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Spy vs. Spy: Selecting the best reporter for ^{19}F NMR competition experiments

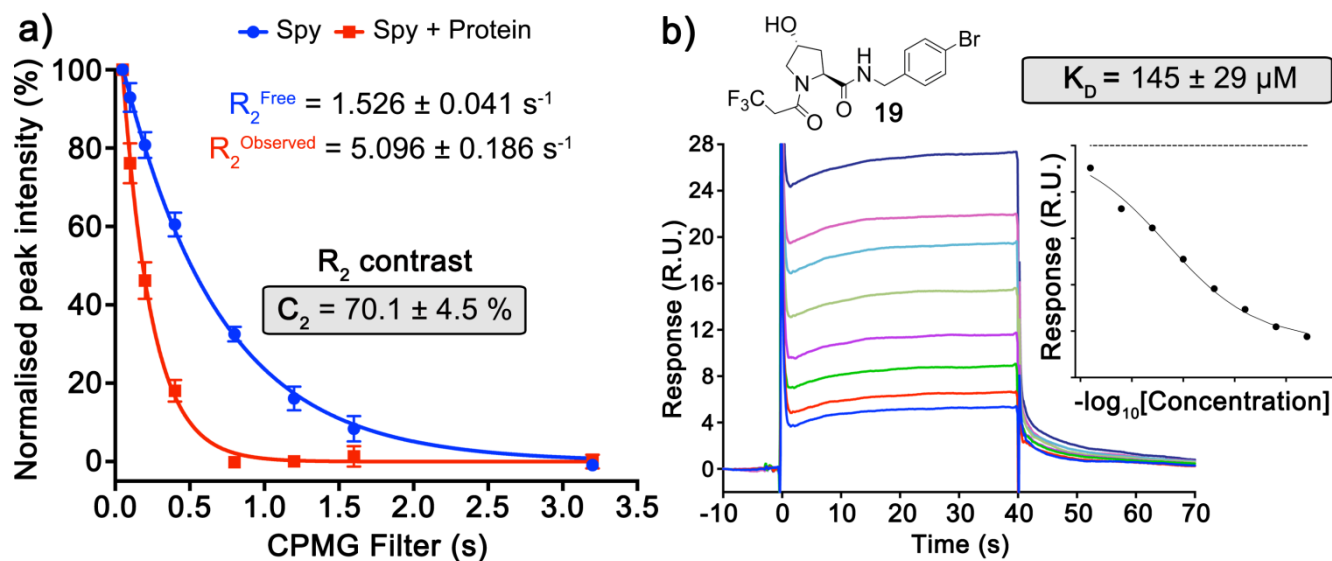
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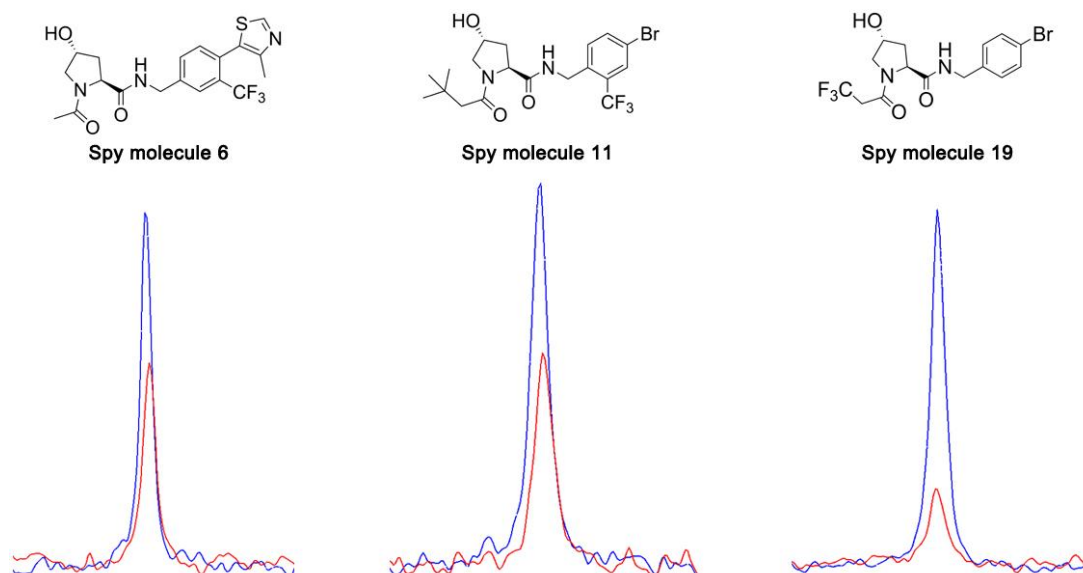
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1. Supplementary figures

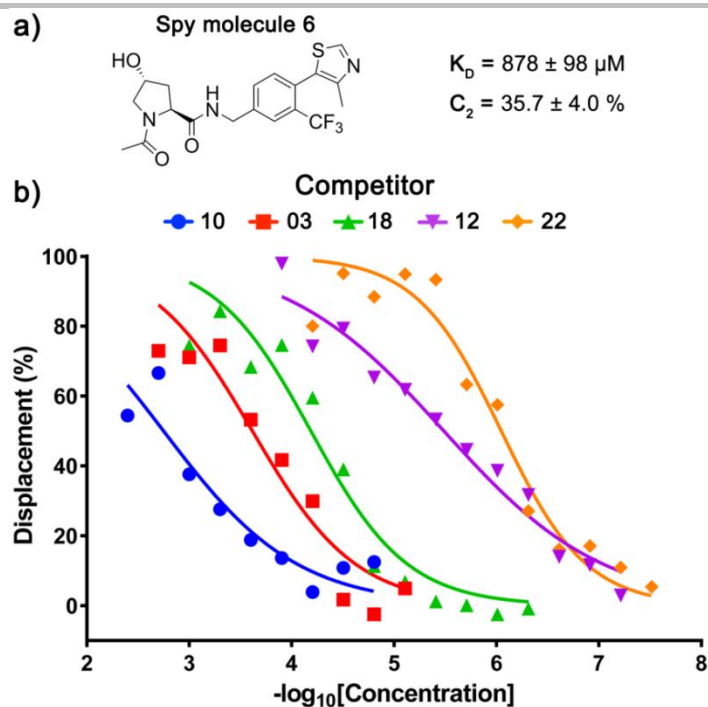


Supplementary Figure 1. Biophysical characterization of spy molecule 19 by ^{19}F NMR and SPR. (a) Measurement of the transverse relaxation rate (R_2) of 19 at 100 μM in absence (blue) and in presence (red) of protein at 1 μM . (b) Measurement of the K_D of 19 by SPR.

