

Supplemental Data

***De Novo* SOX4 Variants Cause a Neurodevelopmental
Disease Associated with Mild Dysmorphism**

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Supplemental Note: Case Reports

Subject 1 was a 4-year-and-8-month-old male. Array comparative genomic hybridization (array-CGH) excluded pathogenic copy-number variants, and analysis for fragile-X syndrome was negative. Trio-based exome sequencing revealed no plausible pathogenic mutation beside a heterozygous *de novo* *SOX4* missense variant: hg19 chr6:21594963C>A, p.Phe66Leu. This subject was the son of healthy non-consanguineous Italian parents. He had two older healthy half-sisters (one paternal and one maternal). His mother was 33-year-old at conception and his father 41. Pregnancy was complicated by intrauterine growth retardation and he was born after 37 weeks of gestation by C-section. Birth weight was 1970 g (10th percentile) and length was 43 cm (<3rd percentile). On initial examination at 3 years and 2 months of age, his height was 85 cm (1st percentile), his weight was 11.0 kg (1st percentile) and his head circumference was 46.5 cm (3rd percentile). At 4 years and 8 months of age, his height was 100.7 cm (10th percentile), his weight was 15.0 kg (8th percentile), and his head circumference was 47.6 cm (1st percentile), indicating progressive microcephaly. He had global developmental delay: independent walking was only possible at 27 months and he spoke his first words at 4 years. He had seizures that started at 3.5 years of age. Treatment with valproic acid was started, but the response was unsatisfactory, with the child maintaining brief, daily crises. He was hypotonic and had a perimembranous ventricular septal defect. He had feeding difficulties, was unable to eat solid food, and had dysphagia and severe constipation. Eye examination revealed strabismus and keratoconus. Brain MRI demonstrated a delay in myelination. His IQ score was 68 (mild ID). He had slight facial dysmorphism: microbrachycephaly, epicanthus of the left eye, a stellate iris pattern, a short nose with anteverted nares, a wide mouth with a cupid bow, and posteriorly rotated ears. He also had bilateral clinodactyly of the 5th finger, but hands and feet were otherwise normal. Submission of *SOX4* to GeneMatcher¹ facilitated the present study by identifying Subjects 2 to 4 in the DDD database of *de novo* variants in 4296 children with developmental disorders.²

Subject 2 was an 11-year-and-10-month-old male of Scottish-Hungarian ancestry. He was found by trio-based exome sequencing to carry a heterozygous *de novo* SOX4 missense variant: hg19 chr6:21595099G>C, p.Ala112Pro. No other plausible pathogenic variant was found. CGH revealed a duplication of Yp11.31-11.222 not present in his father and likely unrelated to his phenotype. This child was the fourth of five children of healthy non-consanguineous parents. His father was 66-year-old at conception and his mother 34. Pregnancy was complicated by a bleed in the first trimester which was thought to be associated with the loss of a twin sibling. Born at 40 weeks of gestation, the baby weighed 3.5 kg (46th percentile), had bilateral, vertical talus, which was surgically corrected in infancy, and also had laryngomalacia. There were initial concerns about his weight gain and he had problems weaning. He otherwise reached his developmental milestones, but lost his ability to roll and bear weight almost overnight when he was 10-month-old. His parents associated this with a homeopathic treatment that he was receiving at the time. He developed a spastic quadriparesis, most prominent in the lower limbs. Brain MRI at age 15 months and 6 years demonstrated progressive cerebellar atrophy. There were also some patchy cerebral white matter changes. At 6 years and 8 months of age, his weight was 14.8 kg (1st percentile) and his head circumference was 48.3 cm (<0.4th percentile). His OFC has remained at this size ever since. When he was last seen, at 11 years and 10 months of age, his neurodevelopmental disorder had been static for many years. He was still a slow eater, had problems chewing and requires mashed food. He had never walked, had no speech and had severe intellectual disability. However, he was very alert and aware of his surroundings. He demonstrated bruxism and drooling. Secondary dentition was markedly delayed as he still had several primary teeth. In addition to microcephaly, he had trigonocephaly with a mild metopic ridge, bilateral epicanthic folds, infra-orbital folds, a wide mouth and a defined cupid's bow of the upper lip. He also had mild 5th finger clinodactyly.

