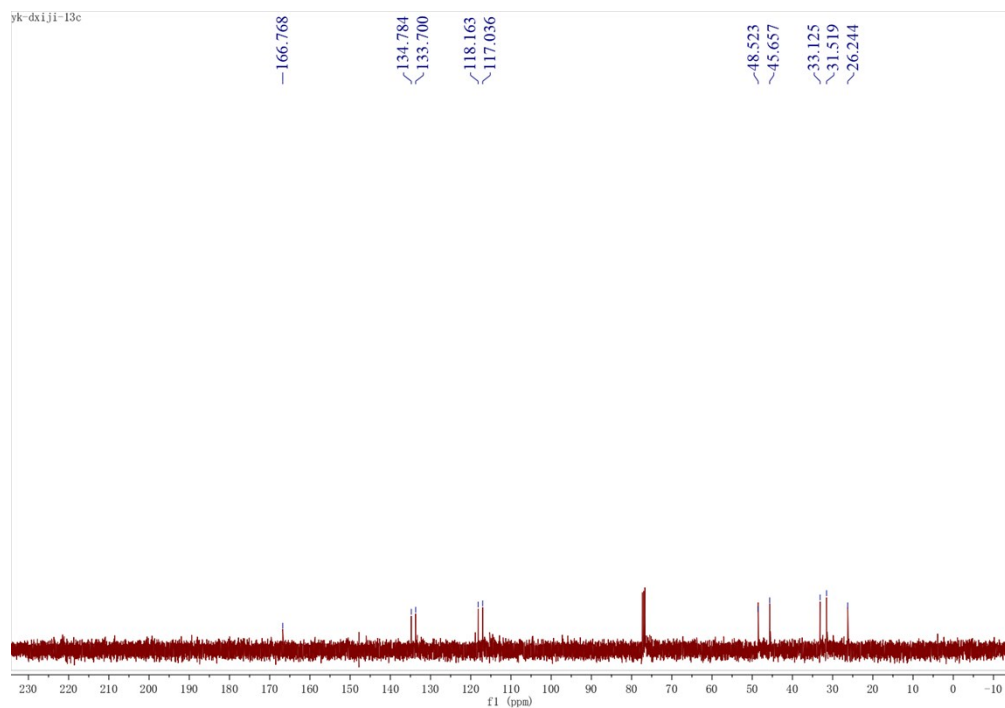
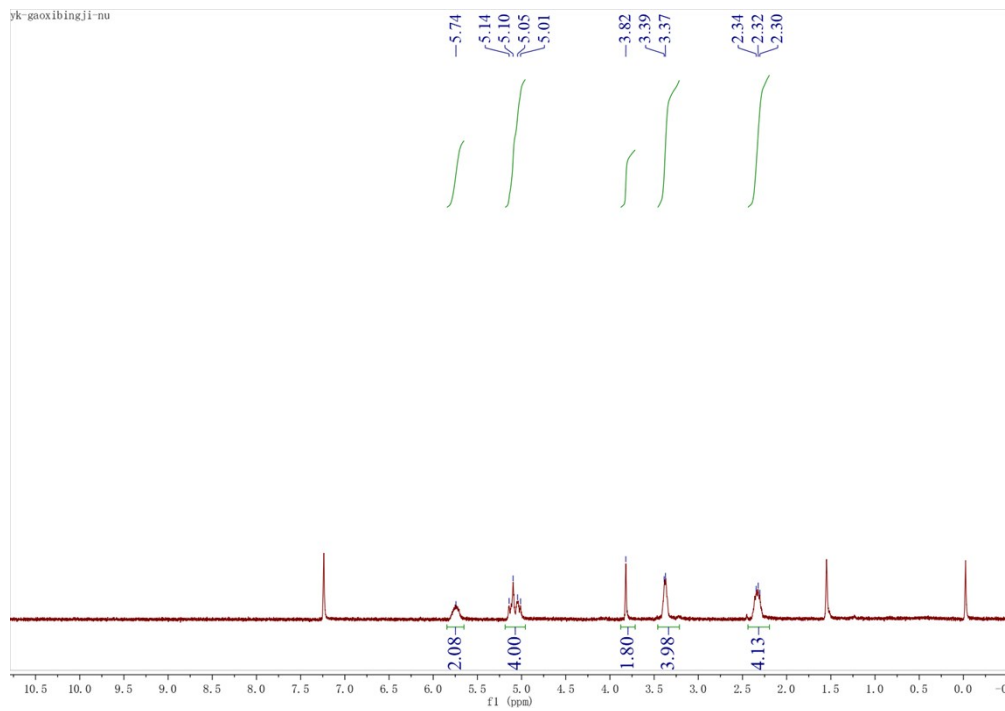
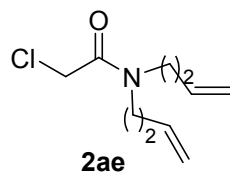
To an oven-dried glassware were added $[\text{Pd}(\text{C}_3\text{H}_5\text{Cl})_2]$ (0.9 mg, 2.5 mol%), **L1** (4.8 mg, 5.5 mol%) and anhydrous DCE (1.0 mL). To another dry glassware were added **13** (0.1 mmol), **3a** (0.2 mmol), and anhydrous DCE (0.5 mL). Then the first glassware was stirred at RT and the second glassware was reflux under a nitrogen atmosphere for 1 h. After cooling to RT, anhydrous K_2CO_3 (0.1 mmol) was added to the second glassware. The solution in the first glassware was added dropwise to another one. The reaction was stirred at RT for 12 h. The solvent was removed in vacuo. THF (1 mL) was added and the resulting mixture was cooled to -20 °C. Then methanol (1 mL) was added and the mixture was stirred for 5 min. NaBH_4 (0.5 mmol) was added in one portion and the reaction was stirred for 2 h. The reaction was quenched with acetic acid, and then was concentrated in vacuo. The residue was purified by flash column chromatography to give the compound **14** (93%, >20:1 dr, determined by the $^1\text{H-NMR}$ of the crude reaction mixture). $[\alpha]_D^{25}$

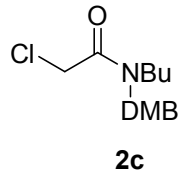
= -18.0 (c 0.10, acetone); ¹H NMR (400 MHz, CDCl₃) δ 6.01–5.50 (m, 3H), 5.06 (d, *J* = 16.4 Hz, 1H), 4.98 (d, *J* = 9.6 Hz, 1H), 4.33–4.18 (m, 1H), 3.90–2.79 (m, 7H), 2.81–2.27 (m, 4H), 2.08–1.80 (m, 4H); ¹³C NMR (101 MHz, CDCl₃) δ 179.1, 170.8, 135.8, 125.5, 125.1, 116.9, 65.6, 62.3, 49.0, 47.5, 46.0, 31.8, 29.3, 26.5, 25.1; IR (KBr) cm⁻¹: 2977, 2923, 1748, 1716 1271, 1045, 879; HRMS (APCI) [M+H]⁺ calcd 279.1703, found 279.1716.

7. References

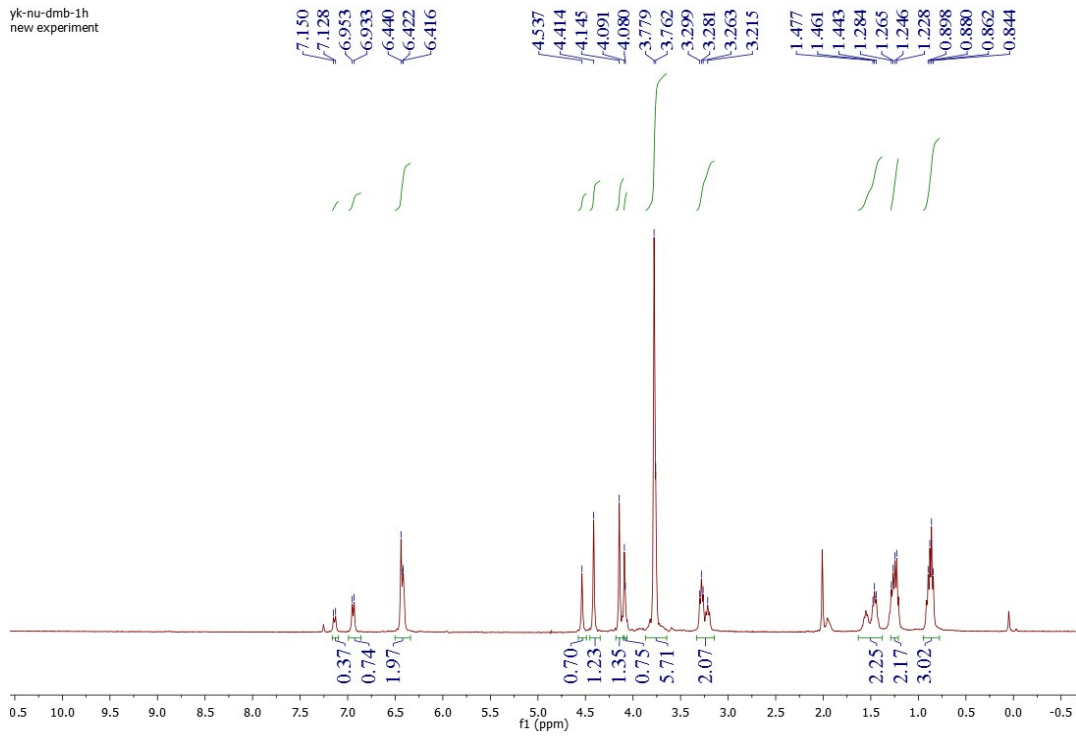
- [1] X. Huo, J. Zhang, J. Fu, R. He and W. Zhang, *J. Am. Chem. Soc.*, 2018, **140**, 2080.
- [2] J. D. Sivey and A. L. Roberts, *Environ. Sci. Technol.*, 2012, **46**, 2187.
- [3] K. Moonen, C. V. Stevens, *Synthesis*, 2005, **20**, 3603.
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8. NMR Spectra

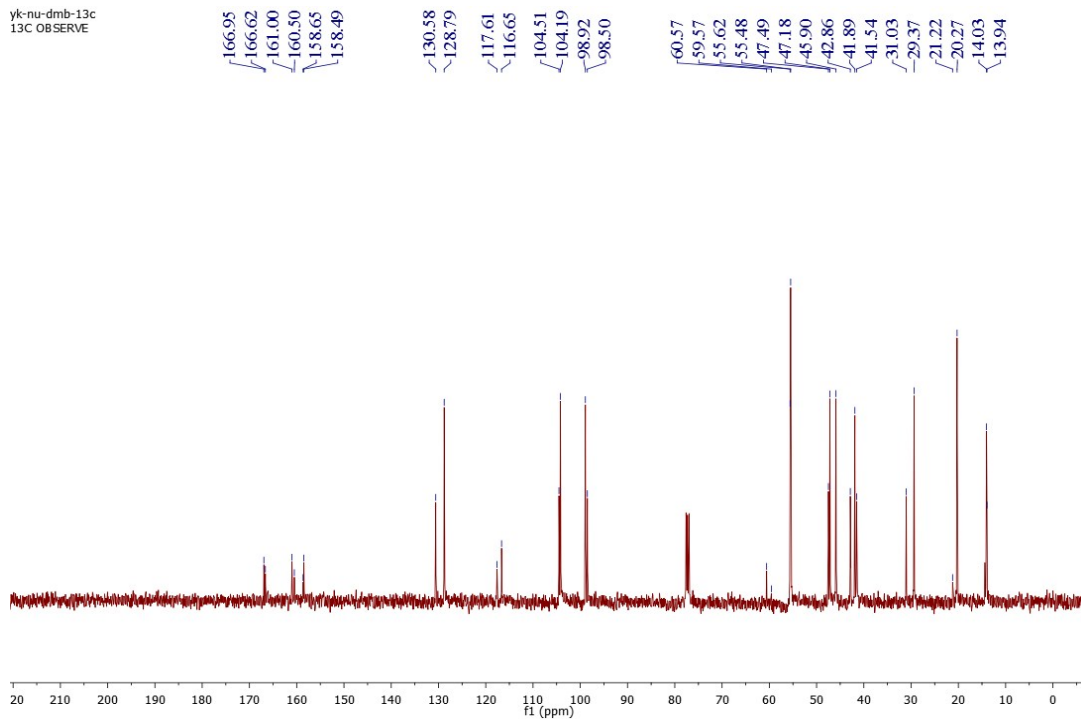


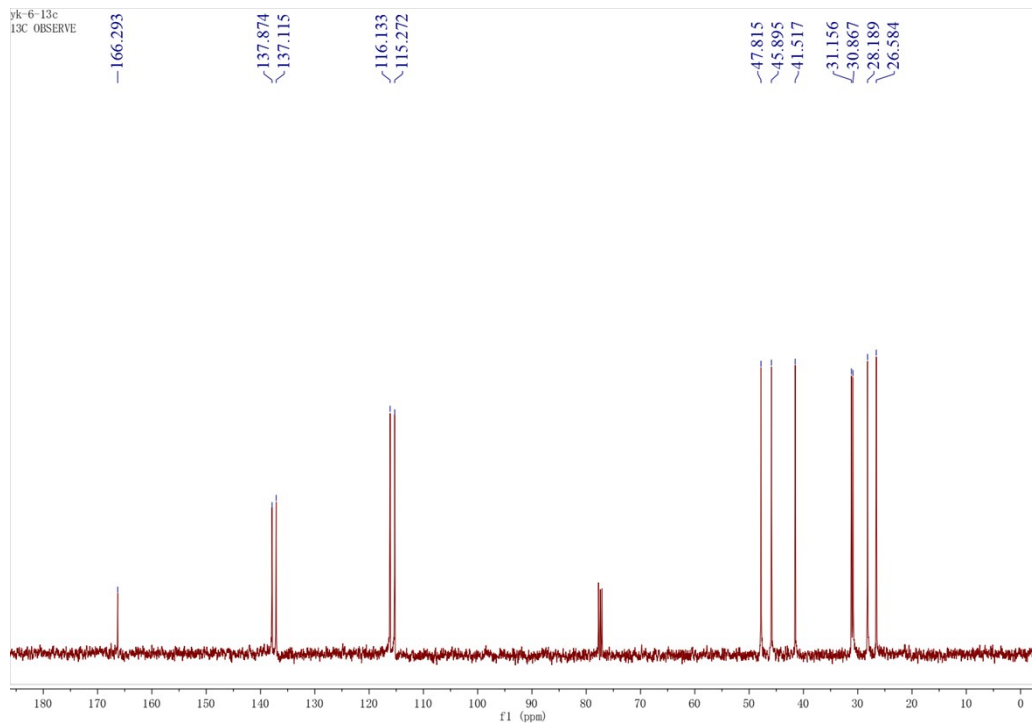
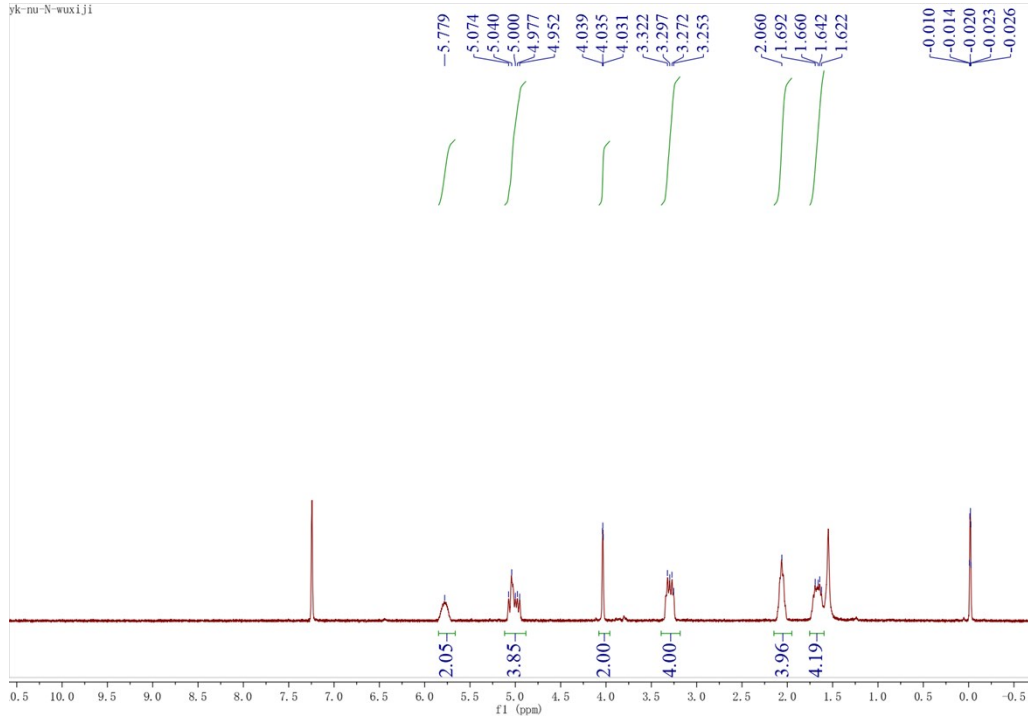
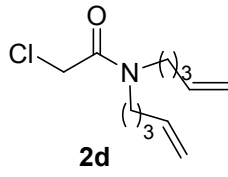


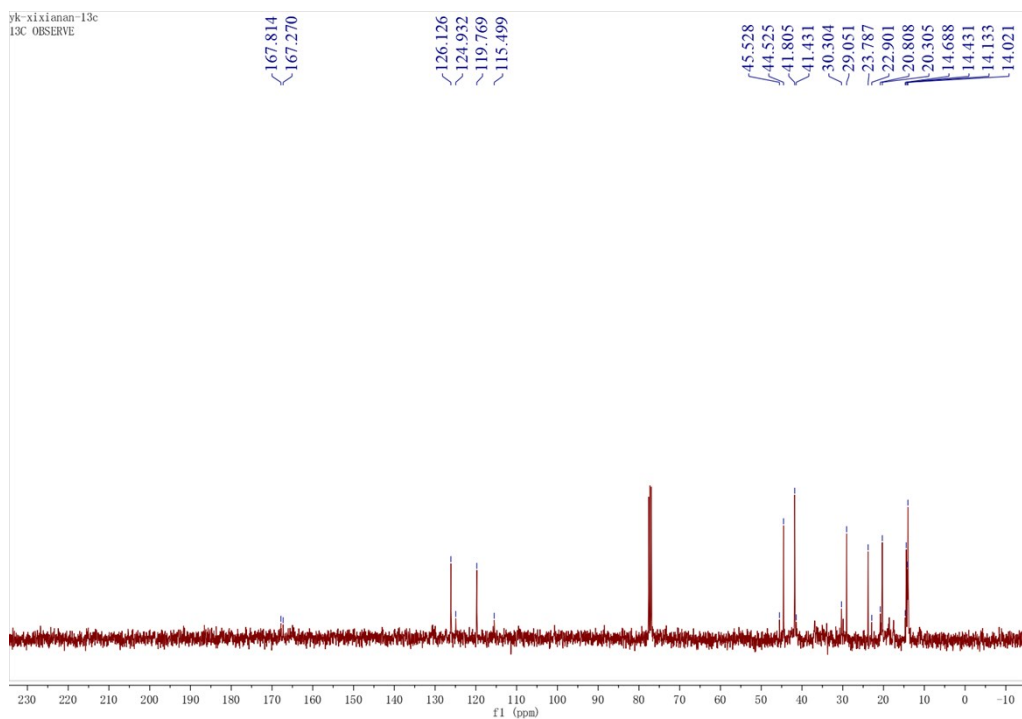
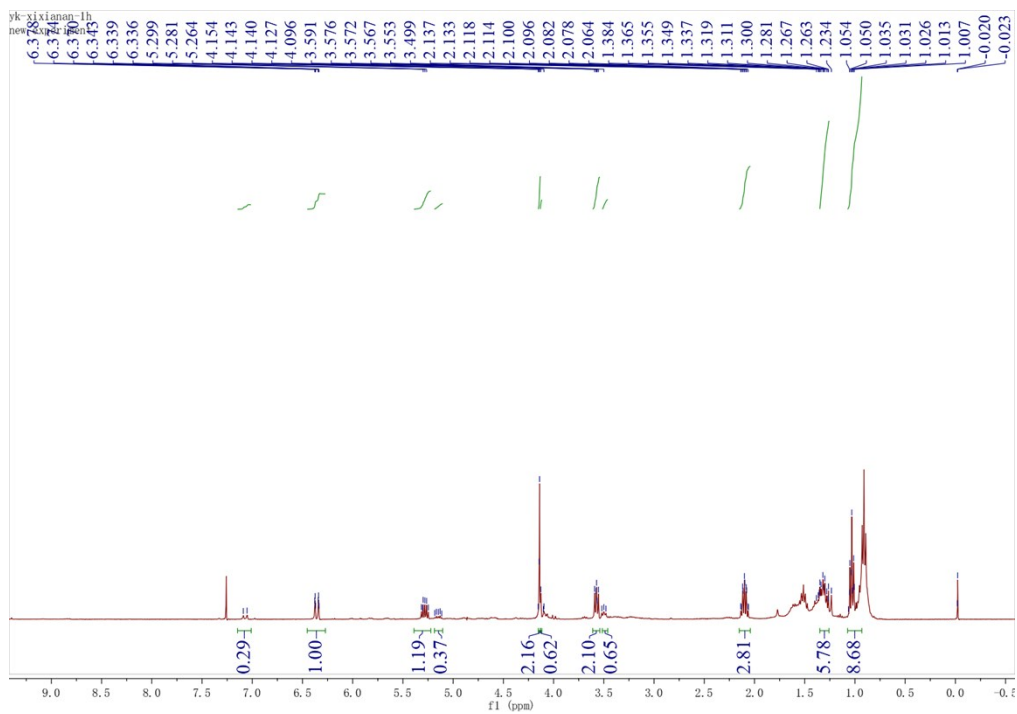
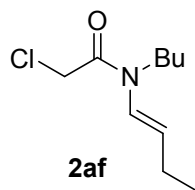
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new experiment

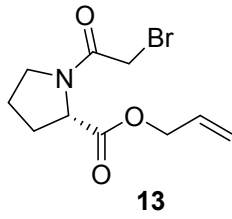


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13C OBSERVE

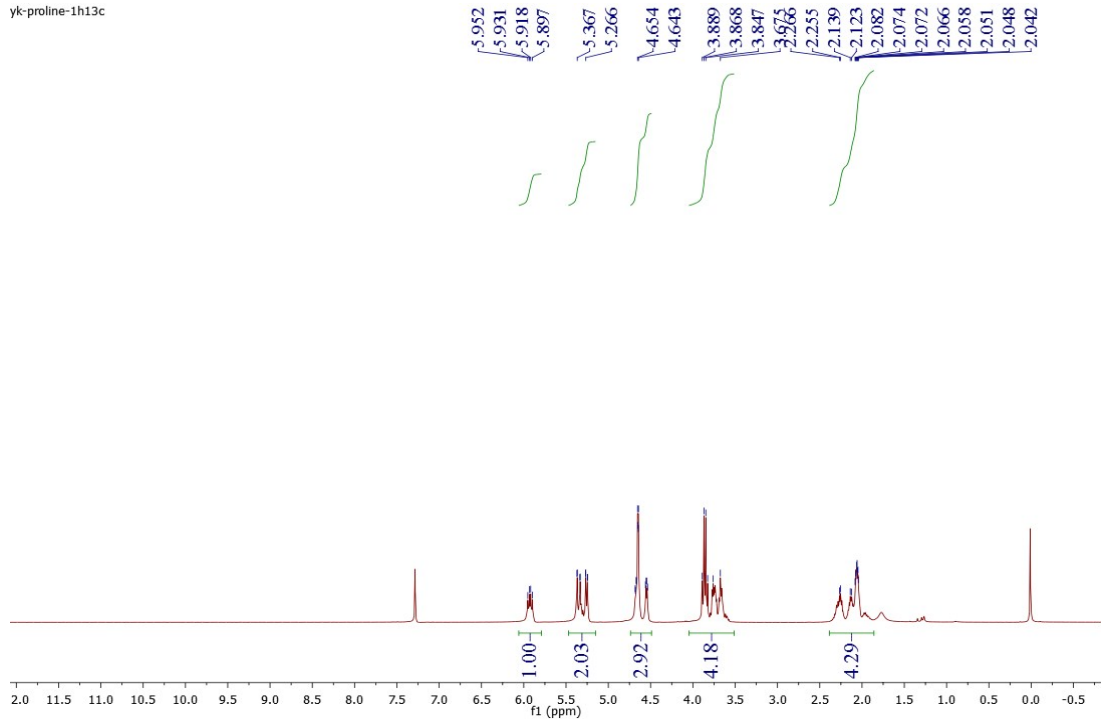




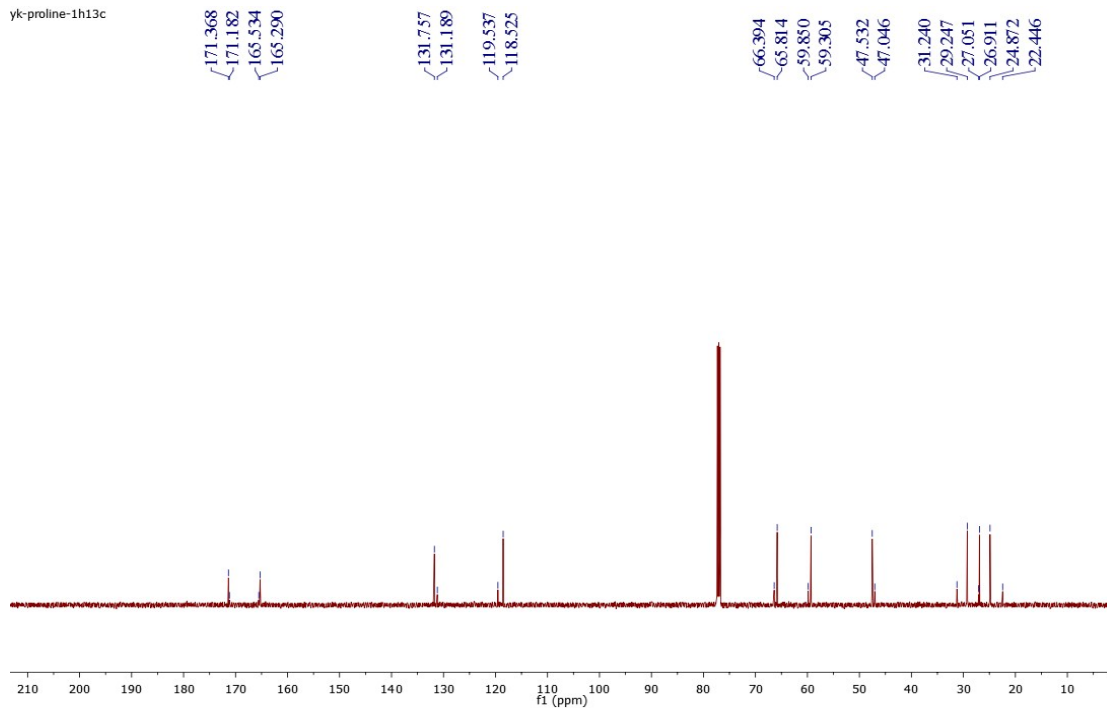


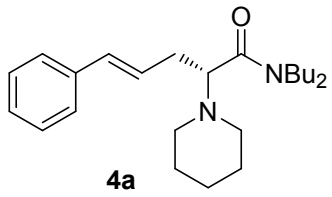


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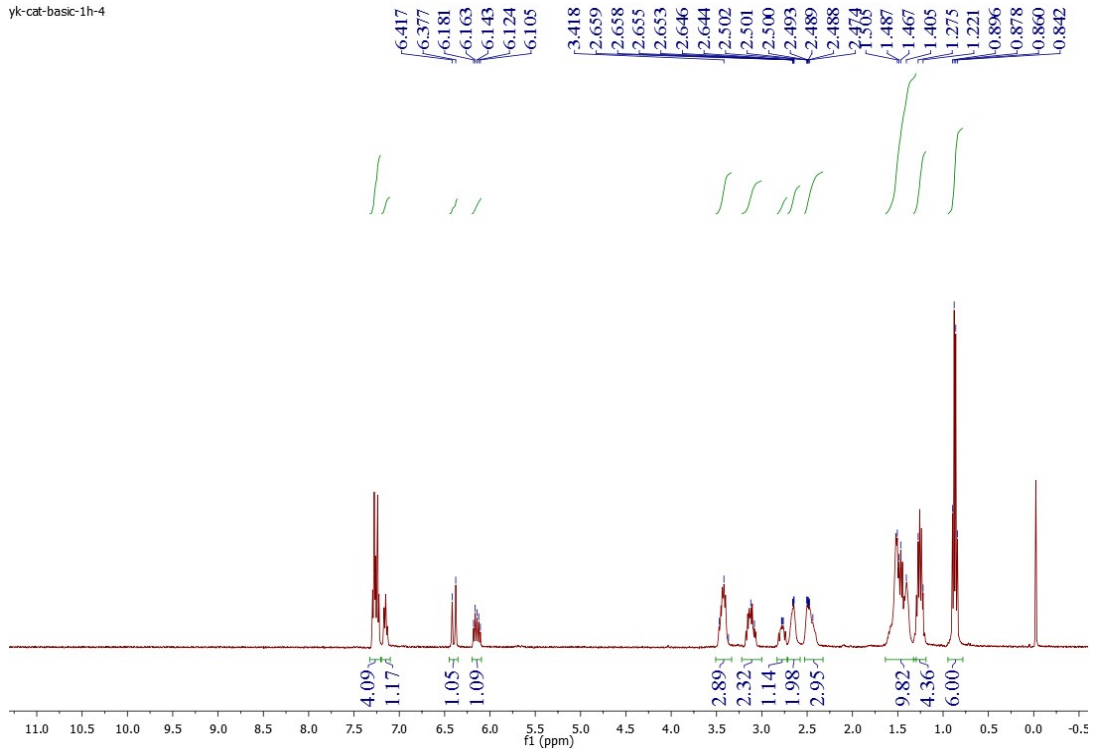


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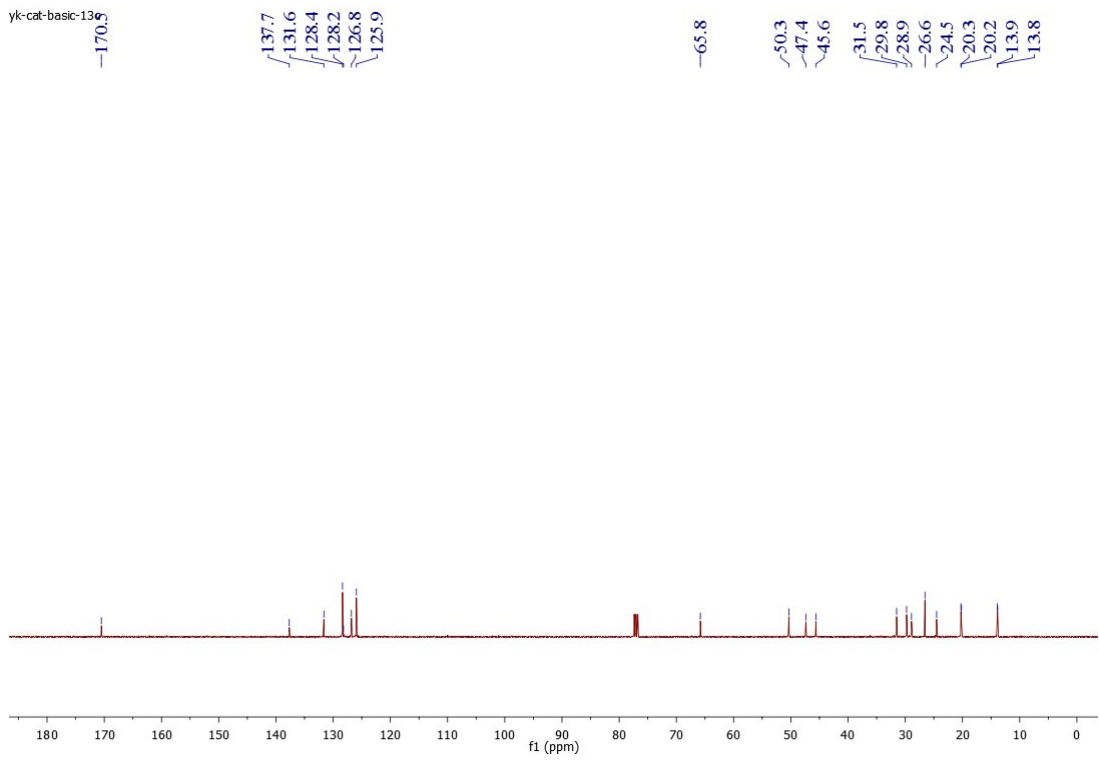


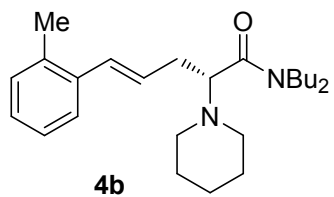


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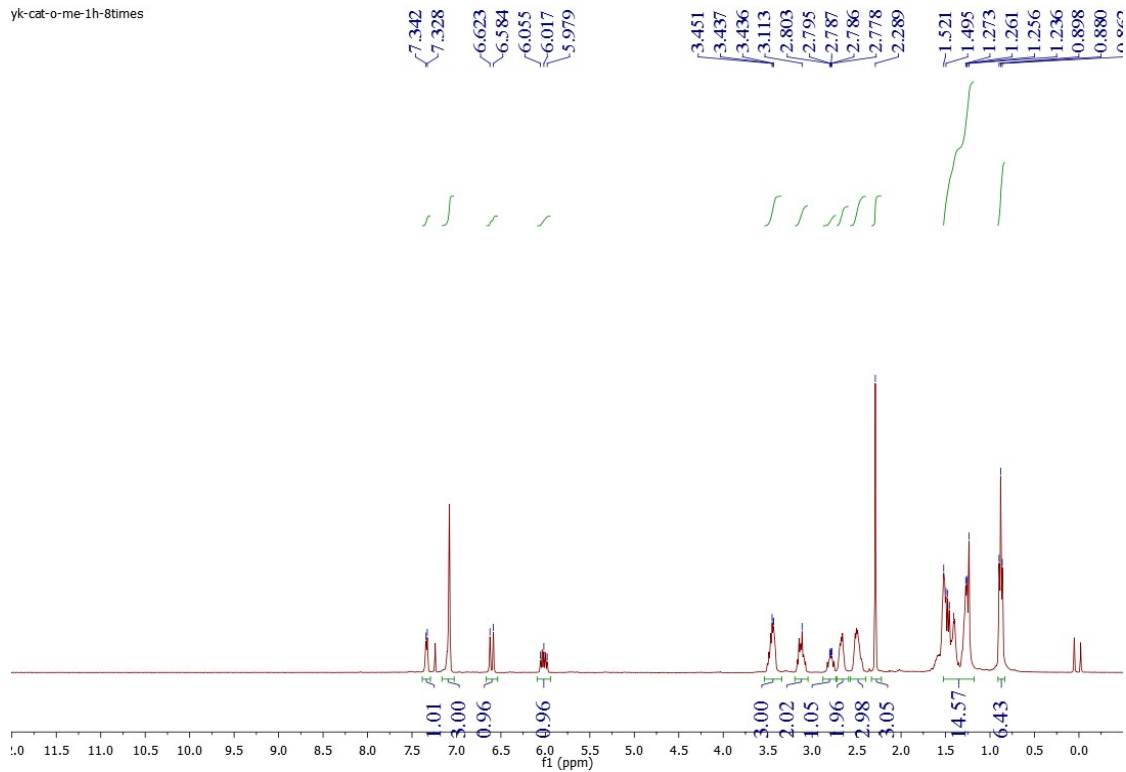


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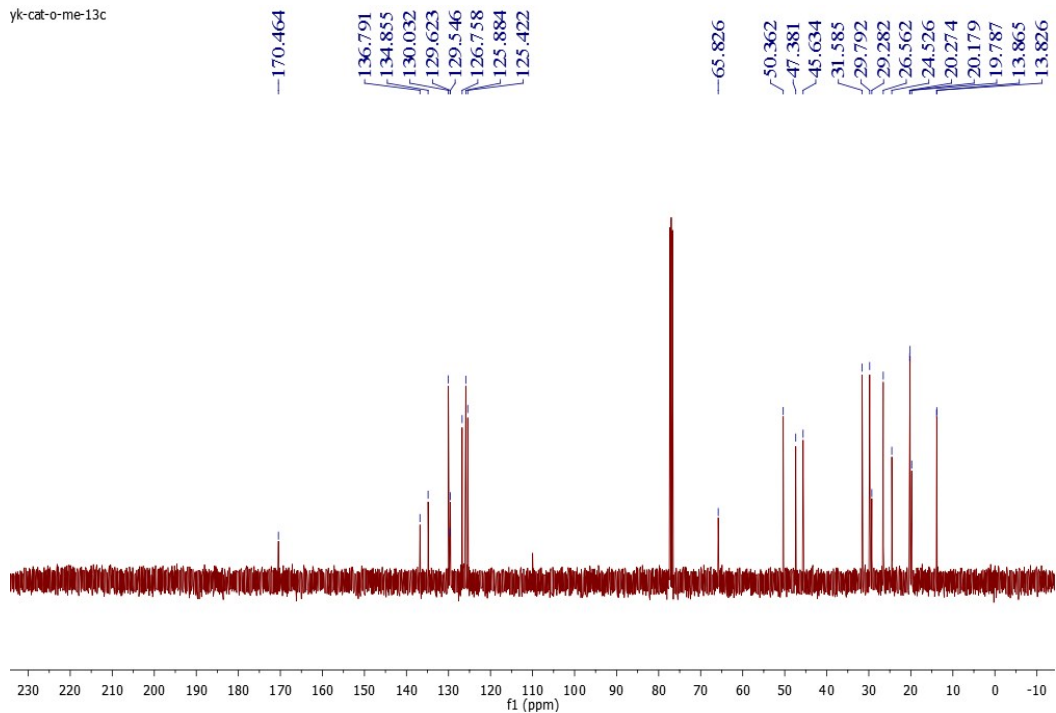


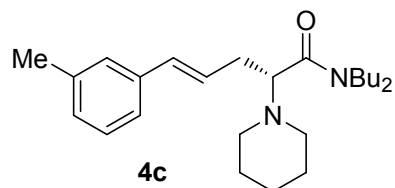


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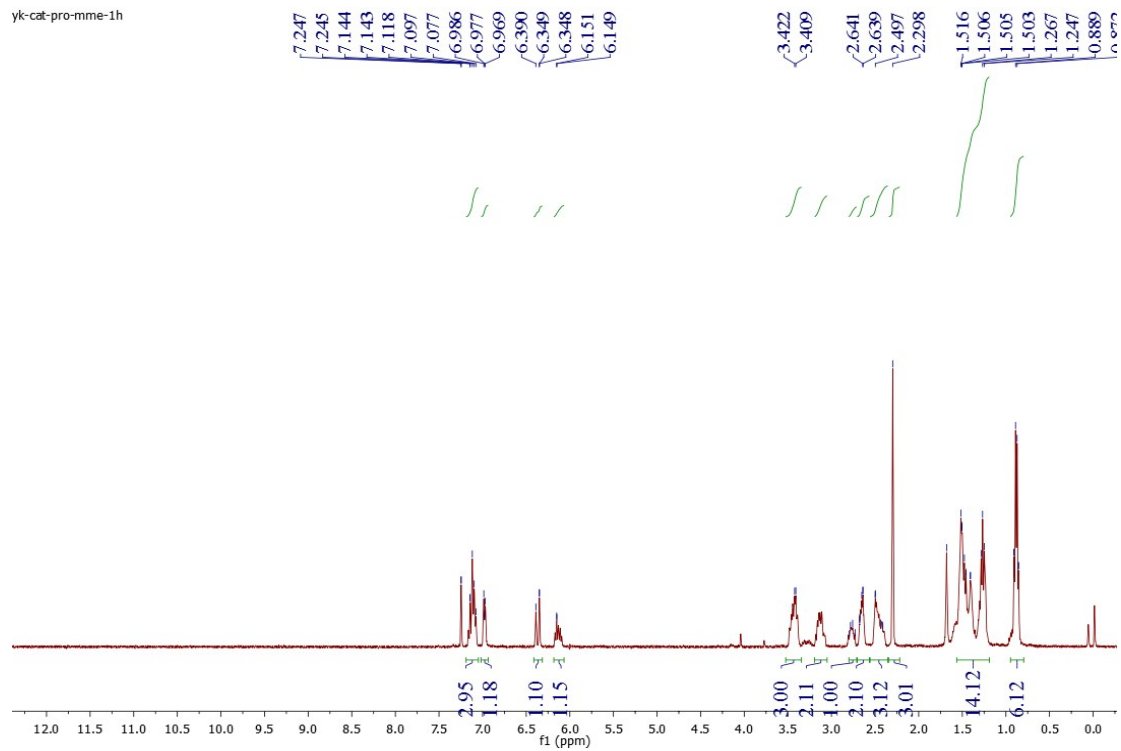


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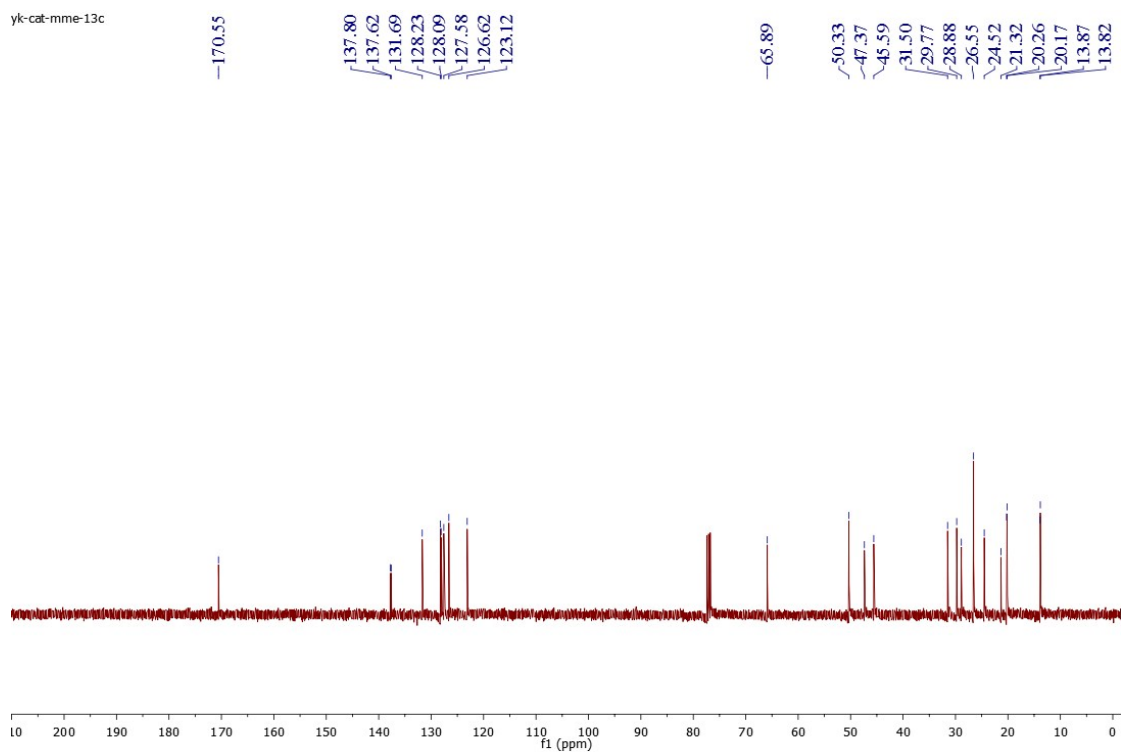


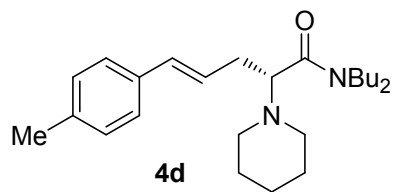


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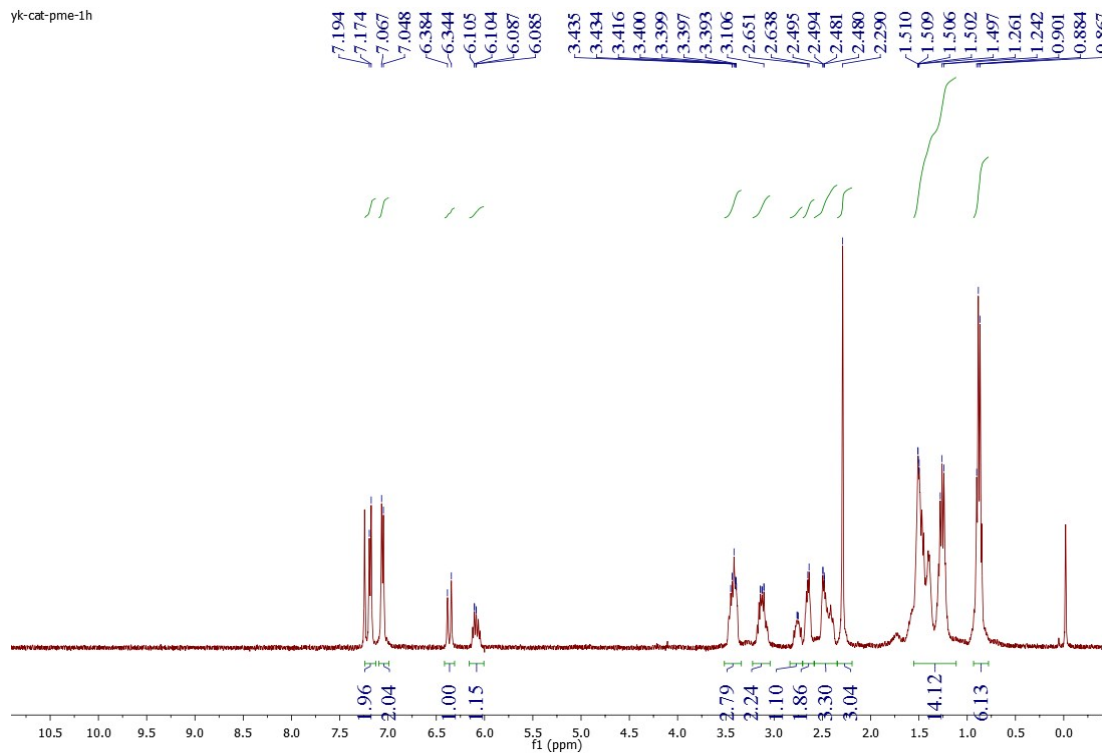