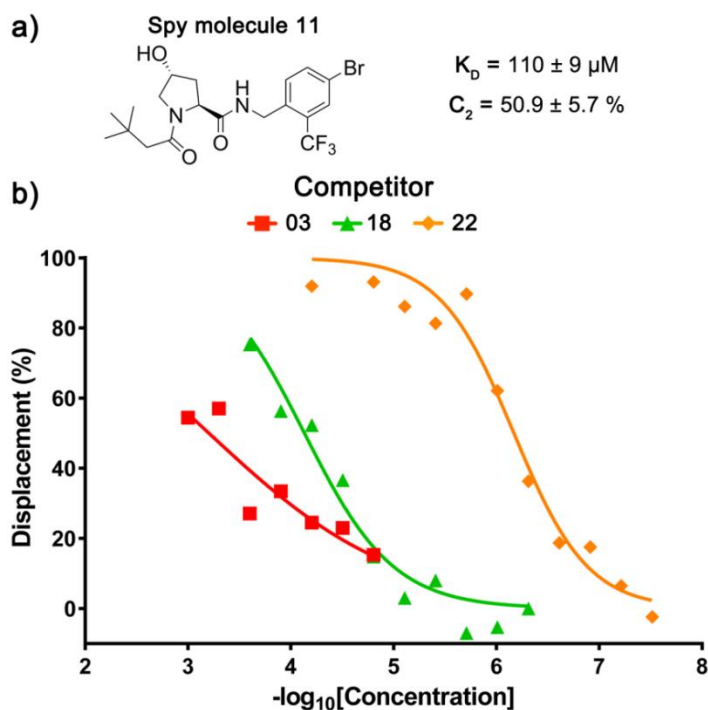


Supplementary Figure 2. Comparison of the assay window of spy molecules 6, 11 and 19 at the same conditions. Overlay of the main ^{19}F CPMG peak (40 scans) of spy molecules 6, 11 and 19 at 50 μM in absence (blue) or in presence of VBC 1 μM , highlighting the wider assay window of spy molecule 19. The CPMG delays used for spy molecules 6, 11 and 19 were, respectively, 634, 447 and 345 ms (see section 6 for details regarding the choice of CPMG delay)

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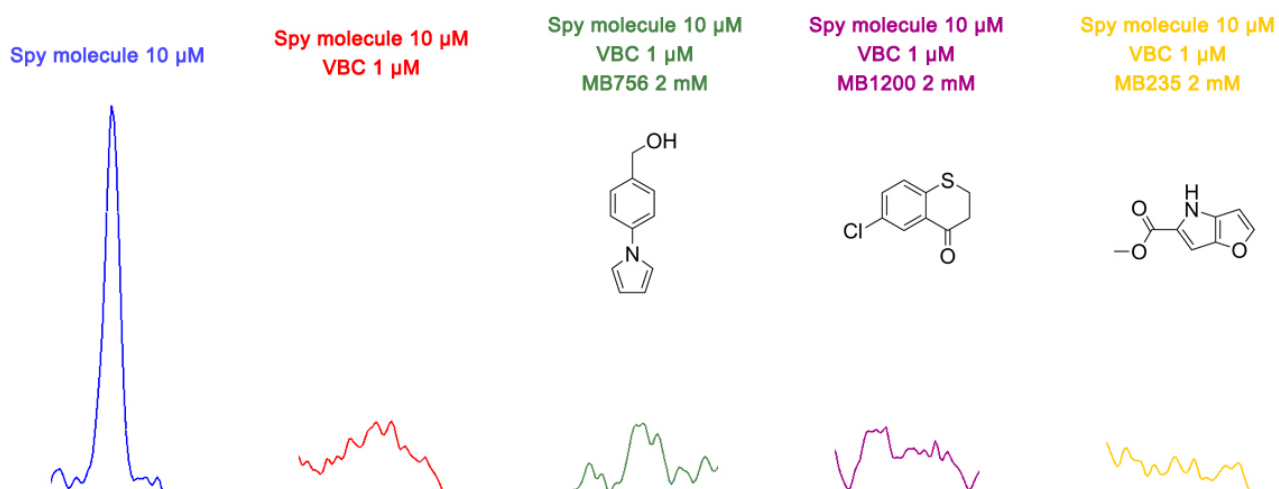


Supplementary Figure 3. Determination of the affinities of VHL binders using spy molecule 6. (a) Structure, dissociation constant and R_2 contrast of spy molecule 6. The C_2 was obtained with spy molecule at 50 μM and VBC at 1 μM . (b) Displacement of spy molecule 6 in presence of different concentrations of five VHL binders (molecules 3, 10, 12, 18 and 22). Data obtained from ^{19}F CPMG experiments using a CPMG delay of 634 ms and 40 scans.



Supplementary Figure 4. Determination of the affinities of VHL binders using spy molecule 11. (a) Structure, dissociation constant and R_2 contrast of spy molecule 11. The C_2 was obtained with spy molecule at 50 μM and VBC at 1 μM . (b) Displacement of spy molecule 11 in presence of different concentrations of five VHL binders (molecules 3, 18 and 22). Competitors 10 and 12 were not used in this case because the chemical shifts of their fluorine peaks overlaid with the spy molecule. Data obtained from ^{19}F CPMG experiments using a CPMG delay of 447 ms and 40 scans.

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Supplementary Figure 5. Binders of other sites present in the VBC complex do not displace spy molecule 19. ^{19}F CPMG NMR peak of spy molecule 19 at different conditions: free in solution (blue), in presence of VBC (red) and in presence of protein and molecules MB756 (green), MB1200 (violet) and MB235 (yellow), binders of other sites present in VBC previously reported.^[1] As these compounds bind to sites not targeted by the spy molecule, no displacement was observed, showing that the competition experiment is site specific. Experiments were performed with a CPMG delay of 258 ms and 120 scans.