Subject 3 was a 6-year-old female. Trio-based exome sequencing indicated a heterozygous *de novo* SOX4 missense variant as sole pathogenic candidate variant: hg19 chr6:21594941 T>G, p.Ile59Ser. This child was small for her age (6th percentile for height, 9th for weight, and 12th for OFC). Her main problem at 3 years was delayed speech, with a limited vocabulary of single words. She responded well to speech therapy, and her speech was normal at 6 years. She had very mild learning disabilities and was not receiving extra help at mainstream school. The school were not concerned, but her mother reported that she was struggling to keep up with peers. She had characteristic facial features, with deep-set eyes, infra-orbital grooves, flat nasal bridge, upturned nares, wide cupid-bow mouth and full lips. She also had mild 5th finger clinodactyly and dysplastic 5th toenails.

Subject 4 was an 8-year-and-10-month-old female. Trio-exome sequencing showed a heterozygous *de novo* SOX4 missense variant: hg19 chr6:21595080 G>T, p.Lys105Asn. No other pathogenic variant candidate was found. She had a normal delivery after 39 weeks of gestation, but had a low birth weight (2.35 kg, 1st percentile). When initially assessed at 6 years and 10 months of age, she was small for her age, but had a normal head circumference (3rd percentile for height and weight, and 45th percentile for OFC). She did not walk until 21 months of age and pronounced her first words at 24 months. At 8 years and 10 months of age, her full-scale IQ (WISC-IV UK) was 52. She was in mainstream school and receiving educational support. She was sociable and had satisfactory speech, despite earlier expressive difficulties. She was yet unable to read or write, and had no sense of danger. She had a characteristic facial appearance, with deep-set eyes, infra-orbital creases, malar flattening, upturned nares, wide cupid-bow mouth and full lips. The ears were low set, and posteriorly rotated. There was mild 5th finger clinodactyly and mild camptodactyly.