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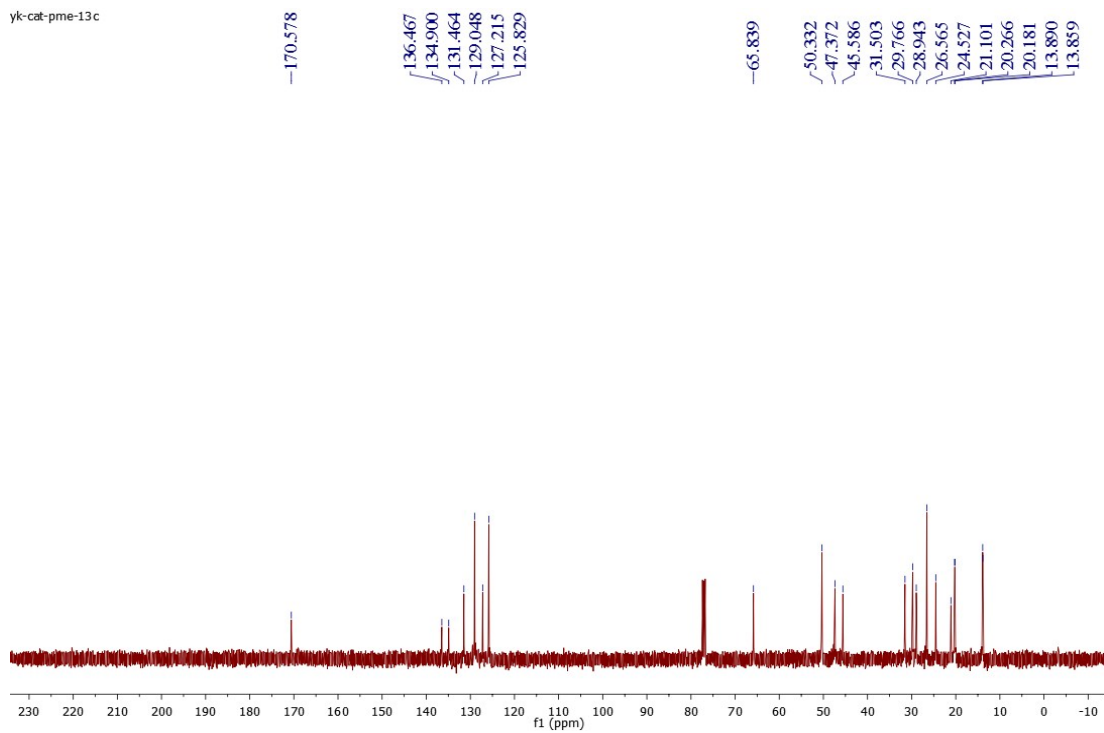


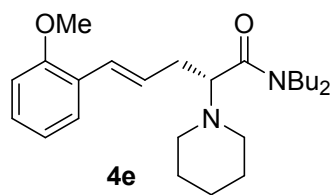


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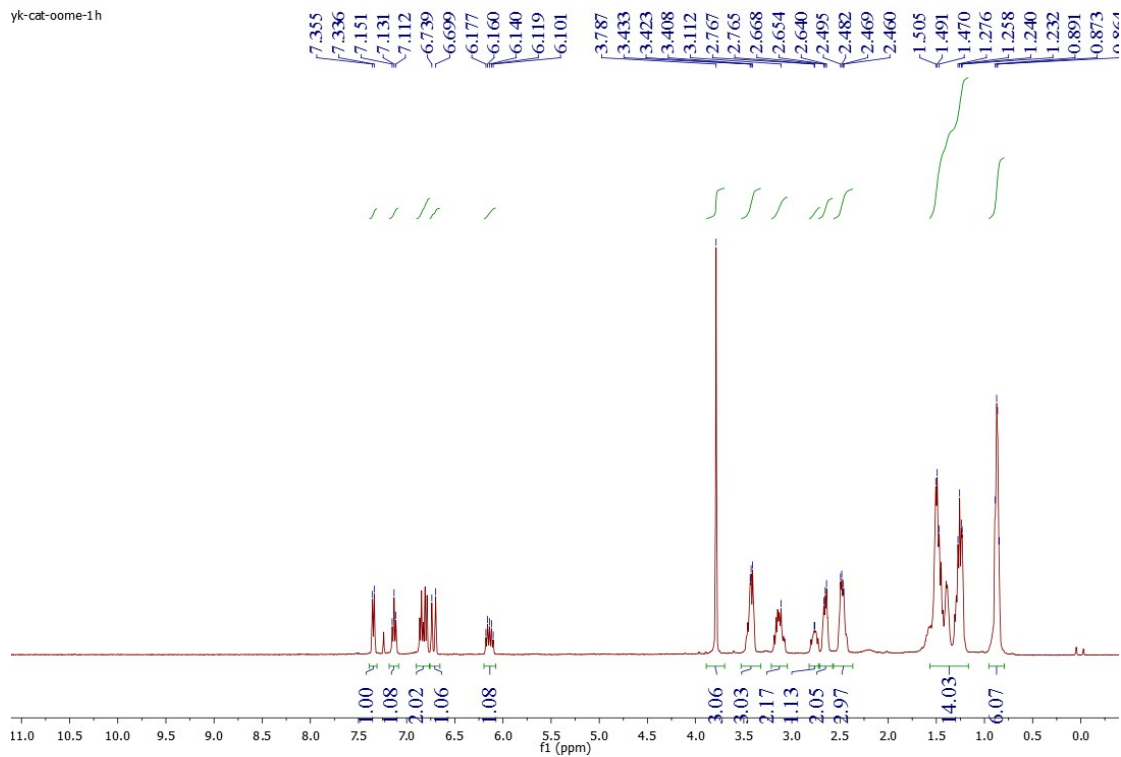


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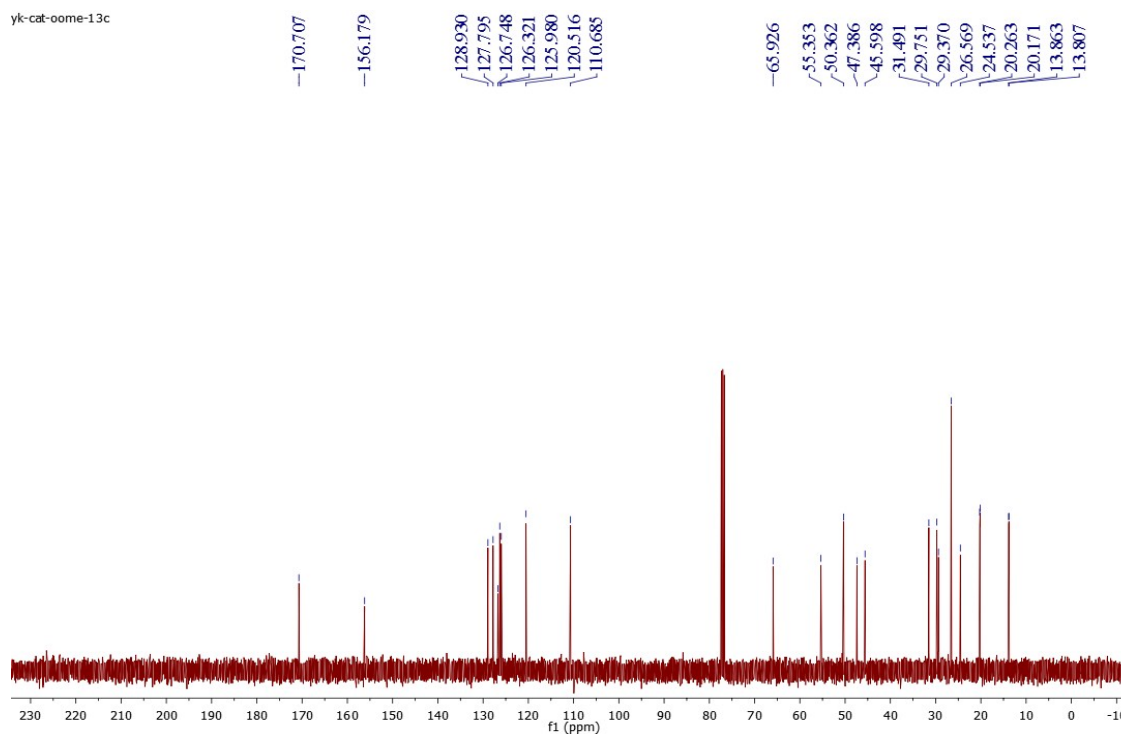


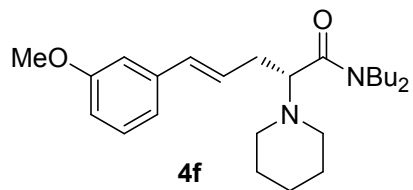


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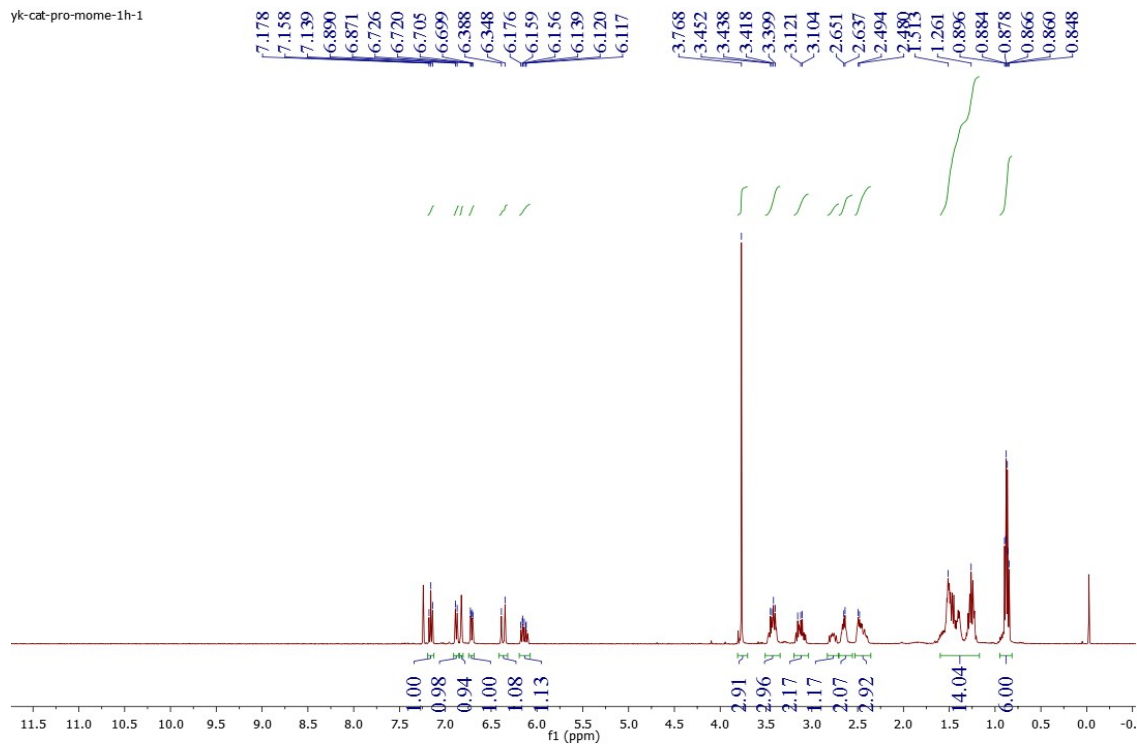


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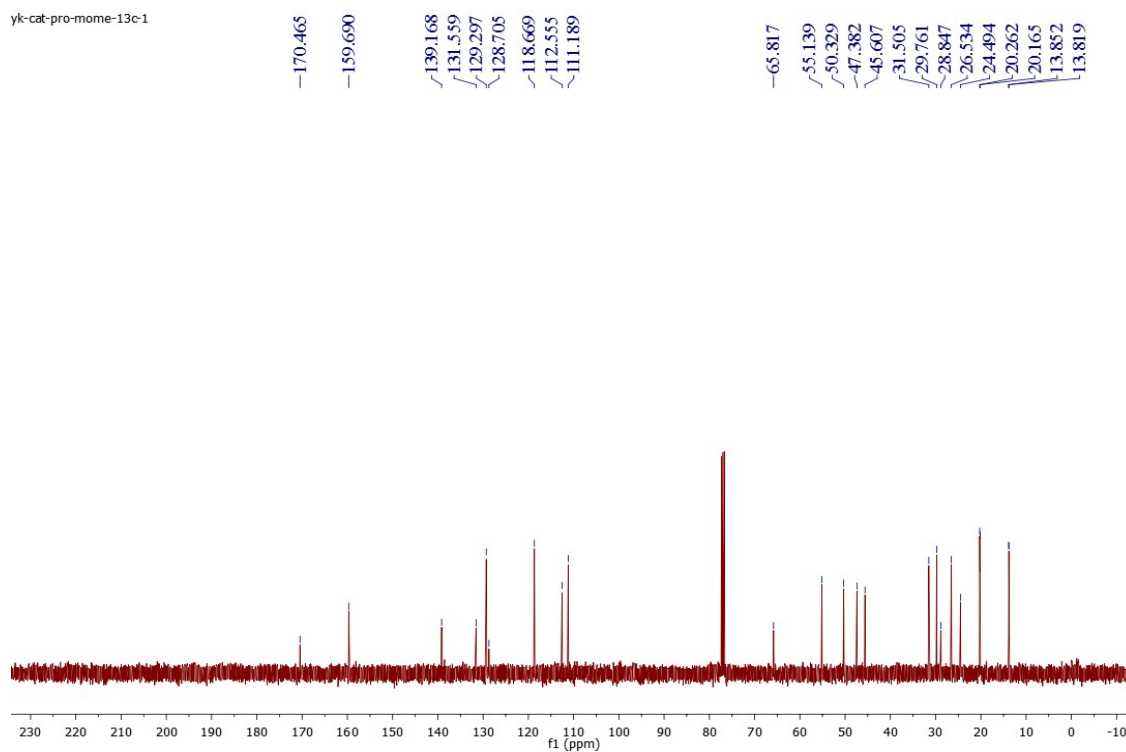


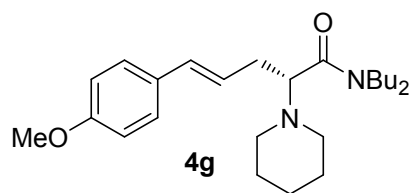


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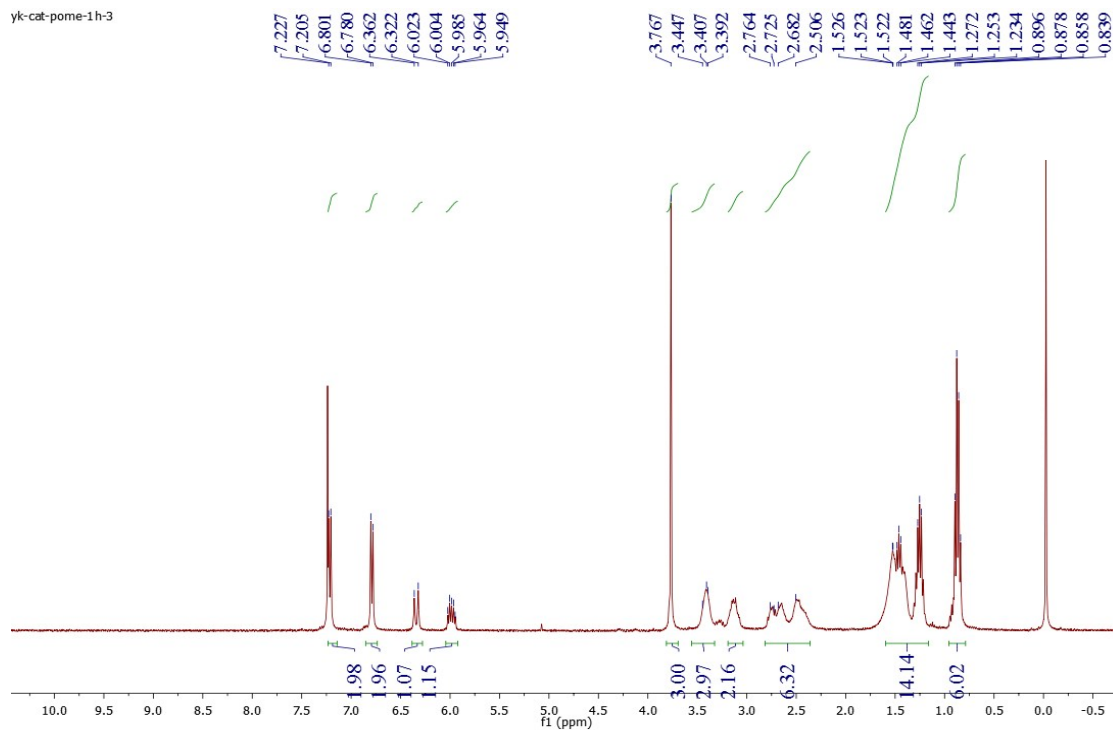


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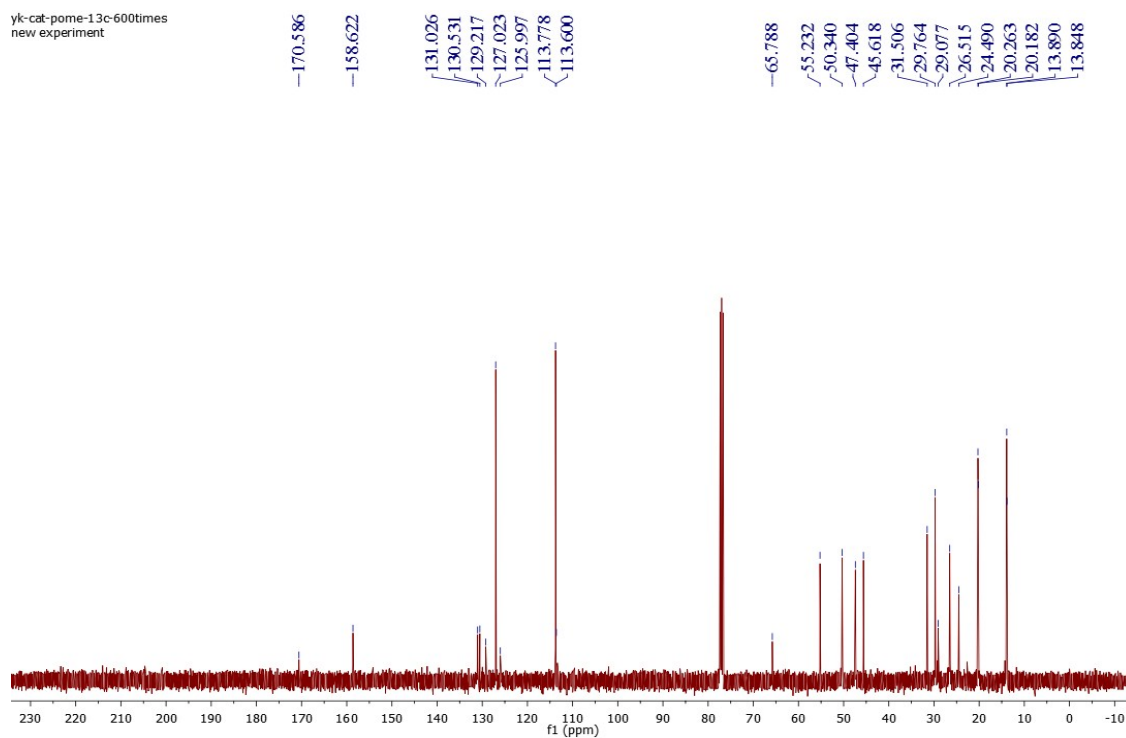


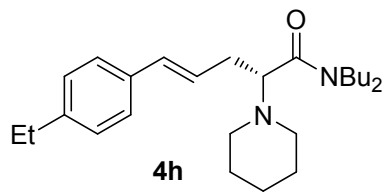


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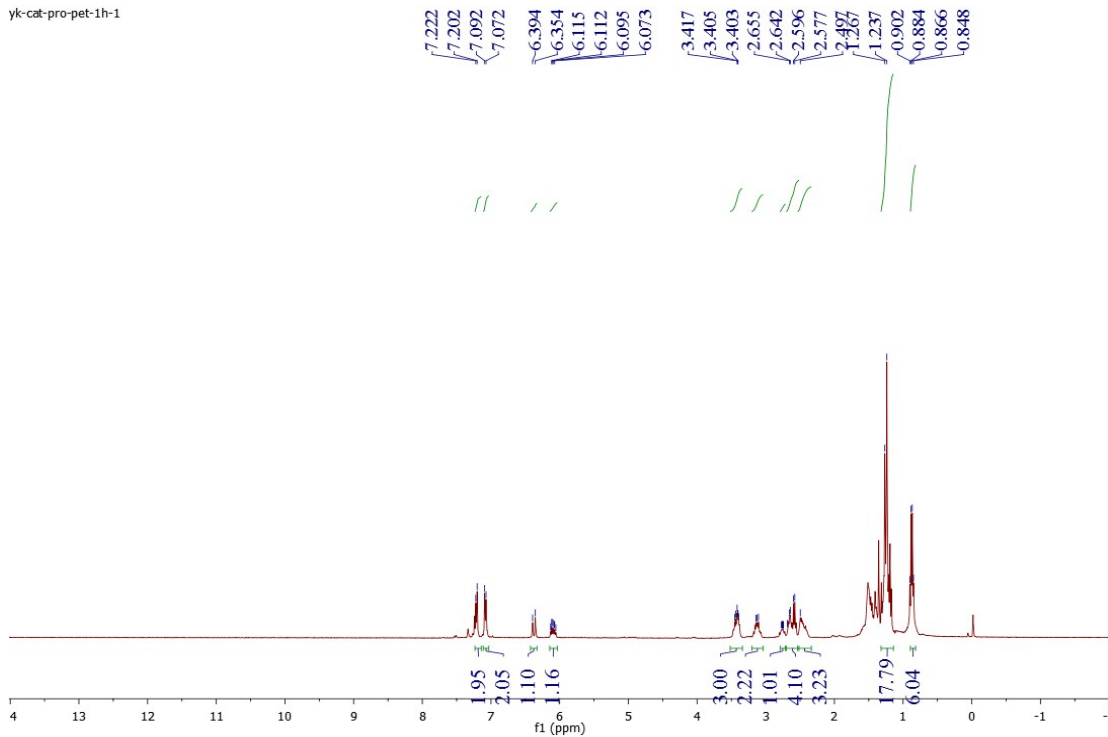


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new experiment

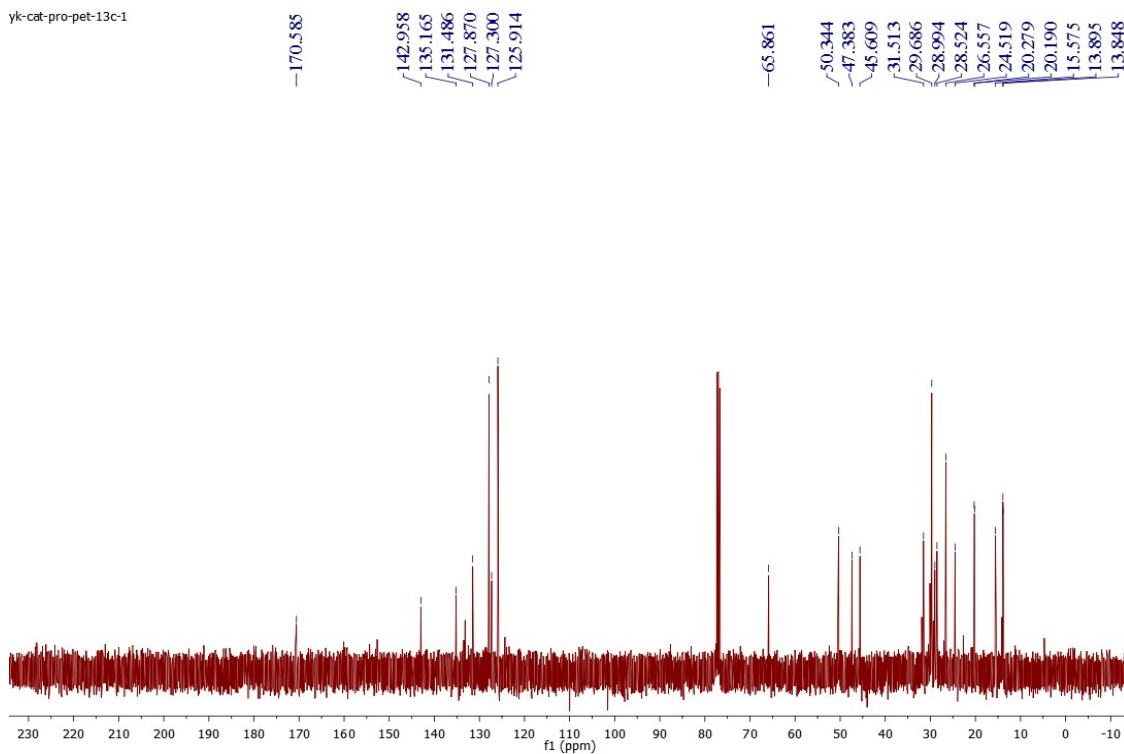


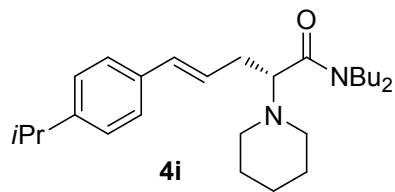


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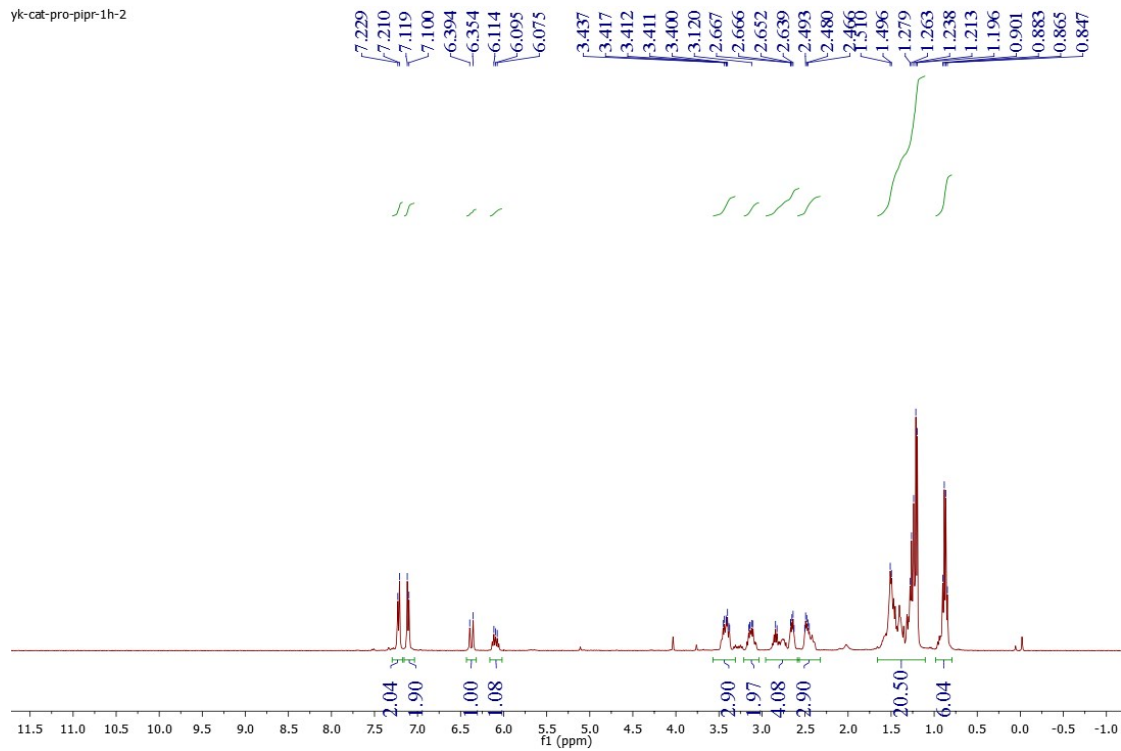


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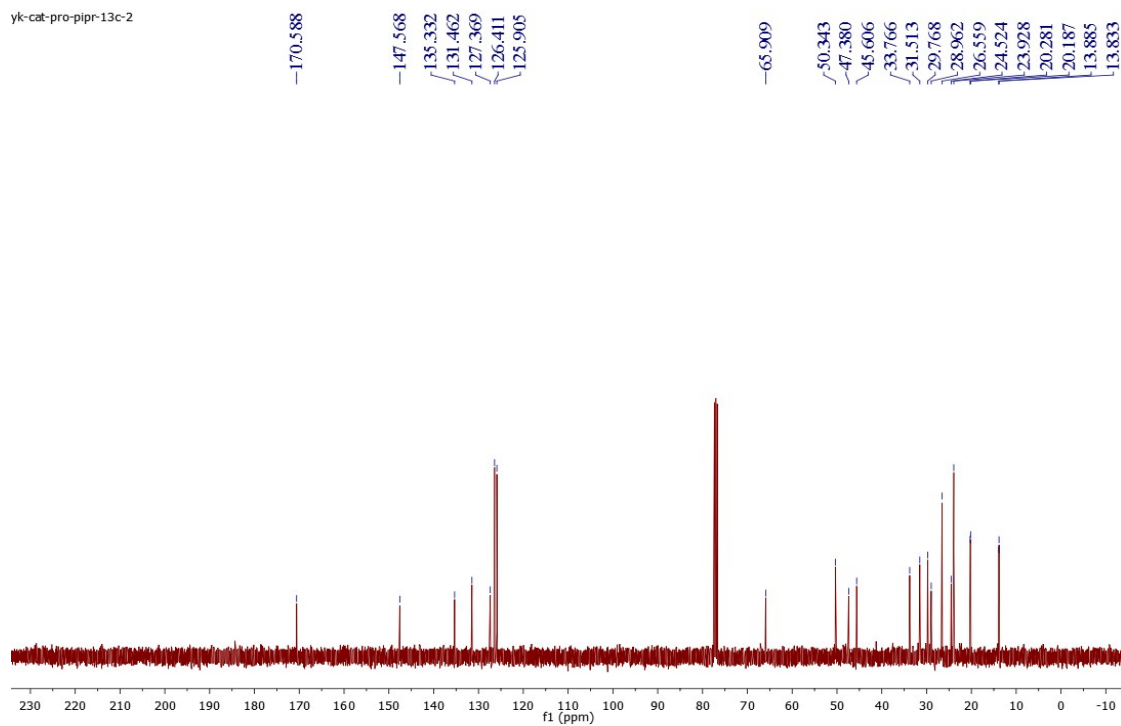


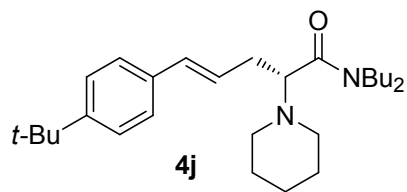


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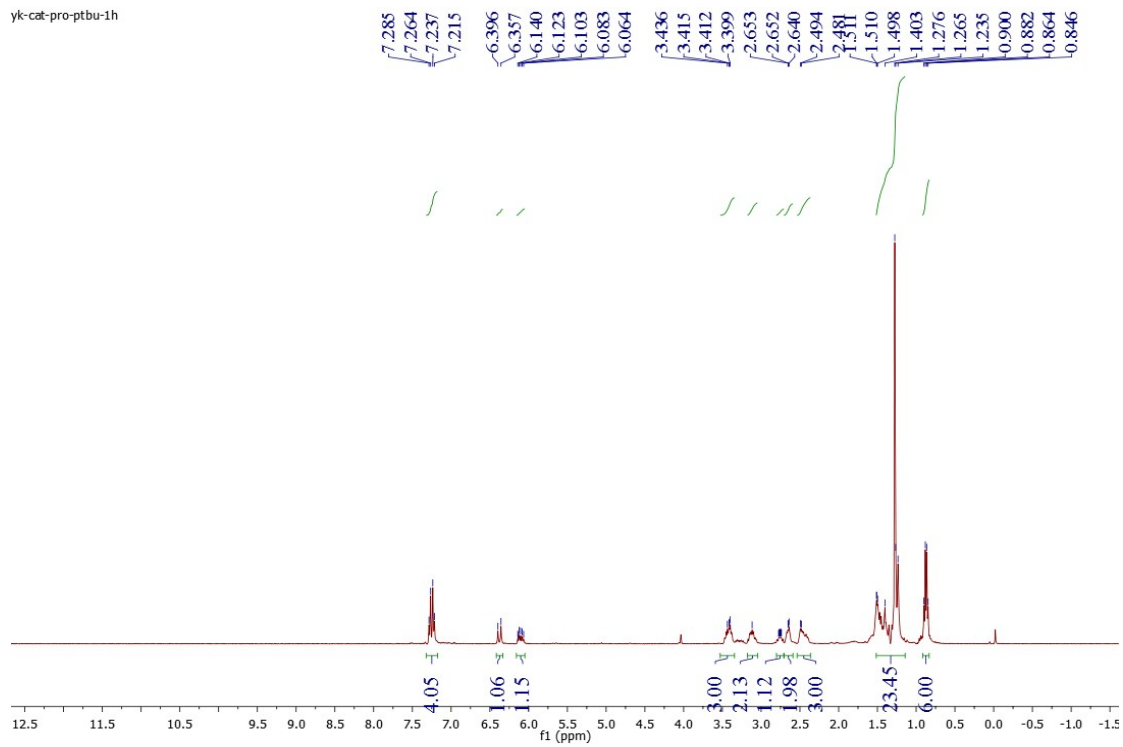


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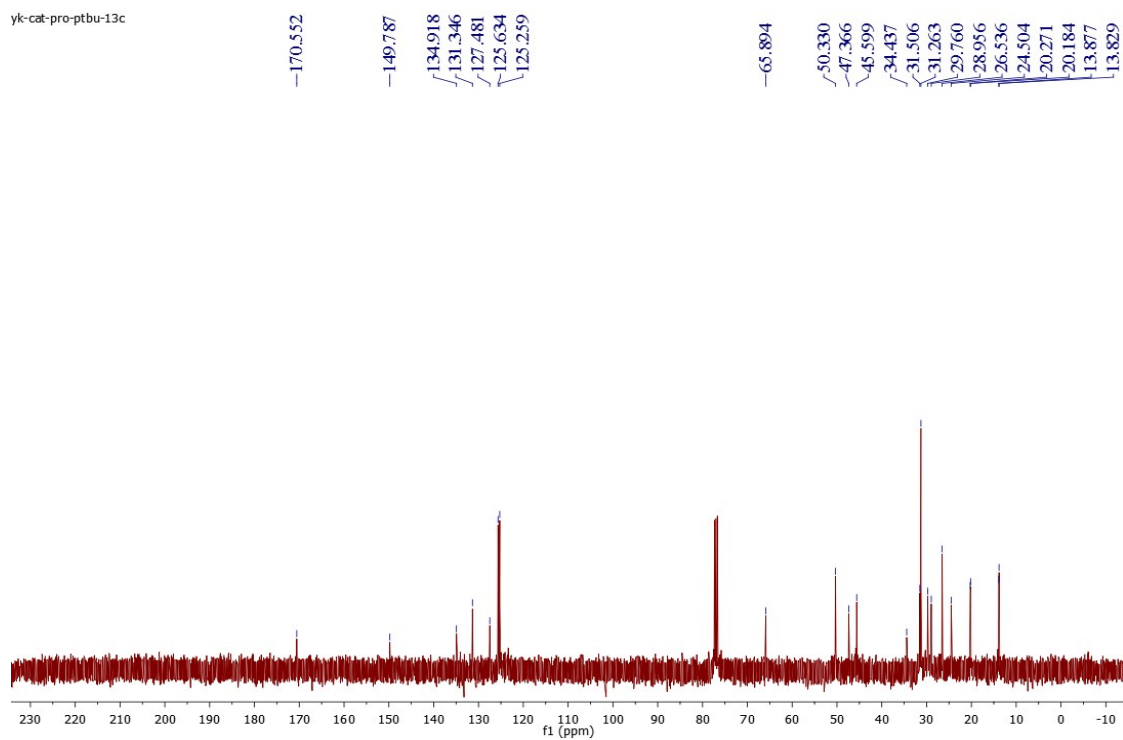


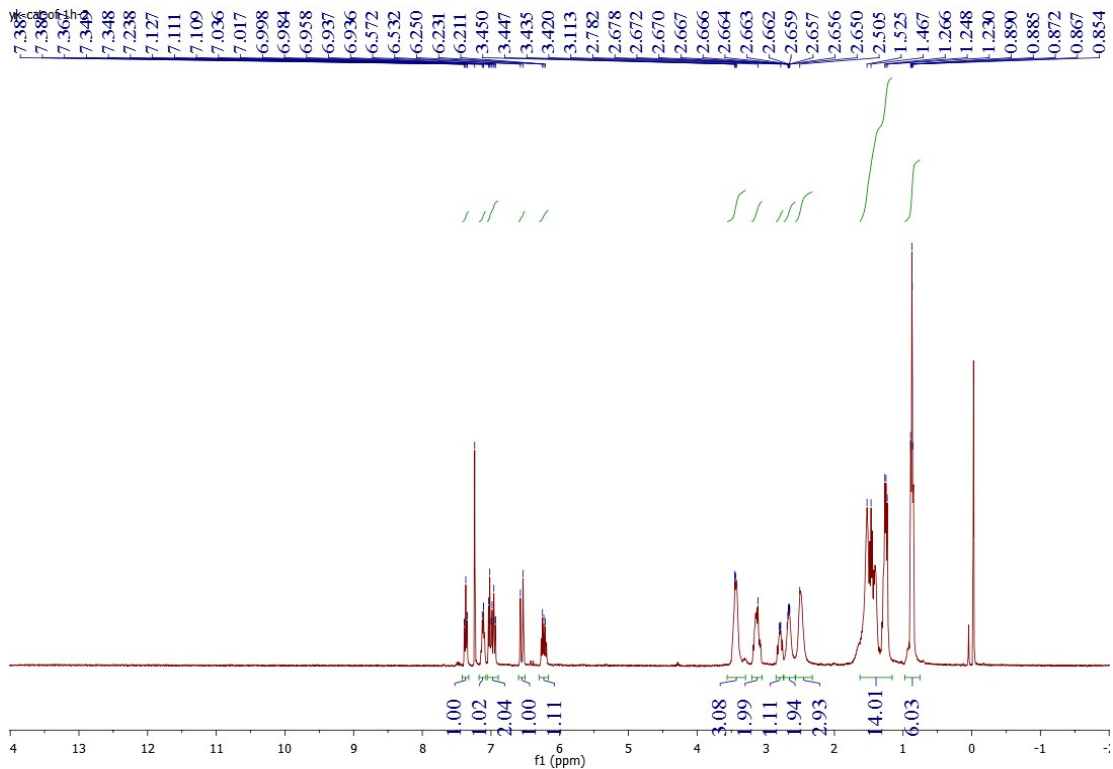
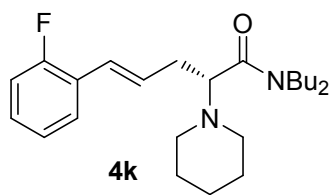


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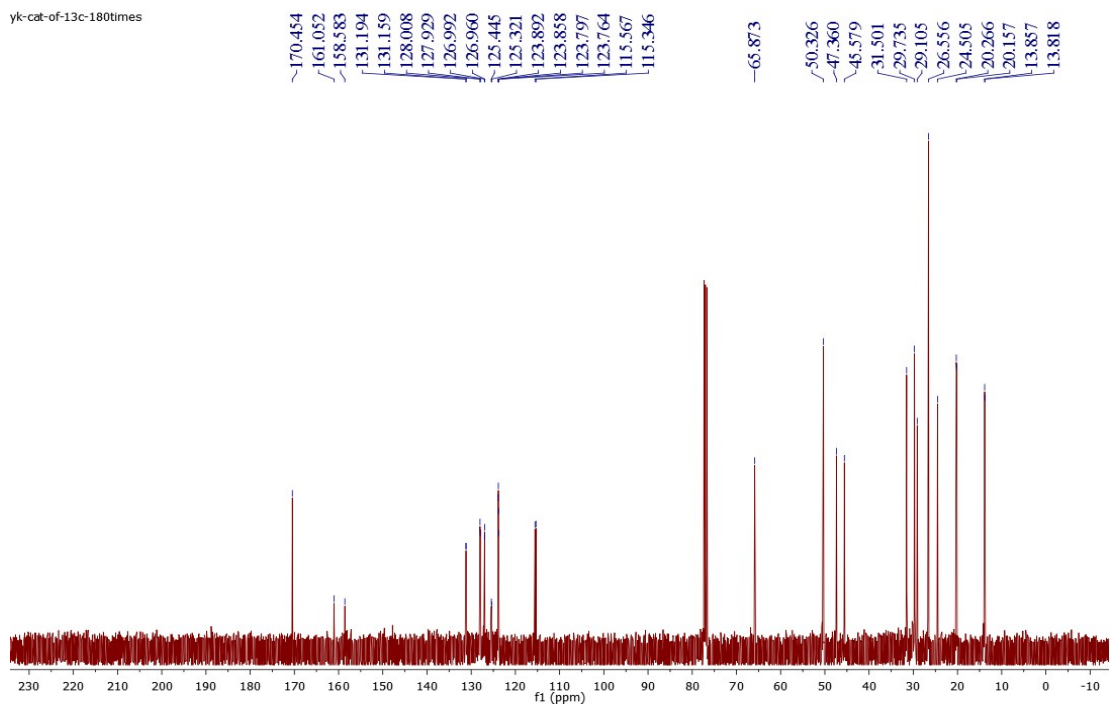


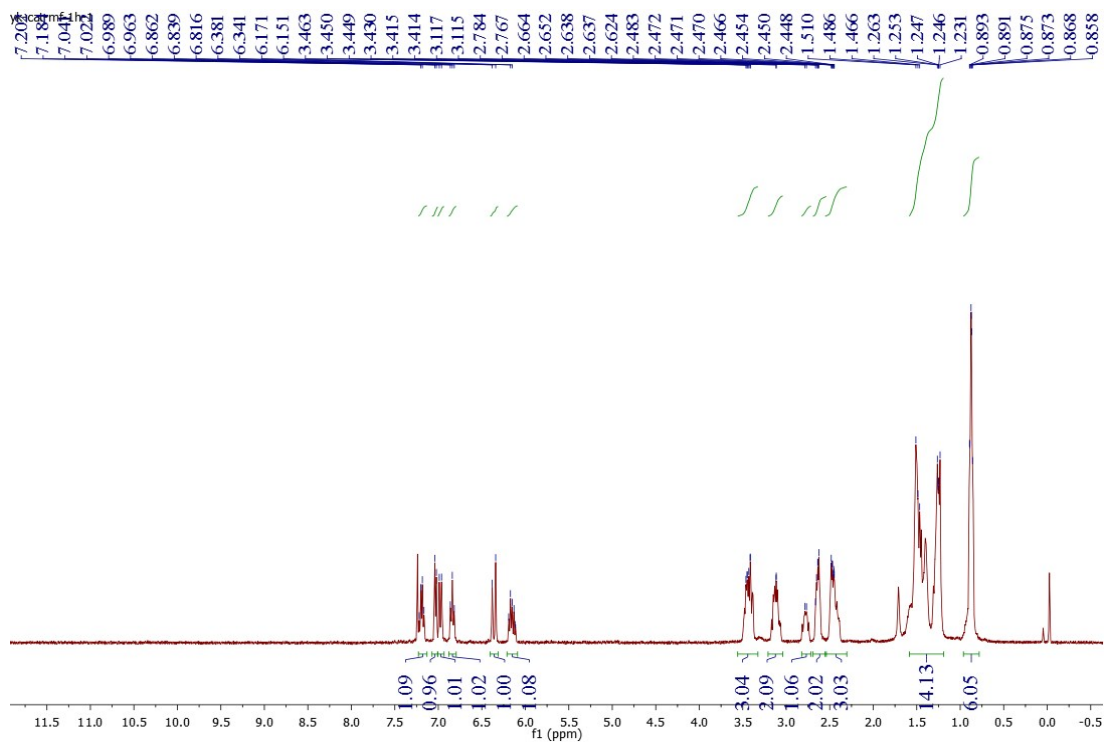
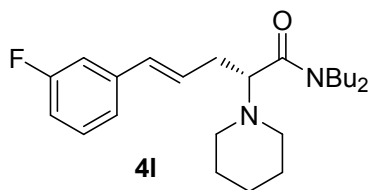
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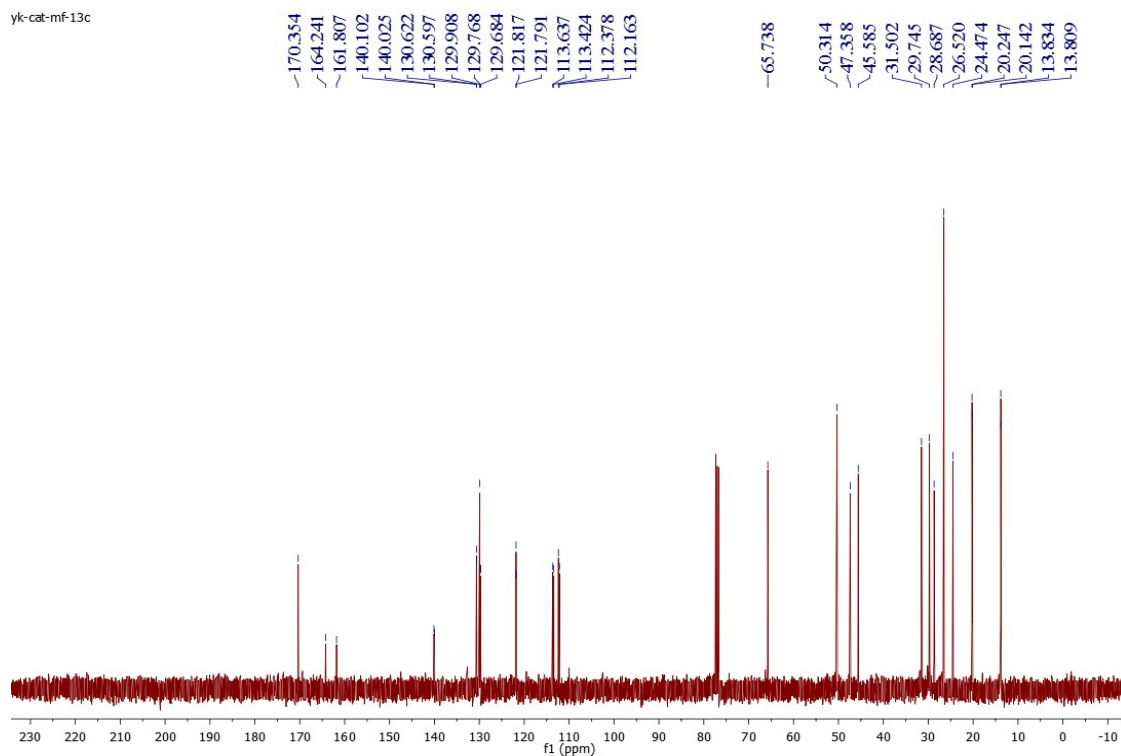