2. Compound synthesis and characterization

All the compounds, reagents and solvents used, aside from the compounds specifically prepared for this work, were obtained from commercial sources and used without further purification. For compound purification of intermediates, flash column chromatography was performed using a Teledyne Isco Combiflash Rf or Rf200i, with RediSep Rf Disposable Columns (Normal phase). Where specified, compounds were purified using a Gilson Preparative HPLC System equipped with a Waters X-Bridge C18 column (100 mm x 19 mm; 5 μ m particle size) using an eleven minutes gradient (25 mL/min) of: 1) 5% to 95% of acetonitrile : 0.1% formic acid, or 2) 5% to 95% of acetonitrile : 0.1% ammonia.

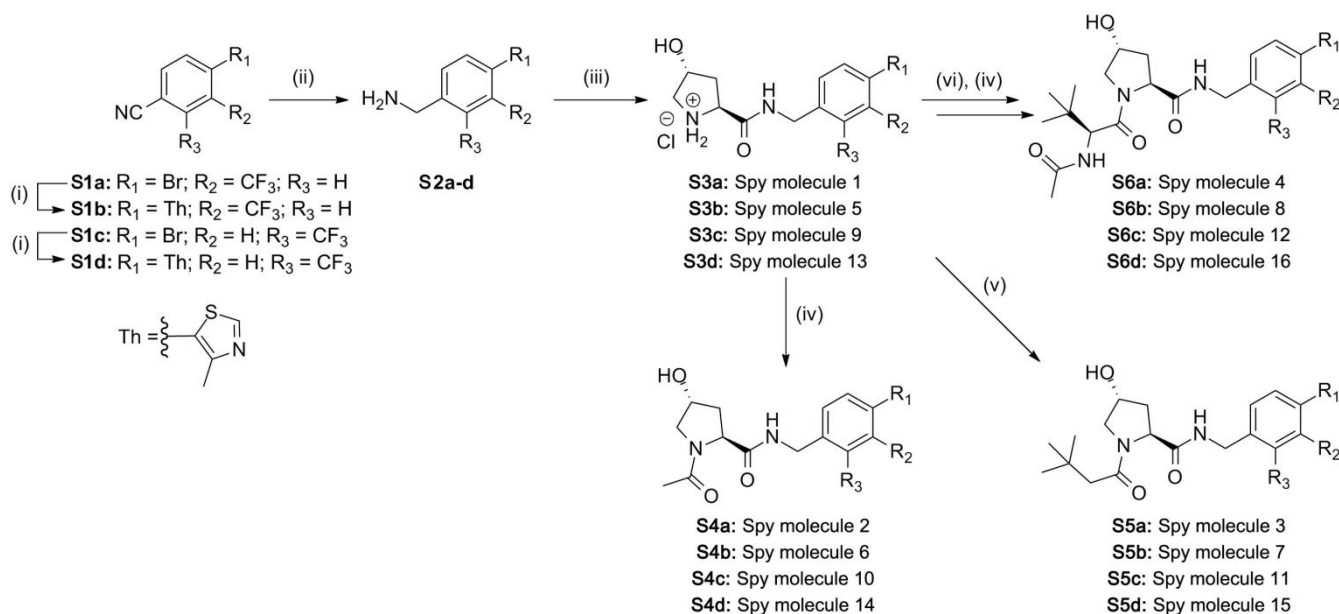
The NMR characterization was performed either on a Bruker 500 Ultrashield or Bruker Ascend 400 spectrometers. The splitting of the NMR signals are described as follows: singlet (s), doublet (d), triplet (t), quartet (q), multiplet (m) and combinations in case of multiple signal splitting. Chemical shifts are described as parts per million (ppm) and coupling constants (*J*) were calculated in hertz (Hz). The proton (^1H) and carbon (^{13}C) spectra were referenced as follows: d6-Chloroform – CDCl_3 ($\delta_{\text{H}} = 7.26$ ppm / $\delta_{\text{C}} = 77.1$ ppm) and d5-Methanol – CD_3OD ($\delta_{\text{H}} = 3.34$ ppm / $\delta_{\text{C}} = 49.1$ ppm). For compounds where amide rotamers could be observed, just the signal of the major rotamer was listed.

Reactions were monitored using an Agilent Technologies 1200 series analytical HPLC connected to an Agilent Technologies 6130 quadrupole LC/MS containing an Agilent diode array detector and a Waters XBridge column (50 mm \times 2.1 mm, 3.5 μ m particle size) for separation of the compounds. Samples were eluted with a 3 minutes gradient of 5% to 95% acetonitrile : 0.1% formic acid.

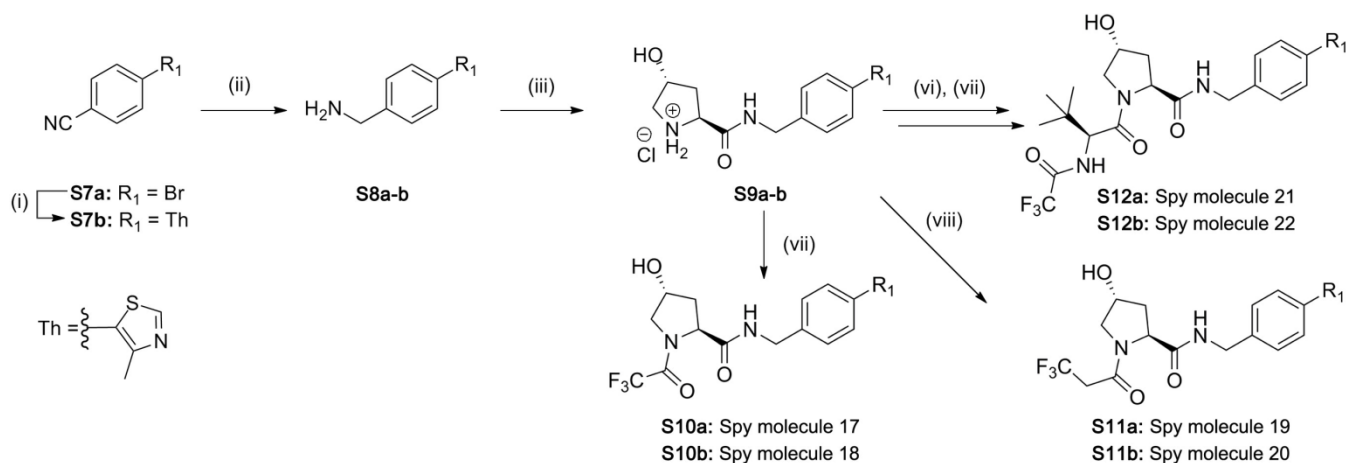
Abbreviations: ACN (acetonitrile), DCM (dichloromethane), DIPEA (N,N-diisopropylethylamine), DMA (dimethylacetamide), DMF (N,N-dimethylformamide), EtOAc (ethyl acetate), Et₂O (diethyl ether), HATU (1-[bis(dimethylamino)methylene]-1H-1,2,3-triazolo[4,5-b]pyridinium 3-oxide hexafluorophosphate), MgSO_4 (Magnesium sulphate), MeOH (methanol), THF (Tetrahydrofuran) and TEA (triethylamine).