Supplemental Figures

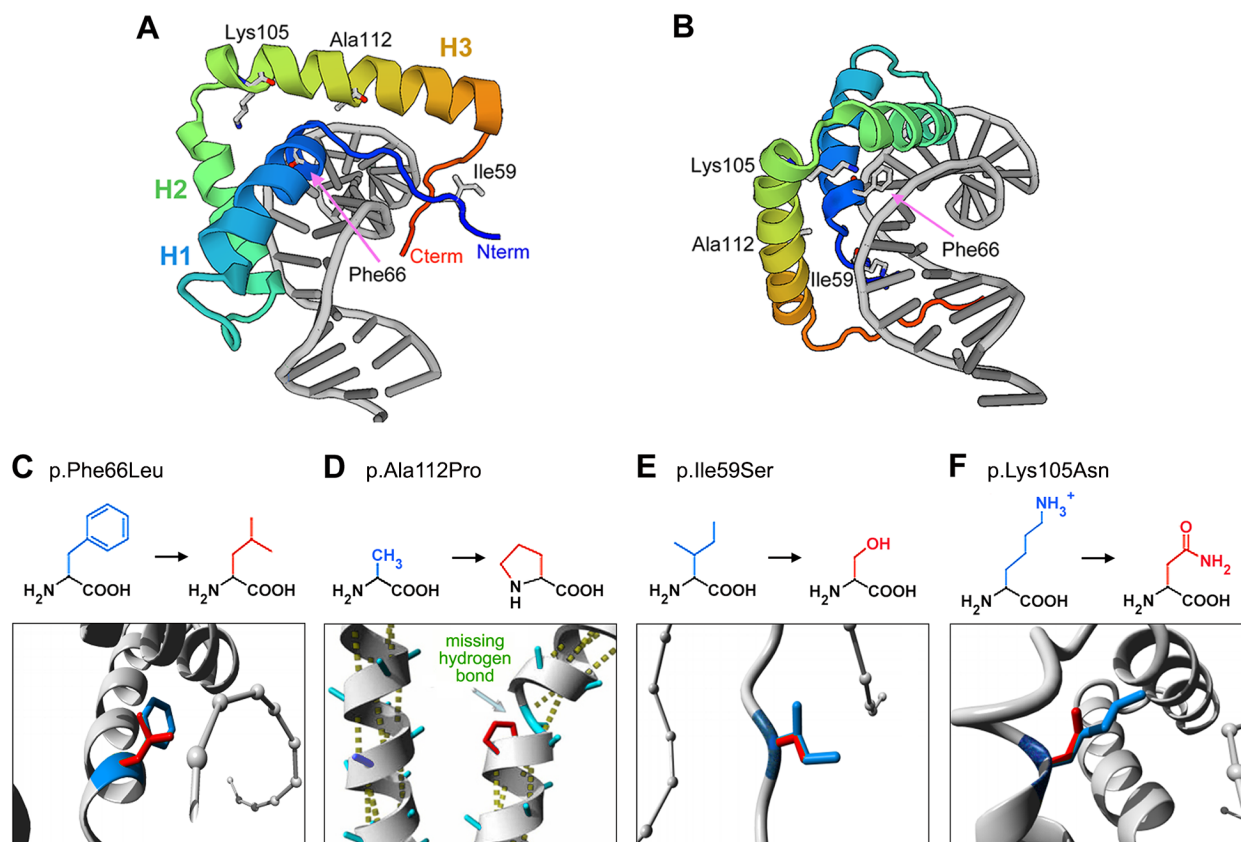


Figure S1. *In Silico* Prediction of Changes in the HMG Domain Structure Induced by the Four Subjects' SOX4 Missense Variants.

(A) Rendering of the wild-type SOX4 HMG domain–DNA complex. DNA is shown in grey and the SOX4 HMG domain is rainbow-colored. Its α -helices are labeled H1, H2, and H3, and its N- and C-termini are indicated. The residues mutated in our subjects are indicated and their side chains are depicted. The HMG domain wraps around the DNA helix and induces a strong bend. This cartoon was generated by SWISS-MODEL using published data.³

(B) Same protein–DNA model as in panel A, but oriented differently to show the intercalation of the Phe66 side chain into the minor groove of the DNA helix.

(C to F) Top, schematics of wild-type and variant amino acids, with side chains colored in blue and red, respectively. Bottom, ribbon presentation of the SOX4 protein structure generated by HOPE to show structural changes caused by the variants. The protein is presented in grey and the side chains of the wild-type and mutant amino acids are colored in blue and red, respectively.