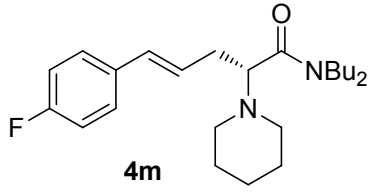
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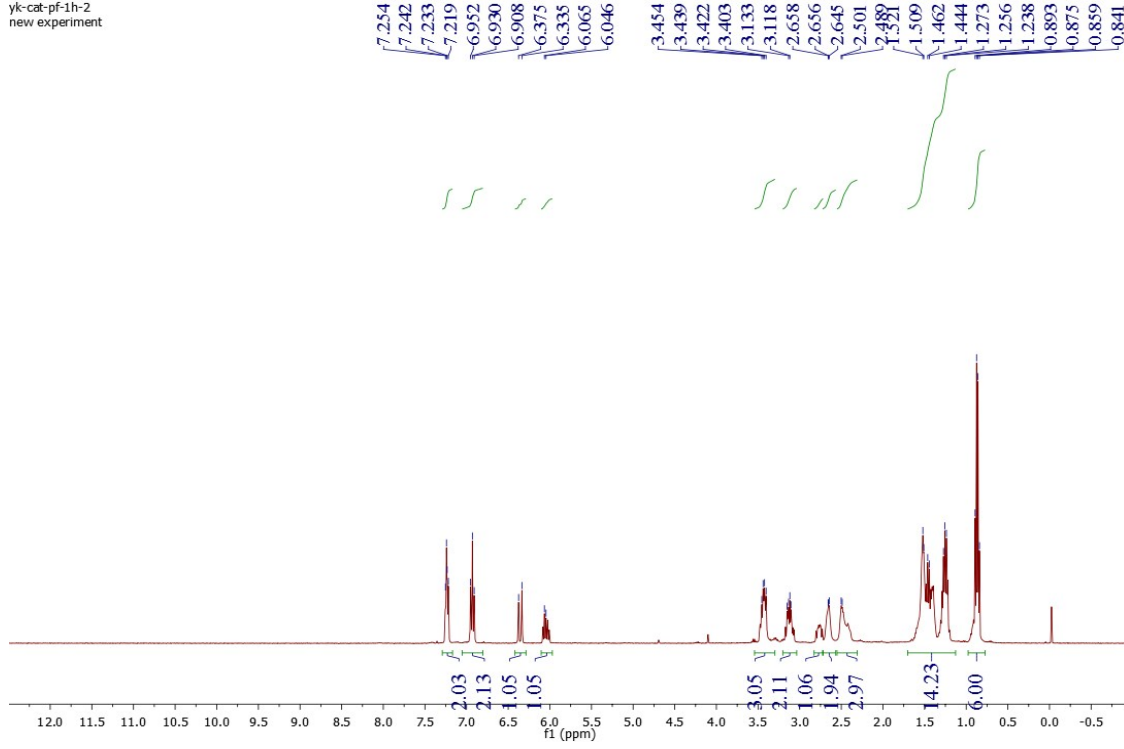


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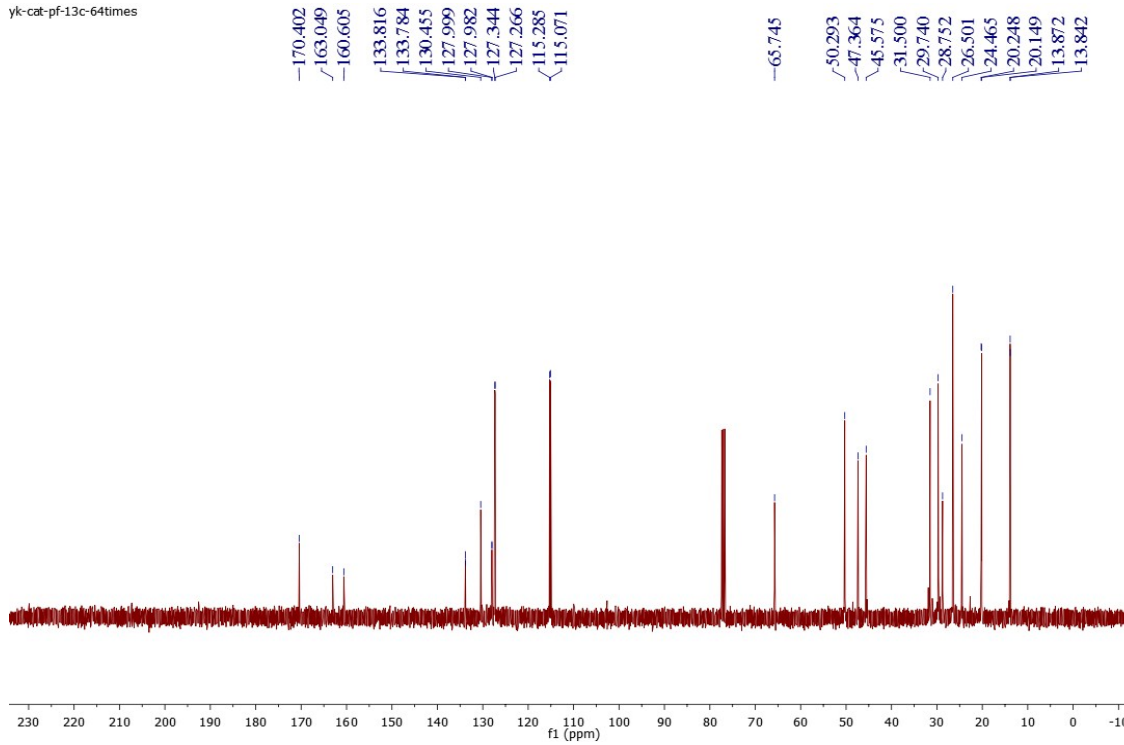


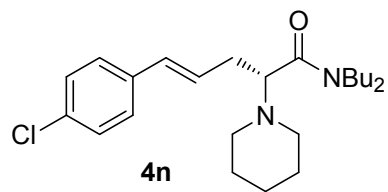


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new experiment

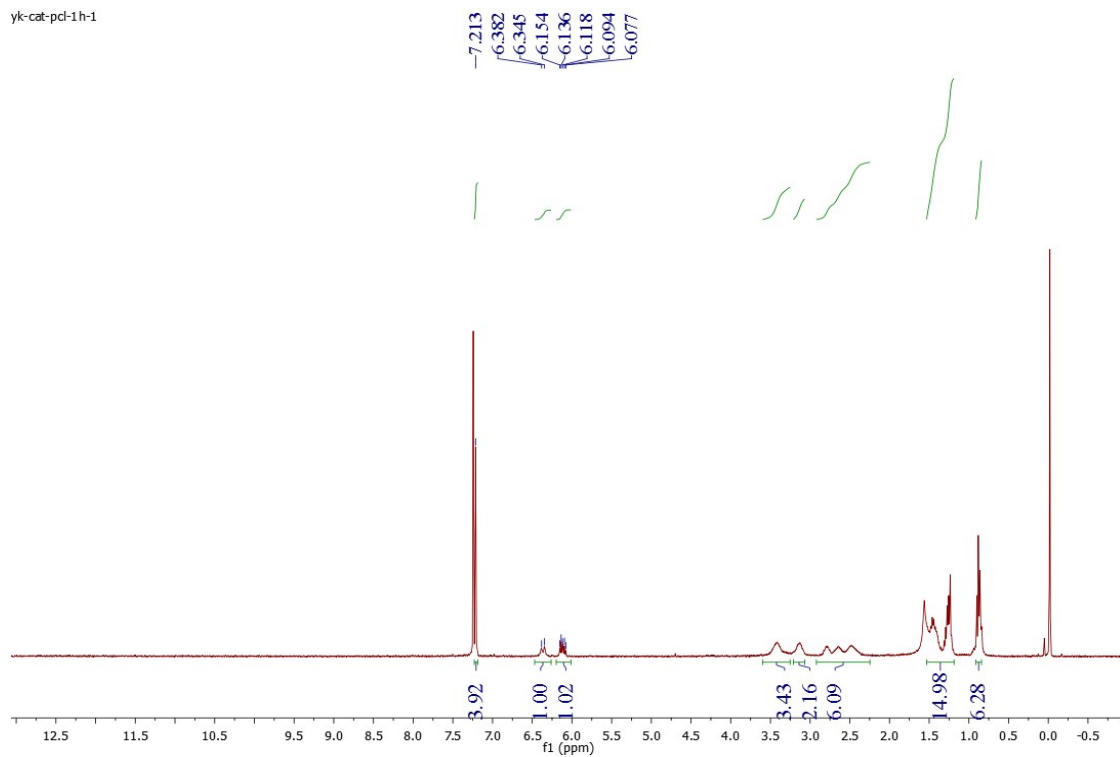


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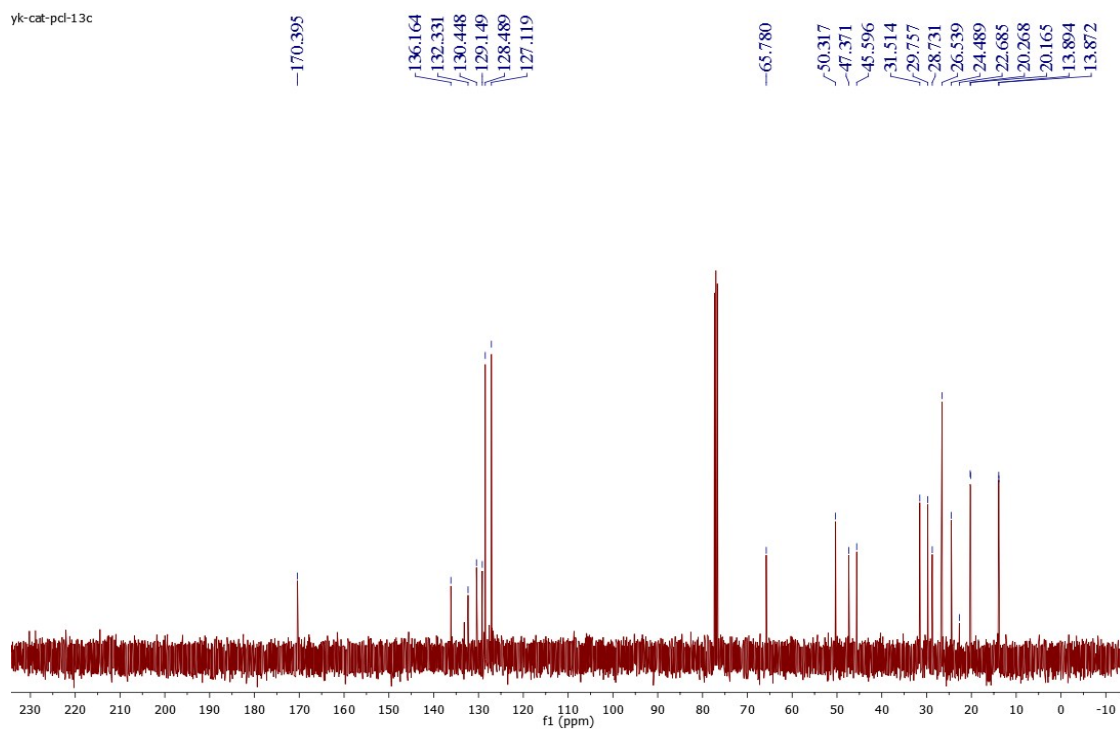


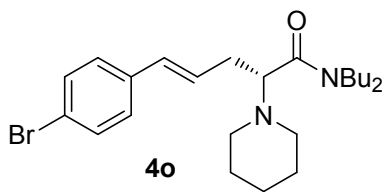


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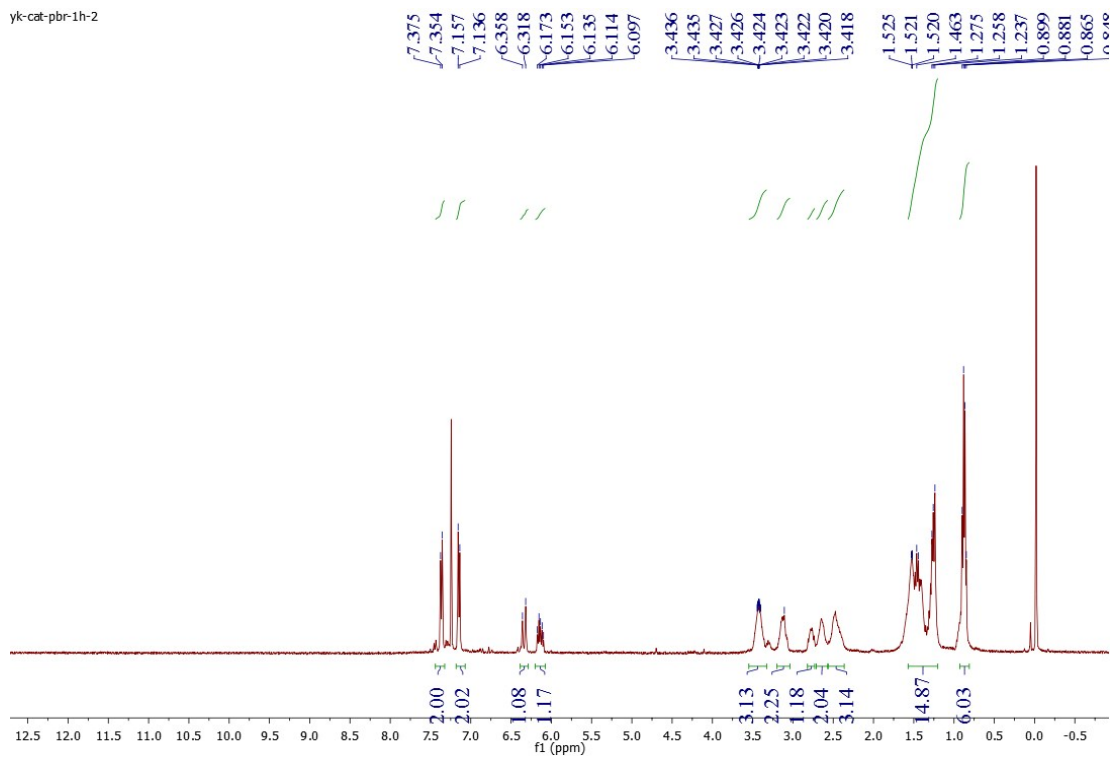


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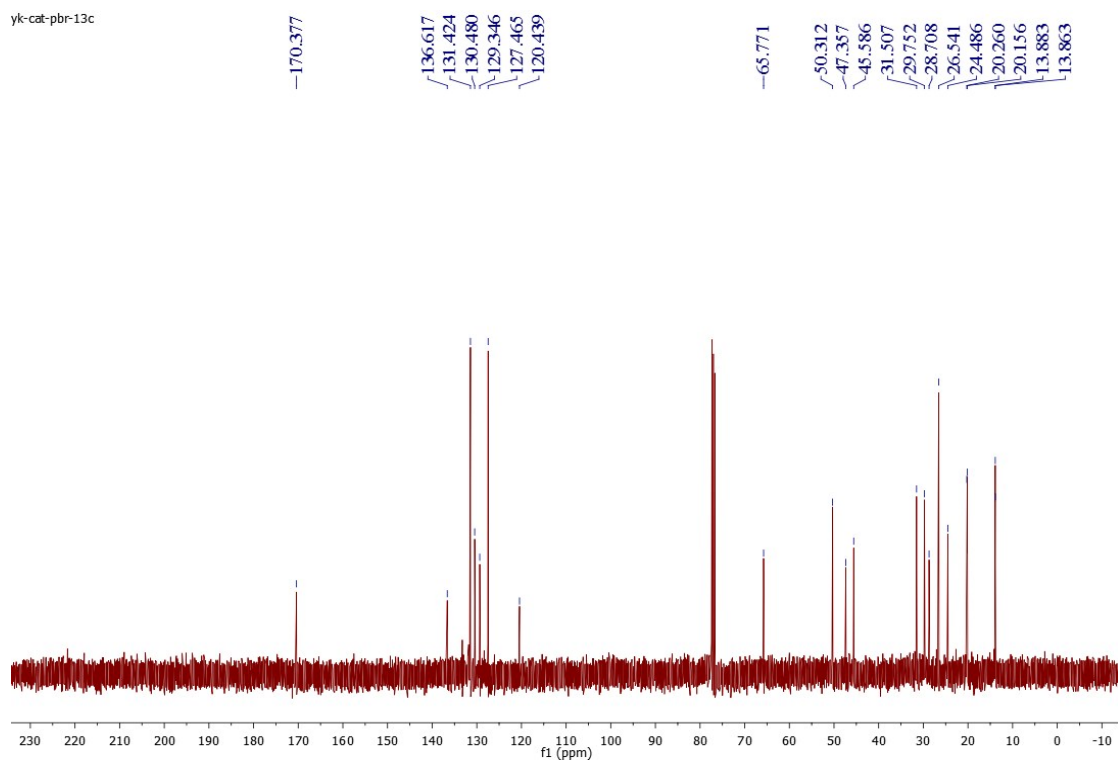


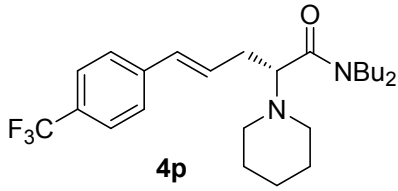


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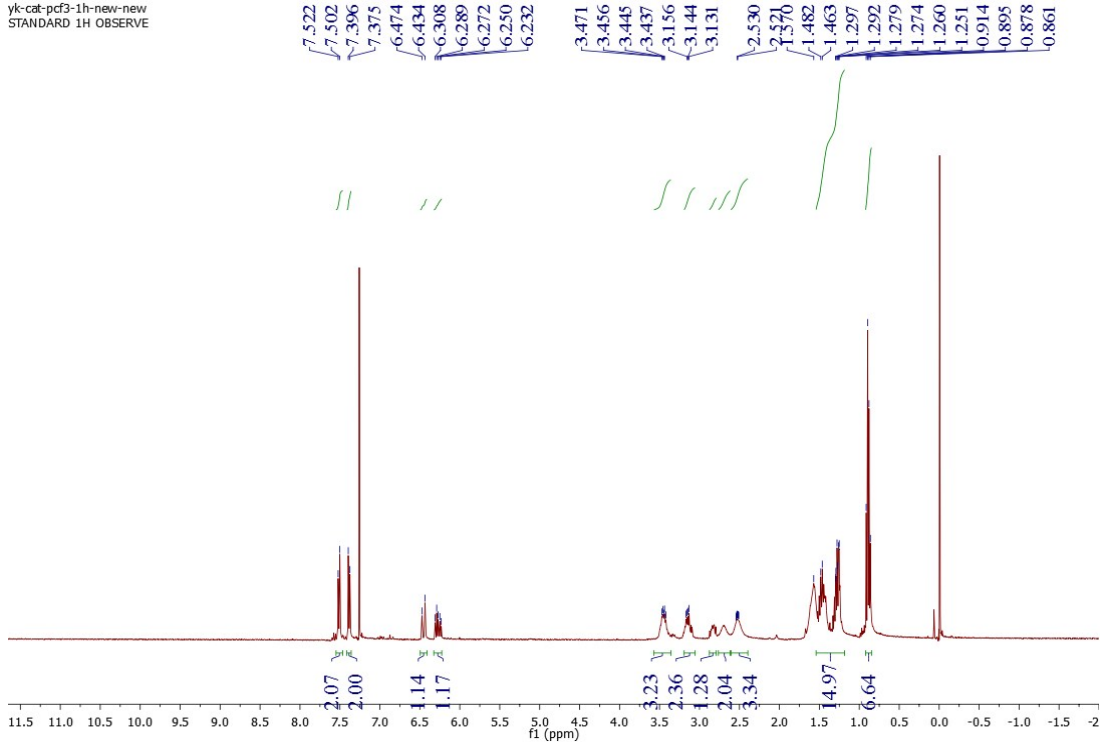


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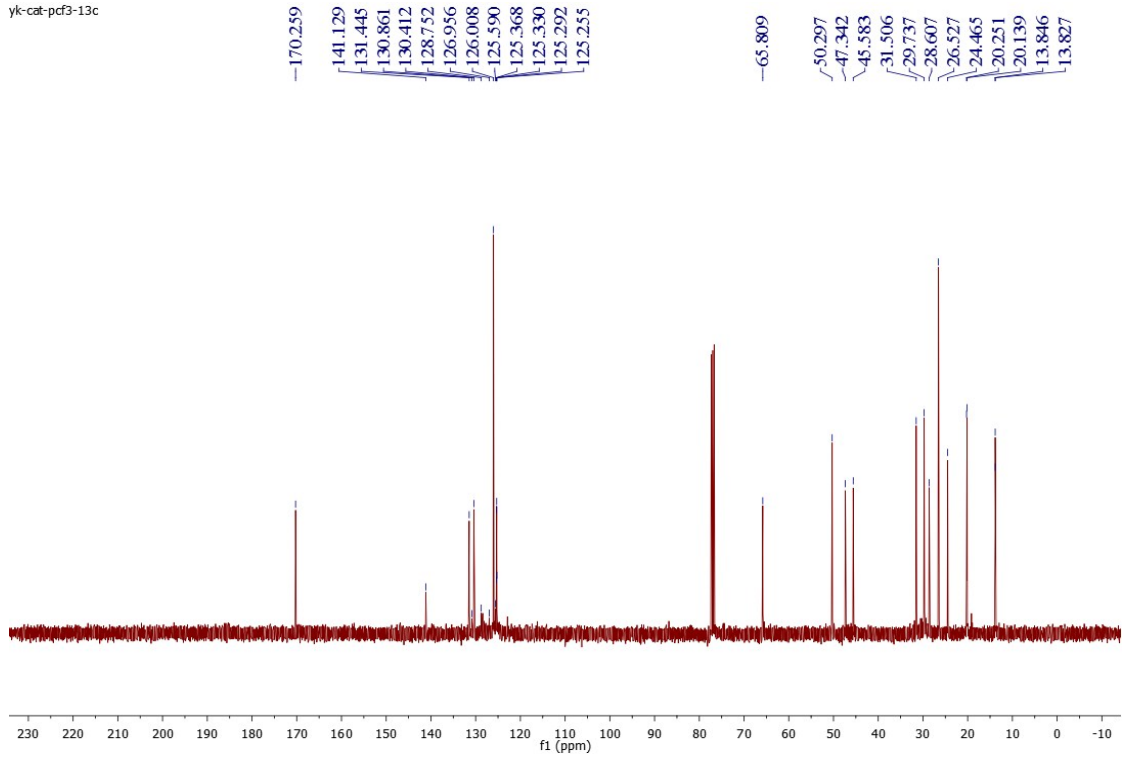


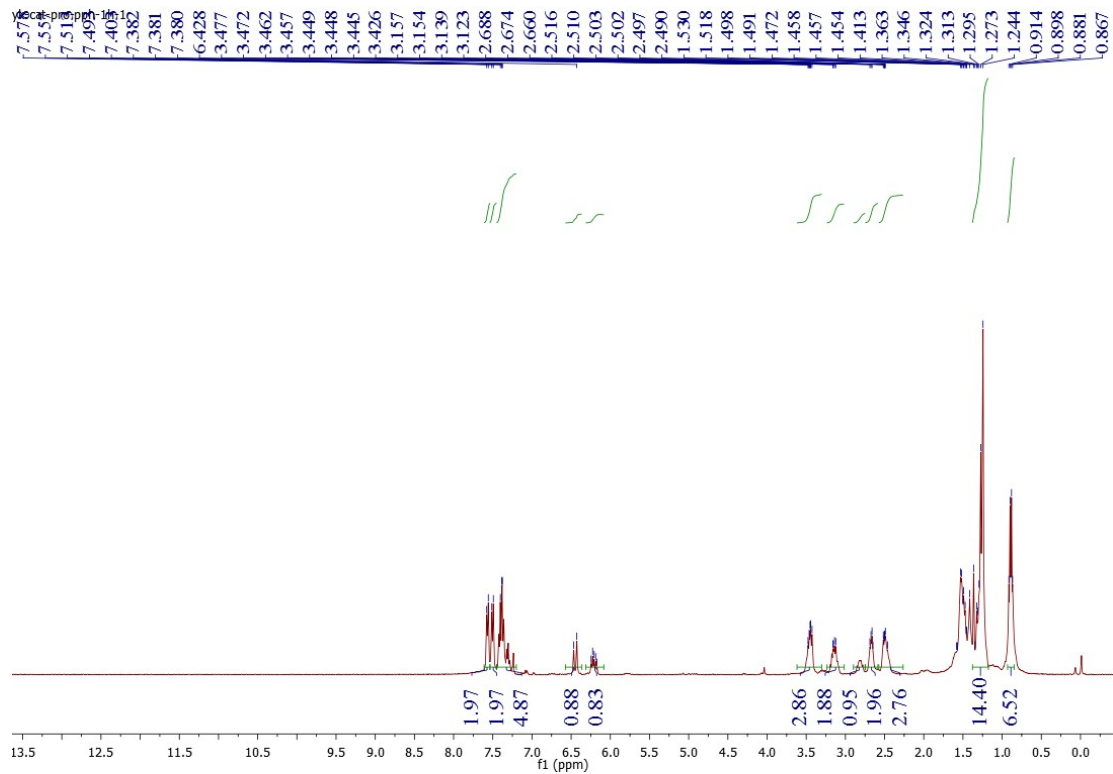
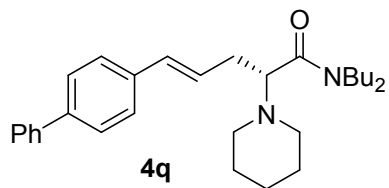


yk-cat-pcf3-1h-new-new
STANDARD 1H OBSERVE



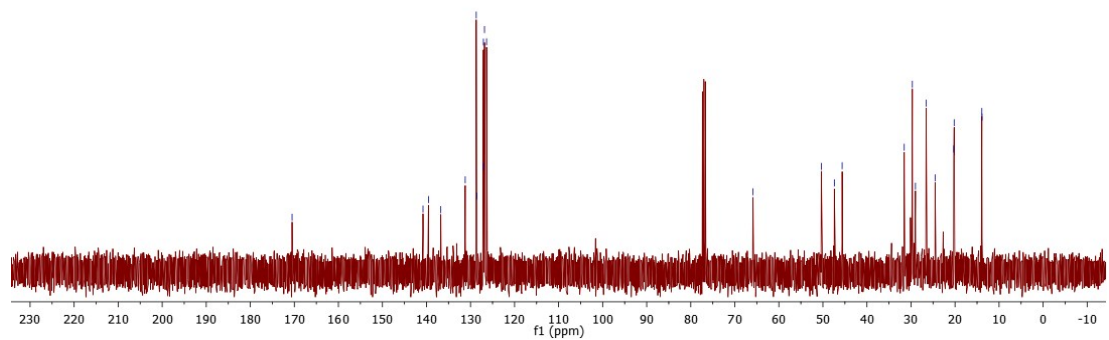
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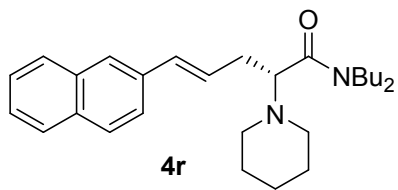




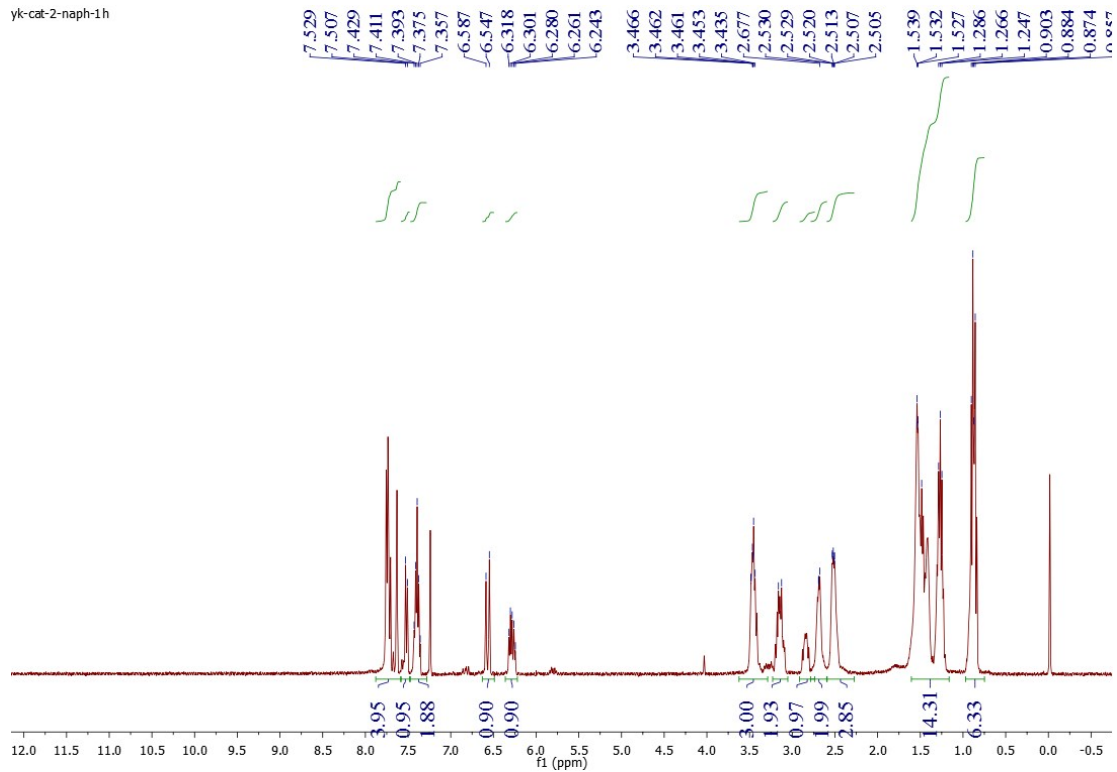
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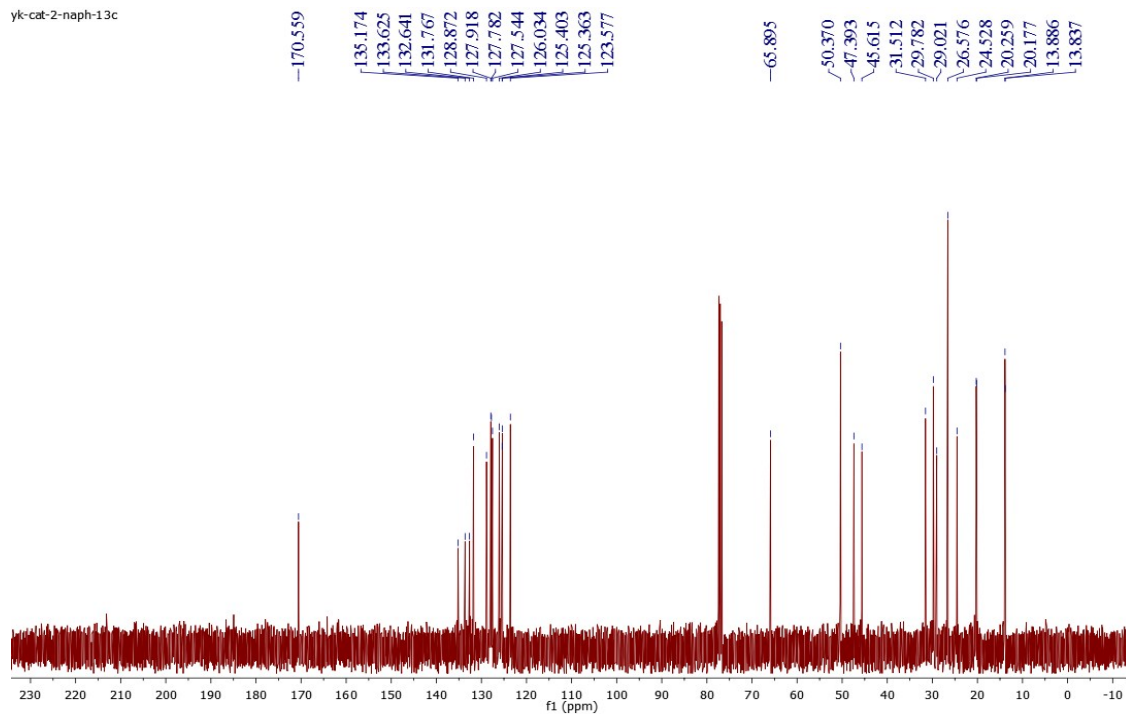


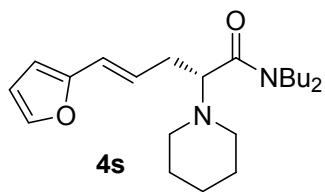


yk-cat-2-naph-1h

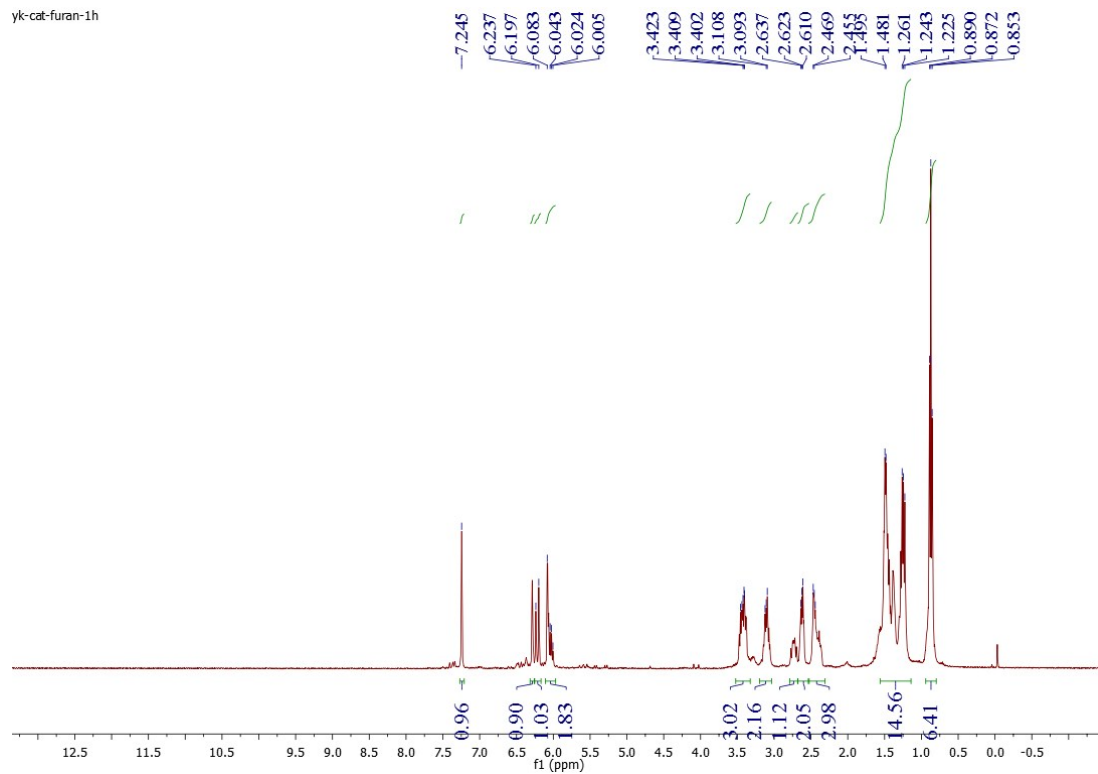


yk-cat-2-naph-13c

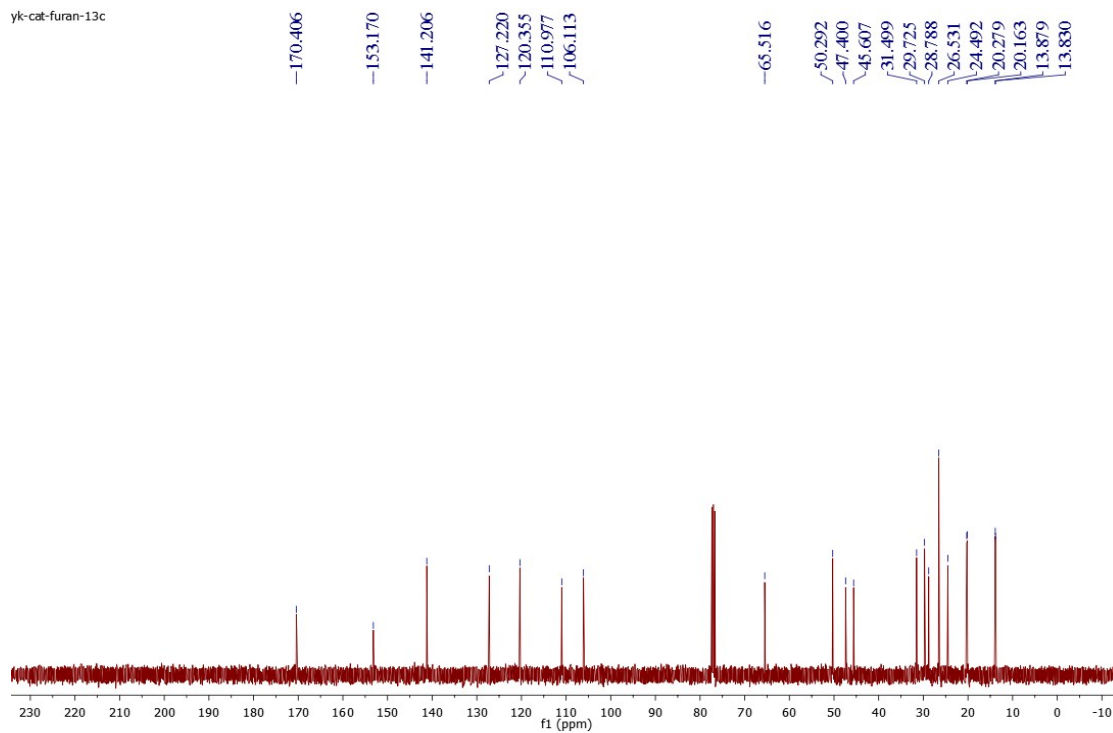


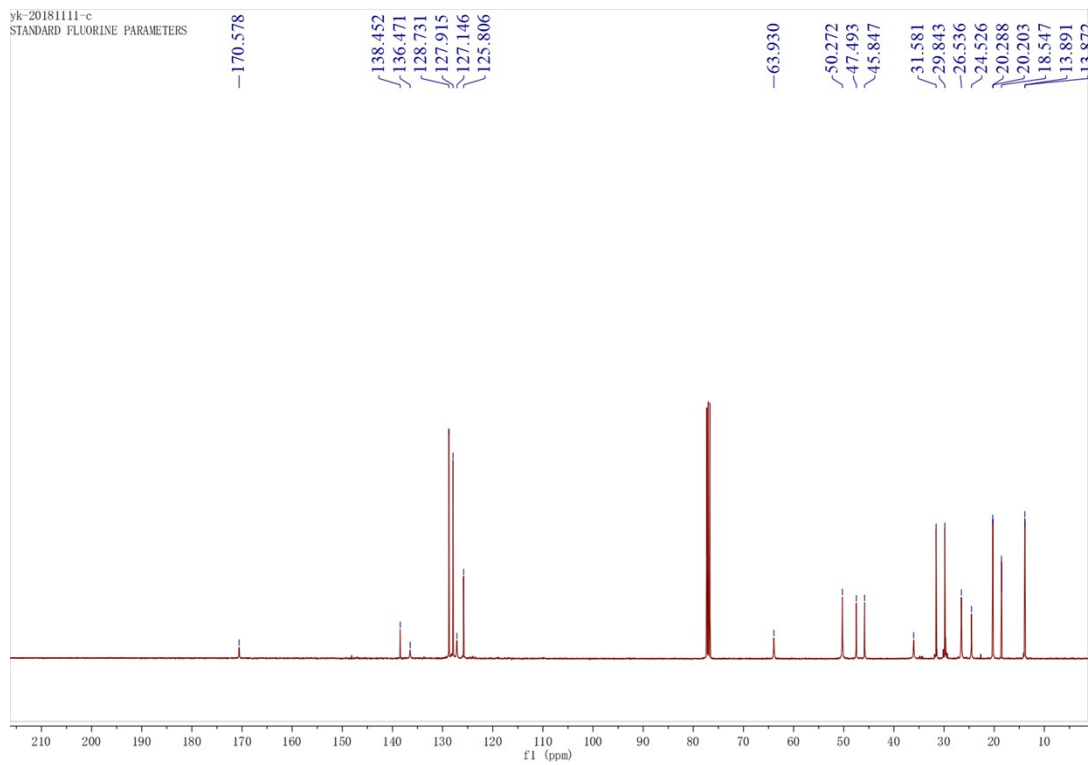
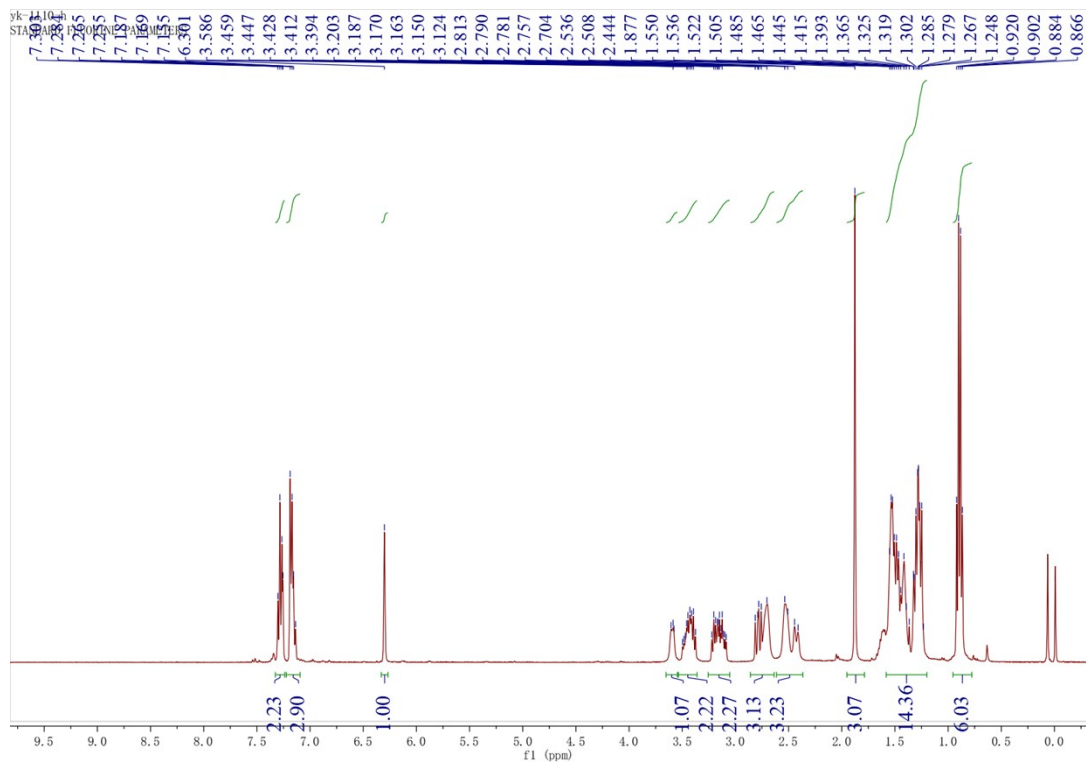
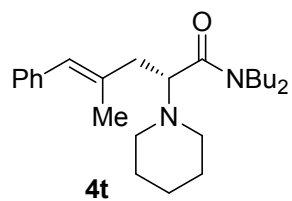