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For describing the synthesis and characterisation of spy molecules and intermediates, compounds were numbered as shown in the two schemes below. The respective compound numbering in the main text is indicated where applicable (*e.g.* compound **S3a** corresponds to **spy molecule 1** in the main text, and so on).



Scheme 2.1 Synthesis of spy molecules with a trifluoromethyl modification on an aromatic position.

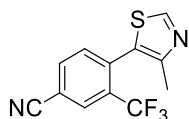


Scheme 2.2 Synthesis of spy molecules with a trifluoromethyl modification on an aliphatic position.

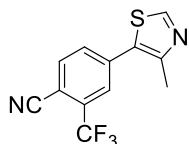
Compounds **S1a**, **S1c** and **S8a** were purchased from commercial sources and used without further purification. The spectroscopic characterization and yields for intermediates **S7b**, **S8b** and **S9b** can be found elsewhere, as these were previously prepared by our group,^[2] while all the remaining compounds were synthesized and characterized as described below. For the NMR characterization of compounds **S3a-d** and **S9a-b**, either a chloride or a formate salt was obtained depending on the purification strategy used.

General procedure i. Coupling of aryl bromides with 4-methylthiazole – Synthesis of S1b, S1d and S7b

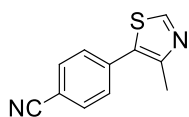
The aryl bromide (1 equiv.) was dissolved in dimethylacetamide (3 mL per mmol of bromide), followed by the sequential addition of 4-methylthiazole (2 equiv.), potassium acetate (2 equiv.) and palladium (II) acetate (0.02 equiv.). The reaction was stirred for approximately two hours at 150 °C under nitrogen atmosphere. The mixture was extracted with brine and dichloromethane. Combined organic phases were concentrated and DMA was removed in the vacuum pump. The desired product was then purified by flash column chromatography with an increasing elution of ethyl acetate (0-100%) in heptane.

4-(4-methylthiazol-5-yl)-3-(trifluoromethyl)benzonitrile (S1b):

Prepared from 0.3163 g (1.3 mmol) of the respective bromide (**S1a**), resulting in 0.221 g (0.8 mmol, 65%) of the desired product as a white solid. ¹H NMR (CDCl₃, 400 MHz) δ: 8.84 (s, 1H); 8.08 (d, *J*_{H-H} = 1.4 Hz, 1H); 7.89 (dd, *J*_{H-H} = 7.9, 1.4 Hz, 1H); 7.54 (d, *J*_{H-H} = 7.9 Hz, 1H); 2.25 (s, 3H); ¹³C NMR (CDCl₃, 100 MHz) δ: 152.75, 152.15, 136.04 (q, ³*J*_{C-F} = 1.7 Hz), 135.06, 134.93, 132.37 (q, ²*J*_{C-F} = 31.3 Hz), 130.36 (q, ³*J*_{C-F} = 5.4 Hz), 125.06, 122.56 (q, ¹*J*_{C-F} = 274.6 Hz), 117.16, 113.76, 15.72; ¹⁹F NMR (CDCl₃, 470 MHz) δ: -60.10.

4-(4-methylthiazol-5-yl)-2-(trifluoromethyl)benzonitrile (S1d):

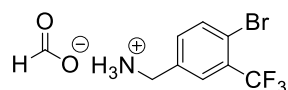
Prepared from 0.8932 g (3.6 mmol) of the respective bromide (**S1c**), resulting in 0.555 g (2.2 mmol, 61%) of the desired product as a bright yellow solid. ¹H NMR (CDCl₃, 400 MHz): 8.81 (s, 1H); 7.91 (d, *J*_{H-H} = 8.08 Hz, 1H); 7.86 (d, *J*_{H-H} = 1.5 Hz, 1H); 7.76 (dd, *J*_{H-H} = 1.5, 8.08 Hz, 1H); 2.59 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz): 152.49, 151.25, 137.69, 135.29, 133.61 (q, ²*J*_{C-F} = 32.8 Hz), 132.50, 128.96, 127.27 (q, ³*J*_{C-F} = 4.5 Hz), 122.28 (q, ¹*J*_{C-F} = 273.8 Hz), 115.32, 109.14, 16.58; ¹⁹F NMR (CDCl₃, 470 MHz) δ: -62.07.

4-(4-methylthiazol-5-yl)benzonitrile (S7b):

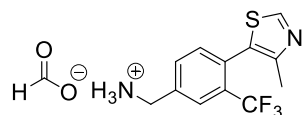
Previously prepared and characterized.^[2]

General procedure ii. Reduction of nitriles to amines – Synthesis of S2a-d and S8b

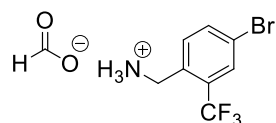
To a stirring solution of the nitrile (1 equiv.) in THF under nitrogen atmosphere, was added a solution of LiAlH₄ (1 equiv. from a 1M solution in THF). After an overnight period, the mixture was cooled down in an ice bath and diluted with diethyl ether. Water was then slowly added (1 μ L per mg of LiAlH₄ added), followed by the addition of 15% NaOH (1 μ L per mg of LiAlH₄ added) and once more water (3 μ L per mg of LiAlH₄ added). The ice bath was removed and the mixture. MgSO₄ was added and after 15 additional minutes stirring the mixture was filtered, then extracted with HCl solution (pH \approx 2). The organic phase was discarded and the pH of the aqueous phase was raised to 12 by adding NaOH 1 M. This solution was extracted three times with DCM and the organic phase was concentrated and purified (purification strategy specified for each compound).