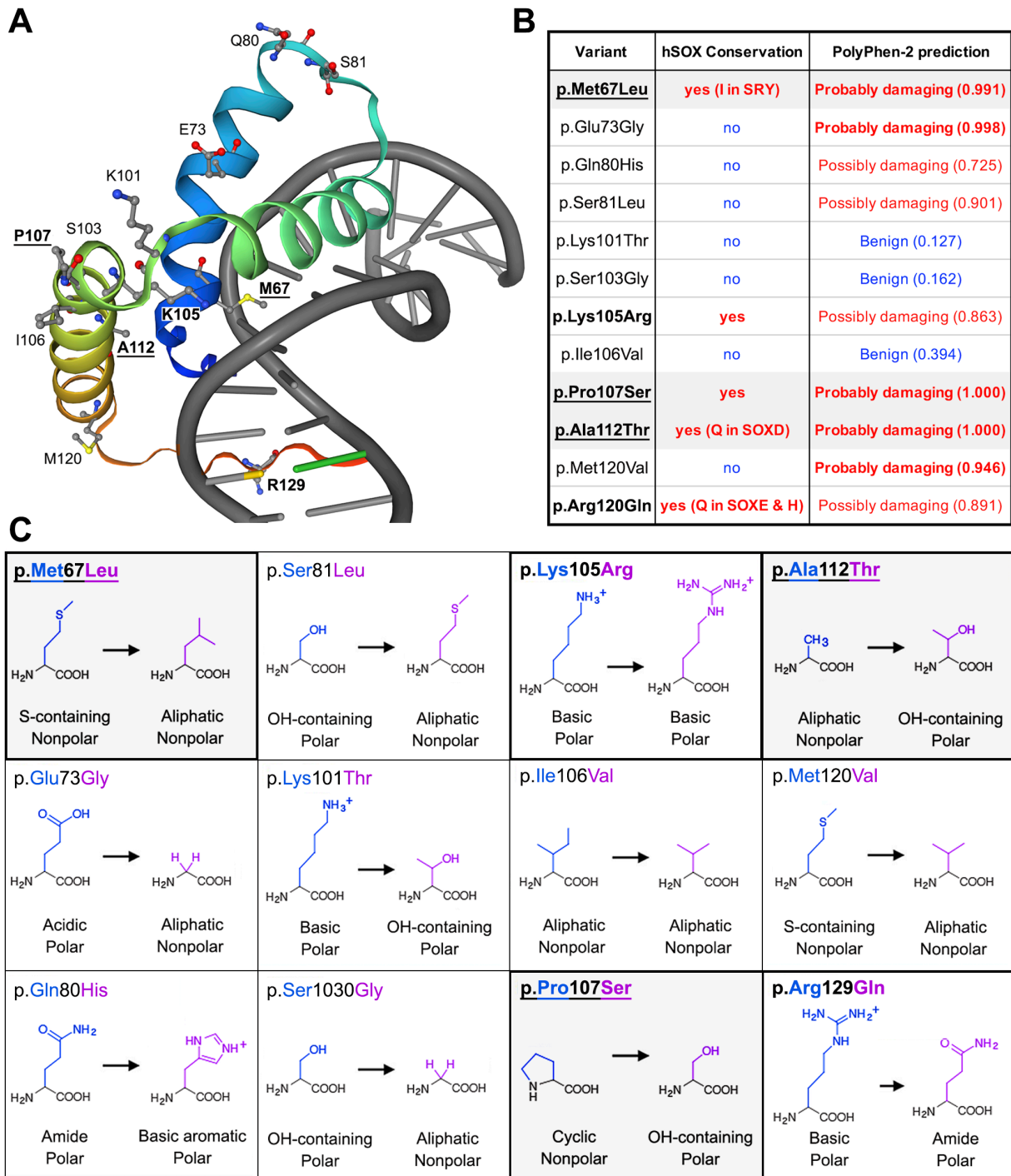


Figure S2. *In Silico* Prediction of Changes in the HMG Domain Structure Induced by SOX4 Missense Variants Identified in gnomAD.

(A) Rendering of the wild-type SOX4 HMG domain–DNA complex. DNA is shown in grey and the SOX4 HMG domain is rainbow-colored. Its α -helices are labeled H1, H2, and H3, and its N- and

C-termini are indicated. The residues showing variants in gnomAD are indicated and their side chains are depicted. Bold characters are used for five residues that are highly conserved in human SOX proteins. Three of these residues are also underlined because their mutation is predicted by PolyPhen-2 to be probably damaging. This cartoon was generated by SWISS-MODEL using published data.³

(B) Table summarizing for each SOX residue that features a variant in gnomAD (left column) the degree of conservation in human SOX proteins (middle column) and the prediction score attributed by PolyPhen-2 for the consequence of the variant (right column). In the middle column, “yes” means conservation in all 20 SOX proteins; “no” means poor conservation; parentheses indicate residues found in other SOX proteins. Bold characters are used for five residues that are highly conserved in human SOX proteins. Three of these residues are also underlined and their boxes shaded in light grey because their mutation is predicted by PolyPhen-2 to be probably damaging.