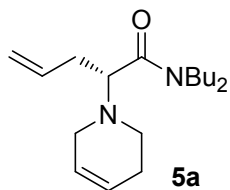
yk-cat-furan-1h



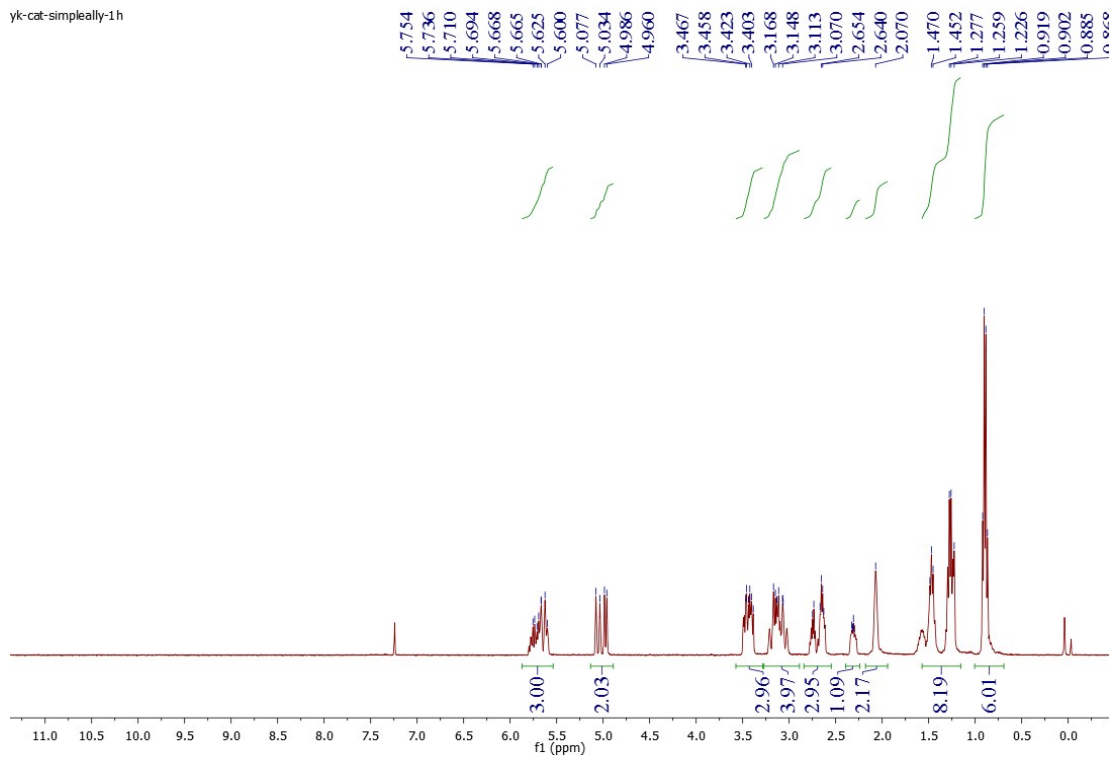
yk-cat-furan-13c



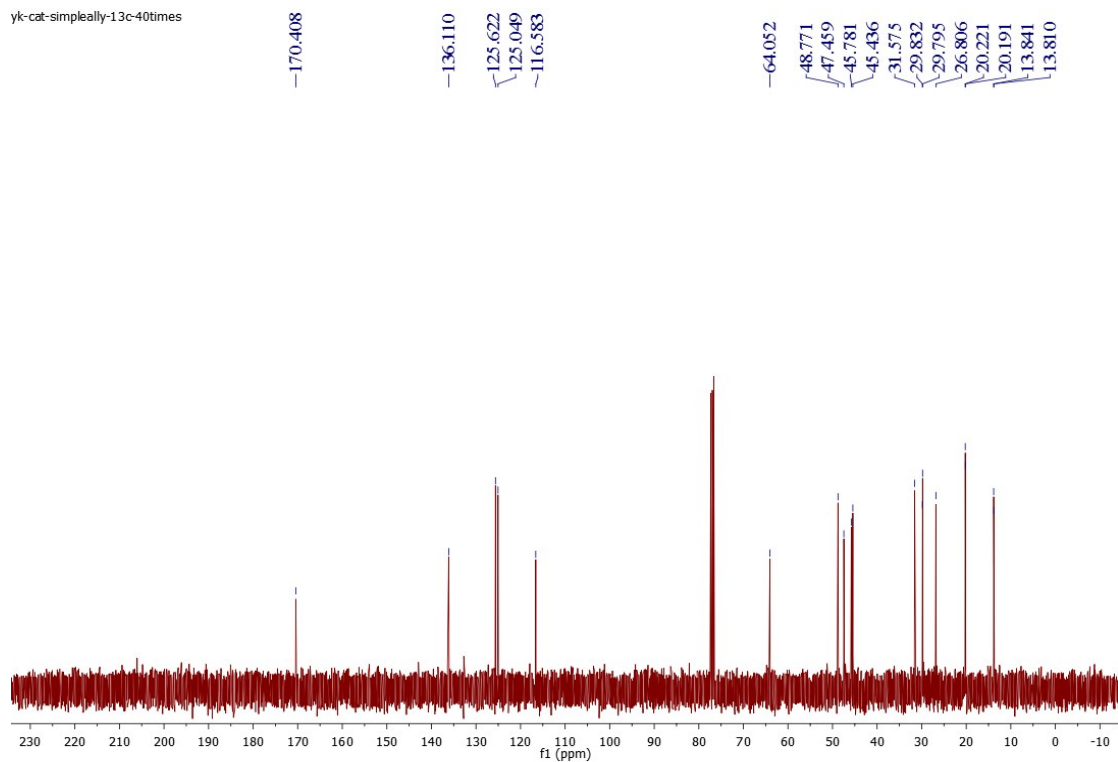


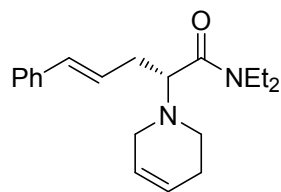


yk-cat-simply-1h

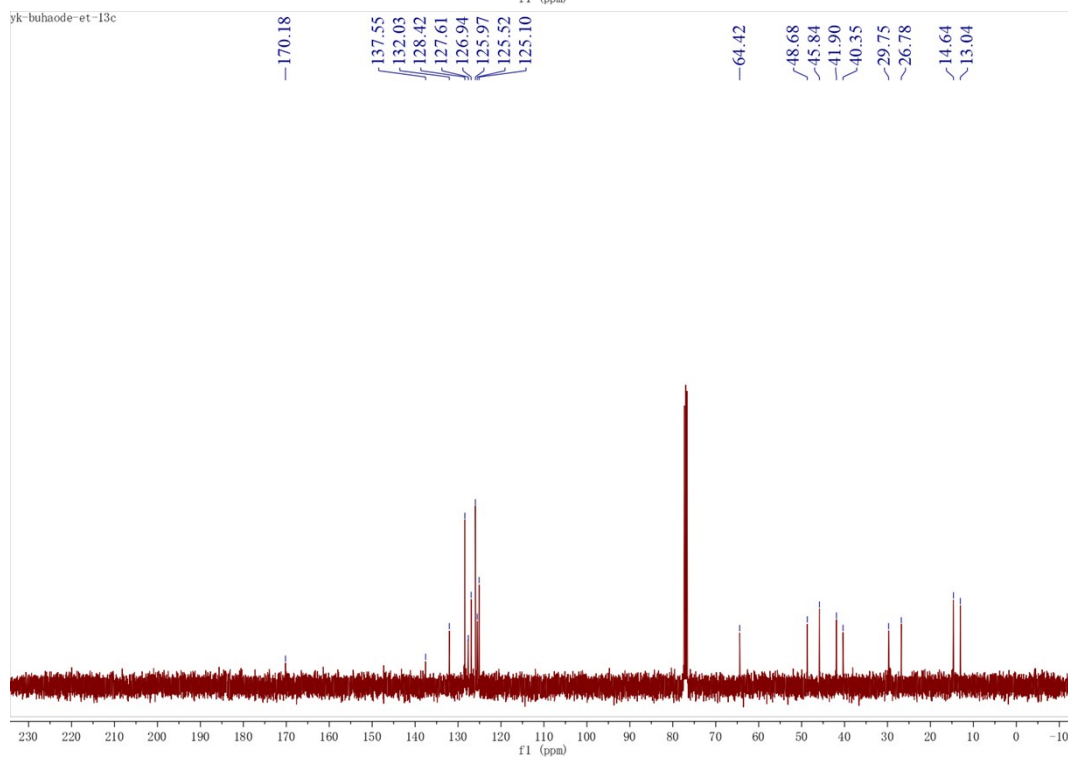
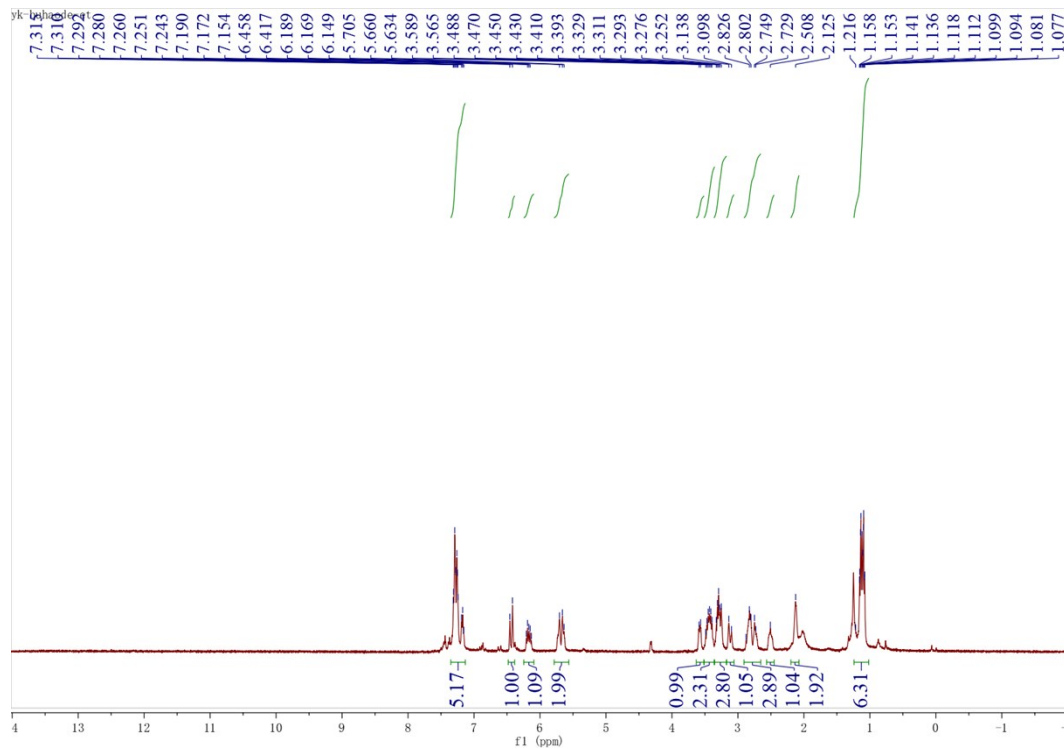


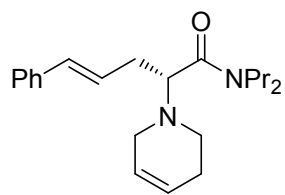
yk-cat-simply-13c-40times



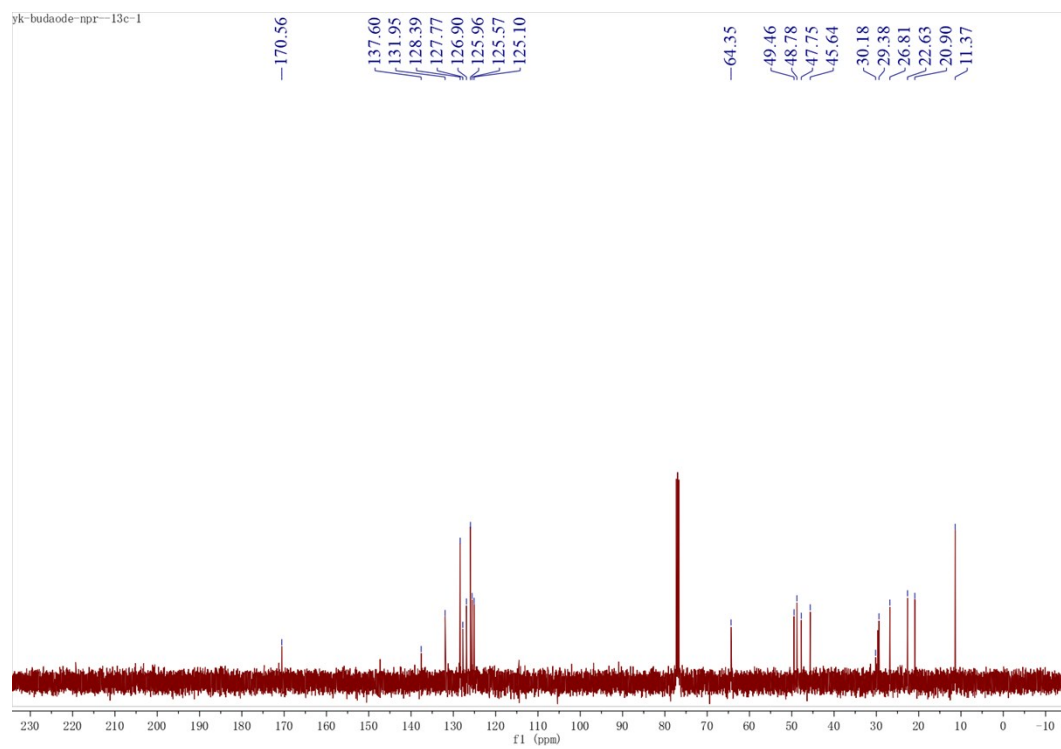
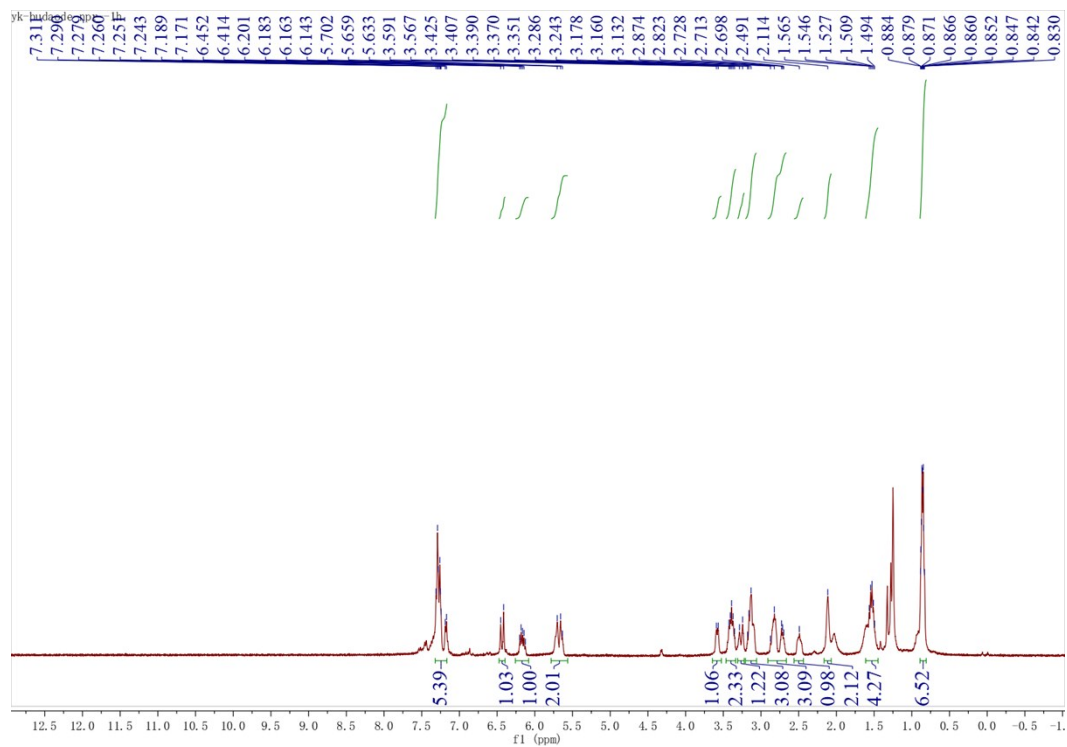


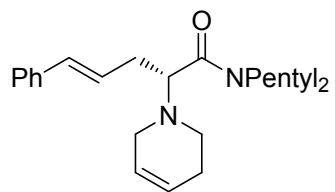
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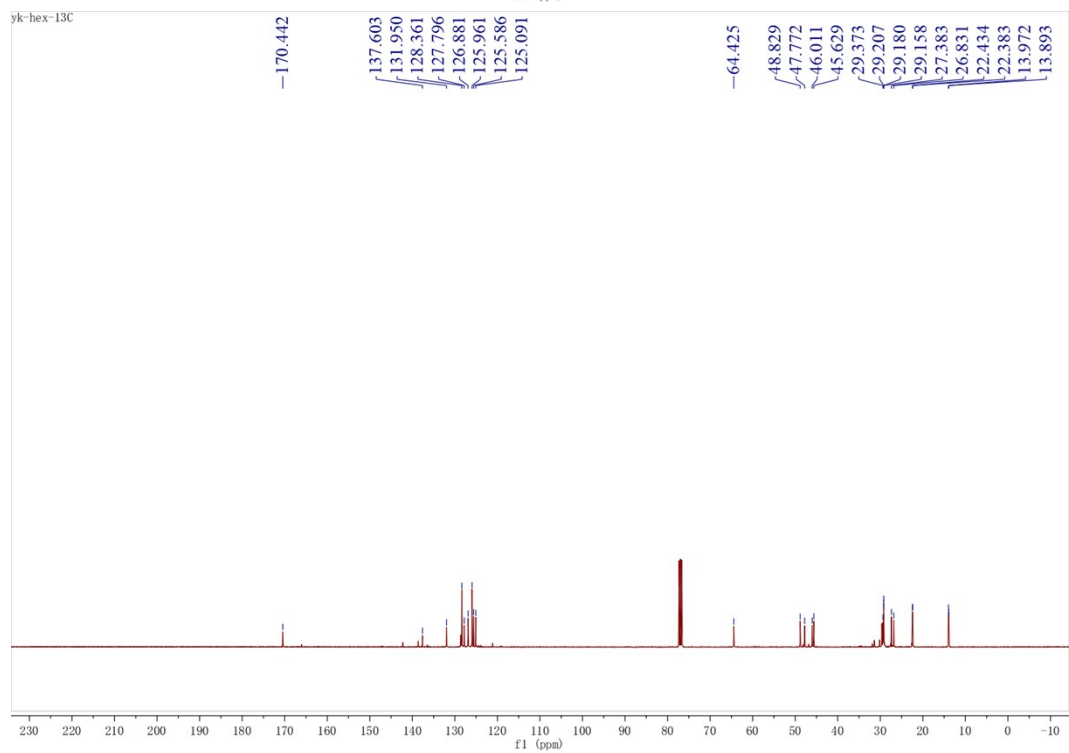
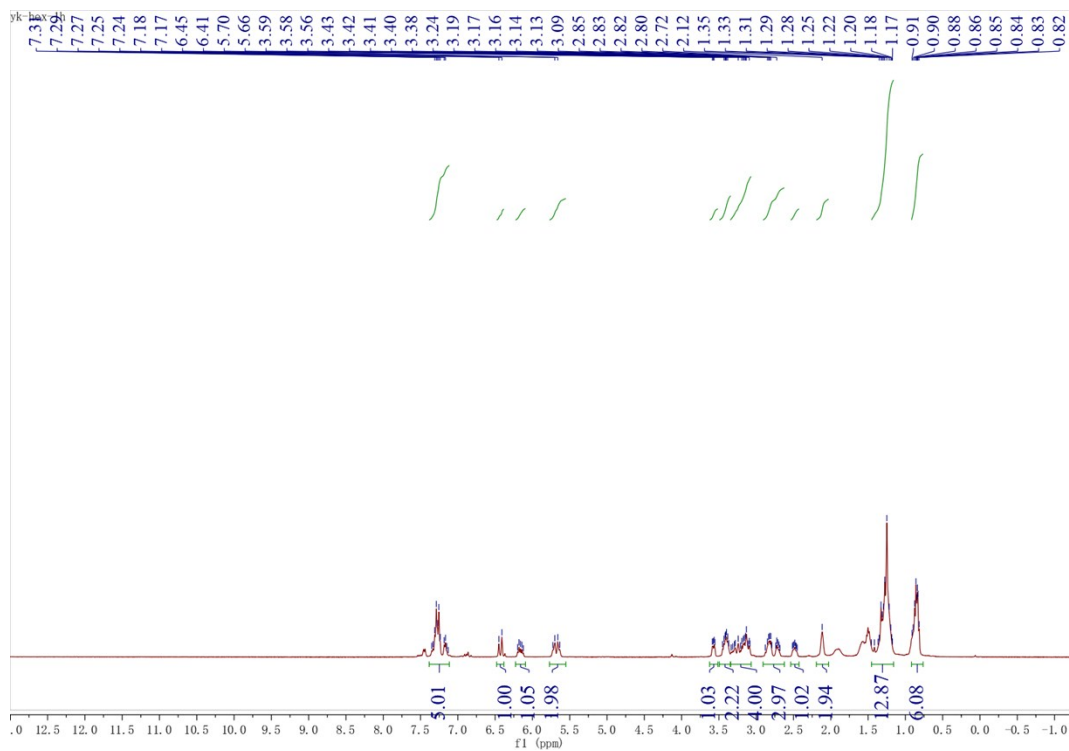


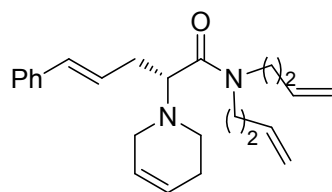
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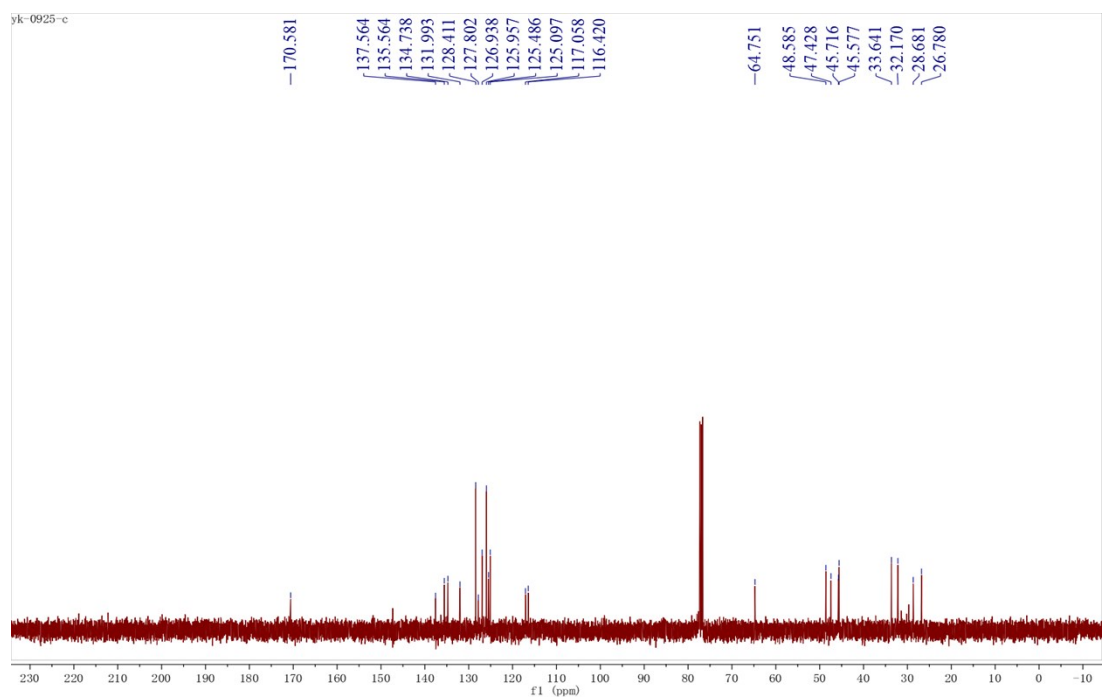
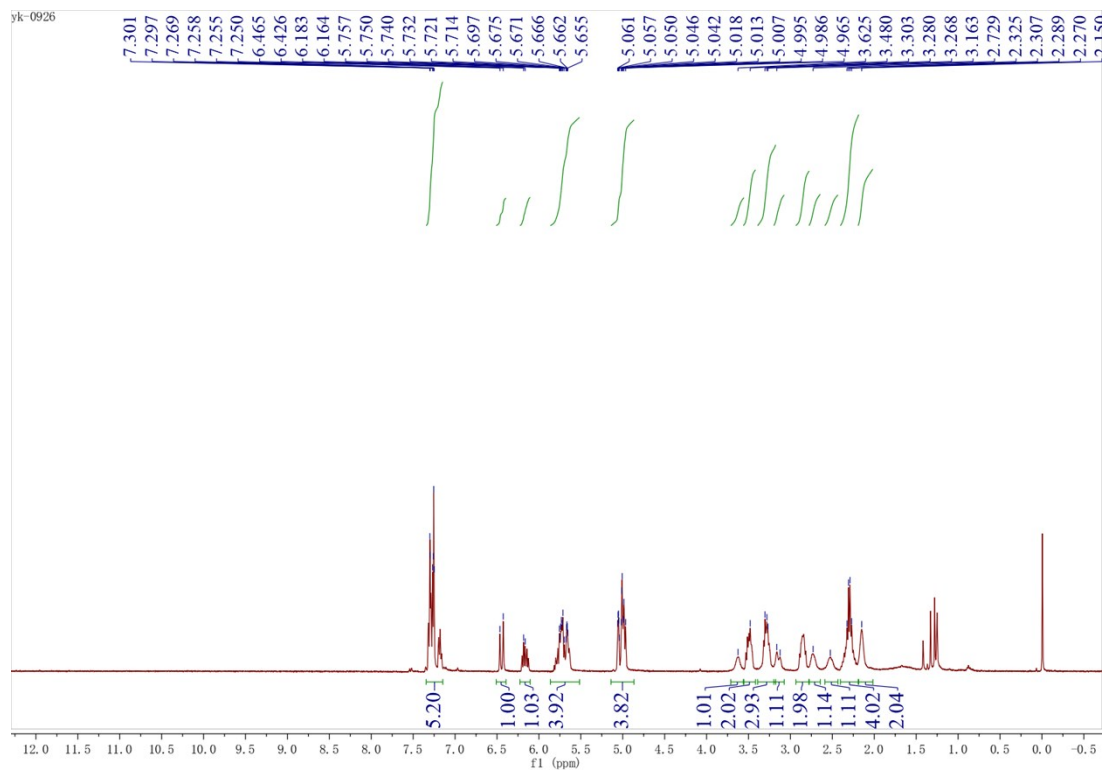


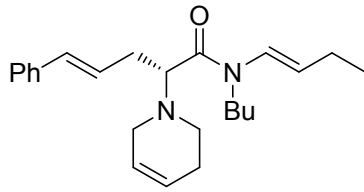
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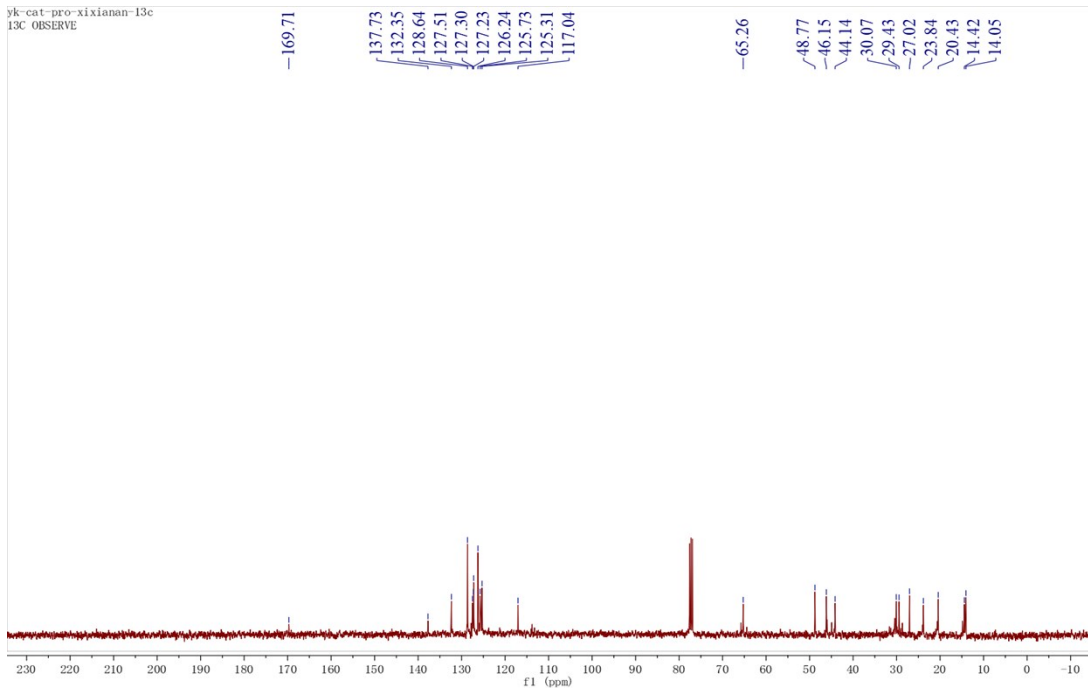
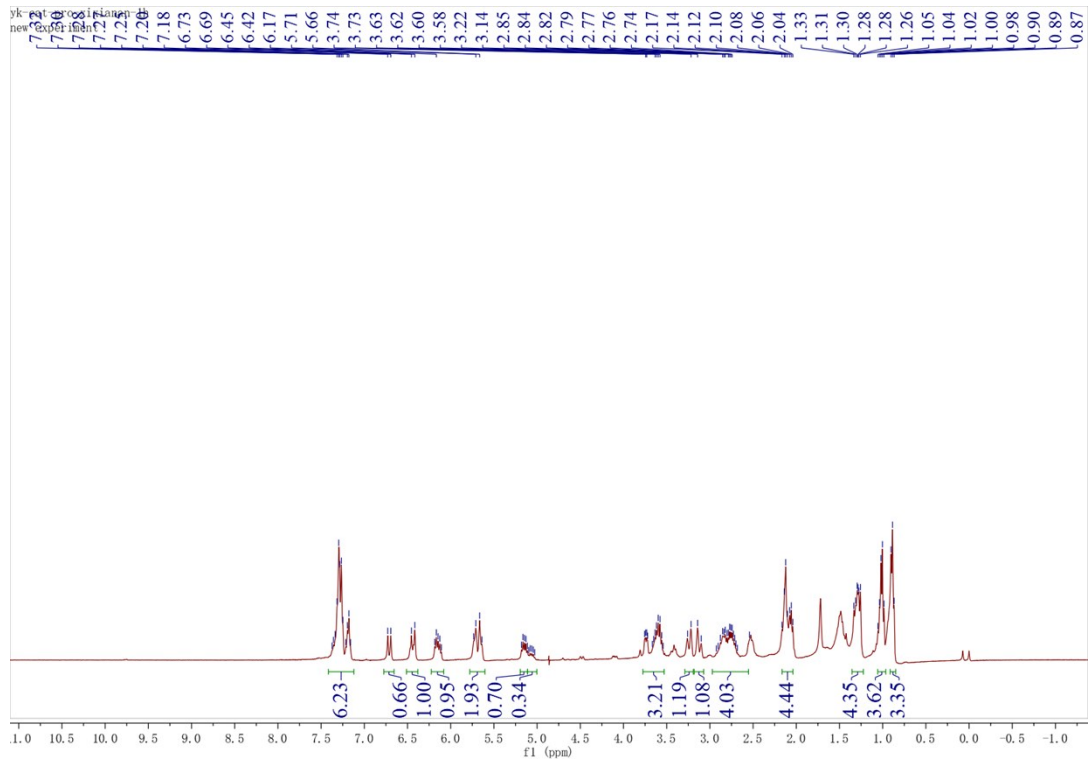


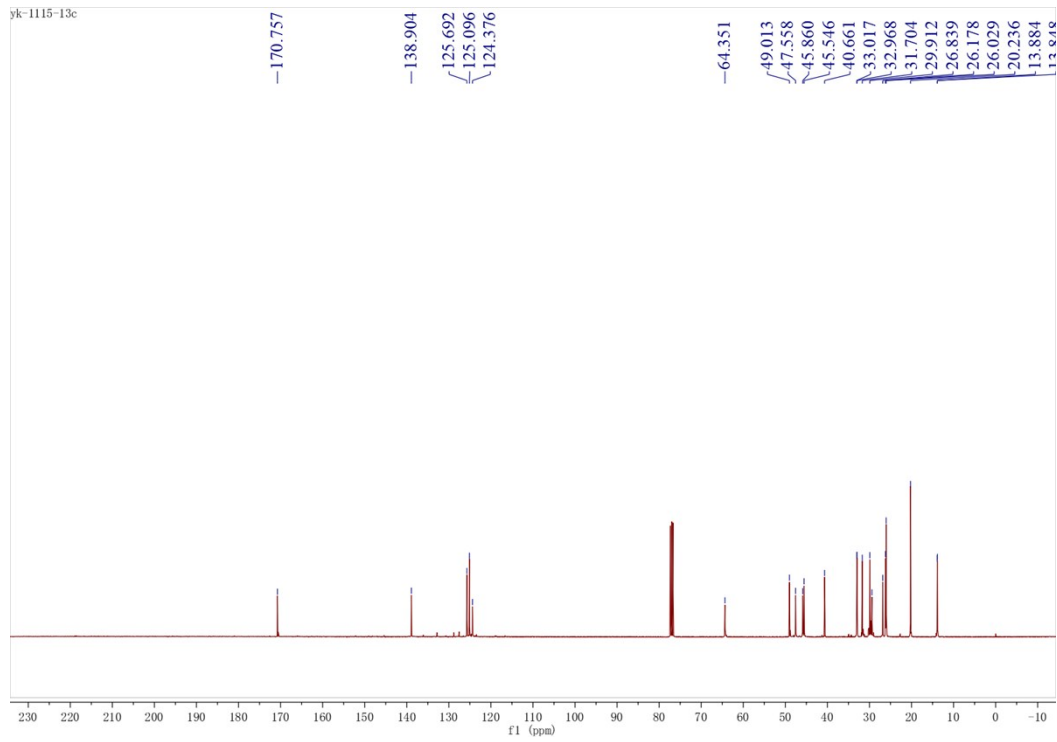
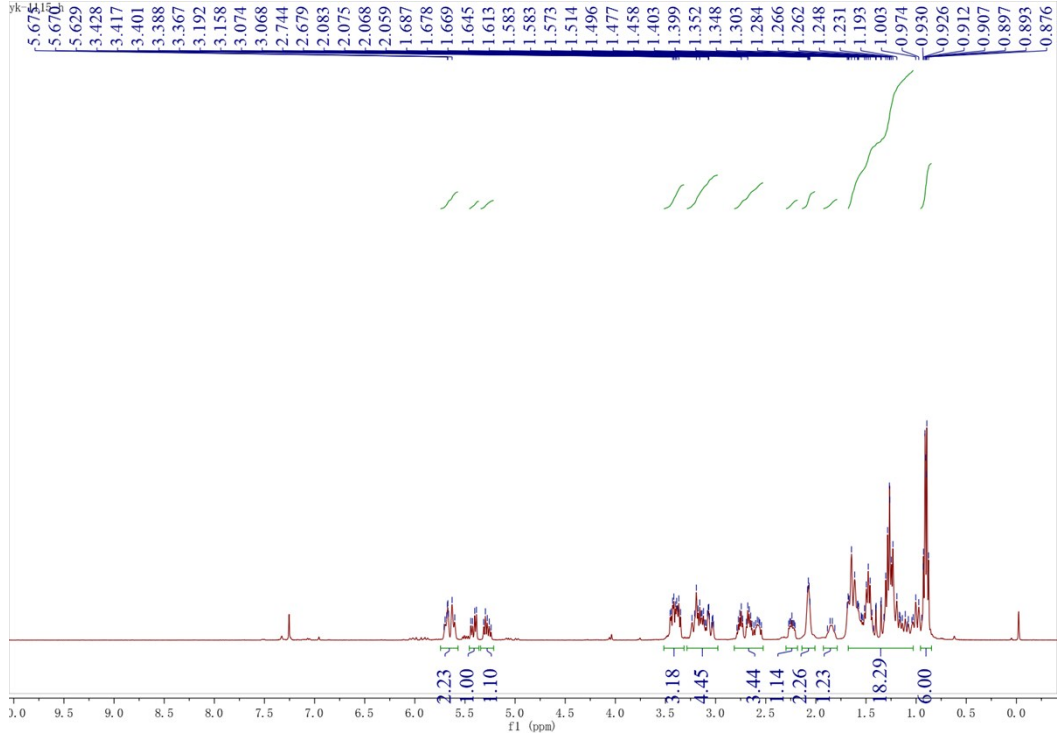
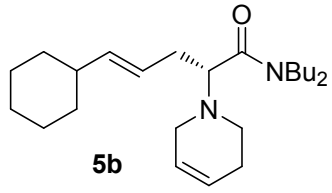
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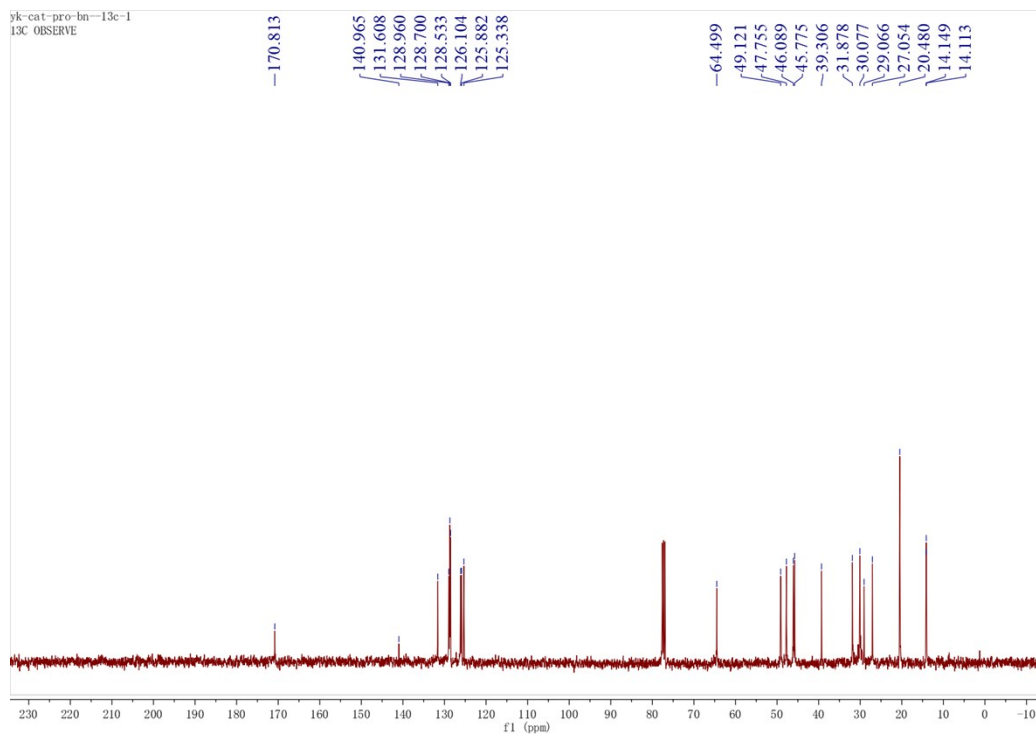
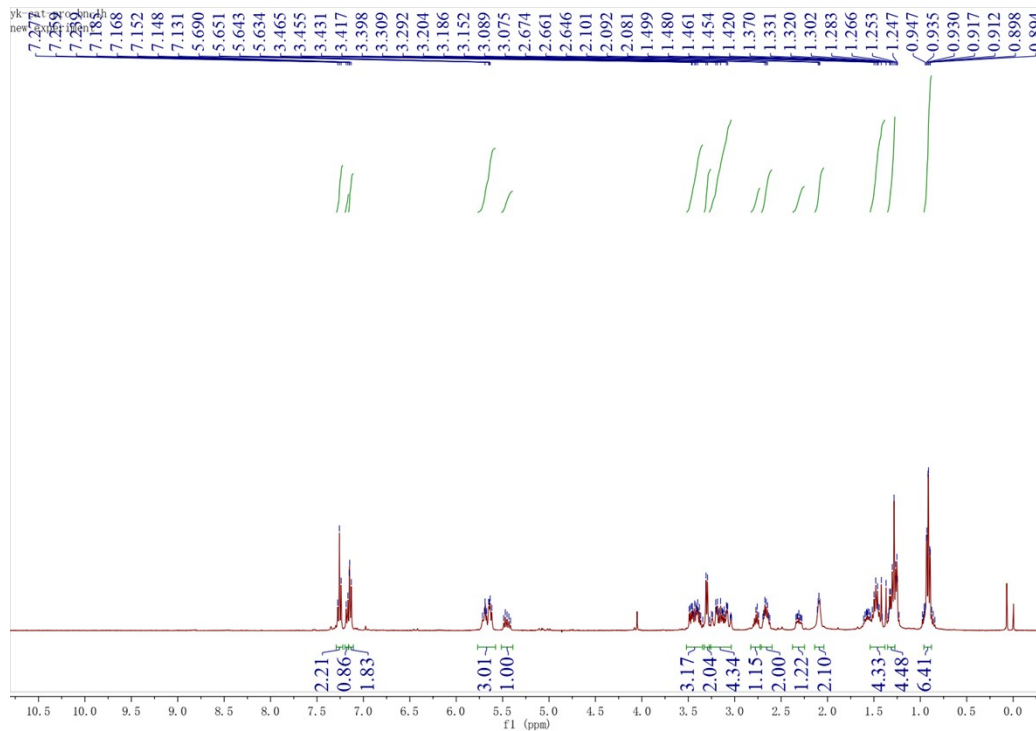
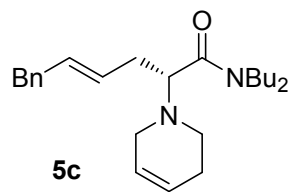


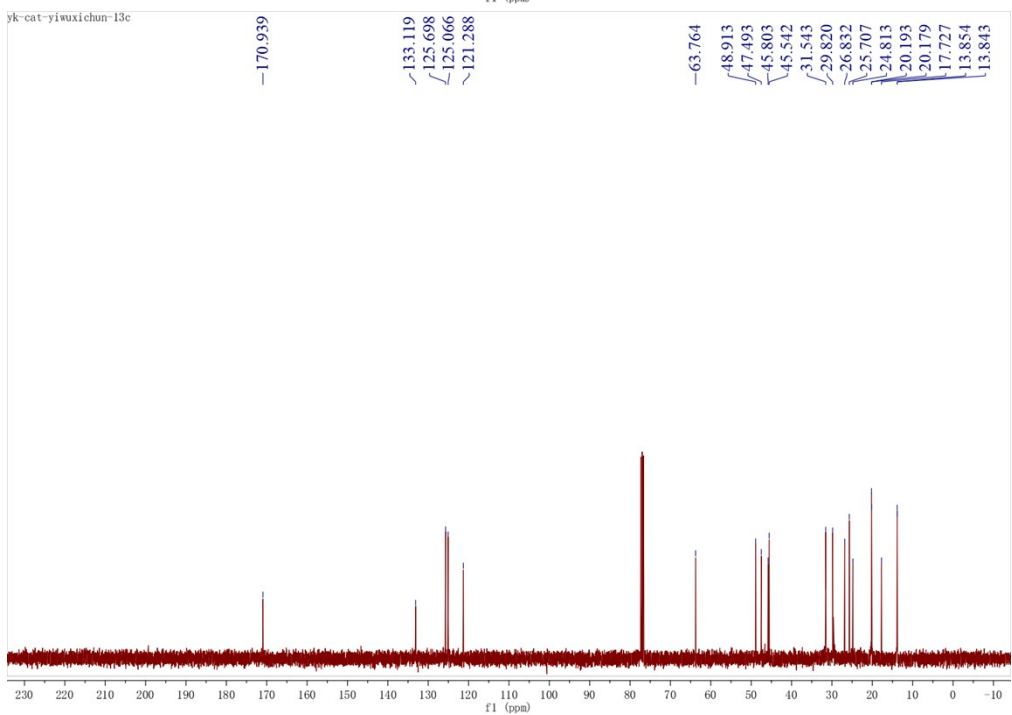
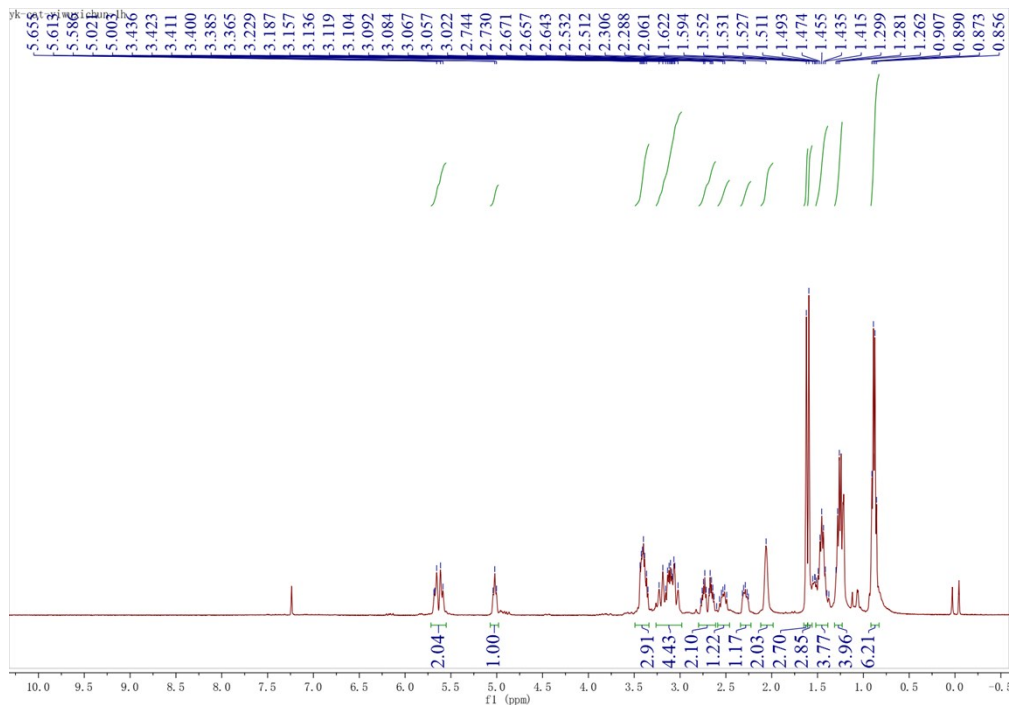
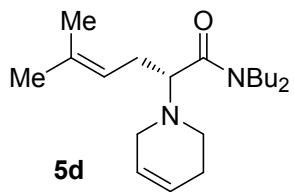