(4-bromo-3-(trifluoromethyl)phenyl)methanaminium formate (S2a):

Prepared from 0.264 g (1.06 mmol) of the respective nitrile (**S1a**), resulting in 0.110 g (0.35 mmol, 35%) of the desired product as a white solid. Compound obtained as a formate salt after purification in reverse phase HPLC using a 5 to 95% gradient of formic acid 0.1% and acetonitrile. ¹H NMR (CD₃OD, 400 MHz) δ : 8.55 (Formic acid), 7.95-7.89 (m, 2H), 7.64-7.60 (m, 1H), 4.19-4.16 (s, 2H); ¹³C NMR (CD₃OD, 100 MHz) δ : 170.08 (Formic acid), 137.05, 136.10, 135.20, 131.73 (q, ²J_{C-F} = 31.2 Hz), 129.66 (q, ³J_{C-F} = 5.2 Hz), 124.34 (q, ¹J_{C-F} = 273.3 Hz), 121.31, 43.57; ¹⁹F NMR (CD₃OD, 470 MHz) δ : -64.05 (CF₃).

(4-(4-methylthiazol-5-yl)-3-(trifluoromethyl)phenyl)methanaminium formate (S2b):

Prepared from 0.221 g (0.82 mmol) of the respective nitrile (**S1b**), resulting in 0.092 g (0.29 mmol, 36%) of the desired product as a yellow solid. Compound obtained as a formate salt after purification in reverse phase HPLC using a 5 to 95% gradient of formic acid 0.1% and acetonitrile. ¹H NMR (CD₃OD, 500 MHz) δ : 9.04 (s, 1H), 8.55 (Formic acid), 8.02-7.99 (s, 1H), 7.82-7.79 (d, *J*_{H-H} = 7.9 Hz, 1H), 7.58-7.55 (d, *J*_{H-H} = 7.9 Hz, 1H), 4.26 (s, 2H), 2.22 (s, 3H); ¹⁹F NMR (CD₃OD, 470 MHz) δ : -62.36 (CF₃).

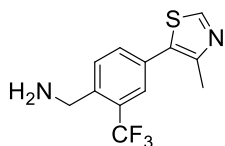
(4-bromo-2-(trifluoromethyl)phenyl)methanaminium formate (S2c):

Prepared from 0.523 g (2.09 mmol) of the respective nitrile (**S1c**), resulting in 0.195 g (0.65 mmol, 31%) of the desired product as a white solid. Compound obtained as a formate salt after purification in reverse phase HPLC

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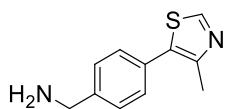
using a 5 to 95% gradient of formic acid 0.1% and acetonitrile. ^1H NMR (CD_3OD , 500 MHz) δ : 8.52 (Formic acid), 7.98-7.97 (d, $J_{\text{H-H}} = 1.6$ Hz, 1H), 7.95-7.92 (dd, $J_{\text{H-H}} = 1.6, 8.3$ Hz, 1H), 7.65-7.62 (d, $J_{\text{H-H}} = 8.3$ Hz, 1H), 4.24 (s, 2H); ^{13}C NMR (CD_3OD , 125 MHz) δ : 169.65 (Formic acid), 137.32, 133.94, 133.75, 131.55 (q, $^2J_{\text{C-F}} = 31.2$), 130.71 (q, $^3J_{\text{C-F}} = 5.8$), 124.76 (q, $^1J_{\text{C-F}} = 273.62$), 124.04, 40.71 (q, $^4J_{\text{C-F}} = 2.68$); ^{19}F NMR (CD_3OD , 470 MHz) δ : -61.08 (CF_3).

(4-(4-methylthiazol-5-yl)-2-(trifluoromethyl)phenyl)methanamine (S2d):



Prepared from 0.398 g (1.49 mmol) of the respective nitrile (**S1d**), resulting in 0.157 g (0.49 mmol, 33%) of the desired product as a yellow solid. Compound obtained as a free amine after purification in reverse phase HPLC using a 5 to 95% gradient of ammonia 0.1% and acetonitrile. ^1H NMR (CD_3OD , 500 MHz) δ : 8.96 (s, 1H), 7.84-7.76 (m, 3H), 4.05 (s, 2H), 2.53 (s, 3H); ^{13}C NMR (CD_3OD , 100 MHz) δ : 153.72, 150.26, 142.43, 134.38, 132.24, 131.86, 131.77, 129.35 (q, $^2J_{\text{C-F}} = 30.2$ Hz), 127.50 (q, $^3J_{\text{C-F}} = 5.8$ Hz), 125.81 (q, $^1J_{\text{C-F}} = 273.7$ Hz), 42.83, 15.87; ^{19}F NMR (CD_3OD , 470 MHz) δ : -59.67 (CF_3).

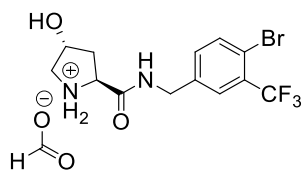
(4-(4-methylthiazol-5-yl)phenyl)methanamine (S8b):



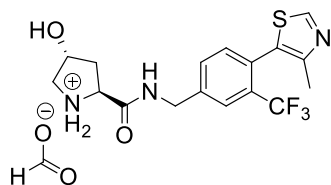
Previously prepared and characterized.^[2]

General procedure iii. Amide coupling with Boc-L-hydroxyproline and deprotection – Synthesis of S3a-d and S9a-b

To a solution of amine (1 equiv.) in DMF, Boc-L-hydroxyproline (1 equiv.) was added and the mixture was stirred at room temperature. DIPEA (2 equiv.) was added dropwise and the mixture was stirred for 5 minutes at room temperature. HATU (1.1 equiv.) was added and the mixture was stirred at room temperature for 30-90 minutes (LCMS monitoring). Water was added and the mixture was extracted with ethyl acetate. The combined organic phases were washed with brine, dried over MgSO_4 and evaporated under reduced pressure to give the corresponding crude, which was purified by flash column chromatography with an increasing gradient of DCM and 20% MeOH in DCM to yield the desired product. The Boc protected compound was dissolved in DCM, followed by the dropwise addition of a 4M HCl solution in dioxane (at least 3 equiv.). Often an insoluble precipitate starts to be formed. A few drops of MeOH were added to make the solution homogeneous, being kept stirring for approximately one hour. The DCM and the HCl were removed by flushing nitrogen into the solution and residual solvents were evaporated under reduced pressure. To remove traces of impurities, compounds S3a, S3b, S3d and S9a were furtherly purified by preparative HPLC, being obtained as a formate salt, while the remaining compounds were obtained as a chloride salt.