(C) Schematics of wild-type and gnomAD variant amino acids, with side chains colored in blue and red, respectively. The features of the side chains are indicated. Bold characters are used for five residues that are highly conserved in human SOX proteins. Three of these residues are also underlined and their boxes shaded in light grey because their mutation is predicted by PolyPhen-2 to be probably damaging.

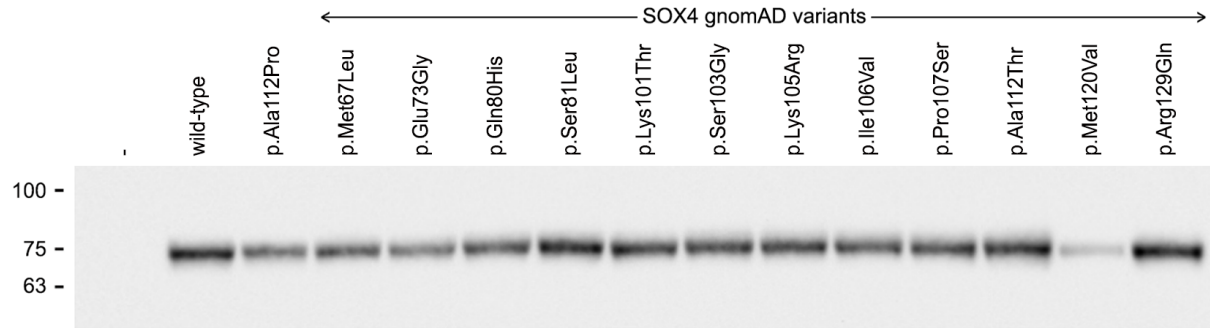


Figure S3. Assessment of SOX4 gnomAD Variant Levels Achieved in COS-1 Cells.

Western blot of lysates from COS-1 cells transiently transfected with expression plasmids for no protein (-), wild-type SOX4 (WT), the case-2 variant (p.Ala112Pro), and the twelve SOX4 gnomAD variants, as indicated. The blot was hybridized with an antibody recognizing the FLAG epitope added at the N-terminus of SOX4. The Mr of protein standards is indicated on the left (in k units). All SOX4 proteins migrate with an expected Mr of 70 k. This experiment complements the reporter assay shown in Figure 6D.

Supplemental Tables

Table S1. NCBI Accession Numbers for the HMG-Domain Sequences of Vertebrate SOX4 Orthologs Aligned in Figure 5B.

Vertebrate Species	Reference Sequence
Human (<i>Homo sapiens</i>)	NM_003107.2
Mouse (<i>Mus musculus</i>)	NM_009238.3
Cow (<i>Bos taurus</i>)	NM_001078128.1
Horse (<i>Equus caballus</i>)	XM_023624515.1
Bat (<i>Eptesicus fuscus</i>)	XM_008146131.1
Hedgehog (<i>Erinaceus europaeus</i>)	XM_007525152.2
Elephant (<i>Loxodonta africana</i>)	XM_010596185.2
Opossum (<i>Monodelphis domestica</i>)	XM_007487879.2
Platypus (<i>Ornithorhynchus anatinus</i>)	XM_007659031.1
Chicken (<i>Gallus gallus</i>)	AY249864.1
Frog (<i>Xenopus laevis</i>)	NM_001172201.1
Zebrafish (<i>Danio rerio</i>)	AY369082.1

Table S2. NCBI Accession Numbers for the HMG-Domain Sequences of All Human SOX Proteins Aligned in Figure 5C.

SOX protein	Reference Sequence
SOX1	NP_005977.2
SOX2	NP_003097.1
SOX3	NP_005625.2
SOX4	AAH72668.1
SOX5	AAH60773.1
SOX6	AAK26115.1
SOX7	CAC84226.1
SOX8	NP_055402.2
SOX9	CAA86598.1
SOX10	CAG38808.1
SOX11	BAA88122.1
SOX12	AAH67361.1
SOX13	AAD50120.1
SOX14	AAC95380.1
SOX15	AAH72003.1
SOX17	BAB83867.1
SOX18	BAA94874.1
SOX21	AAC95381.1
SOX30	BAA37146.1
SRY	CAA37790.1

Table S3. DNA Oligonucleotides Used to Generate SOX4 Variant Expression Plasmids. Wild-type and variant codons are shown in blue and red letters, respectively. Wild-type sequences are shown as references.