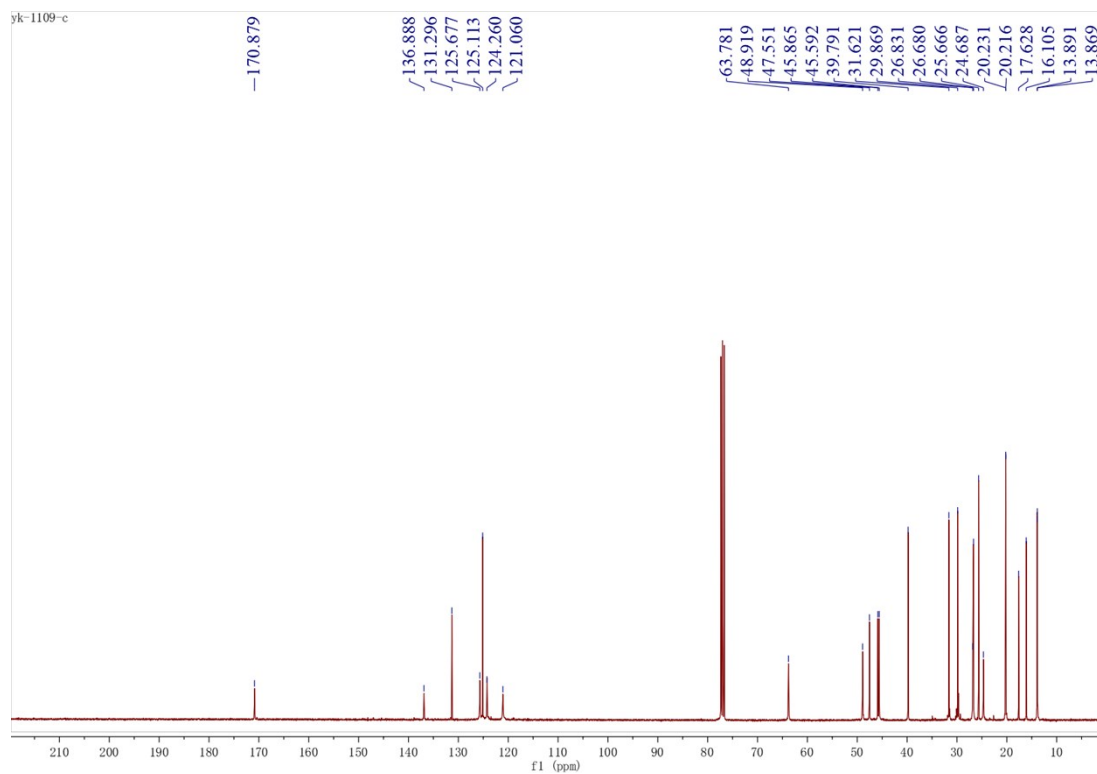
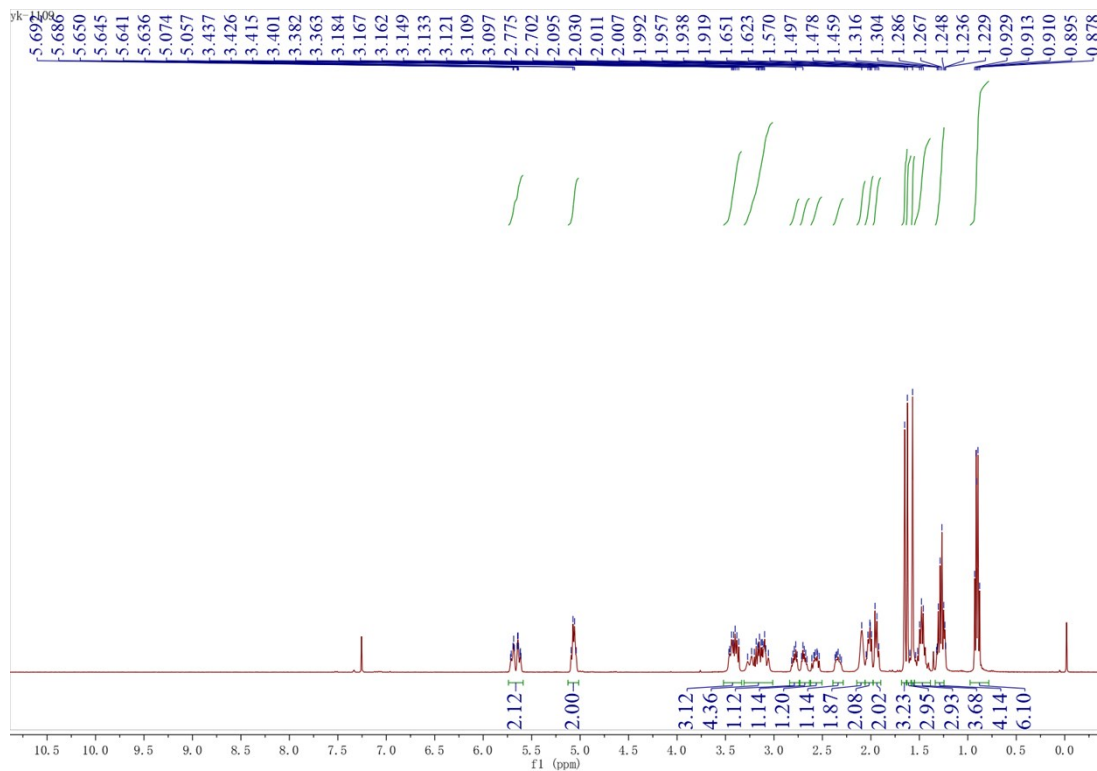
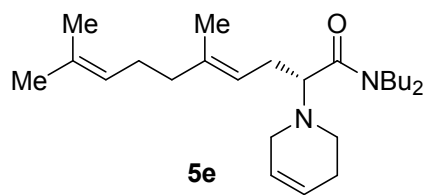
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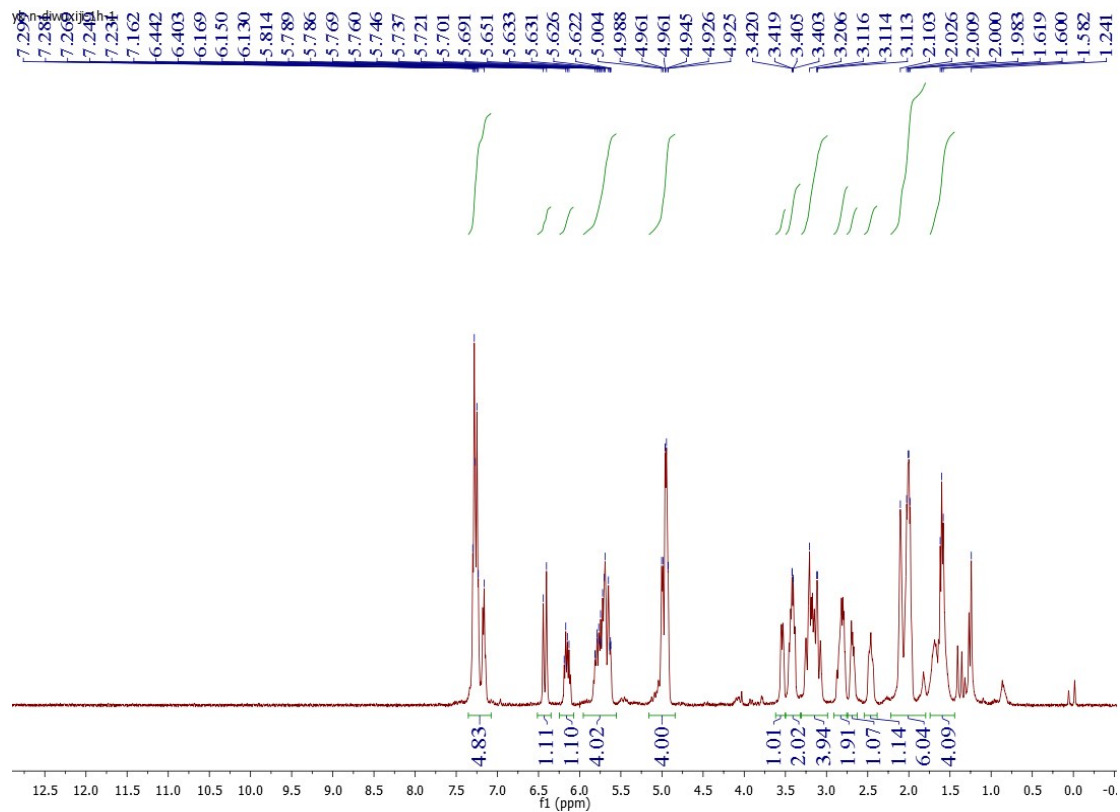
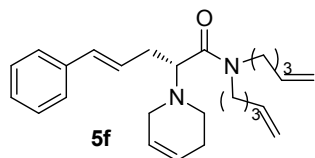




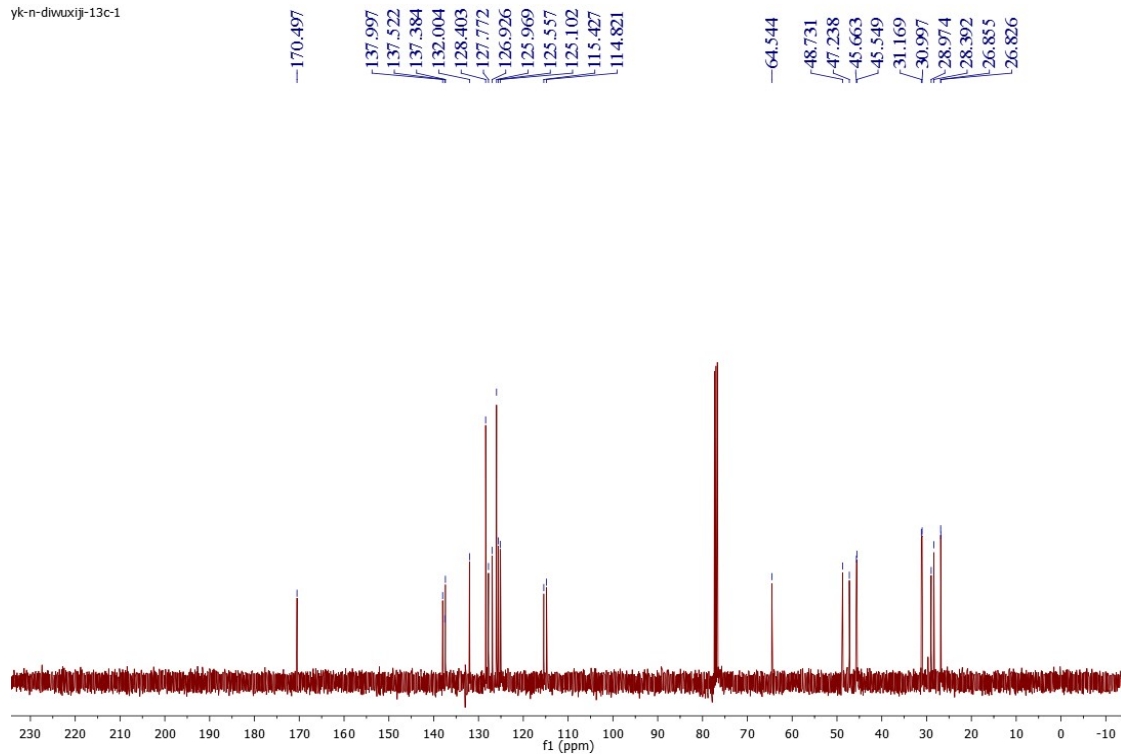


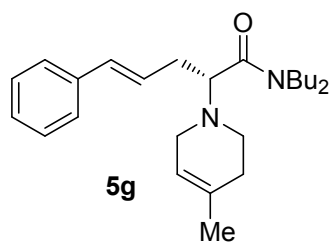




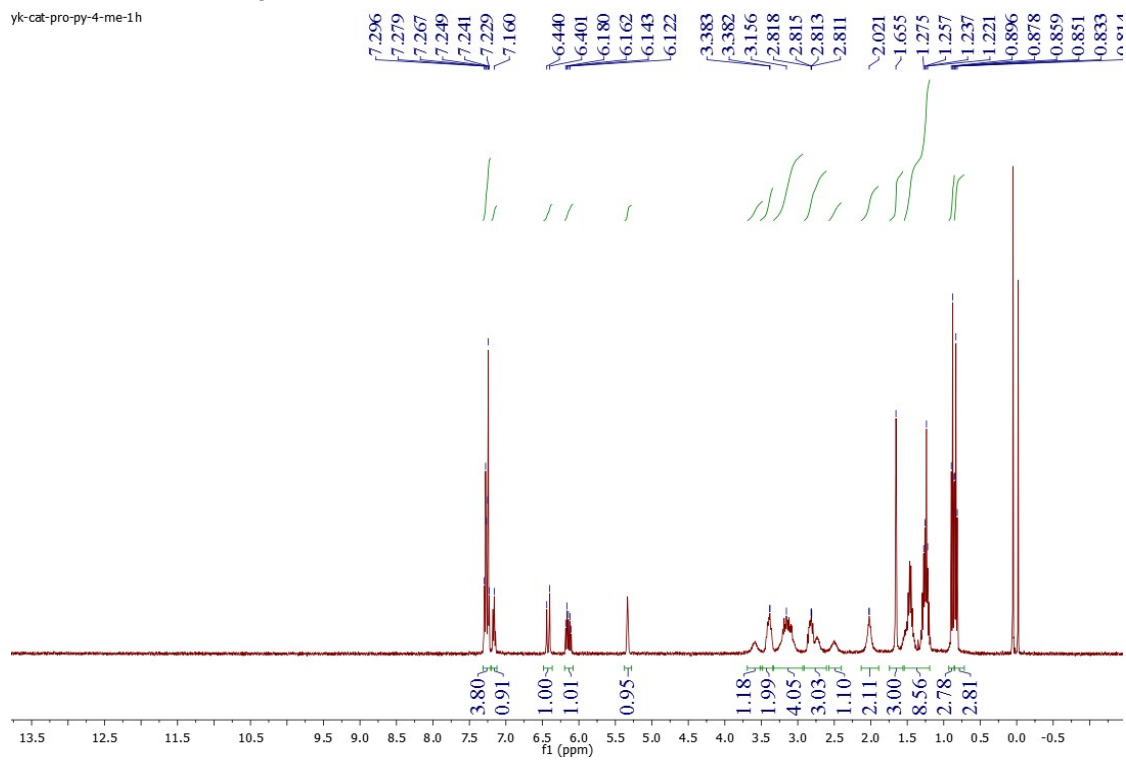


yk-n-divuxij-13c-1

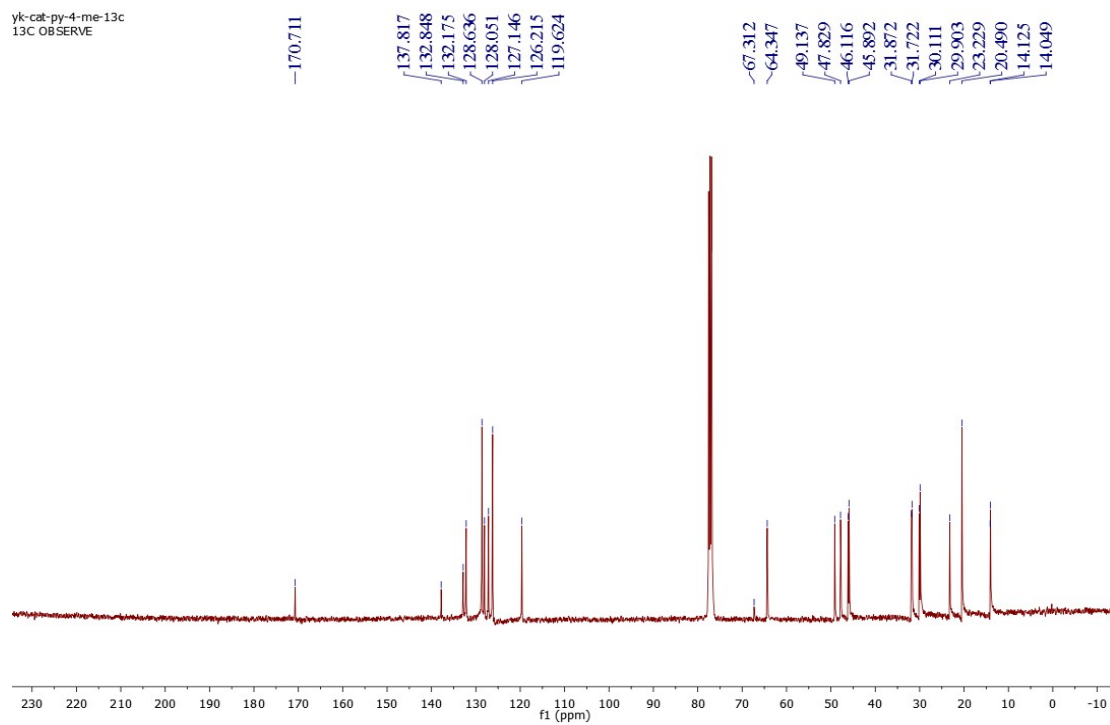


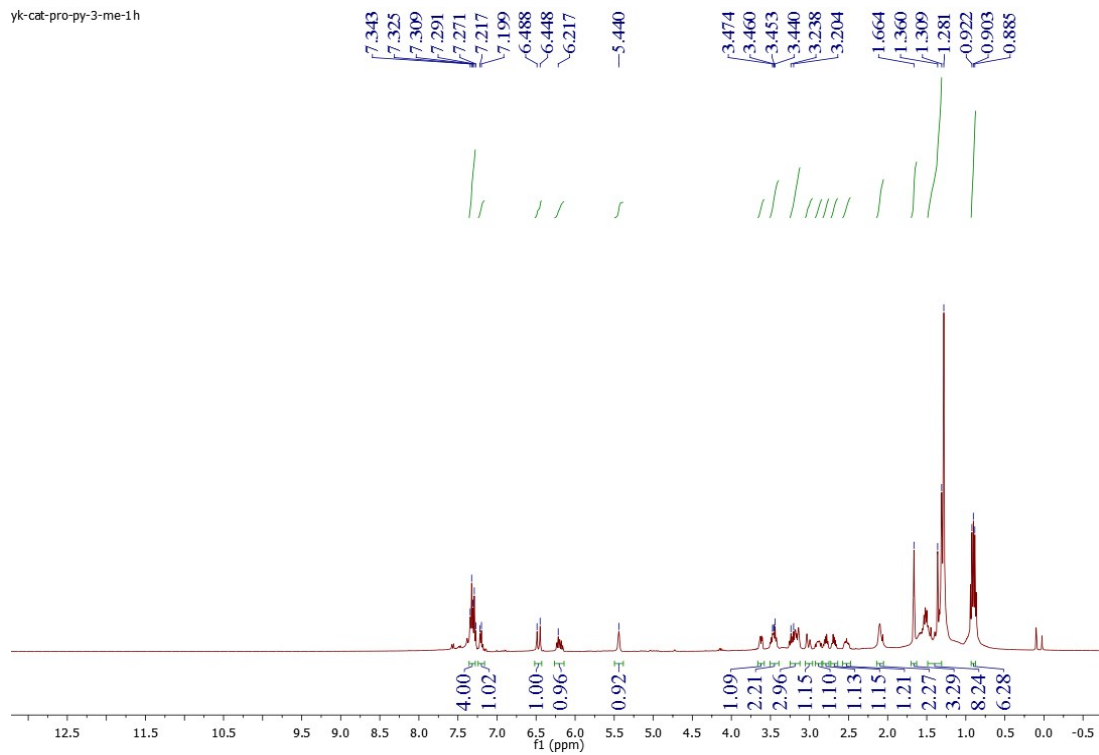
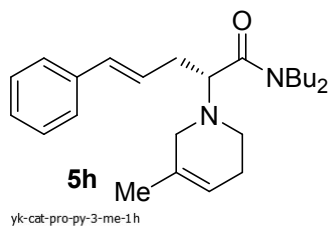


yk-cat-pro-py-4-me-1h

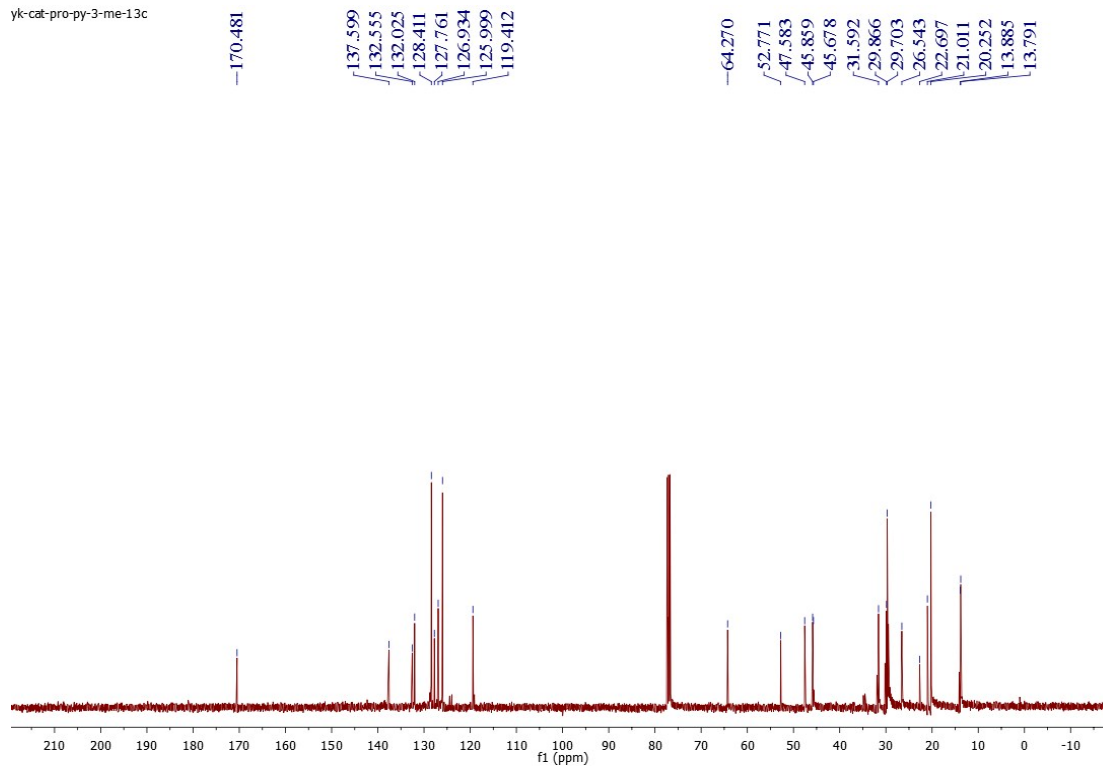


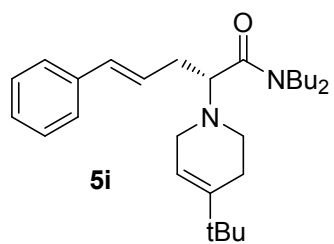
yk-cat-py-4-me-13c
13C OBSERVE



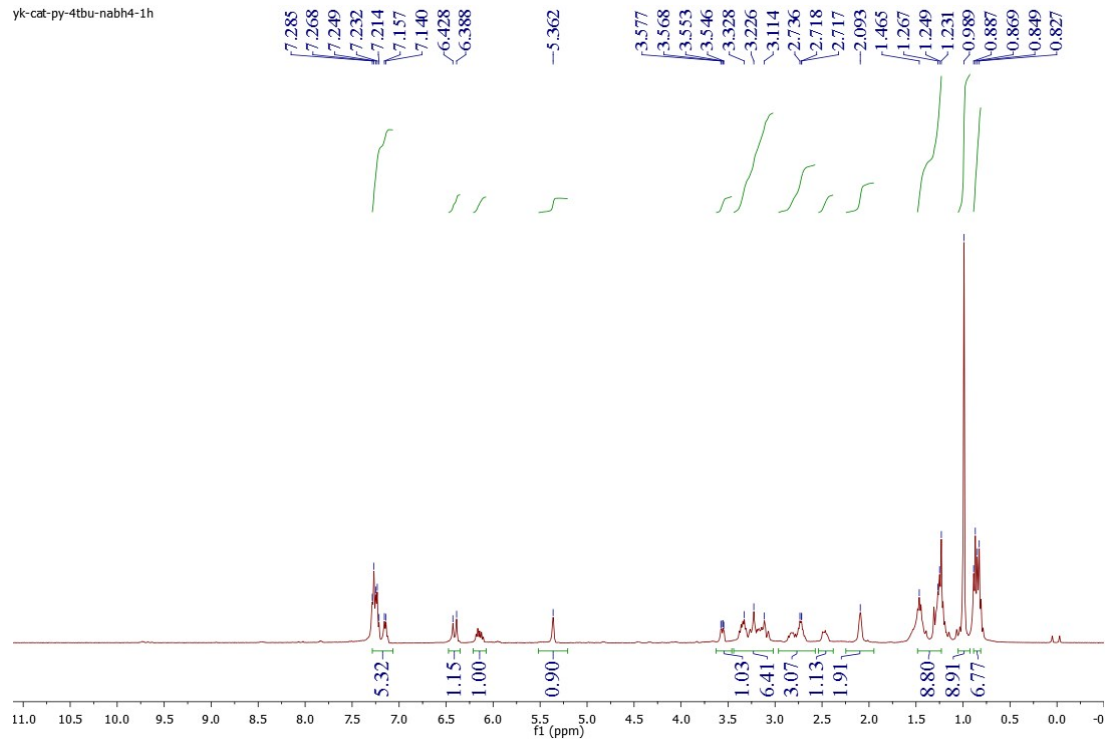


yk-cat-pro-py-3-me-13c

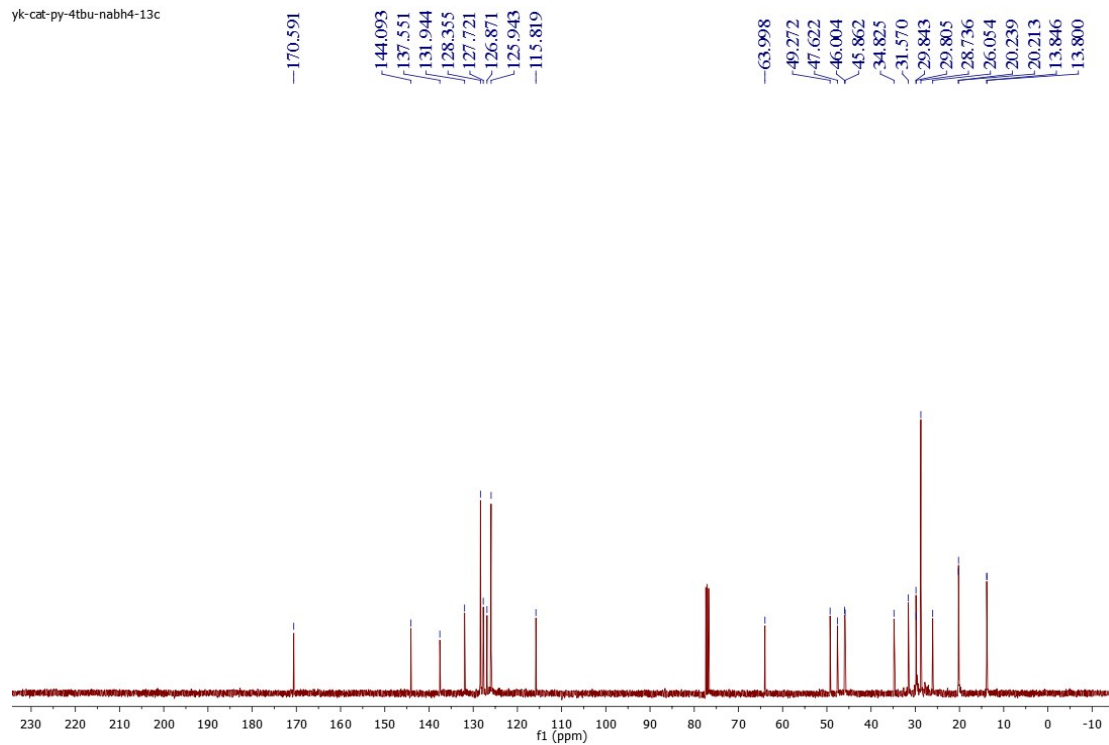


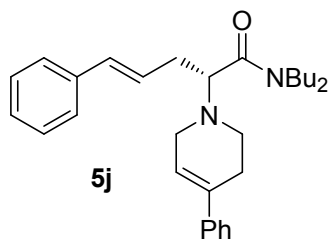


yk-cat-py-4tbu-nabh4-1h

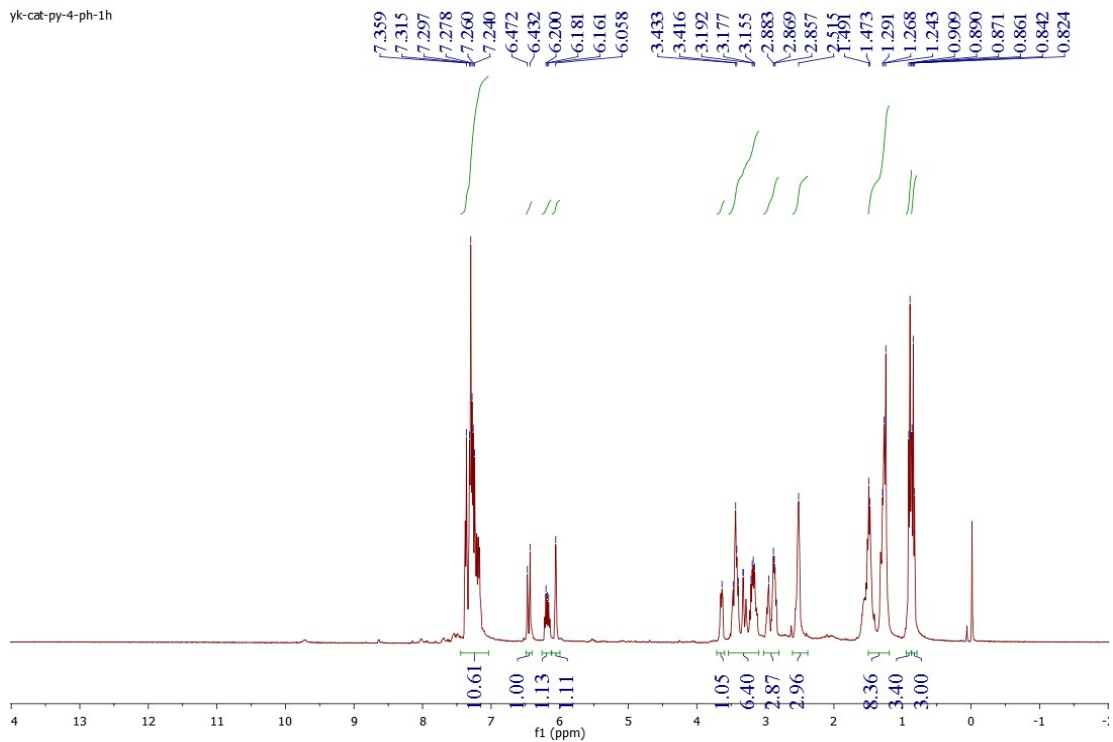


yk-cat-py-4tbu-nabh4-13c

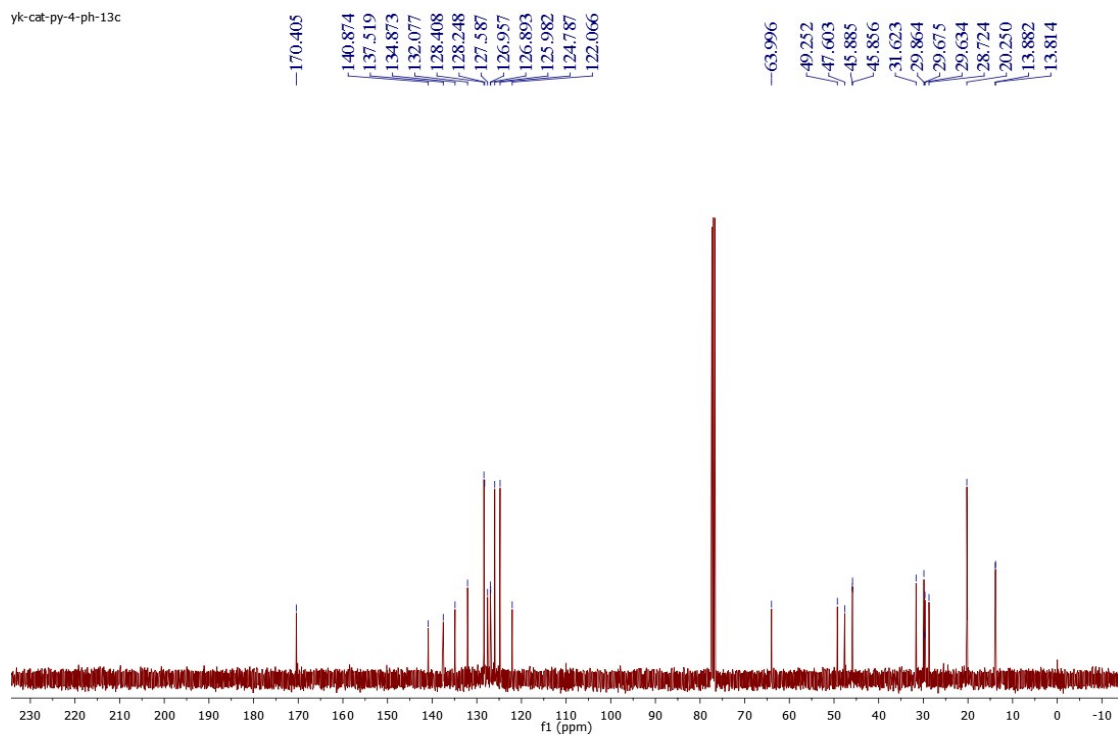


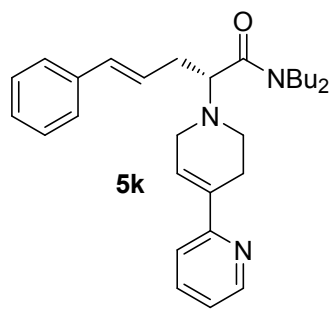


yk-cat-py-4-ph-1h

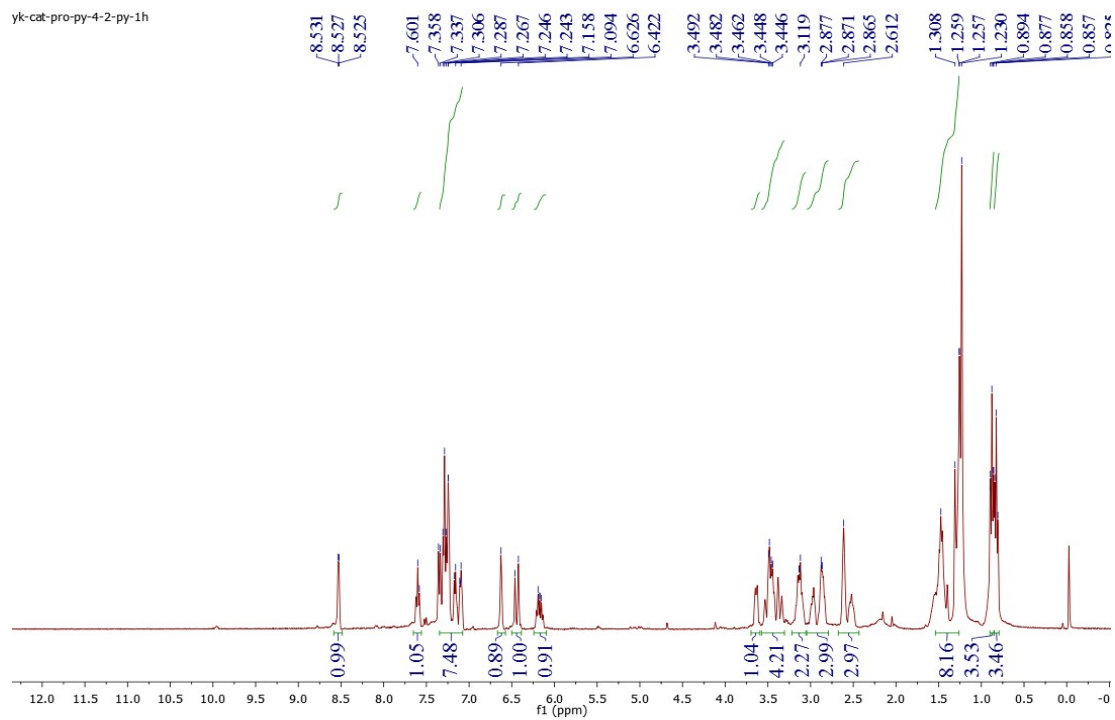


yk-cat-py-4-ph-13c

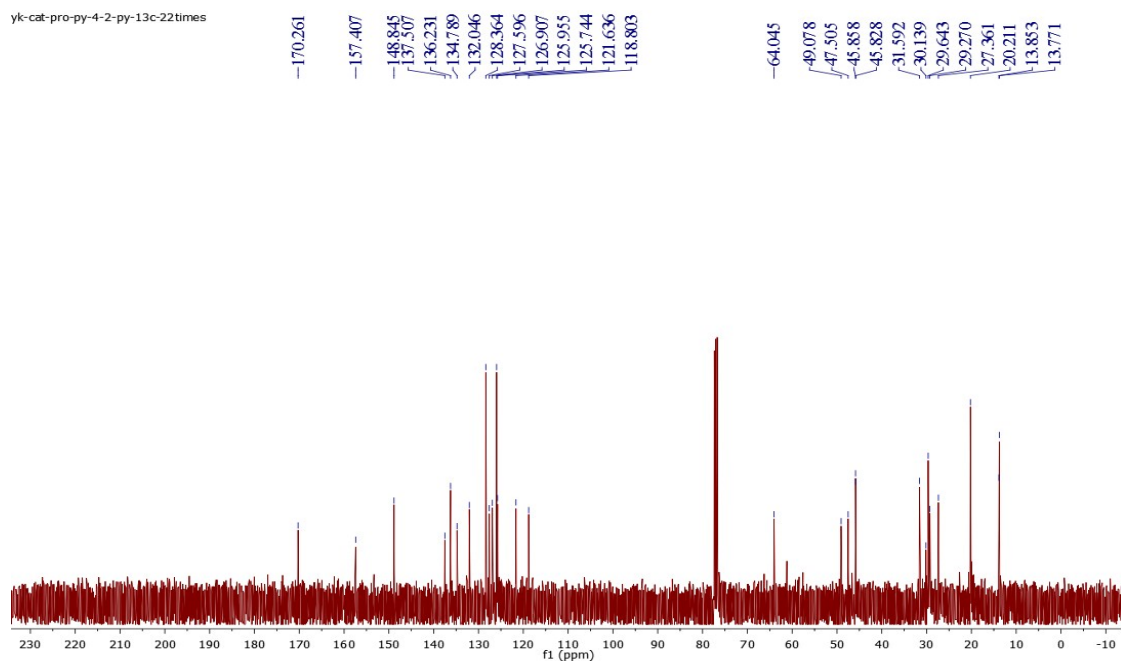


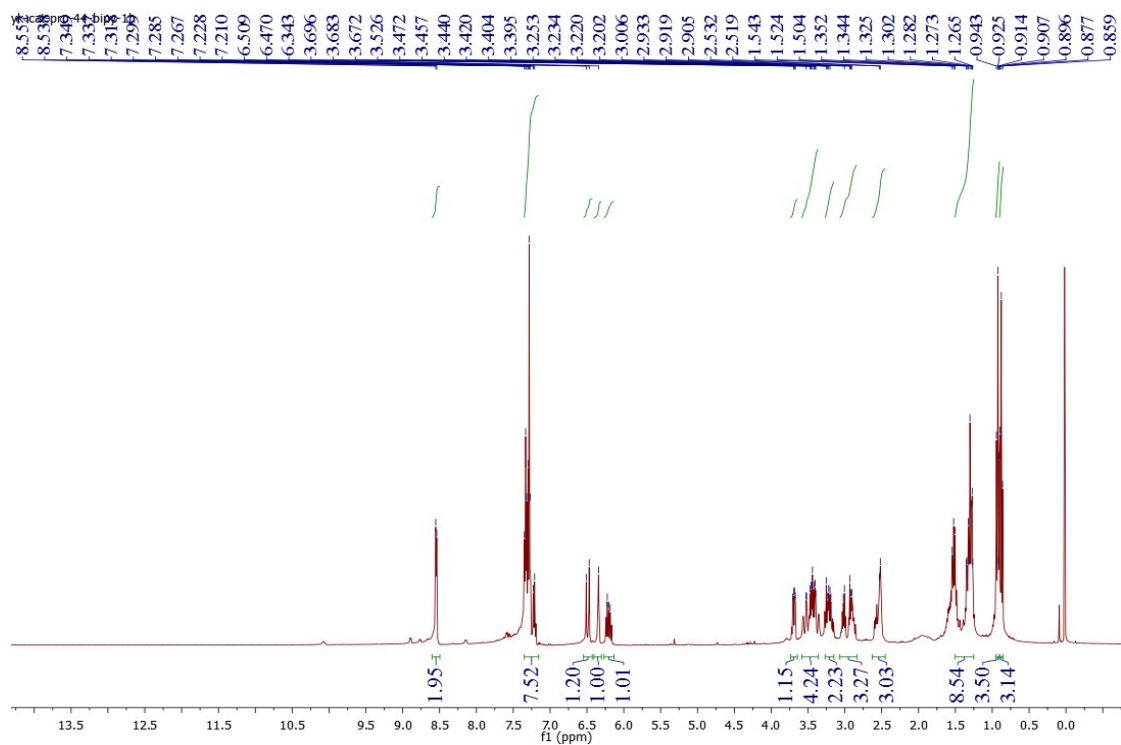
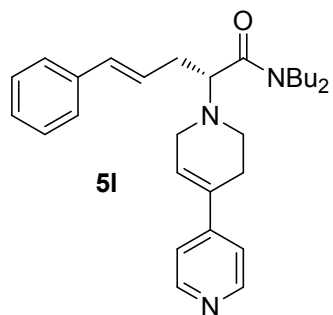