(2S,4R)-2-((4-bromo-3-(trifluoromethyl)benzyl)carbamoyl)-4-hydroxypyrrolidin-1-ium formate (S3a - spy molecule 1):

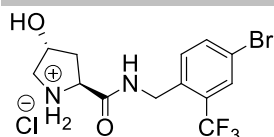
Prepared from 198 mg (0.78 mmol) of amine **S2a**, resulting in 248 mg (0.60 mmol, 78%) of the desired product as a white solid. Compound obtained as a formate salt after purification in reverse phase HPLC using a 5 to 95% gradient of formic acid 0.1% and acetonitrile. HRMS (ESI) $[M+H]^+$ (m/z): Calculated for $C_{13}H_{14}BrF_3N_2O_2$: 367.0264; Observed: 367.0299; 1H NMR (CD_3OD , 400 MHz) δ : 8.50 (Formic acid), 7.80 (d, $J_{H-H} = 8.2$ Hz, 1H), 7.74 (d, $J_{H-H} = 1.7$ Hz, 1H), 7.49 (dd, $J_{H-H} = 1.7, 8.2$ Hz, 1H), 4.58 (m, 1H), 4.49 (s, 2H), 4.42 (dd, $J_{H-H} = 7.6, 10.3$ Hz, 1H), 3.38 (dd, $J_{H-H} = 3.7, 12.1$ Hz, 1H), 3.26 (d, $J_{H-H} = 12.1$ Hz, 1H), 2.44 (ddd, $J_{H-H} = 1.5, 7.6, 13.4$ Hz, 1H), 2.04 (ddd, $J_{H-H} = 4.2, 10.3, 13.4$ Hz, 1H); ^{13}C NMR (CD_3OD , 100 MHz) δ : 171.98, 170.29 (Formic acid), 140.99, 137.34, 134.74, 131.96 (q, $^2J_{C-F} = 31.3$ Hz), 129.06 (q, $^3J_{C-F} = 5.6$ Hz), 125.25 (q, $^1J_{C-F} = 273.3$ Hz), 120.22, 72.47, 60.96, 56.09, 44.19, 40.96; ^{19}F NMR (CD_3OD , 470 MHz) δ : -64.01.

(2S,4R)-2-((4-(4-methylthiazol-5-yl)-3-(trifluoromethyl)benzyl)carbamoyl)-4-hydroxypyrrolidin-1-ium formate (S3b - spy molecule 5):

Prepared from 179 mg (0.56 mmol) of amine **S2b**, resulting in 203 mg (0.47 mmol, 84%) of the desired product as a white solid. Compound obtained as a formate salt after purification in reverse phase HPLC using a 5 to 95% gradient of formic acid 0.1% and acetonitrile. MS $[M+H]^+$ (m/z): 386.1; 1H NMR (CD_3OD , 400 MHz) δ : 9.02 (s, 1H), 8.51 (Formic acid), 7.81 (s, 1H), 7.66 (d, $J_{H-H} = 7.9$ Hz, 1H), 7.46 (d, $J_{H-H} = 7.9$ Hz, 1H), 4.58 (s, 2H), 4.55 (m, 1H), 4.35 (dd, $J_{H-H} = 7.7, 9.9$ Hz, 1H), 3.30 (m, 1H), 3.20 (dt, $J_{H-H} = 1.4, 12.0$ Hz, 1H), 2.41 (ddt, $J_{H-H} = 1.5, 7.6, 13.3$ Hz, 1H), 2.20 (s, 3H), 2.03 (ddd, $J_{H-H} = 4.3, 9.9, 13.4$ Hz, 1H); ^{13}C NMR (CD_3OD , 100 MHz) δ : 172.51, 169.65 (Formic acid), 154.51, 152.18, 142.12, 135.37, 132.41, 131.77 (q, $^2J_{C-F} = 30.1$ Hz), 130.17, 128.59, 126.72 (q, $^3J_{C-F} = 5.6$ Hz), 125.19 (q, $^1J_{C-F} = 273.7$ Hz), 72.11, 60.40, 55.55, 43.62, 40.41, 15.33; ^{19}F NMR (CD_3OD , 470 MHz) δ : -59.37.

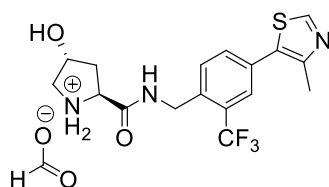
(2S,4R)-2-((4-bromo-2-(trifluoromethyl)benzyl)carbamoyl)-4-hydroxypyrrolidin-1-ium chloride (S3c - spy molecule 9):

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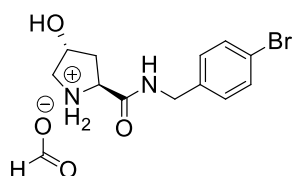
Prepared from 11 mg (0.043 mmol) of amine **S2c**, resulting in 14 mg (0.035 mmol, 80%) of the desired product as a white solid. HRMS (ESI) $[M+H]^+$ (m/z): Calculated for $C_{13}H_{14}BrF_3N_2O_2$: 367.0264; Observed: 367.0280; 1H NMR (CD_3OD , 500 MHz) δ : 7.88-7.85 (d, $J_{H-H} = 1.8$ Hz, 1H), 7.82-7.79 (dd, $J_{H-H} = 1.8, 8.4$ Hz, 1H), 7.47-7.44 (d, $J_{H-H} = 8.4$ Hz, 1H), 4.61-4.53 (m, 2H), 4.41-4.38 (m, 1H), 4.00-3.76 (t, $J_{H-H} = 8.3$ Hz, 1H), 3.06-3.01 (dd, $J_{H-H} = 4.0, 12.0$ Hz, 1H), 2.96-2.91 (dt, $J_{H-H} = 1.8, 12.0$ Hz, 1H), 2.23-2.17 (ddt, $J_{H-H} = 1.8, 8.0, 13.4$ Hz, 1H), 1.91-1.85 (ddd, $J_{H-H} = 5.0, 8.7, 13.4$ Hz, 1H); ^{13}C NMR (CD_3OD , 125 MHz) δ : 177.58, 137.72, 136.72, 132.33, 130.63 (q, $^2J_{C-F} = 31.5$ Hz), 130.09 (q, $^3J_{C-F} = 5.8$ Hz), 125.02 (q, $^1J_{C-F} = 273.6$ Hz), 122.07, 73.67, 60.82, 56.15, 40.99, 40.19 (q, $^4J_{C-F} = 2.9$ Hz); ^{19}F NMR (CD_3OD , 470 MHz) δ : -61.86 (CF_3).