Subject Variants	Forward Primers	Reverse Primers
Wild-type	TGAACGCC TTT ATGGTGTGGTCGCAGA	ACCACACCAT AAA GGCGTTCATGGGCC
p.Phe66Leu	TGAACGCC TTA ATGGTGTGGTCGCAGA	ACCACACCAT TAA GGCGTTCATGGGCC
Wild-type	CATCCAGGAG GCG GAGCGGCTGCGCCT	AGGCGCAGCCGCTC CGC CTCCTGGATG
p.Ala112Pro	CATCCAGGAG CCG GAGCGGCTGCGCCT	AGGCGCAGCCGCTC CGG CTCCTGGATG
Wild-type	AGTGCCAC ATC AAGCGGCCCATGAAC	GTTTCATGGGCCGCTT GAT GTGGCCACT
p.Ile59Ser	AGTGCCAC AGC AAGCGGCCCATGAAC	GTTTCATGGGCCGCTT GCT GTGGCCACT
Wild-type	ACAGCGAC AAG ATTCCGTTTCATCCAGG	CCTGGATGAACGGAAT CTT GTGCGTGT
p.Lys105Asn	ACAGCGAC AAT ATTCCGTTTCATCCAGG	CCTGGATGAACGGAAT ATT GTGCGTGT
gnomAD Variants	Forward Primers	Reverse Primers
Wild-type	ACGCCTTT ATG GTGTGGTCGCAGATCG	CCACAC CAT AAAGGCGTTCATGGGCCG
p.Met67Leu	ACGCCTTT CTG GTGTGGTCGCAGATCG	CCACAC CAG AAAGGCGTTCATGGGCCG
Wild-type	GCAGATC GAG CGGCGCAAGATCATGGA	TGCGCCG CTC GATCTGCGACCACACCA
p.Glu73Gly	GCAGATC GGG CGGCGCAAGATCATGGA	TGCGCCG CCC GATCTGCGACCACACCA
Wild-type	ATGGAG CAG TCGCCCGACATGCACAAC	TCGGGCGA CTG CTCCATGATCTTGCGC
p.Gln80His	ATGGAG CAC TCGCCCGACATGCACAAC	TCGGGCGA GTG CTCCATGATCTTGCGC
Wild-type	GGAGCAG TCG CCCGACATGCACAACGC	TGTCGGG CGA CTGCTCCATGATCTTGC
p.Ser81Leu	GGAGCAG TTG CCCGACATGCACAACGC	TGTCGGG CAA CTGCTCCATGATCTTGC
Wild-type	GCTGCTC AAG GACAGCGACAAGATTCC	CGCTGTC CTT GAGCAGCTTCCAGCGTT
p.Lys101Thr	GCTGCTC ACG GACAGCGACAAGATTCC	CGCTGTC CGT GAGCAGCTTCCAGCGTT
Wild-type	TCAAGGAC AGC GACAAGATTCCGTTCA	CTTGTC GCT GTCCCTTGAGCAGCTTCCA
p.Ser103Gly	TCAAGGAC GGC GACAAGATTCCGTTCA	CTTGTC GCC GTCCCTTGAGCAGCTTCCA
Wild-type	CAGCGAC AAG ATTCCGTTTCATCCAGGA	ACGGAAT CTT GTGCTGTCTTCCAGCA
p.Lys105Arg	CAGCGAC AGG ATTCCGTTTCATCCAGGA	ACGGAAT CCT GTGCTGTCTTCCAGCA
Wild-type	GCGACAAG ATT CCGTTTCATCCAGGAGG	GAACGG AAT CTTGTGCTGTCTTCCAG
p.Ile106Val	GCGACAAG GTT CCGTTTCATCCAGGAGG	GAACGG AAC CTTGTGCTGTCTTCCAG
Wild-type	ACAAGATT CCG TTCATCCAGGAGGCGG	GATGAA CGG AATCTTGTGCTGTCTTCC
p.Pro107Ser	ACAAGATT TCG TTCATCCAGGAGGCGG	GATGAA CGA AATCTTGTGCTGTCTTCC
Wild-type	TCCAGGAG GCG GAGCGGCTGCGCCTCA	CCGCTC CGC CTCCTGGATGAACGGAAT
p.Ala112Thr	TCCAGGAG ACG GAGCGGCTGCGCCTCA	CCGCTC CGT CTCCTGGATGAACGGAAT
Wild-type	TCAAGCAC ATG GCTGACTACCCTGACT	TCAAG CAT GTGGCTGACTACCCTGACT
p.Met120Val	TCAAGCAC GTG GCTGACTACCCTGACT	TCAAG CAC GTGGCTGACTACCCTGACT
Wild-type	CAAGTAC CGG CCGCGAAAGAAGGTGAA	TTCGCGG CCG GTAAGTGTAGTCAGGGT
p.Arg129Gln	CAAGTAC CAG CCGCGAAAGAAGGTGAA	TTCGCGG CTG GTAAGTGTAGTCAGGGT