yk-cat-pro-py-4-2-py-1h

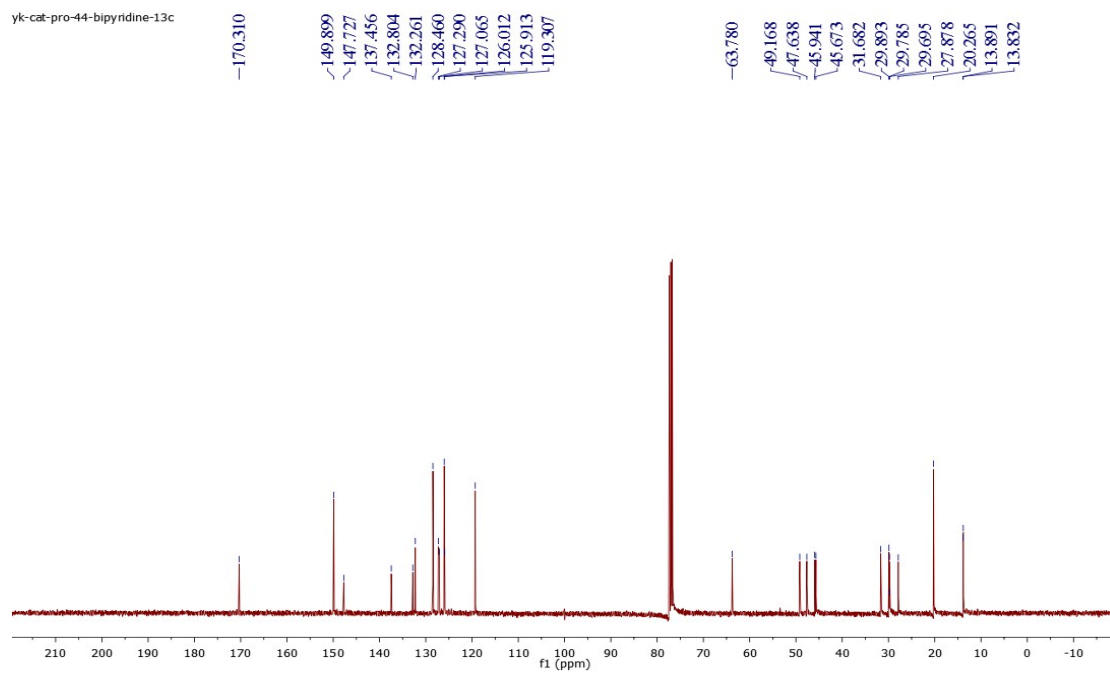


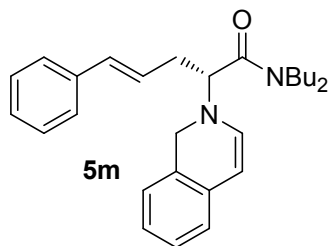
yk-cat-pro-py-4-2-py-13c-22times



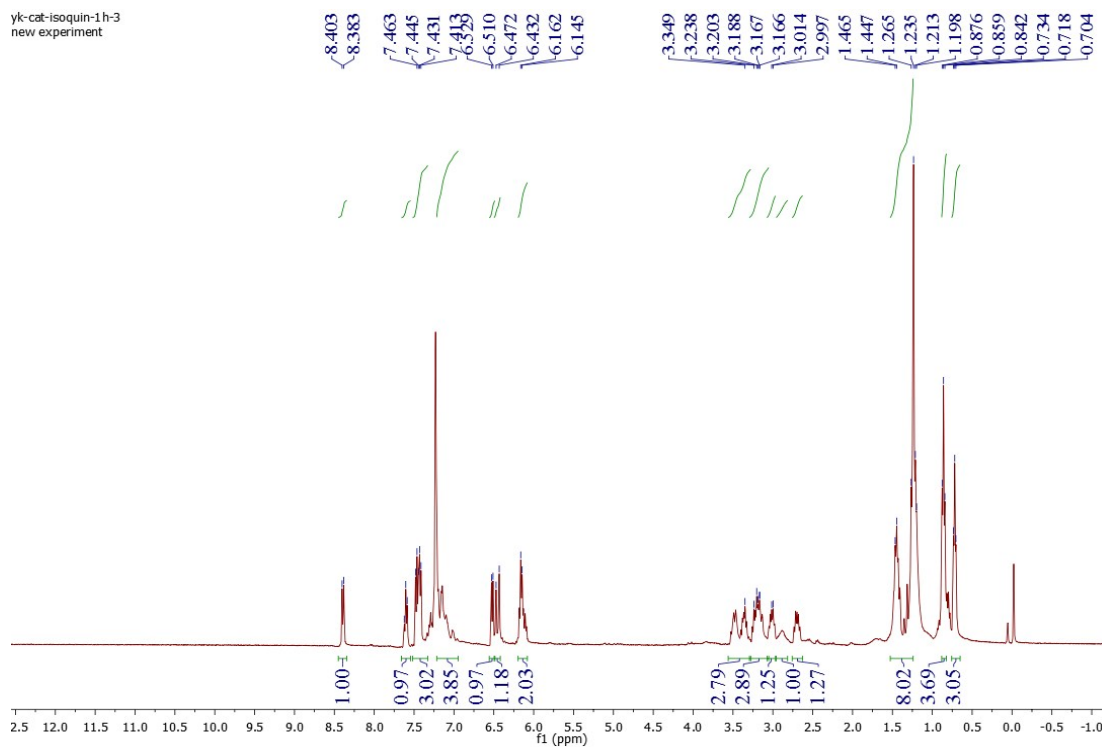


yk-cat-pro-44-bipyridine-13c

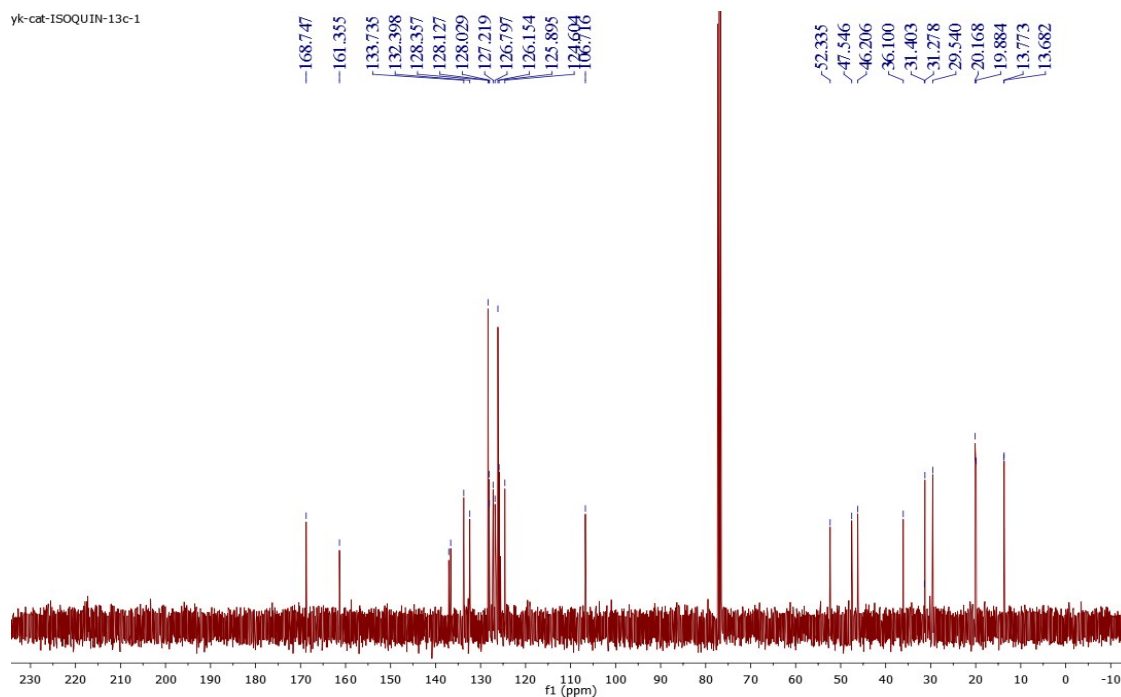


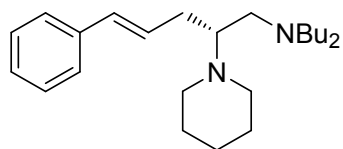


yk-cat-isoquin-1-h-3
new experiment



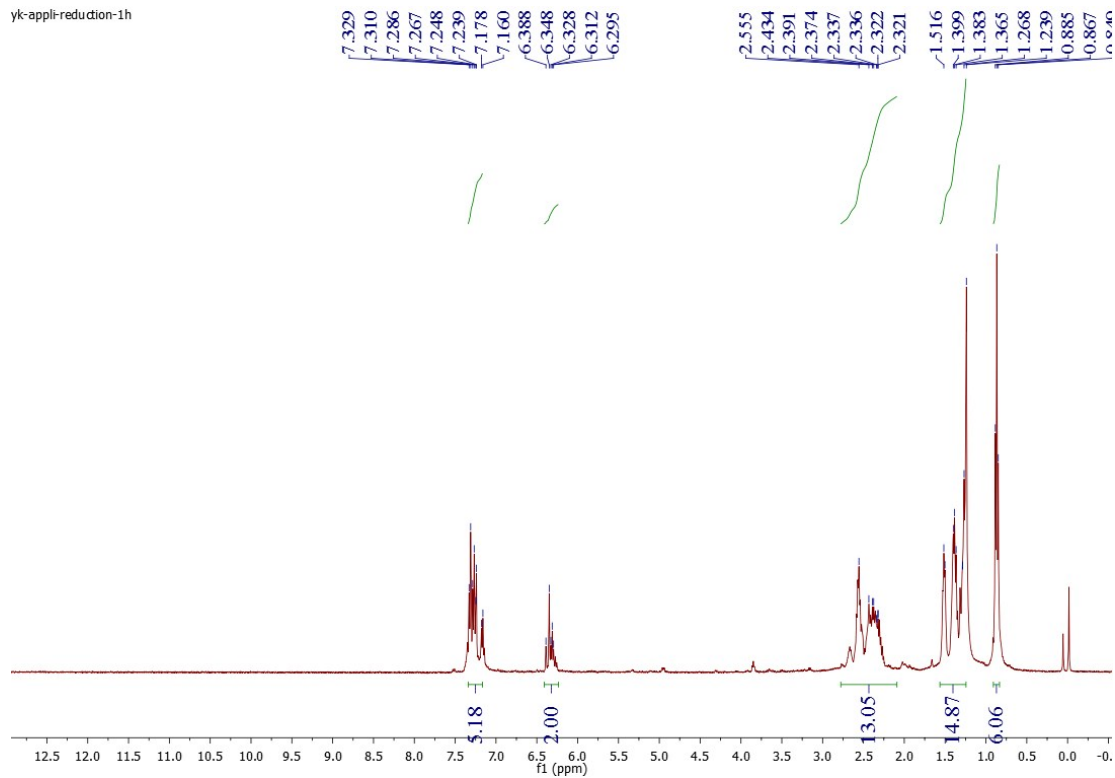
yk-cat-ISOQUIN-13c-1



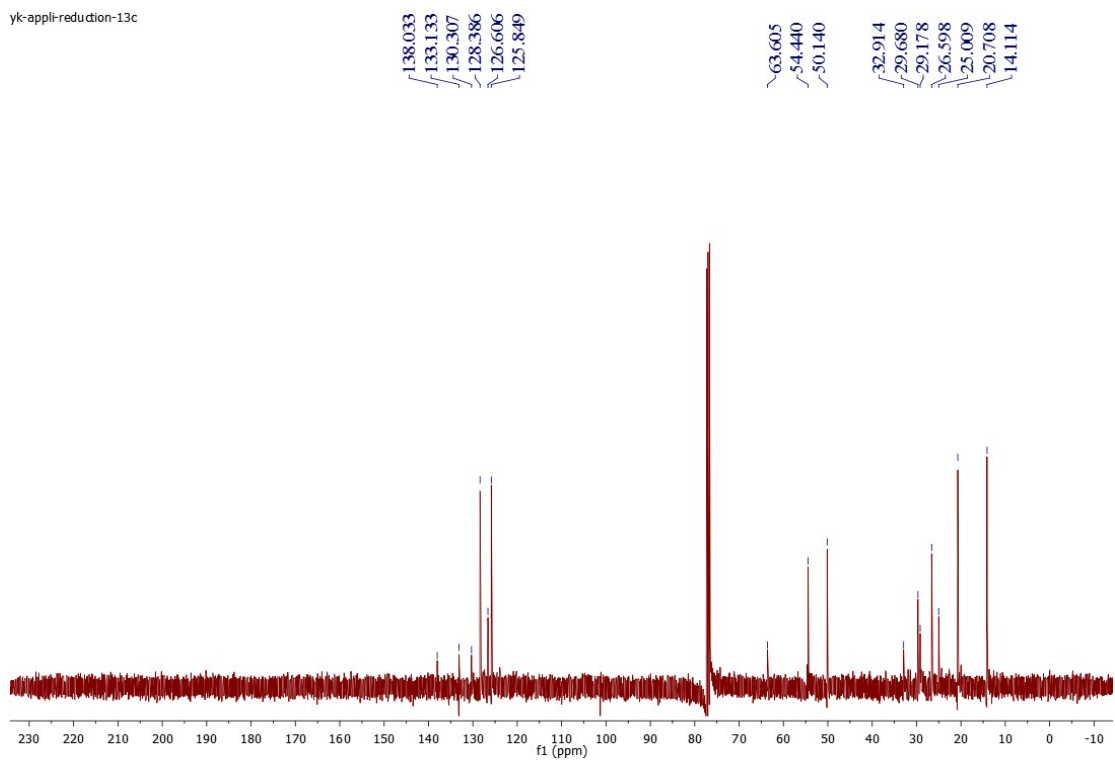


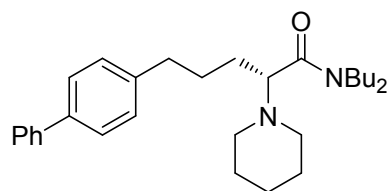
6

yk-appli-reduction-1h

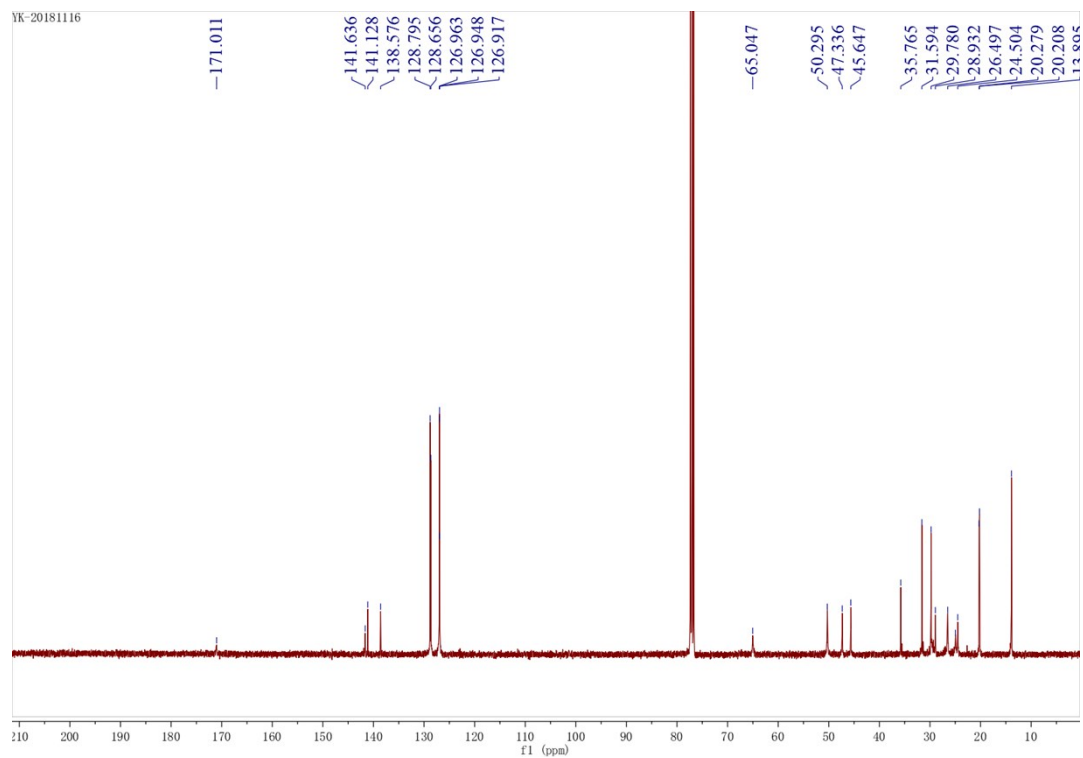
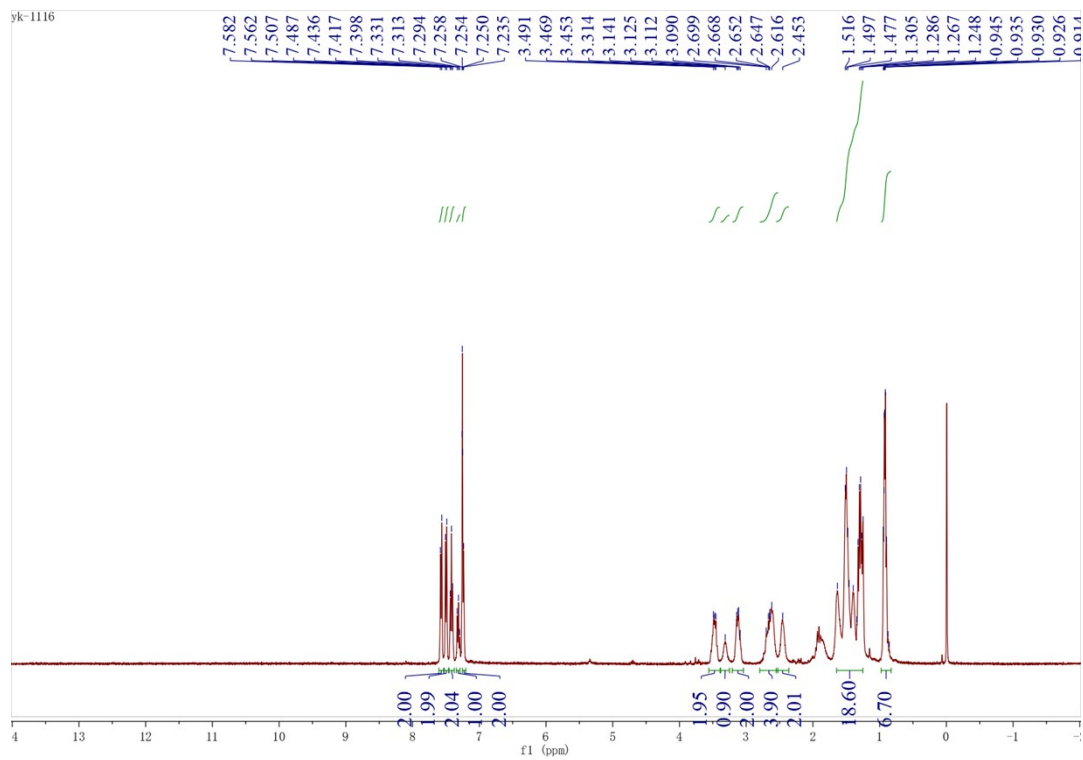


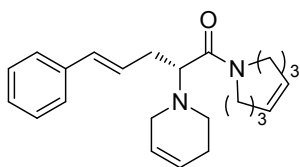
yk-appli-reduction-13c





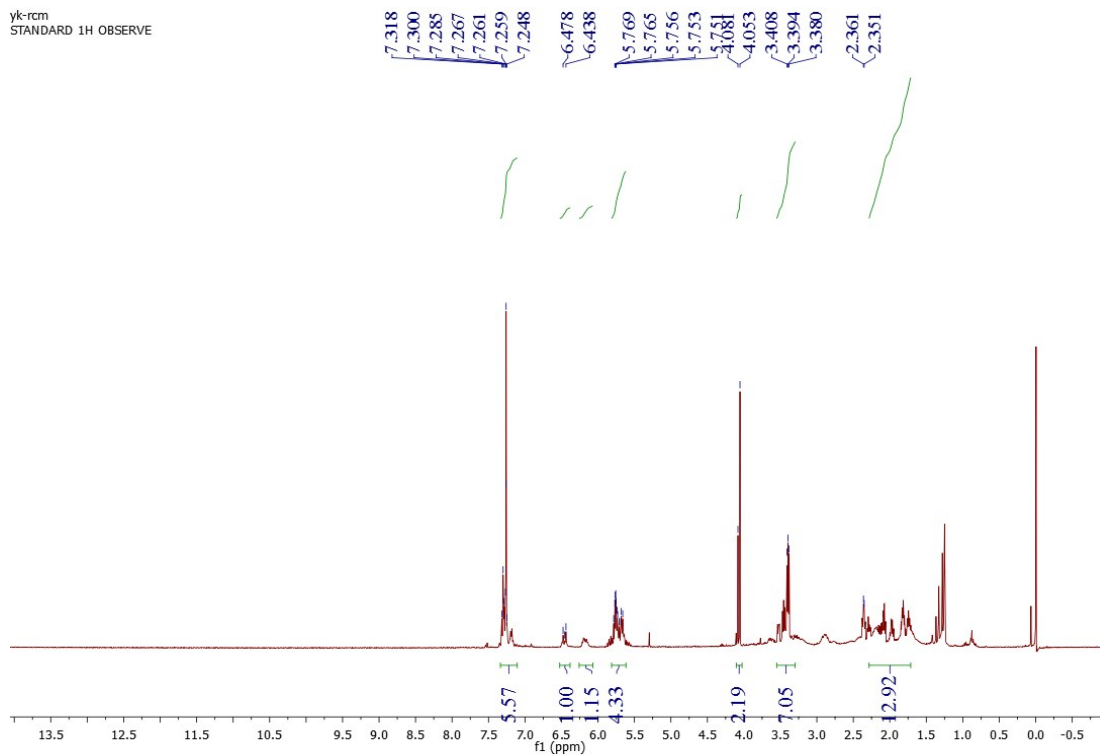
7



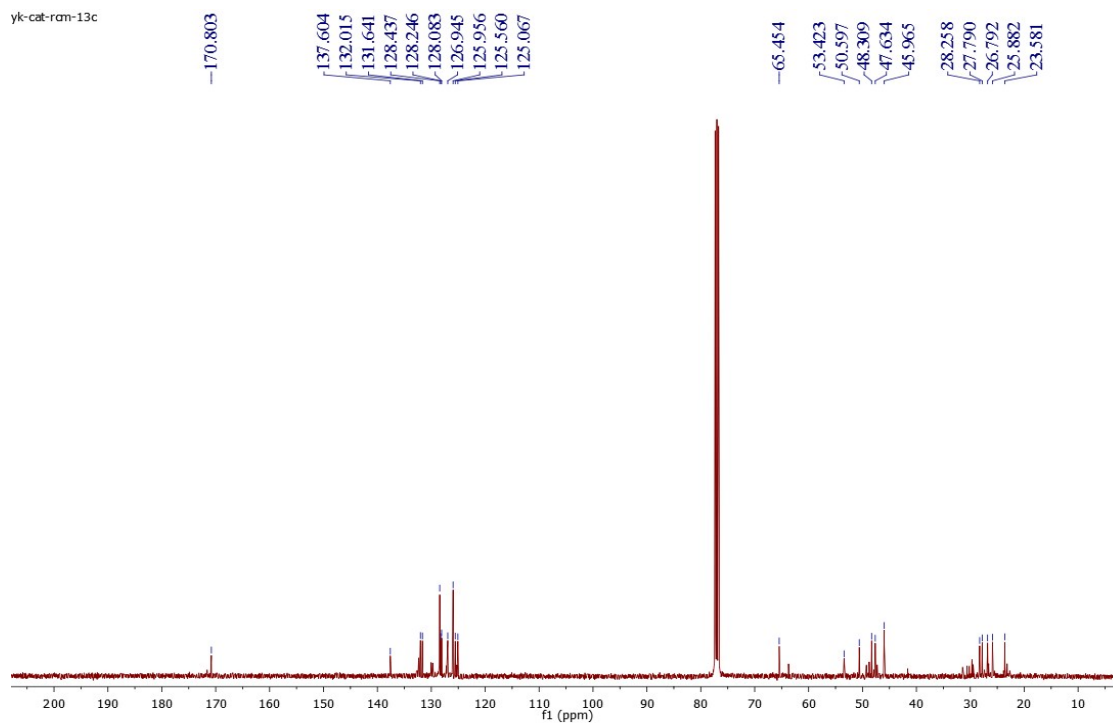


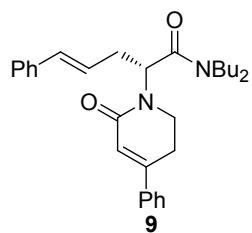
8

yk-rcm
STANDARD 1H OBSERVE

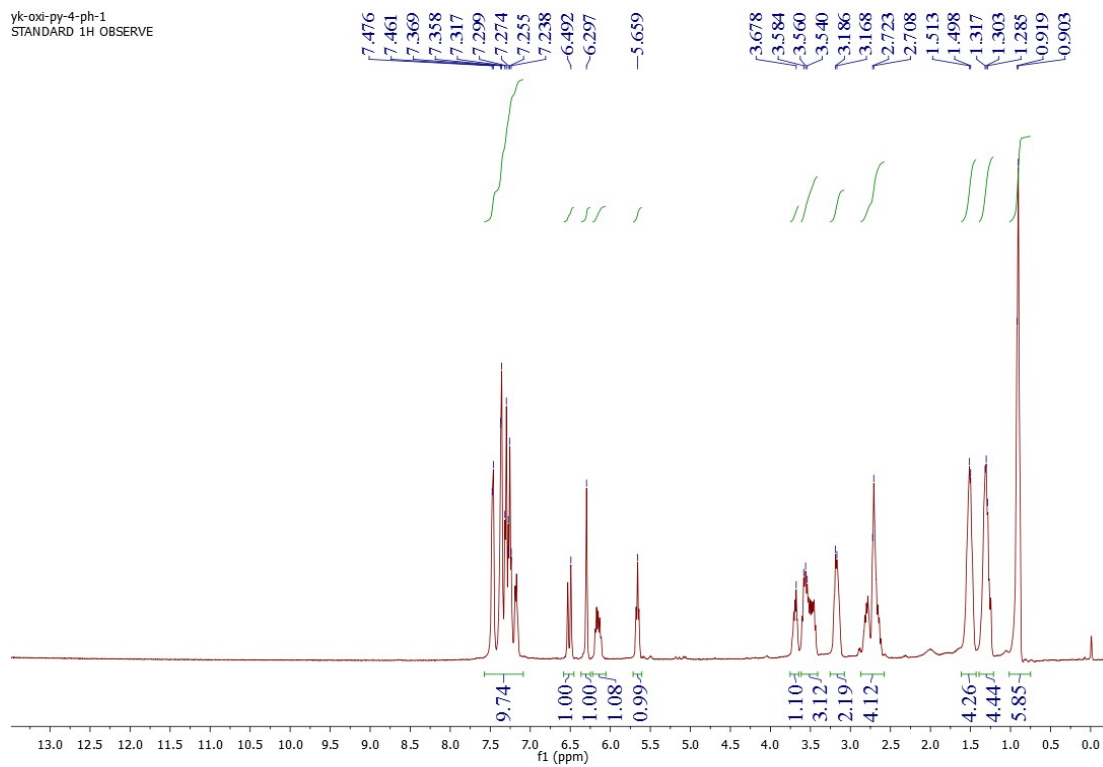


yk-cat-rcm-13c

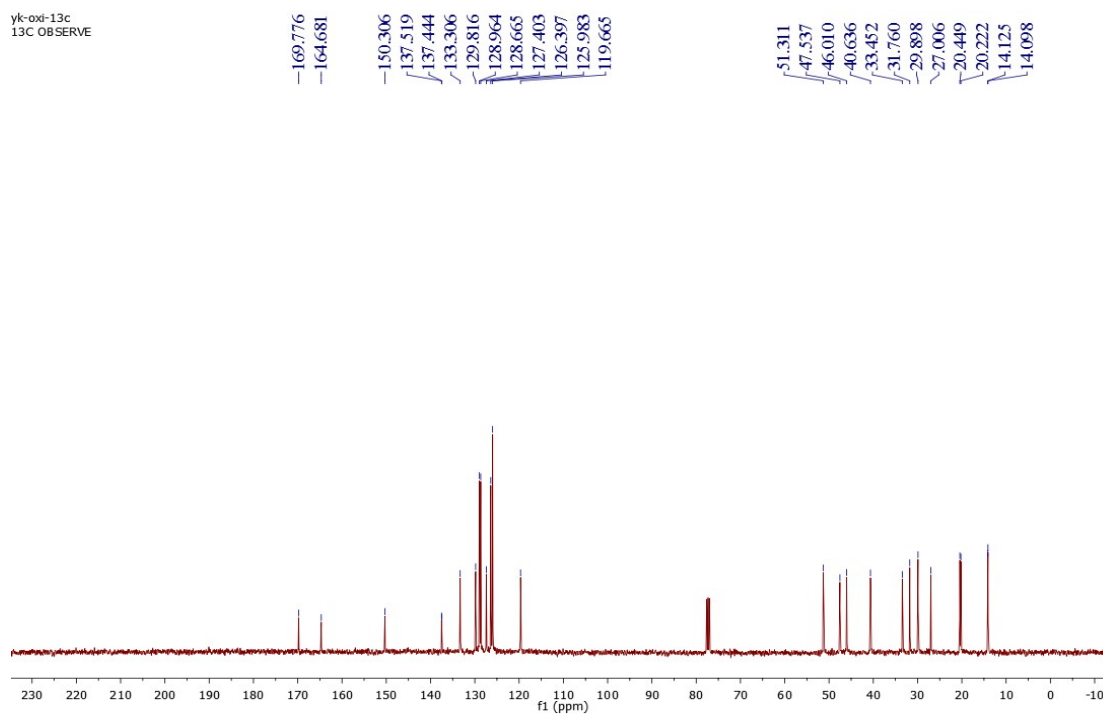


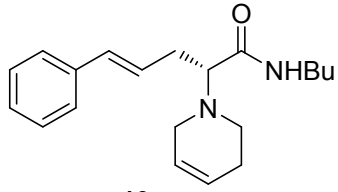


yk-oxi-py-4-ph-1
STANDARD 1H OBSERVE



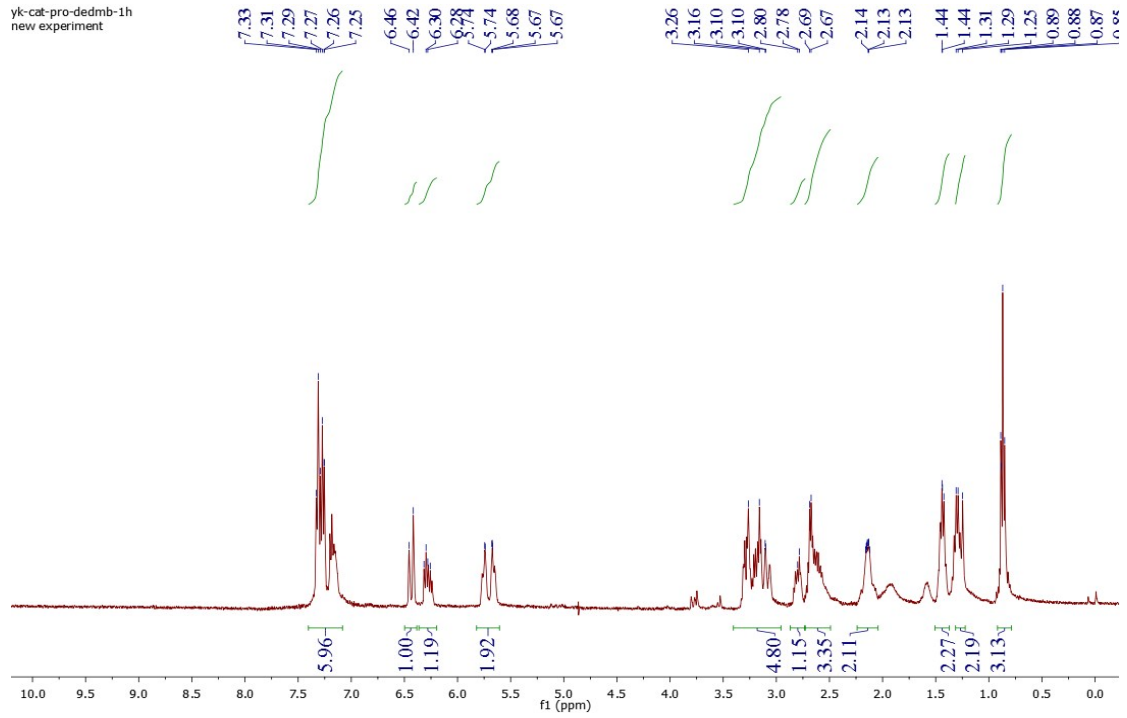
yk-oxi-13c
13C OBSERVE



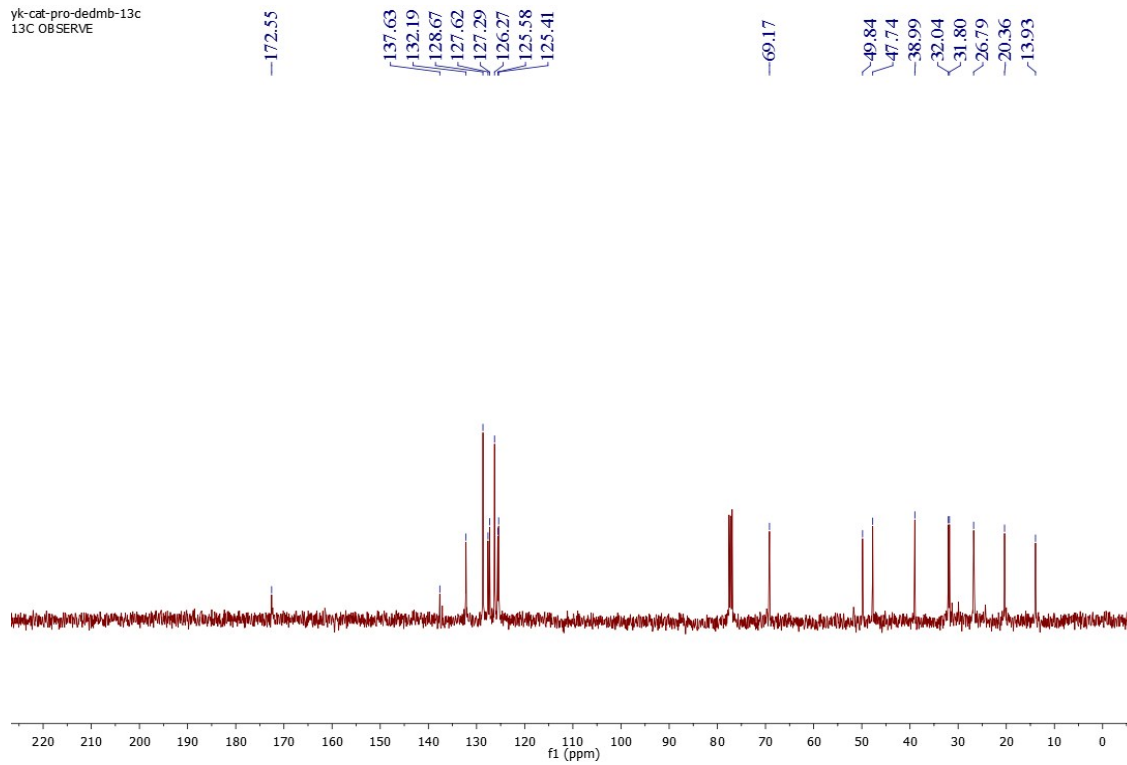


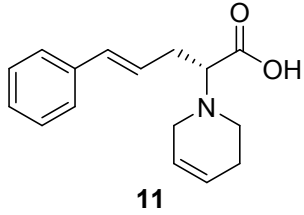
10

yk-cat-pro-dedmb-1h
new experiment

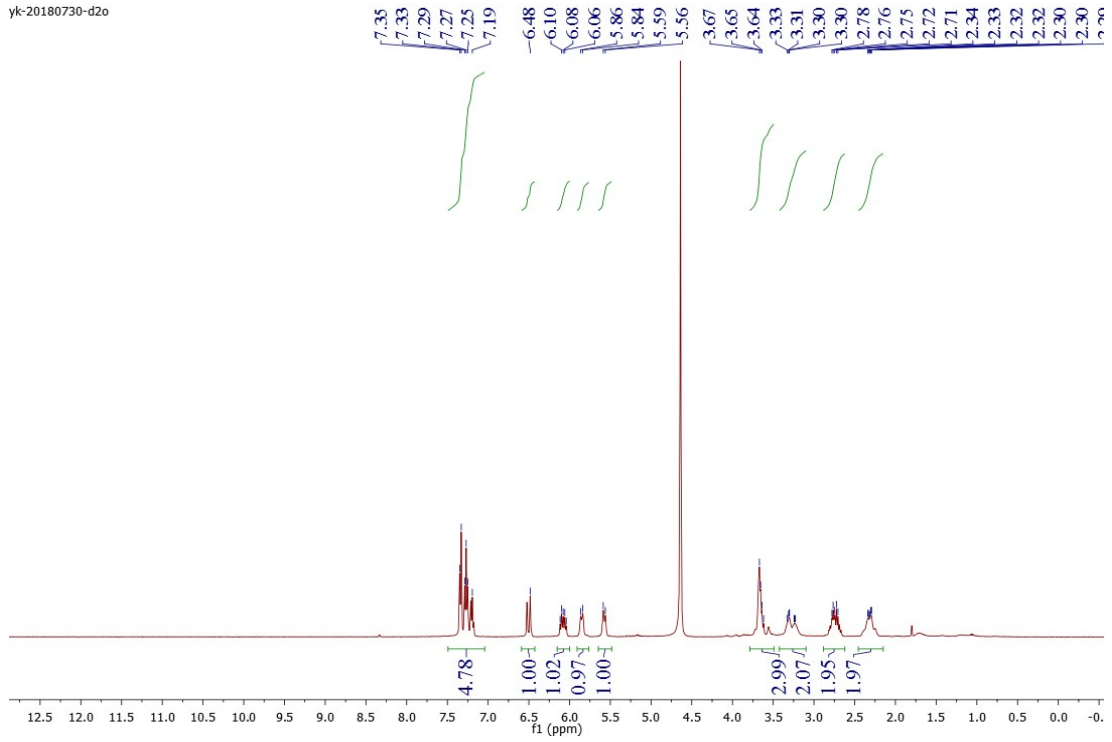


yk-cat-pro-dedmb-13c
13C OBSERVE

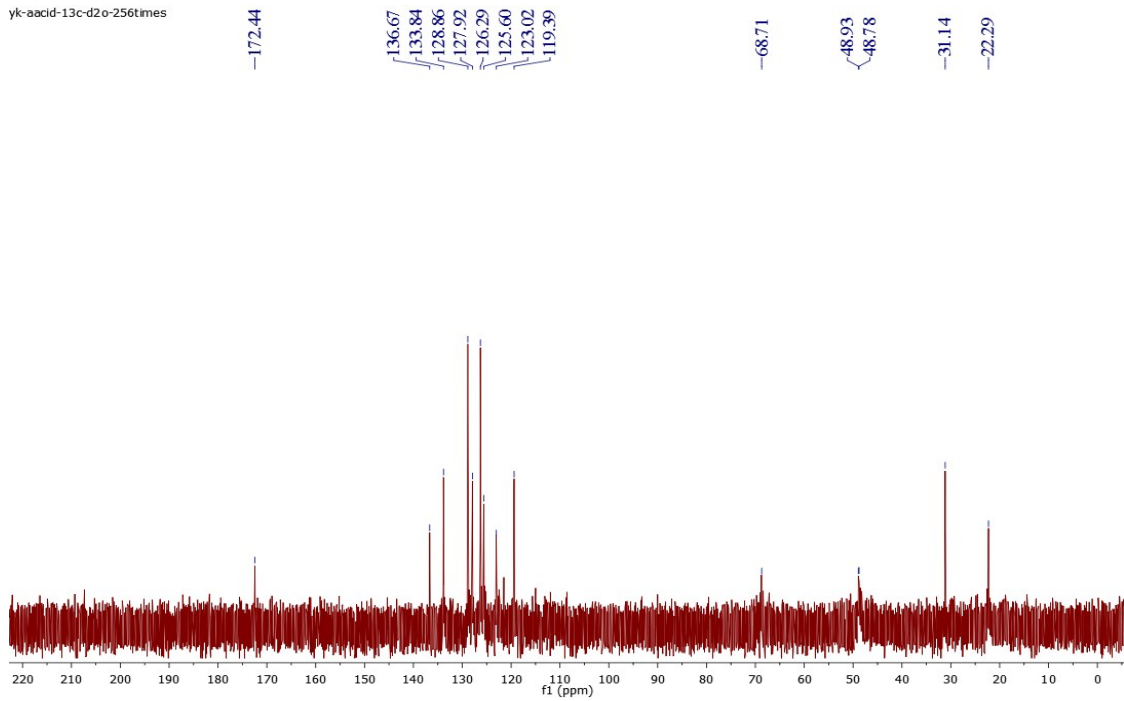


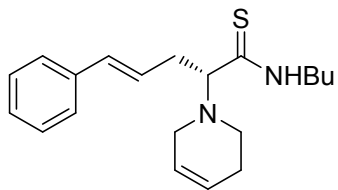


yk-20180730-d2o

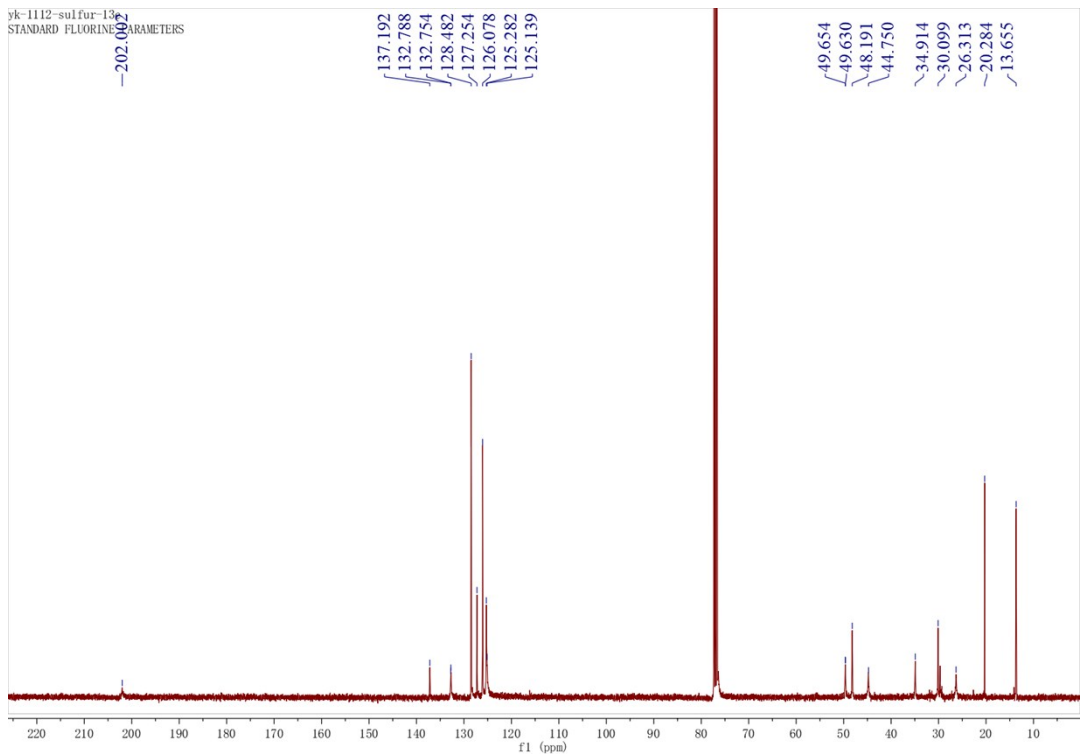
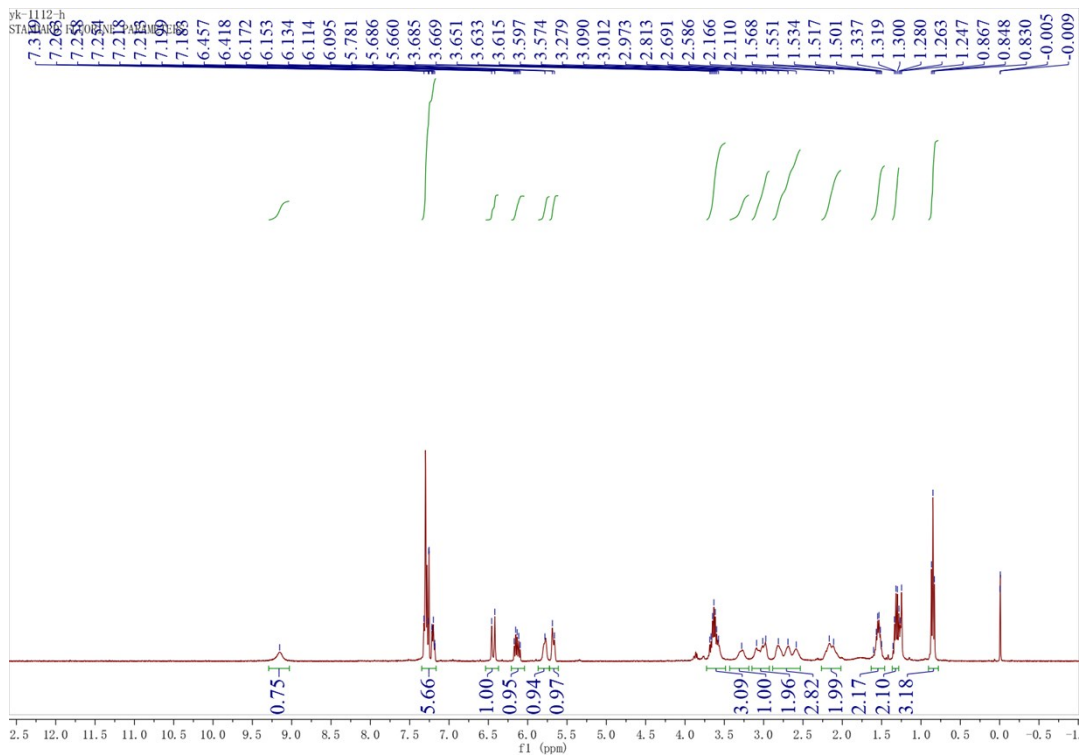