(2S,4R)-2-((4-(4-methylthiazol-5-yl)-2-(trifluoromethyl)benzyl)carbamoyl)-4-hydroxypyrrolidin-1-ium chloride (**S3d** - spy molecule 13):



Prepared from 164 mg (0.60 mmol) of amine **S2d**, resulting in 239 mg (0.55 mmol, 92%) of the desired product as a white solid. Compound obtained as a formate salt after purification in reverse phase HPLC using a 5 to 95% gradient of formic acid 0.1% and acetonitrile. HRMS (ESI) $[M+H]^+$ (m/z): Calculated for $C_{17}H_{18}F_3N_3O_2S$: 386.1145; Observed: 386.1168; 1H NMR (CD_3OD , 500 MHz) δ : 8.98 (s, 1H), 8.55 (Formic acid), 7.80 (s, 1H), 7.77 (dd, $J_{H-H} = 1.4, 8.1$ Hz, 1H), 7.66 (d, $J_{H-H} = 8.1$ Hz, 1H), 4.69 (m, 2H), 4.48 (m, 1H), 4.18 (t, $J_{H-H} = 8.7$ Hz, 1H), 3.18 (dd, $J_{H-H} = 3.5, 11.7$ Hz, 1H), 3.07 (d, $J_{H-H} = 12.1$ Hz, 1H), 2.53 (s, 3H), 2.32 (ddt, $J_{H-H} = 1.6, 7.8, 13.5$ Hz, 1H), 1.98 (ddd, $J_{H-H} = 4.7, 9.3, 13.5$ Hz, 1H); ^{13}C NMR (CD_3OD , 125 MHz) δ : 174.57, 170.18 (Formic acid), 153.88, 150.39, 138.01, 134.30, 132.82, 131.65, 131.37, 129.60 (q, $^2J_{C-F} = 30.9$ Hz), 127.78 (q, $^3J_{C-F} = 5.9$ Hz), 125.64 (q, $^1J_{C-F} = 273.4$ Hz), 72.77, 60.56, 55.82, 40.64, 15.87; ^{19}F NMR (CD_3OD , 470 MHz) δ : -60.11.

(2S,4R)-2-((4-bromobenzyl)carbamoyl)-4-hydroxypyrrolidin-1-ium chloride (**S9a**):

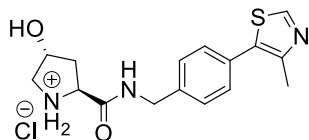


Prepared from 182 mg (1.00 mmol) of amine **S8a**, resulting in 294 mg (0.47 mmol, 84%) of the desired product as a white solid. Compound obtained as a formate salt after purification in reverse phase HPLC using a 5 to 95% gradient of formic acid 0.1% and acetonitrile. 1H NMR (CD_3OD , 500 MHz) δ : 7.51-7.47 (m, 2H), 7.26-7.23 (m, 2H), 4.52-4.49 (m, 1H), 4.41-4.38 (s, 2H), 4.27-4.23 (dd, $J_{H-H} = 7.9, 9.4$ Hz, 1H), 3.27-3.23 (dd, $J_{H-H} = 3.8, 12.1$

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Hz, 1H), 3.16-3.13 (m, 1H), 2.37-2.32 (m, 1H), 2.01-1.95 (ddd, $J_{\text{H-H}} = 4.5, 9.6, 13.5$ Hz, 1H); ^{13}C NMR (CD_3OD , 100 MHz) δ : 173.09, 170.38 (Formic acid), 139.11, 132.76, 130.63, 122.13, 72.35, 60.39, 55.57, 43.57, 40.50.

(2*S*,4*R*)-4-hydroxy-2-((4-(4-methylthiazol-5-yl)benzyl)carbamoyl)pyrrolidin-1-ium chloride (S9b):

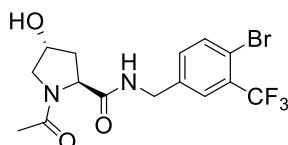


Previously prepared and characterized.^[2]

General procedure iv. Amine acetylation – Synthesis of S4a-d and S6a-d

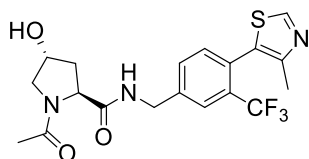
The amine (1 equiv.) was dissolved in DMF, followed by the addition of triethylamine (2.0 equiv.) and 1-acetylimidazole (1.0 equiv.). The mixture was kept stirring overnight, followed by extraction with ethyl acetate and brine. Combined organic phases were dried with MgSO_4 , concentrated and purified in the acidic Gilson HPLC system, yielding the desired product.

(2*S*,4*R*)-1-acetyl-N-(4-bromo-3-(trifluoromethyl)benzyl)-4-hydroxypyrrolidine-2-carboxamide (S4a - spy molecule 2):



Prepared from 50.3 mg (0.125 mmol) of hydrochloride salt of amine **S3a**, resulting in 47.7 mg (0.117 mmol, 93%) of the desired product as a white solid. HRMS (ESI) $[\text{M}+\text{H}]^+$ (m/z): Calculated for $\text{C}_{15}\text{H}_{17}\text{BrF}_3\text{N}_2\text{O}_3$: 409.0369; Observed: 409.0387; ^1H NMR (CD_3OD , 400 MHz) δ : 7.79 (d, $J_{\text{H-H}} = 8.3$ Hz, 1H), 7.73 (d, $J_{\text{H-H}} = 1.6$ Hz, 1H), 7.49 (dd, $J_{\text{H-H}} = 1.6, 8.3$ Hz, 1H), 4.48 (m, 4H), 3.80 (dd, $J_{\text{H-H}} = 4.2, 11.0$ Hz, 1H), 3.58 (dd, $J_{\text{H-H}} = 1.8, 11.0$ Hz, 1H), 2.27 (dddd, $J_{\text{H-H}} = 1.6, 3.0, 8.0, 13.1$ Hz, 1H), 2.12 (s, 3H), 2.07 (ddd, $J_{\text{H-H}} = 4.6, 8.3, 13.1$ Hz, 1H); ^{13}C NMR (CD_3OD , 100 MHz) δ : 175.88, 173.49, 141.40, 137.19, 134.38, 131.77 (q, $^2J_{\text{C-F}} = 31.3$ Hz), 128.72 (q, $^3J_{\text{C-F}} = 5.5$ Hz), 125.32 (q, $^1J_{\text{C-F}} = 273.1$ Hz), 119.77, 71.70, 61.21, 58.21, 43.87, 40.35, 23.18; ^{19}F NMR (CD_3OD , 470 MHz) δ : -63.86 (CF_3).