Table S4. Missense Variants in SRY and Other SOX Protein Residues Matching SOX4 HMG-domain Variants Identified in gnomAD. Mutations of interest, i.e., resulting in the same variants as in SOX4 gnomAD individuals, are written in blue if they were identified in gnomAD and in red if they were identified in disease. Note that the last SOX4 variant matches wild-type SOX10 and is therefore also written in blue.

Protein	Variant	Phenotype	Reference
SOX4	p.Met67Leu	Unknown	gnomAD
SRY	p.Ile68Thr	Gonadal dysgenesis	4
SOX9	p.Met113Thr	Campomelic dysplasia	5
SOX9	p.Met113Val	Acampomelic Dysplasia	6
SOX10	p.Met112Ile	Waardenburg syndrome	(Pingault, 2014)
SOX4	p.Glu73Gly	Unknown	gnomAD
SRY	p.Gln74His	XY sex reversal	4
SOX9	p.Ala119Val	Campomelic dysplasia	7
SOX11	p.Glu63Gln	Unknown	gnomAD
SOX4	p.Gln80His	Unknown	gnomAD
SOX9	p.Gln126His	Unknown	gnomAD
SOX4	p.Ser81Leu	Unknown	gnomAD
SOX10	p.Tyr126His	Unknown	gnomAD
SOX4	p.Lys101Thr	Unknown	gnomAD
SOX9	p.Asn147Thr	Unknown	gnomAD
SOX4	p.Ser103Gly	Unknown	gnomAD
SOX4	p.Lys105Arg	Unknown	gnomAD
SRY	p.Lys106Ile	Gonadal dysgenesis	4
SOX4	p.Ile106Val	Unknown	gnomAD
SOX9	p.Arg152Pro	Campomelic dysplasia	7
SOX11	p.Ile96Thr	Unknown	gnomAD

SOX4	p.Pro107Ser	Unknown	gnomAD
SRY	p.Pro108His	46, XY gonadal dysgenesis	4
SOX4	p.Ala112Thr	Unknown	gnomAD
SRY	p.Ala113Thr	Gonadal dysgenesis	4
SOX9	p.Ala158Val	Campomelic dysplasia	7
SOX9	p.Ala158Thr	Campomelic dysplasia, XY sex reversal	7
SOX10	p.Ala157Val	Waardenburg syndrome type IV	8
SOX11	p.Ala102Val	Coffin-Siris syndrome	9
SOX4	p.Met120Val	Unknown	gnomAD
SOX10	p.Lys165Arg	Unknown	gnomAD
SOX4	p.Arg129Gln	Unknown	gnomAD
SOX10	p.Gln174Pro	Waardenburg syndrome	10

Supplemental References

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