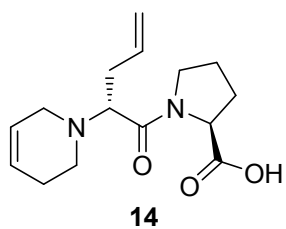
yk-aacid-13c-d2o-256times



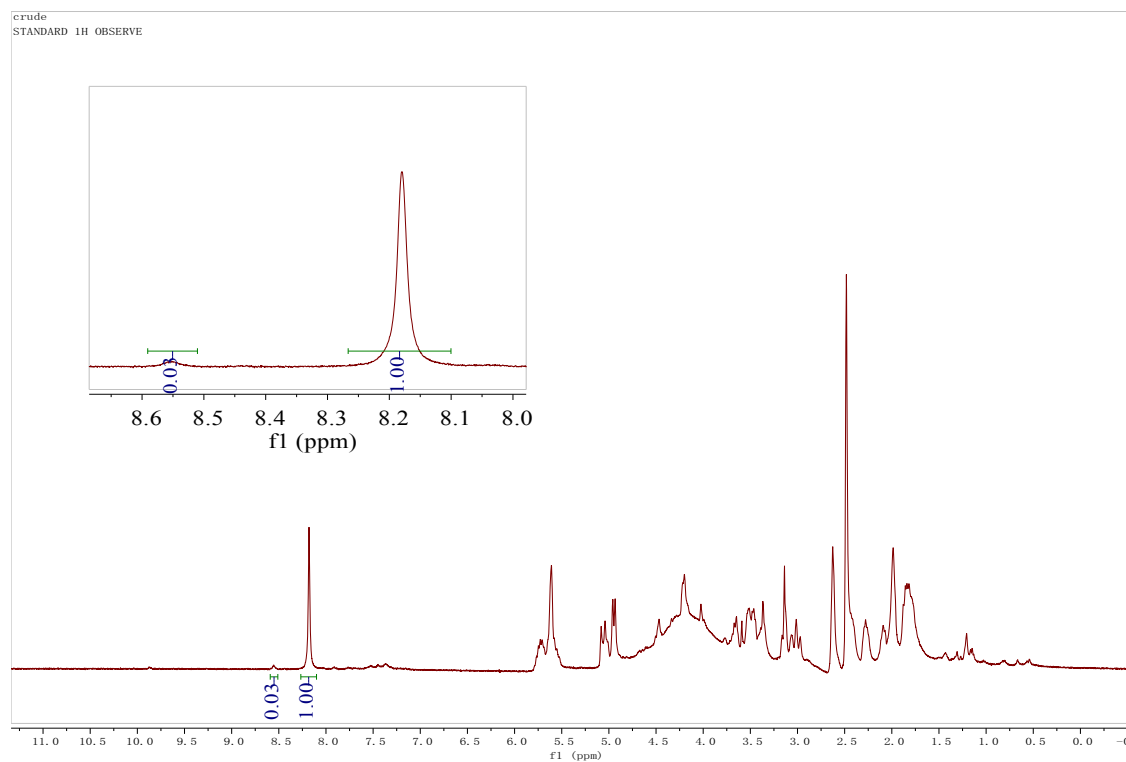


12

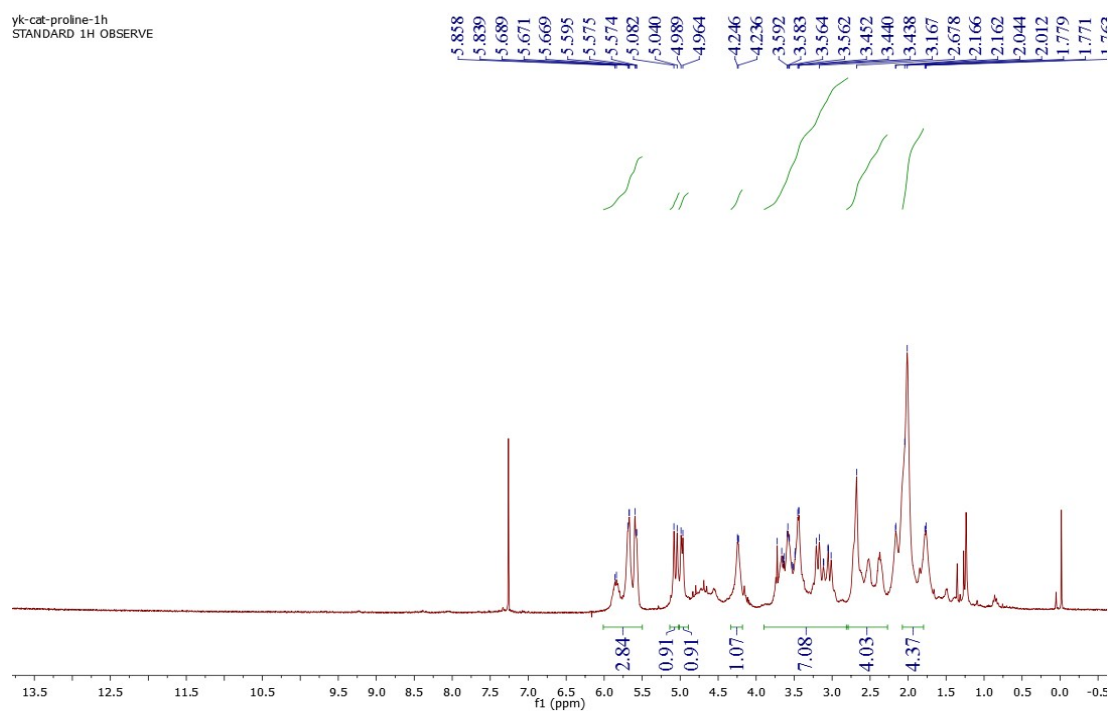




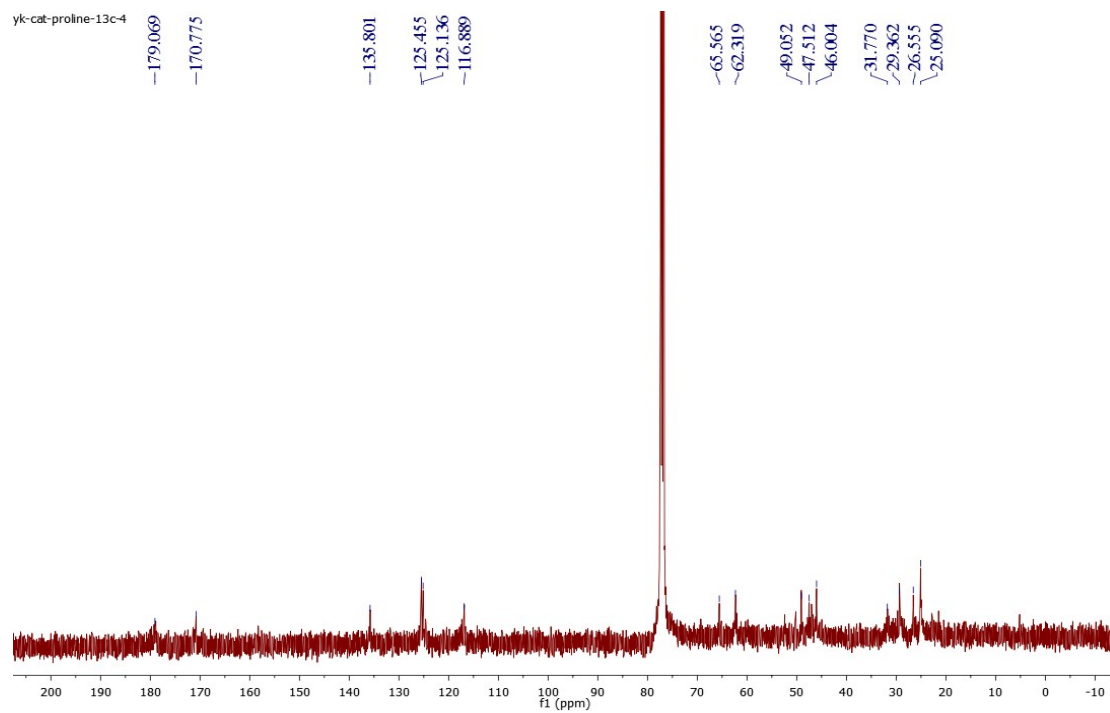
^1H NMR ($\text{DMSO-}d_6$) of the crude reaction mixture



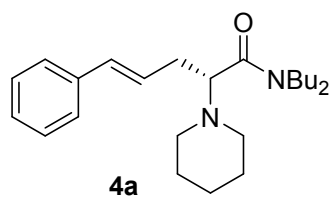
yk-cat-proline-1h
STANDARD 1H OBSERVE



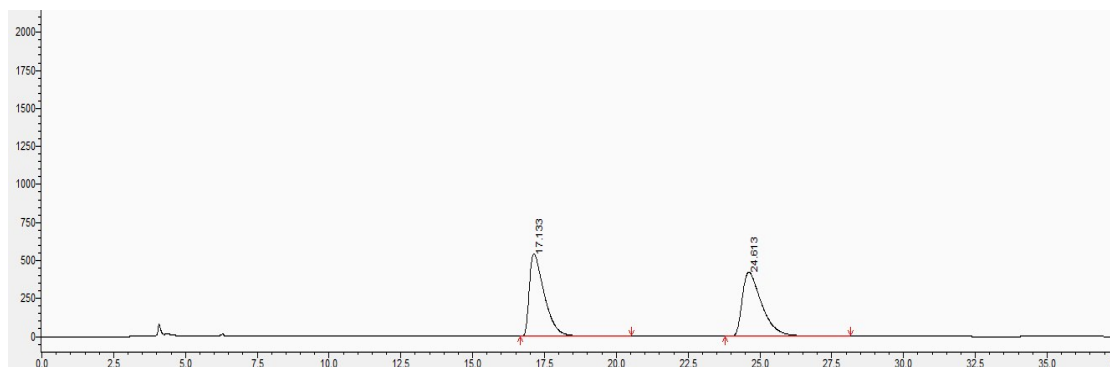
yk-cat-proline-13c-4



9. HPLC Data

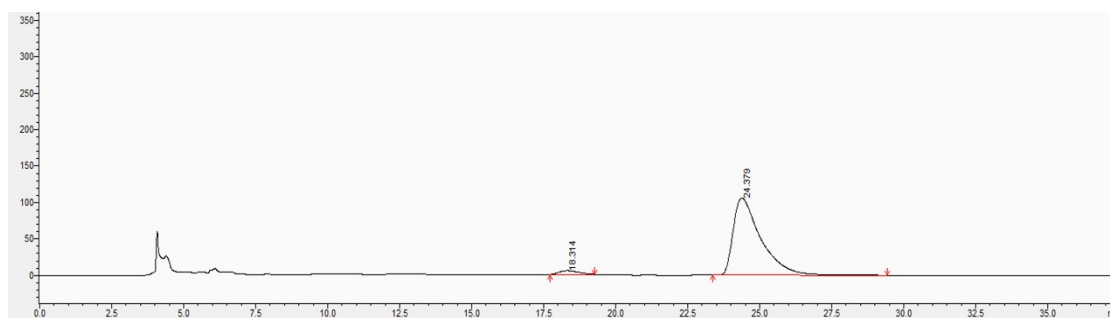


Racemate:

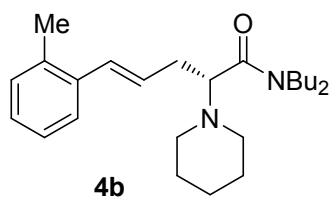


	Retention Time (min)	Area (%)
Peak 1	17.133	49.221
Peak 2	24.613	50.779

Chiral:

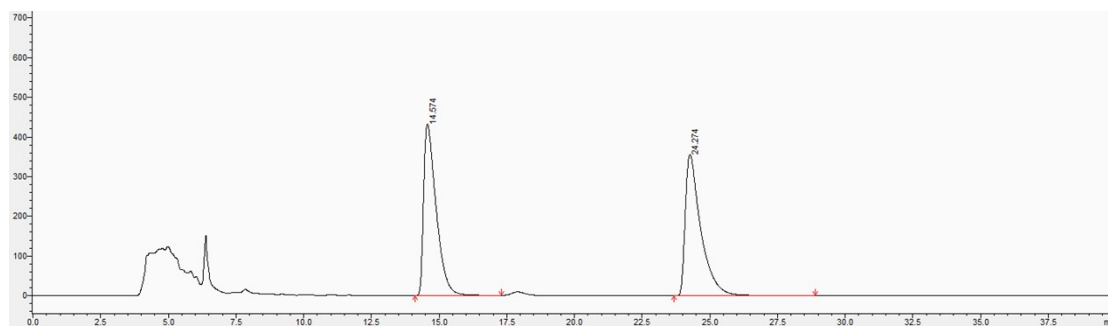


	Retention Time (min)	Area (%)	
Peak 1	18.314	2.608	95% ee
Peak 2	24.379	97.392	



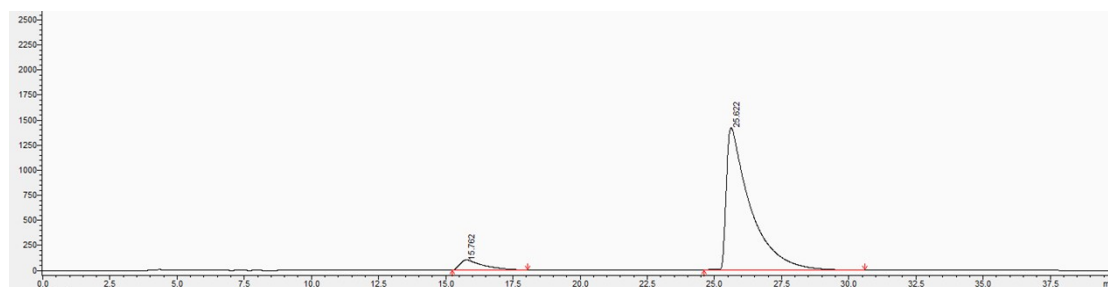
88% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 95/5, 0.8 mL/min, 254 nm.

Racemate:

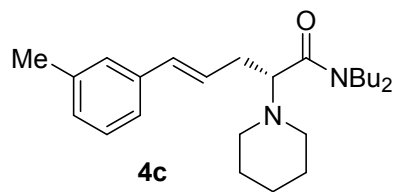


	Retention Time (min)	Area (%)
Peak 1	14.574	49.576
Peak 2	24.274	50.424

Chiral:

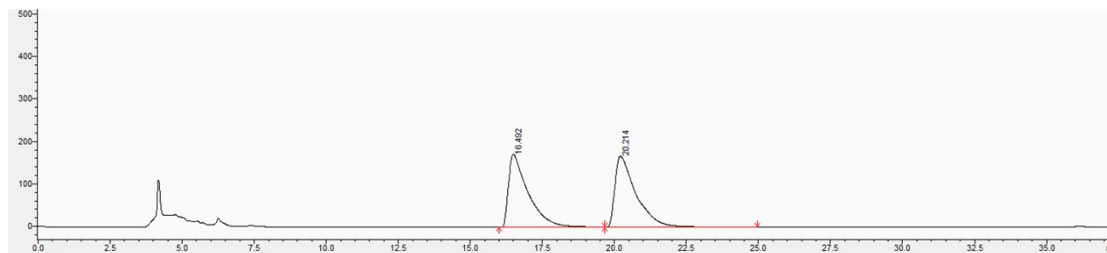


	Retention Time (min)	Area (%)	
Peak 1	14.969	6.242	88% ee
Peak 2	25.141	93.758	



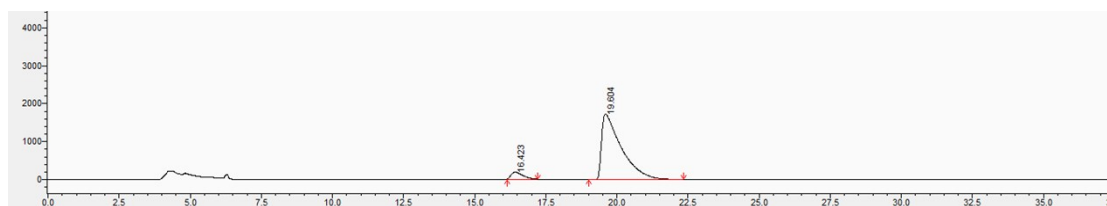
88% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 95/5, 0.8 mL/min, 254 nm.

Racemate:

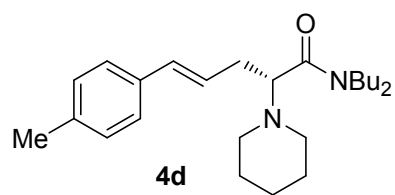


	Retention Time (min)	Area (%)
Peak 1	16.492	48.946
Peak 2	20.214	51.054

Chiral:

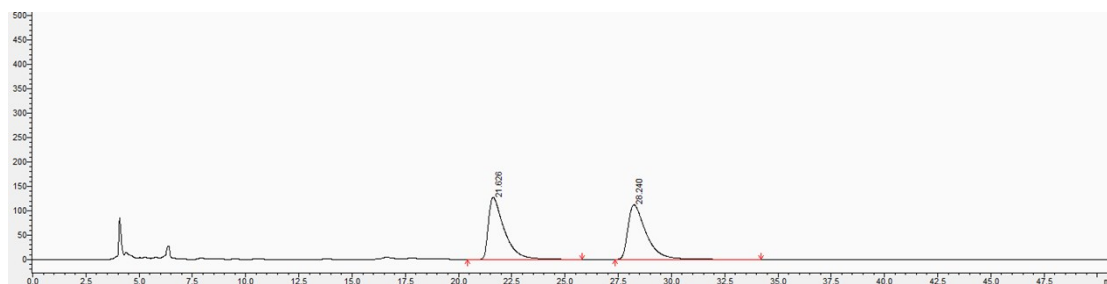


	Retention Time (min)	Area (%)	
Peak 1	16.423	6.119	88% ee
Peak 2	19.604	93.881	



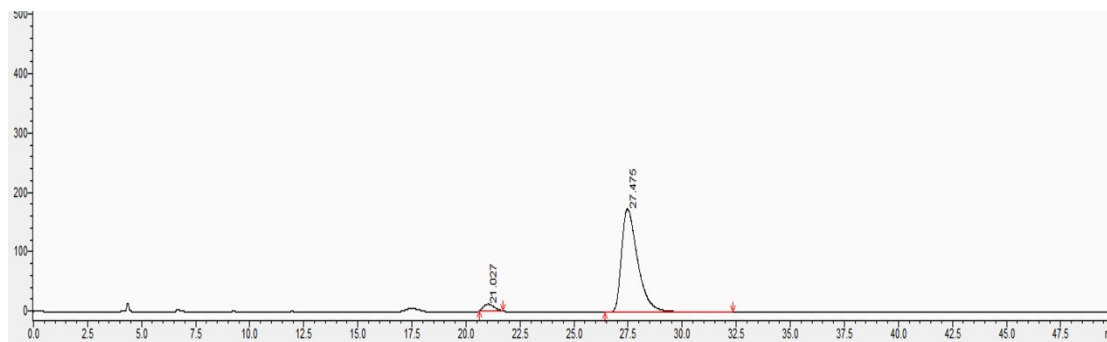
92% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 95/5, 0.8 mL/min, 254 nm.

Racemate:

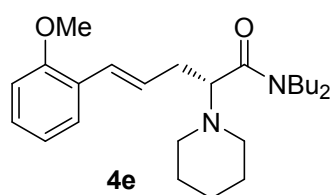


	Retention Time (min)	Area (%)
Peak 1	21.626	49.888
Peak 2	28.240	50.112

Chiral:

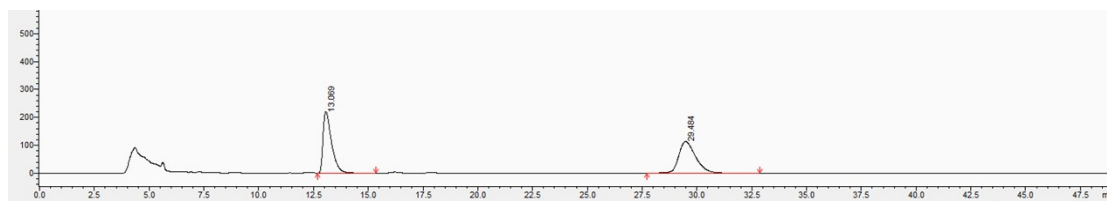


	Retention Time (min)	Area (%)	
Peak 1	21.027	4.121	92% ee
Peak 2	27.475	95.879	



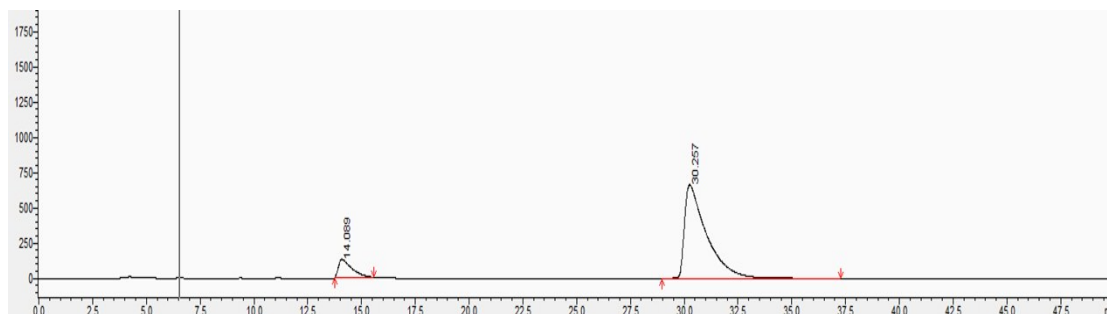
82% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 85/15, 0.8 mL/min, 254 nm.

Racemate:

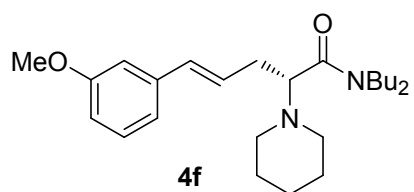


	Retention Time (min)	Area (%)
Peak 1	13.069	49.216
Peak 2	29.484	50.784

Chiral:

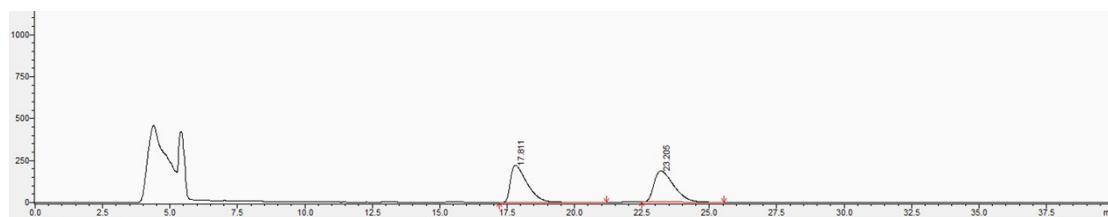


	Retention Time (min)	Area (%)	
Peak 1	14.089	8.902	82% ee
Peak 2	30.257	91.098	



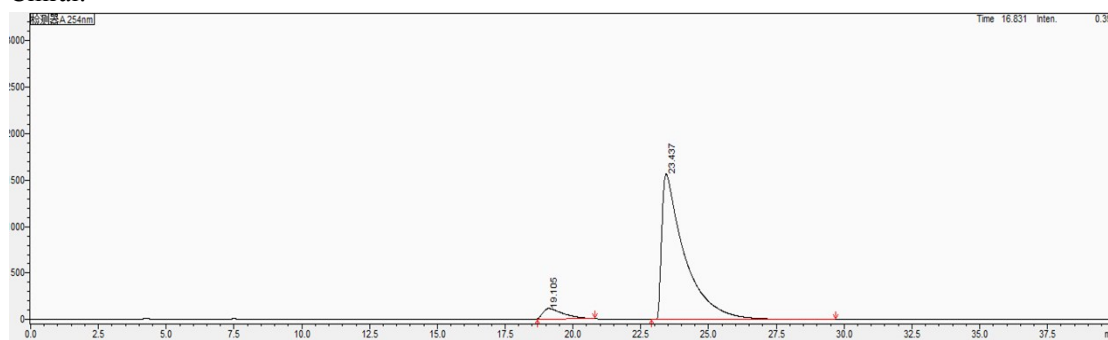
87% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 90/10, 0.8 mL/min, 254 nm.

Racemate:

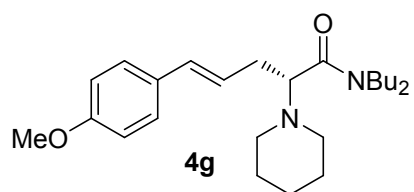


	Retention Time (min)	Area (%)
Peak 1	17.811	50.137
Peak 2	23.205	49.863

Chiral:

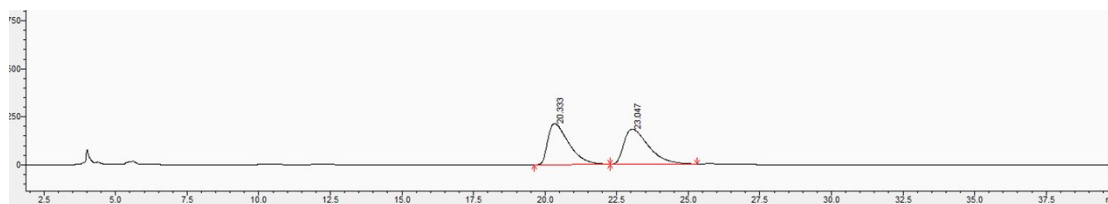


	Retention Time (min)	Area (%)	
Peak 1	19.105	6.264	87% ee
Peak 2	23.437	93.736	



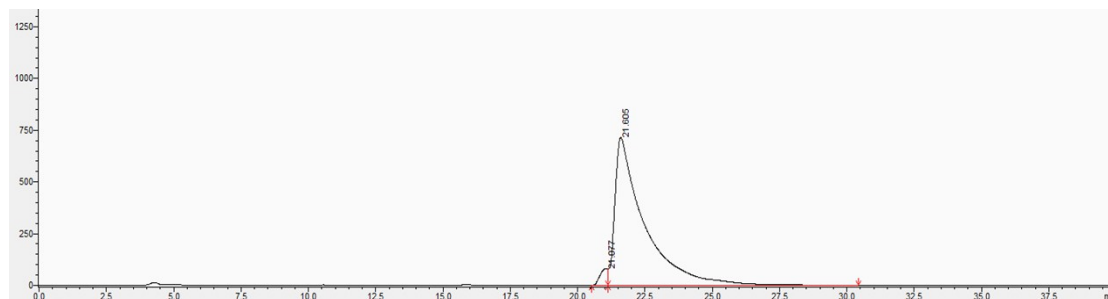
92% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 90/10, 0.8 mL/min, 254 nm.

Racemate:

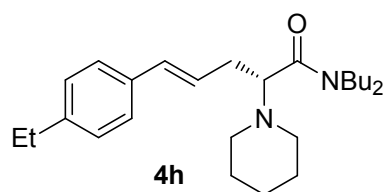


	Retention Time (min)	Area (%)
Peak 1	20.333	49.993
Peak 2	23.047	50.007

Chiral:

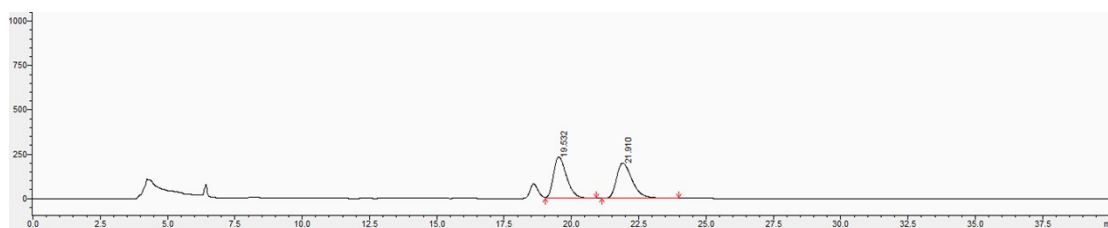


	Retention Time (min)	Area (%)	
Peak 1	21.077	3.879	92% ee
Peak 2	21.605	96.121	



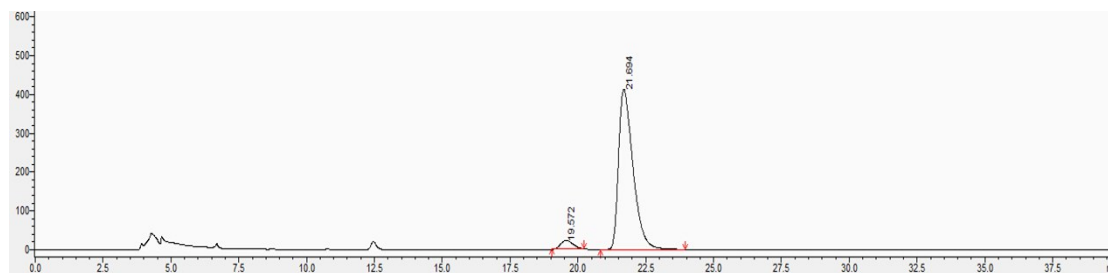
91% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 95/5, 0.8 mL/min, 254 nm.

Racemate:

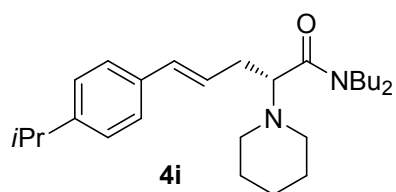


	Retention Time (min)	Area (%)
Peak 1	19.532	49.256
Peak 2	21.910	50.744

Chiral:

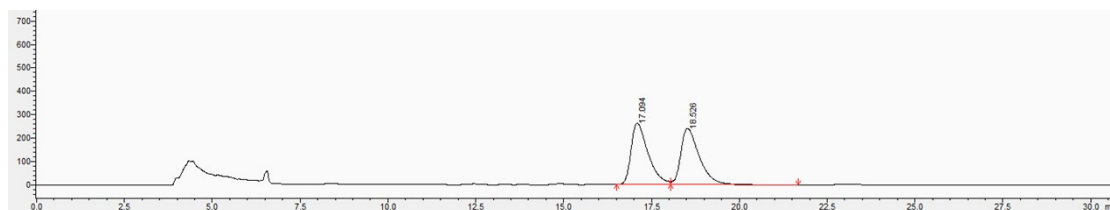


	Retention Time (min)	Area (%)	
Peak 1	19.572	4.262	91% ee
Peak 2	21.694	95.738	



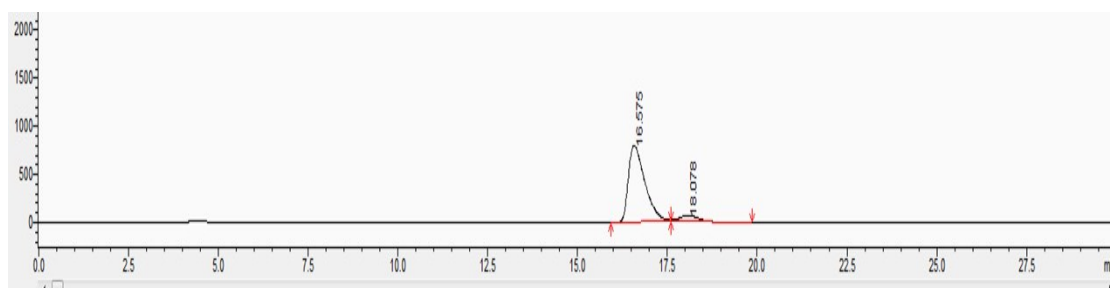
90% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 95/5, 0.8 mL/min, 254 nm.

Racemate:

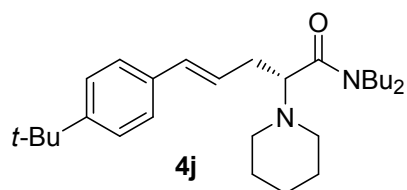


	Retention Time (min)	Area (%)
Peak 1	17.094	50.126
Peak 2	18.526	49.874

Chiral:

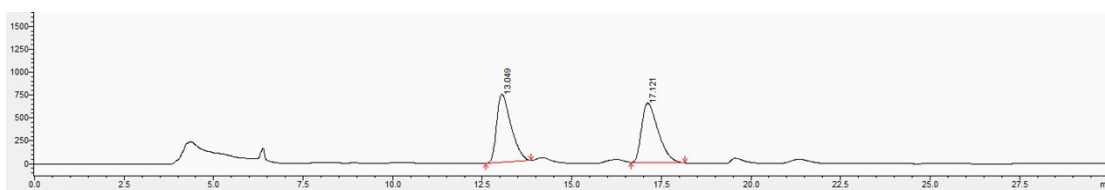


	Retention Time (min)	Area (%)	
Peak 1	16.575	94.809	90% ee
Peak 2	18.078	5.191	



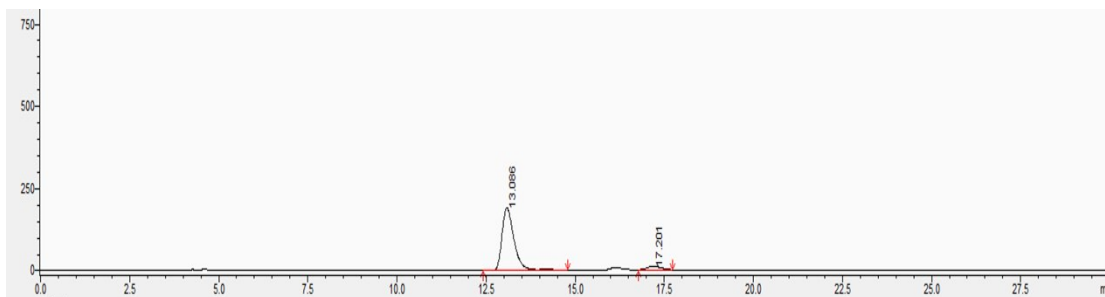
87% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 95/5, 0.8 mL/min, 254 nm.

Racemate:

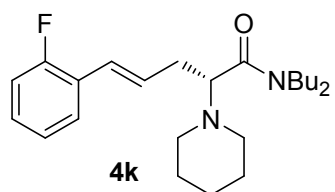


	Retention Time (min)	Area (%)
Peak 1	13.049	49.861
Peak 2	17.121	50.139

Chiral:

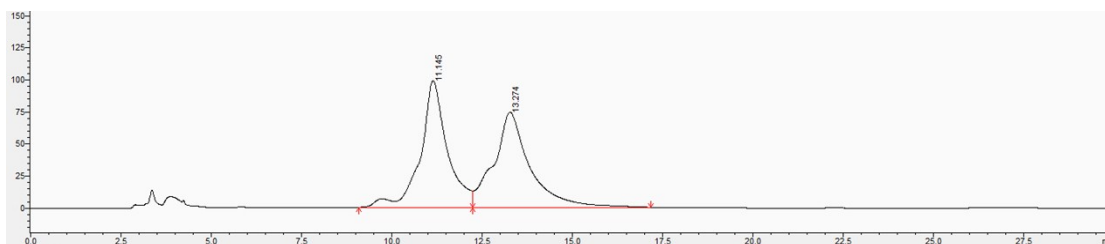


	Retention Time (min)	Area (%)	
Peak 1	13.086	93.692	87% ee
Peak 2	17.201	6.308	



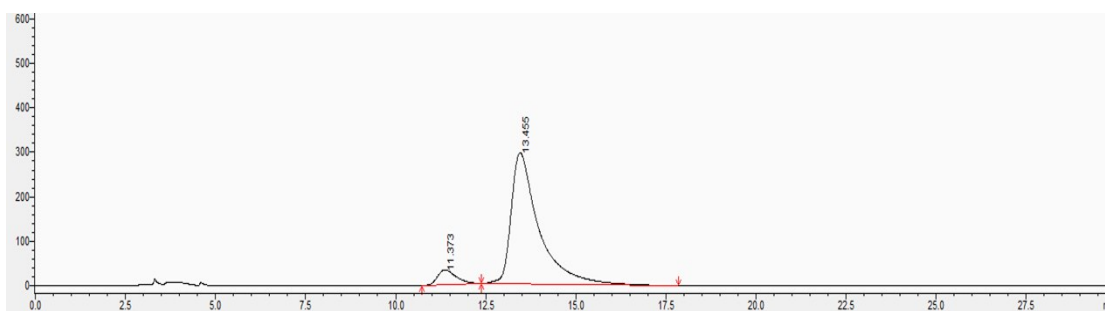
84% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 95/5, 0.8 mL/min, 254 nm.

Racemate:

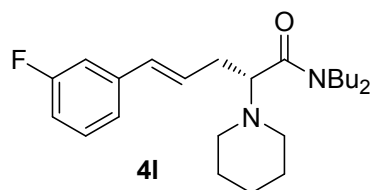


	Retention Time (min)	Area (%)
Peak 1	11.145	49.964
Peak 2	13.274	50.036

Chiral:

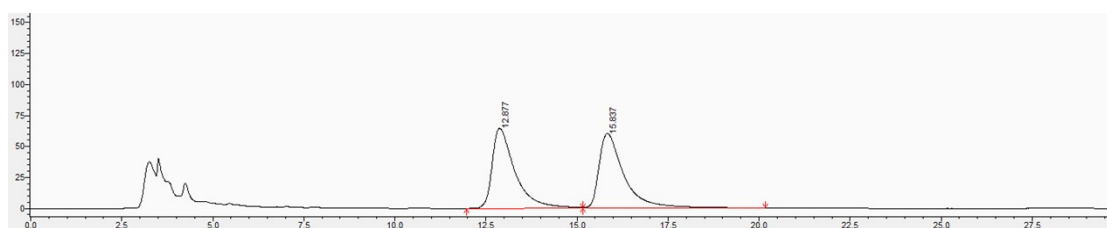


	Retention Time (min)	Area (%)	
Peak 1	11.373	7.947	84% ee
Peak 2	13.455	92.053	



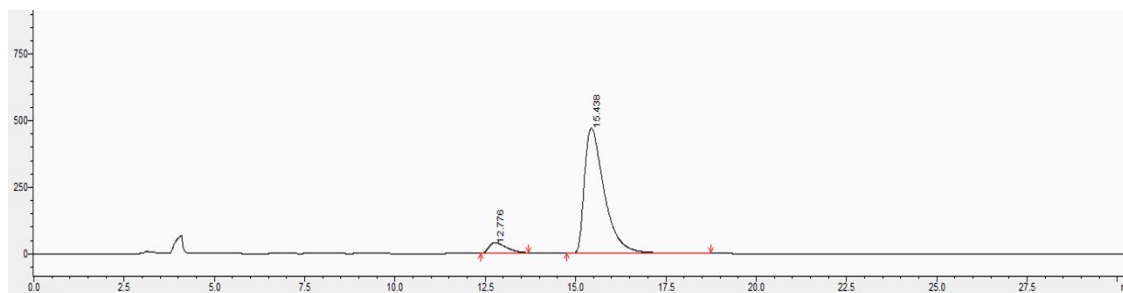
91% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 95/5, 0.8 mL/min, 254 nm.

Racemate:

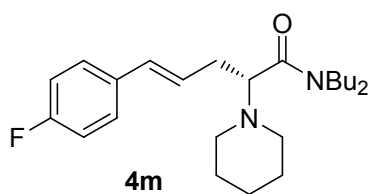


	Retention Time (min)	Area (%)
Peak 1	12.877	50.973
Peak 2	15.837	49.027

Chiral:

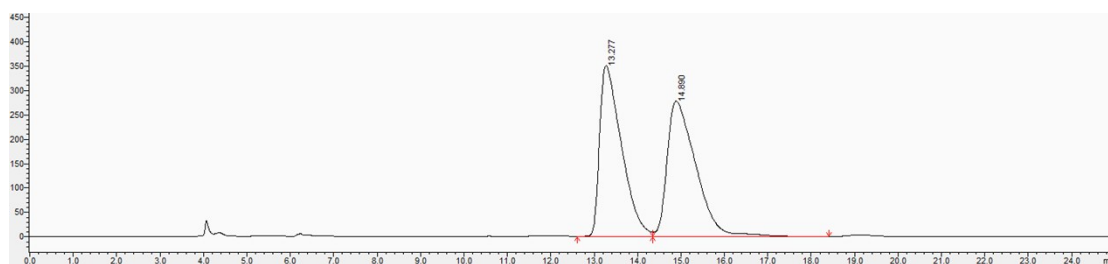


	Retention Time (min)	Area (%)	
Peak 1	12.776	4.577	91% ee
Peak 2	15.438	95.423	



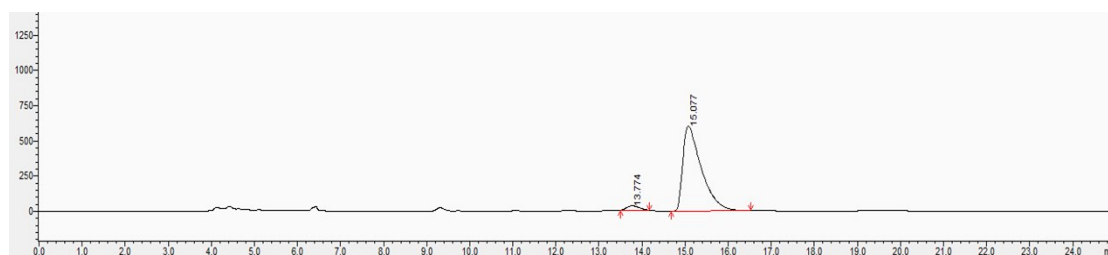
93% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 95/5, 0.8 mL/min, 254 nm.

Racemate:

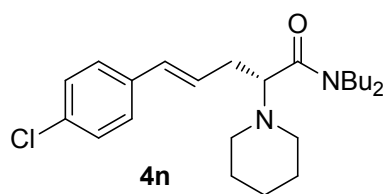


	Retention Time (min)	Area (%)
Peak 1	13.277	49.015
Peak 2	14.890	50.985

Chiral:

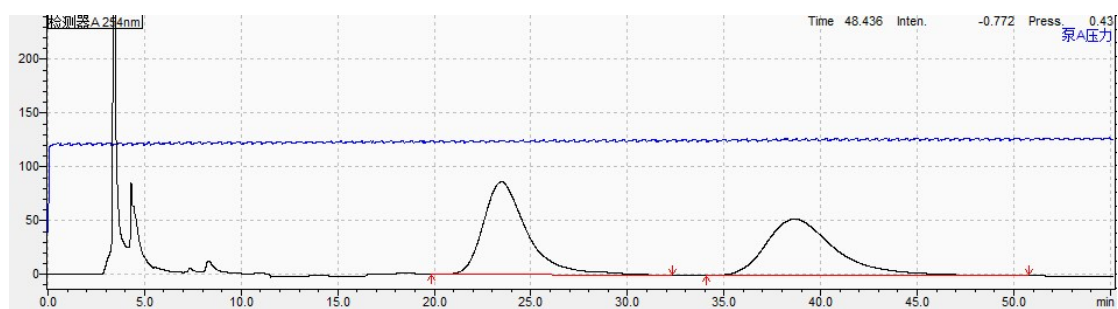


	Retention Time (min)	Area (%)	
Peak 1	13.774	3.597	93% ee
Peak 2	15.077	96.403	



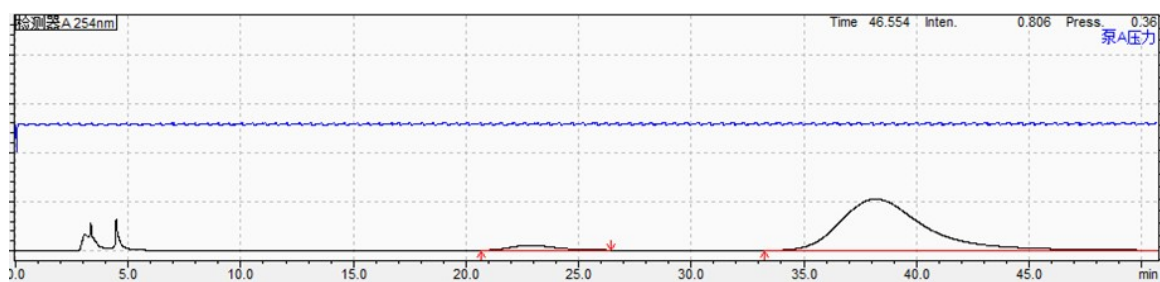
92% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 95/5, 0.4 mL/min, 254 nm.

Racemate:

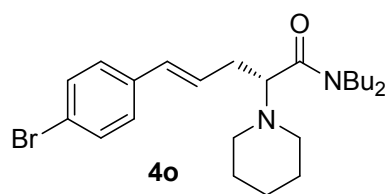


	Retention Time (min)	Area (%)
Peak 1	23.481	49.729
Peak 2	38.651	50.271

Chiral:

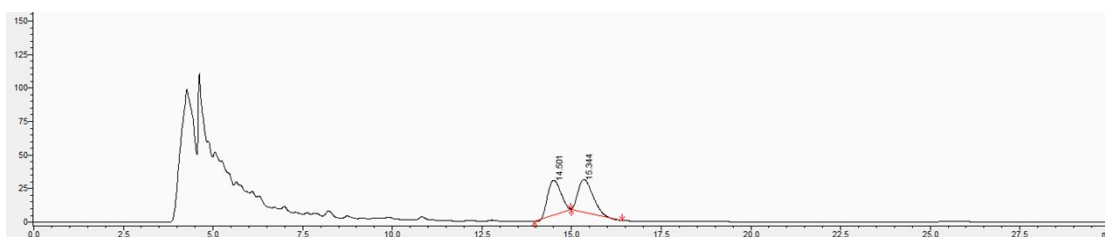


	Retention Time (min)	Area (%)	
Peak 1	22.855	4.014	92% ee
Peak 2	38.173	95.986	



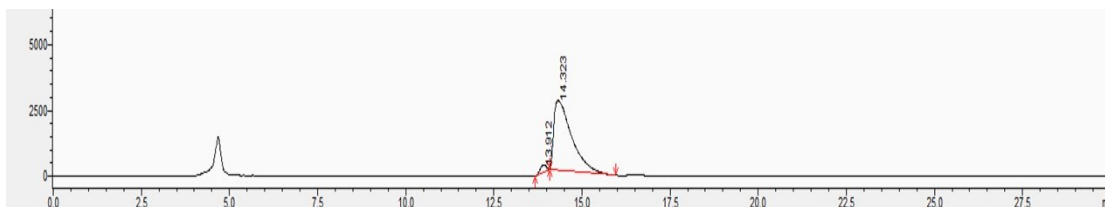
93% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IC-3 column, *n*-hexane/*i*-PrOH = 95/5, 0.8 mL/min, 254 nm.

Racemate:

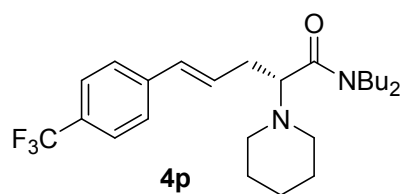


	Retention Time (min)	Area (%)
Peak 1	14.501	48.860
Peak 2	15.344	51.140

Chiral:

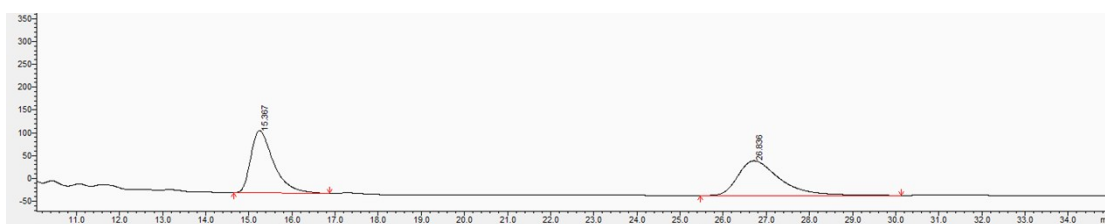


	Retention Time (min)	Area (%)	
Peak 1	17.077	3.543	93% ee
Peak 2	25.302	96.457	



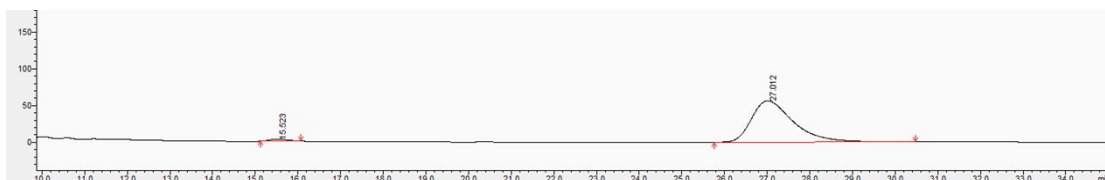
96% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak AD-H column, *n*-hexane/*i*-PrOH = 99.5/0.5, 0.8 mL/min, 254 nm.

Racemate:

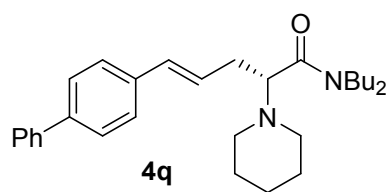


	Retention Time (min)	Area (%)
Peak 1	15.367	49.661
Peak 2	26.836	50.339

Chiral:

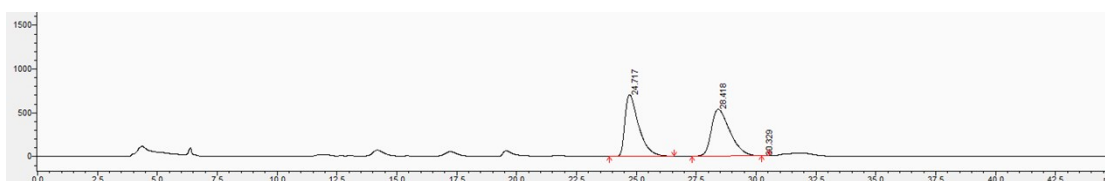


	Retention Time (min)	Area (%)	
Peak 1	15.523	2.065	96% ee
Peak 2	27.012	97.935	



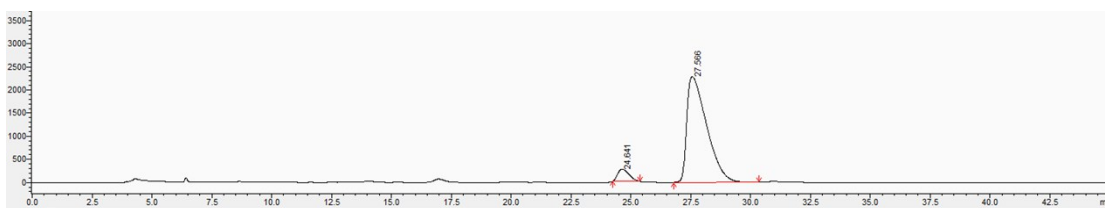
88% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IC-3 column, *n*-hexane /*i*-PrOH = 95/5, 0.8 mL/min, 254 nm.

Racemate:

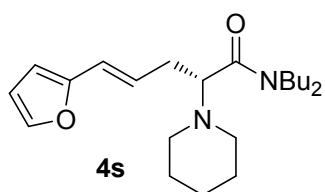


	Retention Time (min)	Area (%)
Peak 1	24.717	49.176
Peak 2	28.418	50.824

Chiral:

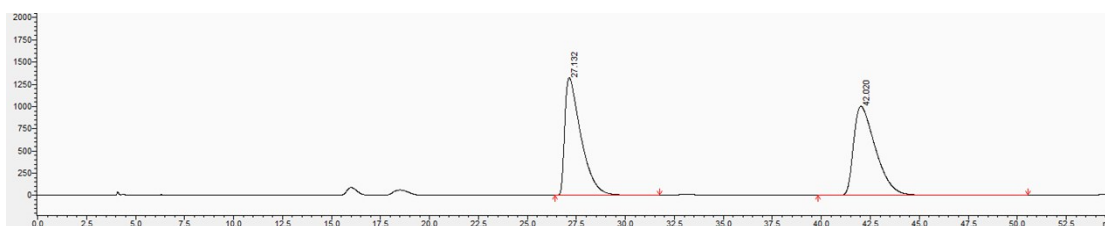


	Retention Time (min)	Area (%)	
Peak 1	24.641	6.073	88% ee
Peak 2	27.566	93.927	



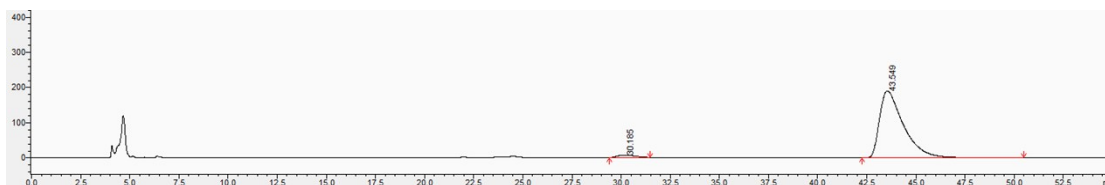
95% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IC-3 column, *n*-hexane /*i*-PrOH = 95/5, 0.8 mL/min, 254 nm.

Racemate:

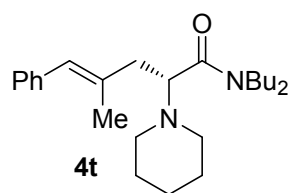


	Retention Time (min)	Area (%)
Peak 1	27.132	49.241
Peak 2	42.020	50.759

Chiral:

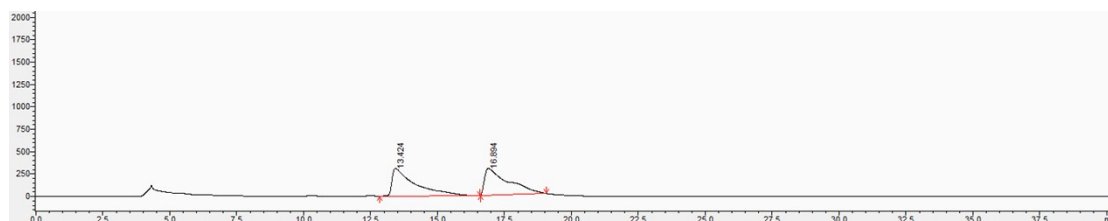


	Retention Time (min)	Area (%)	
Peak 1	30.185	2.728	95% ee
Peak 2	43.549	97.272	



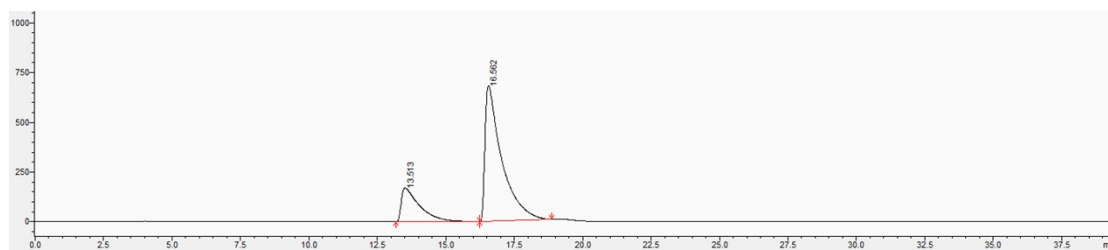
57% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IC-3 column, *n*-hexane /*i*-PrOH = 97/3, 0.8 mL/min, 254 nm.

Racemate:

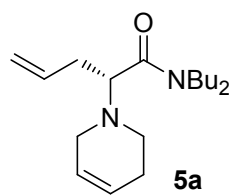


	Retention Time (min)	Area (%)
Peak 1	13.424	50.730
Peak 2	16.894	49.270

Chiral:

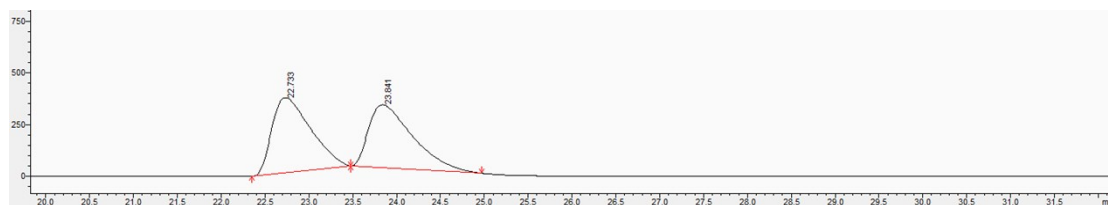


	Retention Time (min)	Area (%)	
Peak 1	13.513	21.427	57% ee
Peak 2	16.562	78.573	



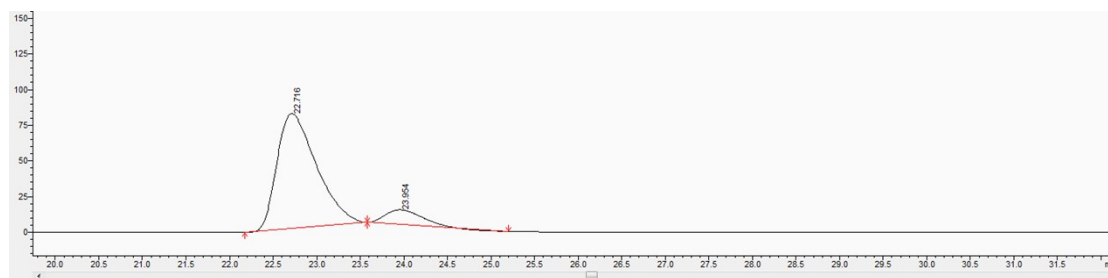
80% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IC-3 column, *n*-hexane /*i*-PrOH = 95/5, 0.8 mL/min, 210 nm.

Racemate:

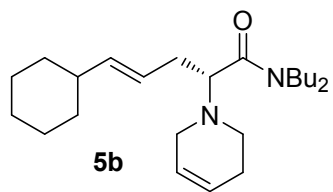


	Retention Time (min)	Area (%)
Peak 1	22.733	51.047
Peak 2	23.841	48.953

Chiral:

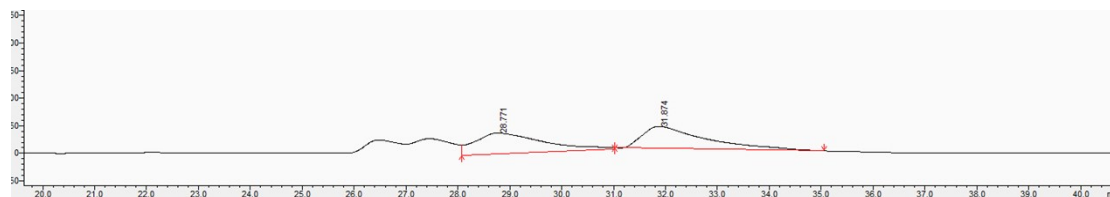


	Retention Time (min)	Area (%)	
Peak 1	22.716	89.816	80% ee
Peak 2	23.954	10.184	



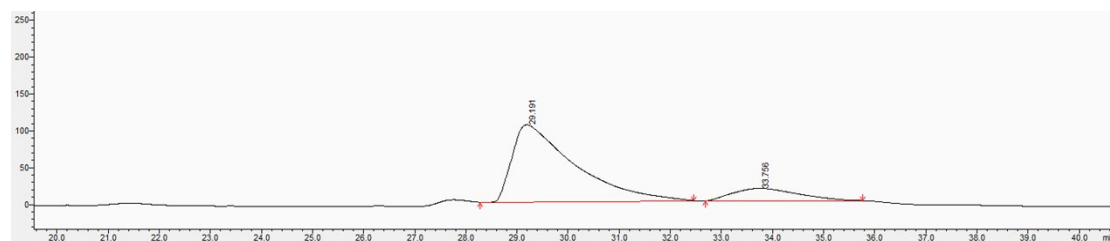
71% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IC-3 column, *n*-hexane /*i*-PrOH = 98/2, 0.8 mL/min, 210 nm.

Racemate:

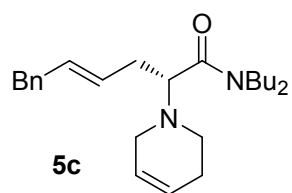


	Retention Time (min)	Area (%)
Peak 1	28.771	50.626
Peak 2	31.874	49.374

Chiral:

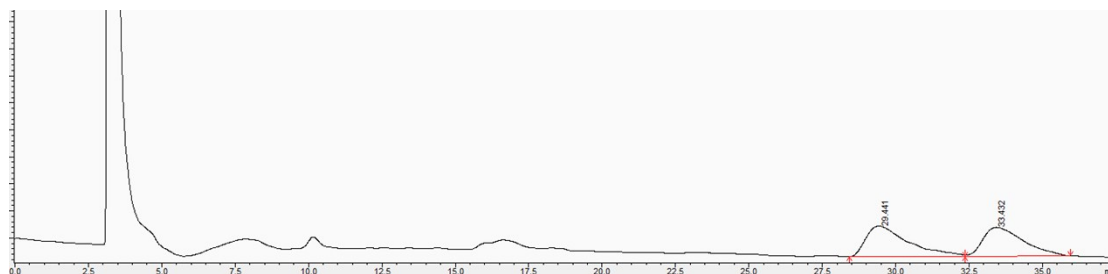


	Retention Time (min)	Area (%)	
Peak 1	29.191	85.607	71% ee
Peak 2	33.756	14.393	



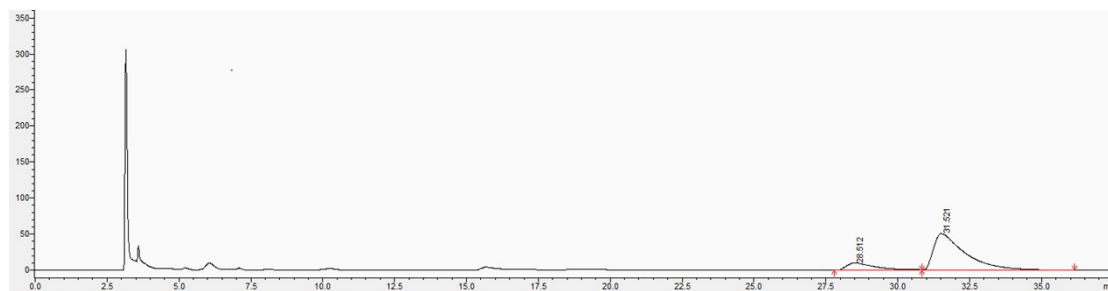
68% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IC-3 column, *n*-hexane /*i*-PrOH = 98/2, 0.8 mL/min, 210 nm.

Racemate:

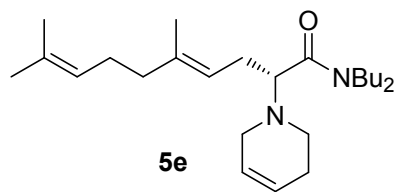


	Retention Time (min)	Area (%)
Peak 1	29.441	51.587
Peak 2	33.432	48.413

Chiral:

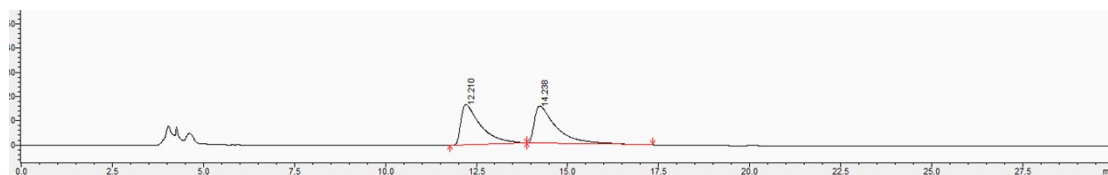


	Retention Time (min)	Area (%)	
Peak 1	28.512	16.026	68% ee
Peak 2	31.521	83.974	



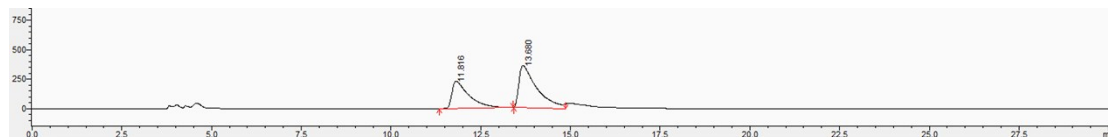
24% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IC-3 column, *n*-hexane /*i*-PrOH = 97/3, 0.8 mL/min, 210 nm.

Racemate:

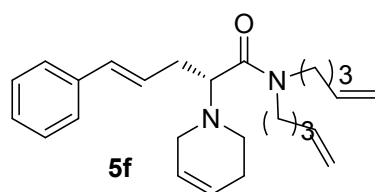


	Retention Time (min)	Area (%)
Peak 1	12.210	49.386
Peak 2	14.238	50.614

Chiral:

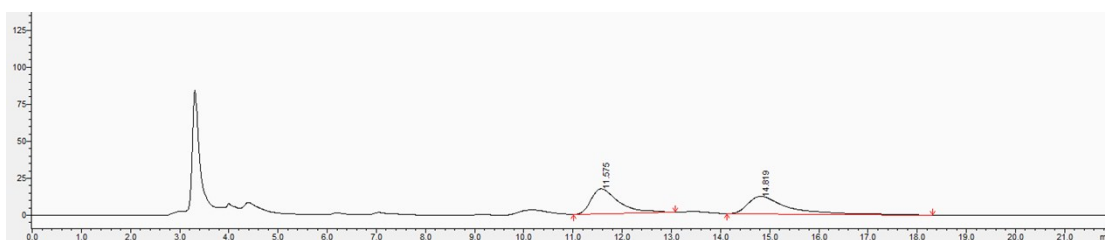


	Retention Time (min)	Area (%)	
Peak 1	11.816	38.002	24% ee
Peak 2	13.680	61.998	



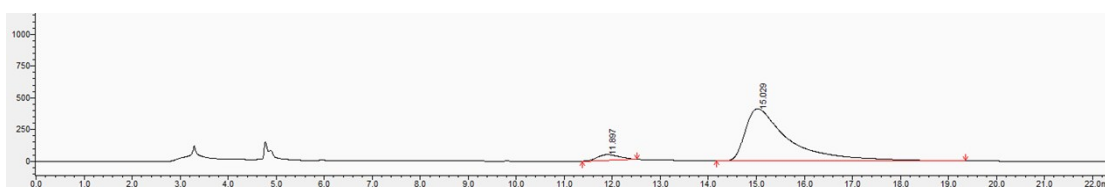
90% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak AD-H column, *n*-hexane /*i*-PrOH = 98/2, 1 mL/min, 254 nm.

Racemate:

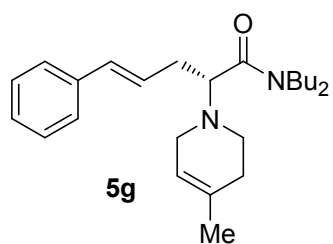


	Retention Time (min)	Area (%)
Peak 1	11.575	50.253
Peak 2	14.819	49.747

Chiral:

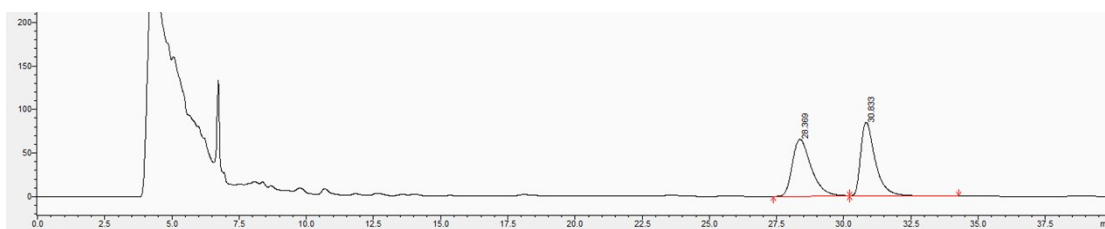


	Retention Time (min)	Area (%)	
Peak 1	11.897	5.215	90% ee
Peak 2	15.029	94.785	



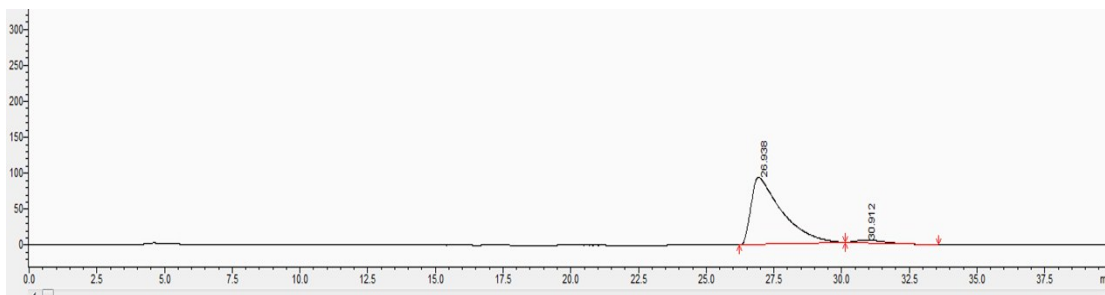
93% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IC-3 column, *n*-hexane /*i*-PrOH = 95/5, 0.8 mL/min, 254 nm.

Racemate:

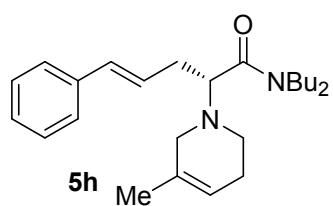


	Retention Time (min)	Area (%)
Peak 1	28.369	49.687
Peak 2	30.833	50.313

Chiral:

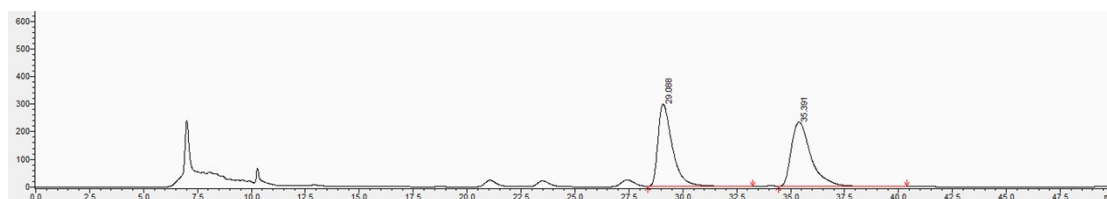


	Retention Time (min)	Area (%)	
Peak 1	26.938	96.530	93% ee
Peak 2	30.912	3.470	



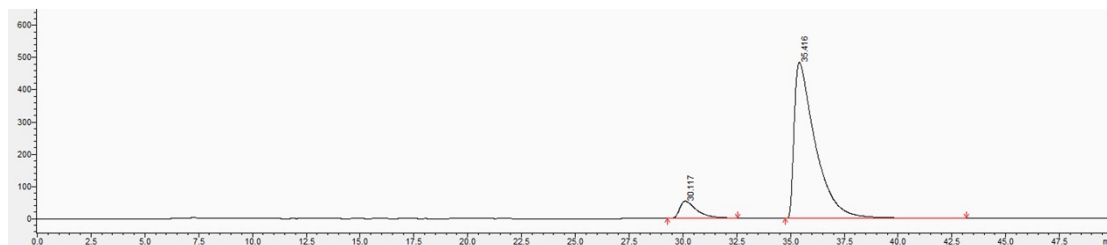
84% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IC-3 column, *n*-hexane /*i*-PrOH = 95/5, 0.8 mL/min, 254 nm.

Racemate:

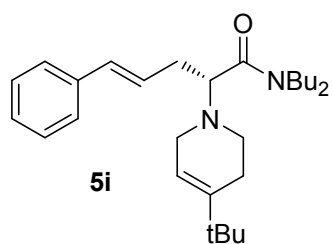


	Retention Time (min)	Area (%)
Peak 1	29.088	48.596
Peak 2	35.391	51.404

Chiral:

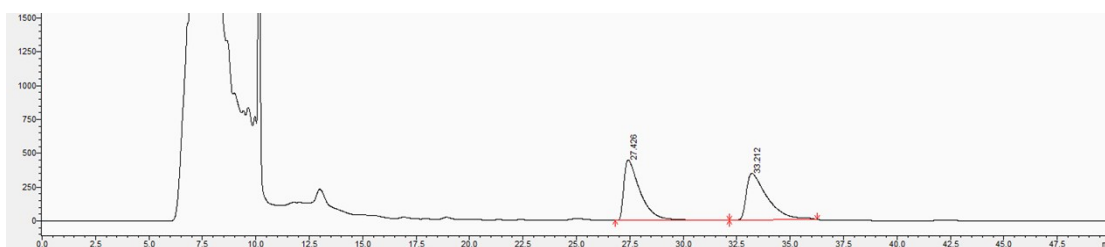


	Retention Time (min)	Area (%)	
Peak 1	30.117	8.147	84% ee
Peak 2	35.416	91.853	



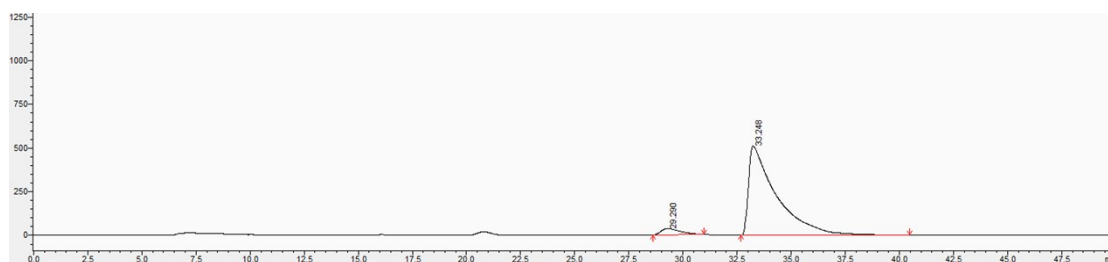
90% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IC-3 column, *n*-hexane /*i*-PrOH = 95/5, 1 mL/min, 254 nm.

Racemate:

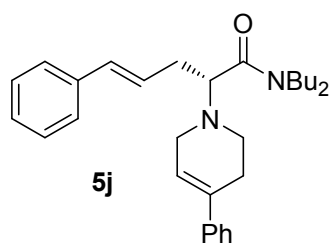


	Retention Time (min)	Area (%)
Peak 1	27.426	50.017
Peak 2	33.212	49.983

Chiral:

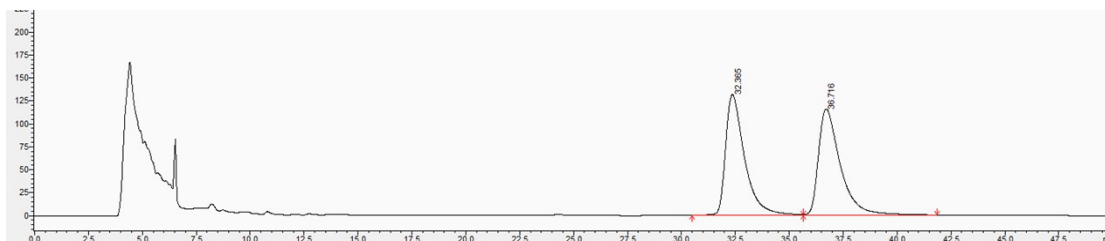


	Retention Time (min)	Area (%)	
Peak 1	29.290	4.801	90% ee
Peak 2	33.248	95.199	



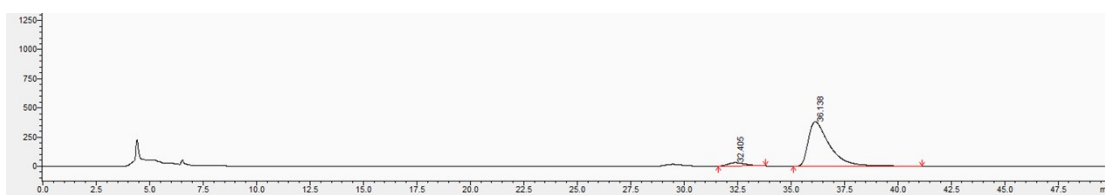
88% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IC-3 column, *n*-hexane /*i*-PrOH = 95/5, 1 mL/min, 254 nm.

Racemate:

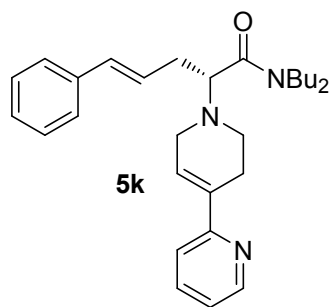


	Retention Time (min)	Area (%)
Peak 1	32.365	49.867
Peak 2	36.716	50.133

Chiral:

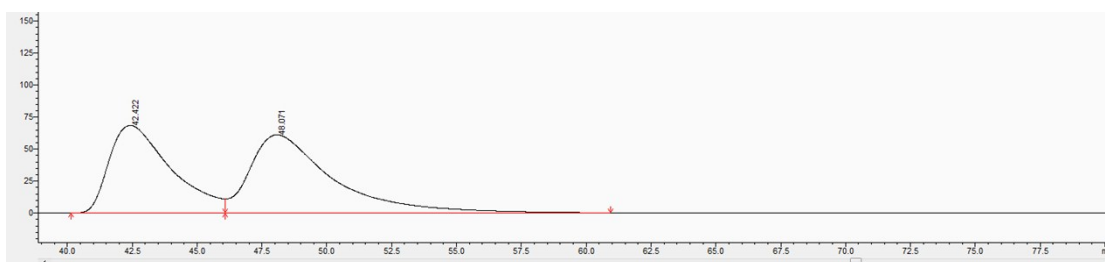


	Retention Time (min)	Area (%)	
Peak 1	32.405	5.962	88% ee
Peak 2	36.138	94.038	



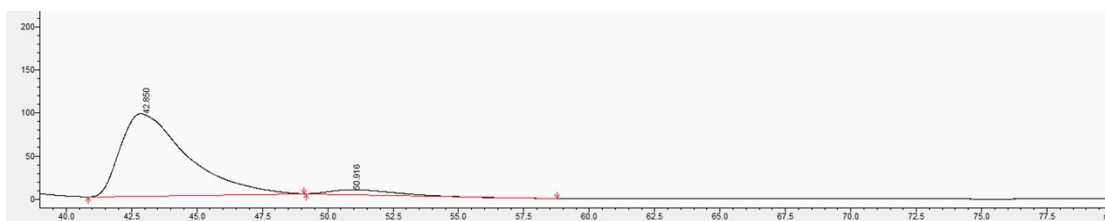
91% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak AD-H column, *n*-hexane /*i*-PrOH = 98.5/1.5, 1 mL/min, 254 nm.

Racemate:

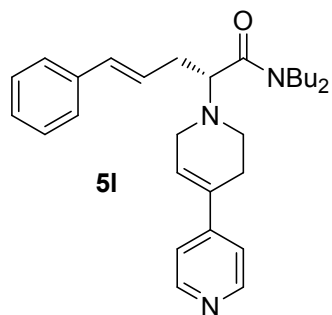


	Retention Time (min)	Area (%)
Peak 1	42.422	48.735
Peak 2	48.071	51.265

Chiral:

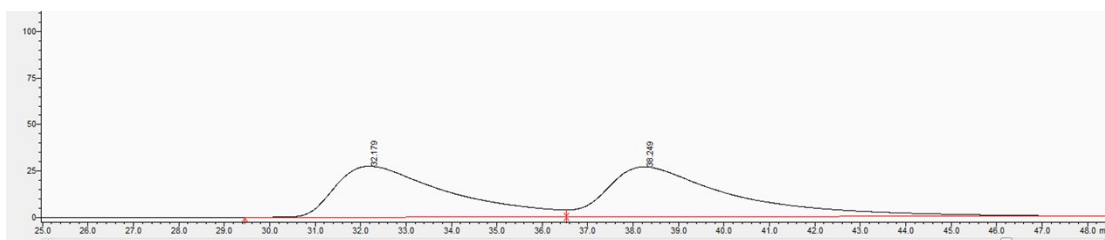


	Retention Time (min)	Area (%)	
Peak 1	42.850	95.371	91% ee
Peak 2	50.916	4.629	



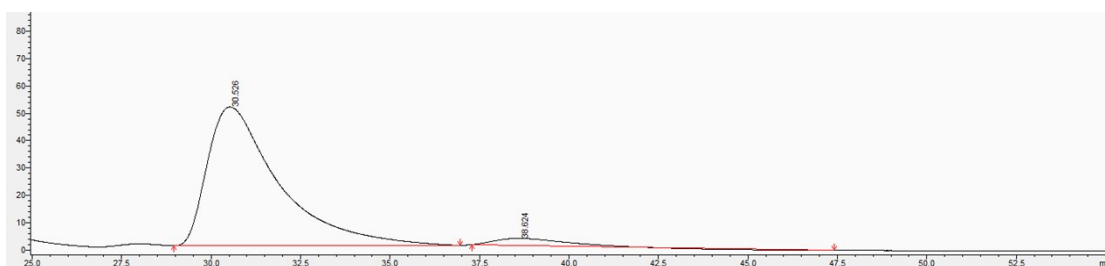
92% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak AD-H column, *n*-hexane /*i*-PrOH = 95/5, 1 mL/min, 254 nm.

Racemate:

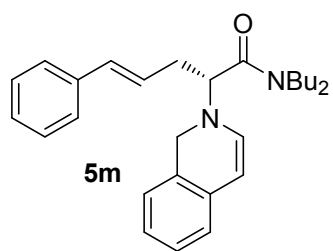


	Retention Time (min)	Area (%)
Peak 1	32.179	47.792
Peak 2	38.249	52.208

Chiral:

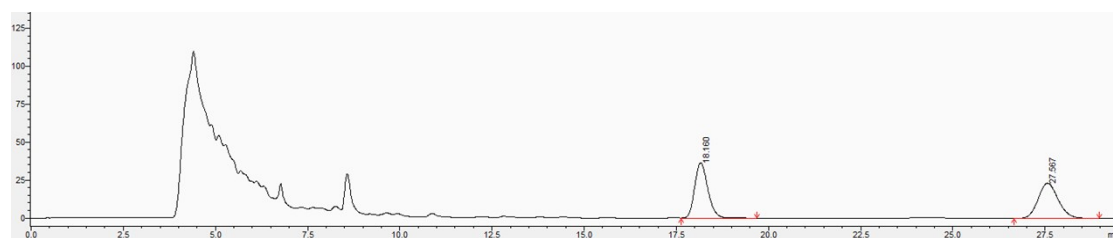


	Retention Time (min)	Area (%)	
Peak 1	30.526	95.910	92% ee
Peak 2	38.624	4.090	



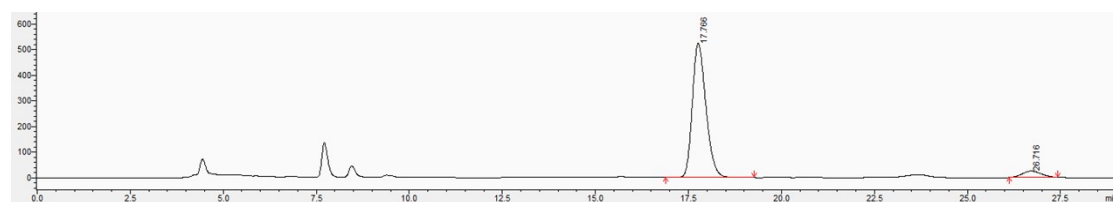
88% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IC-3 column, *n*-hexane /*i*-PrOH = 95/5, 0.8 mL/min, 254 nm.

Racemate:

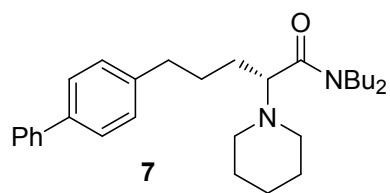


	Retention Time (min)	Area (%)
Peak 1	18.160	50.875
Peak 2	27.567	49.125

Chiral:

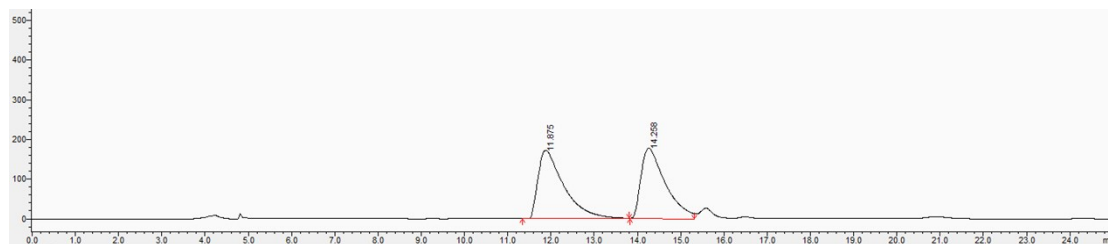


	Retention Time (min)	Area (%)	
Peak 1	17.766	93.794	88% ee
Peak 2	26.716	6.206	



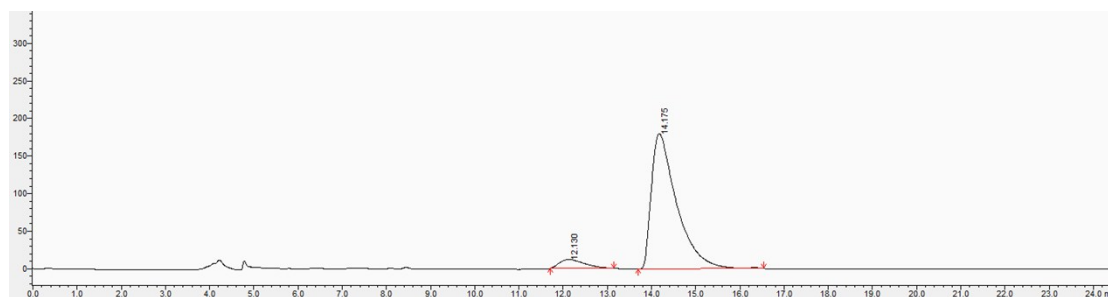
88% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IC-3 column, *n*-hexane /*i*-PrOH = 95/5, 0.8 mL/min, 254 nm.

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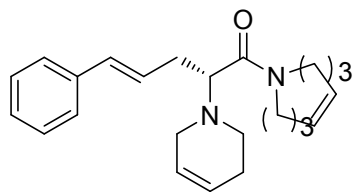


	Retention Time (min)	Area (%)
Peak 1	11.875	50.360
Peak 2	14.258	49.640

Chiral:



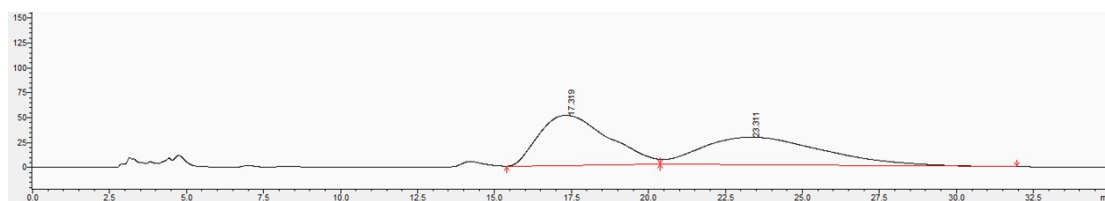
	Retention Time (min)	Area (%)	
Peak 1	12.130	6.148	88% ee
Peak 2	14.175	93.852	



8

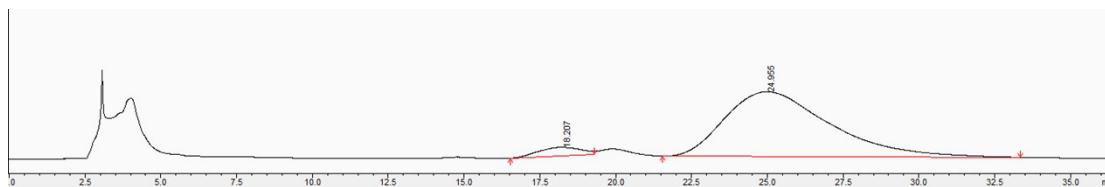
90% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak AD-H column, *n*-hexane /*i*-PrOH = 98/2, 0.8 mL/min, 254 nm.

Racemate:

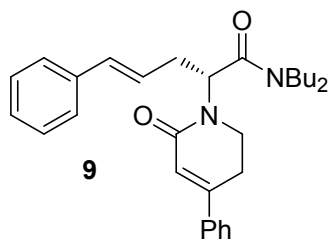


	Retention Time (min)	Area (%)
Peak 1	17.319	51.198
Peak 2	23.311	48.802

Chiral:

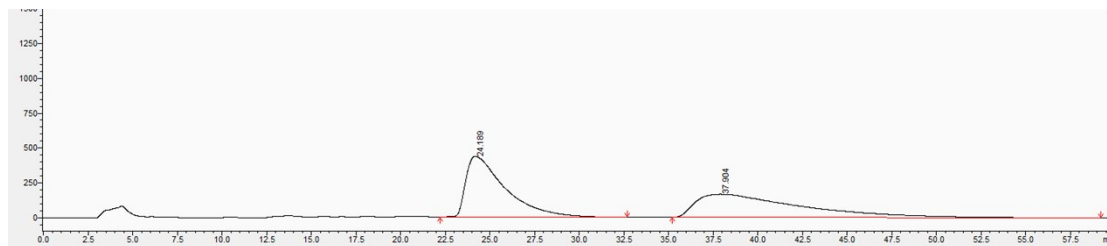


	Retention Time (min)	Area (%)	
Peak 1	18.207	4.950	90% ee
Peak 2	24.955	95.050	



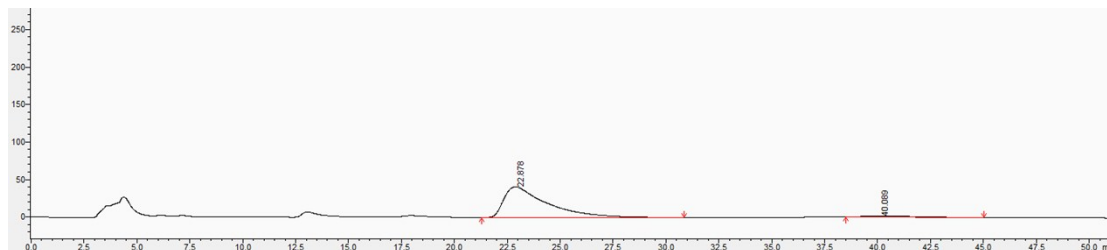
88% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak AD-H column, *n*-hexane /*i*-PrOH = 95/5, 1 mL/min, 254 nm.

Racemate:

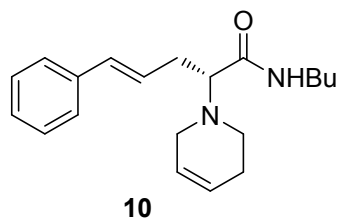


	Retention Time (min)	Area (%)
Peak 1	24.189	50.038
Peak 2	37.904	49.962

Chiral:

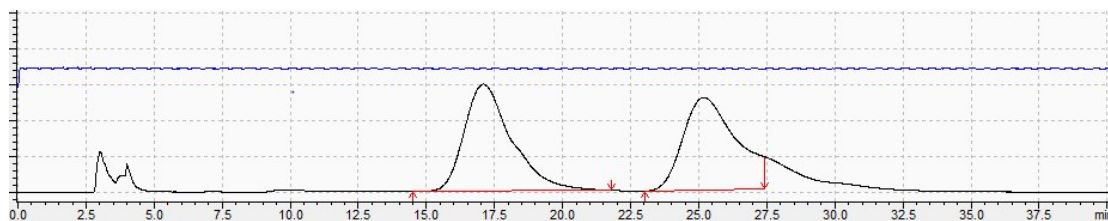


	Retention Time (min)	Area (%)	
Peak 1	22.878	94.076	88% ee
Peak 2	40.089	5.924	



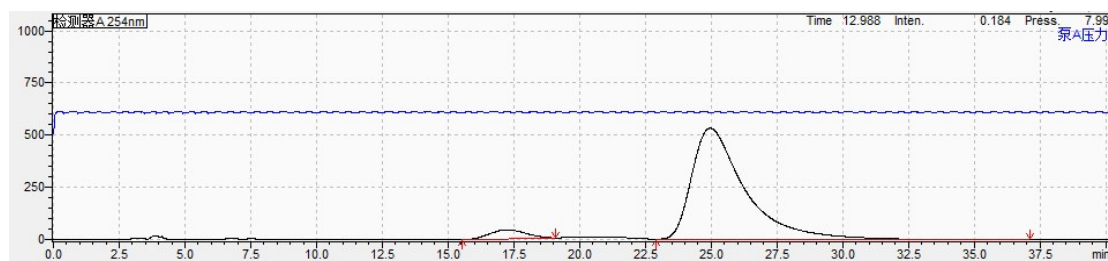
90% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak OD-H column, *n*-hexane /*i*-PrOH = 95/5, 1 mL/min, 254 nm.

Racemate:

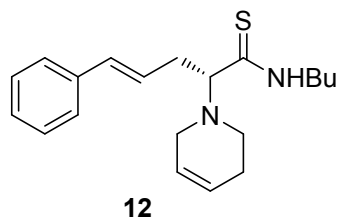


	Retention Time (min)	Area (%)
Peak 1	17.092	50.696
Peak 2	25.188	49.304

Chiral:

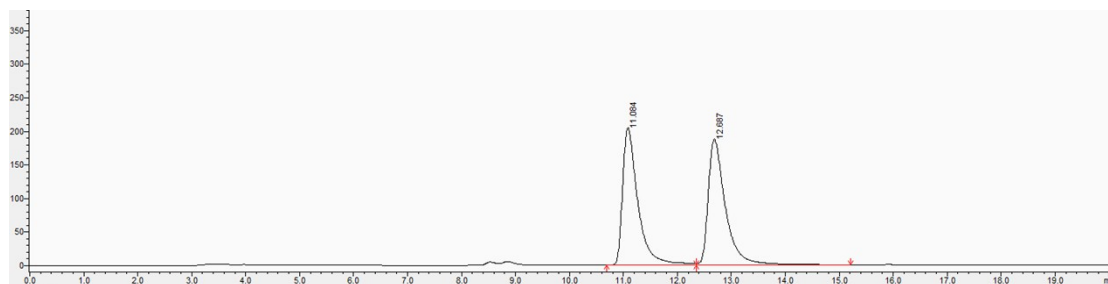


	Retention Time (min)	Area (%)	
Peak 1	17.239	5.169	90% ee
Peak 2	24.966	94.831	



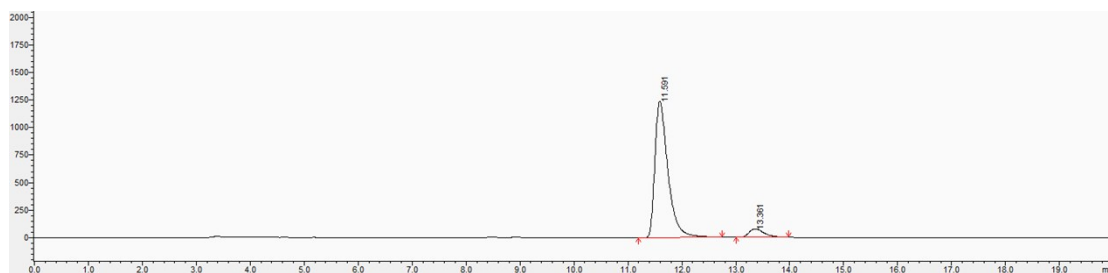
88% *ee*, enantiomeric excess was determined by HPLC, Daicel Chiralpak IC-3 column, *n*-hexane /*i*-PrOH = 95/5, 1 mL/min, 254 nm.

Racemate:



	Retention Time (min)	Area (%)
Peak 1	11.084	49.361
Peak 2	12.687	50.639

Chiral:



	Retention Time (min)	Area (%)	
Peak 1	11.591	94.016	88% ee
Peak 2	13.361	5.984	