(2*S*,4*R*)-1-acetyl-4-hydroxy-N-(4-(4-methylthiazol-5-yl)-3-(trifluoromethyl)benzyl)pyrrolidine-2-carboxamide (S4b - spy molecule 6):

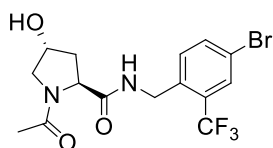


Prepared from 26 mg (0.060 mmol) of formate salt of amine **S3b**, resulting in 20 mg (0.047 mmol, 78%) of the desired product as a white solid. HRMS (ESI) $[\text{M}+\text{H}]^+$ (m/z): Calculated for $\text{C}_{19}\text{H}_{20}\text{F}_3\text{N}_3\text{O}_3\text{S}$: 428.1250;

SUPPORTING INFORMATION

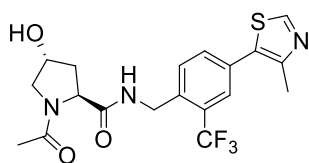
Observed: 428.1272; ^1H NMR (CD_3OD , 500 MHz) δ : 9.04-9.02 (s, 1H), 7.83-7.81 (s, 1H), 7.74-7.66 (d, $J_{\text{H-H}} = 7.9$ Hz, 1H), 7.49-7.44 (d, $J_{\text{H-H}} = 7.9$ Hz, 1H), 4.66-4.44 (m, 4H), 3.83-3.79 (dd, $J_{\text{H-H}} = 4.3, 11.1$ Hz, 1H), 3.61-3.57 (dd, $J_{\text{H-H}} = 1.7, 11.1$ Hz, 1H), 2.48-2.27 (m, 1H), 2.22-2.20 (s, 3H), 2.20-2.07 (m, 1H), 2.14-1.96 (s, 3H); ^{13}C NMR (CD_3OD , 125 MHz) δ : 175.20, 172.75, 154.45, 152.11, 142.44, 135.25, 132.04, 131.57 (q, $^2J_{\text{C-F}} = 29.8$ Hz), 129.70 (q, $^3J_{\text{C-F}} = 1.9$ Hz), 128.81, 126.42 (q, $^3J_{\text{C-F}} = 5.3$ Hz), 125.25 (q, $^1J_{\text{C-F}} = 273.1$ Hz), 70.96, 60.48, 57.48, 43.39, 39.63, 22.44, 15.34; ^{19}F NMR (CD_3OD , 470 MHz) δ : -60.75 (CF_3).

(2S,4R)-1-acetyl-N-(4-bromo-2-(trifluoromethyl)benzyl)-4-hydroxypyrrolidine-2-carboxamide (S4c - spy molecule 10):



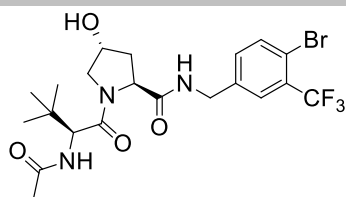
Prepared from 29 mg (0.072 mmol) of amine **S3c**, resulting in 22 mg (0.054 mmol, 75%) of the desired product as a white solid. HRMS (ESI) $[\text{M}+\text{H}]^+$ (m/z): Calculated for $\text{C}_{15}\text{H}_{16}\text{BrF}_3\text{N}_2\text{O}_3$: 409.0369; Observed: 409.0377; ^1H NMR (CD_3OD , 500 MHz) δ : 7.89-7.80 (m, 2H), 7.66-7.47 (d, $J_{\text{H-H}} = 8.4$ Hz, 1H), 4.66-4.43 (m, 4H), 3.83-3.79 (dd, $J_{\text{H-H}} = 4.1, 11.0$ Hz, 1H), 3.61-3.57 (dd, $J_{\text{H-H}} = 1.7, 11.1$ Hz, 1H), 2.45-2.26 (dddd, $J_{\text{H-H}} = 1.7, 2.8, 7.8, 13.2$ Hz, 1H), 2.21-2.07 (ddd, $J_{\text{H-H}} = 4.6, 8.5, 13.2$ Hz, 1H), 2.15-1.97 (s, 3H); ^{13}C NMR (CD_3OD , 125 MHz) δ : 175.33, 172.81, 137.72, 136.75, 132.12, 130.26 (q, $^2J_{\text{C-F}} = 31.3$ Hz), 129.81 (q, $^3J_{\text{C-F}} = 6.1$ Hz), 125.05 (q, $^1J_{\text{C-F}} = 273.5$ Hz), 121.81, 70.98, 60.49, 57.52, 40.29 (q, $^4J_{\text{C-F}} = 3.4$ Hz), 39.57, 22.44; ^{19}F NMR (CD_3OD , 470 MHz) δ : -62.14 (CF_3).

(2S,4R)-1-acetyl-4-hydroxy-N-(4-(4-methylthiazol-5-yl)-2-(trifluoromethyl)benzyl)pyrrolidine-2-carboxamide (S4d - spy molecule 14):



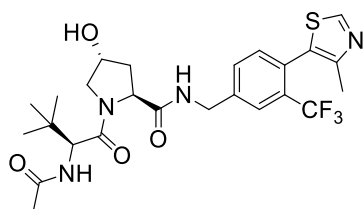
Prepared from 39 mg (0.090 mmol) of amine **S3d**, resulting in 28 mg (0.066 mmol, 72%) of the desired product as a white solid. HRMS (ESI) $[\text{M}+\text{H}]^+$ (m/z): Calculated for $\text{C}_{19}\text{H}_{20}\text{F}_3\text{N}_3\text{O}_3\text{S}$: 428.1250; Observed: 428.1265; ^1H NMR (CD_3OD , 500 MHz) δ : 8.98 (s, 1H), 7.86-7.76 (m, 3H), 4.75-4.51 (m, 4H), 3.84-3.80 (dd, $J_{\text{H-H}} = 4.2, 11.1$ Hz, 1H), 3.63-3.56 (dt, $J_{\text{H-H}} = 1.7, 11.1$ Hz, 1H), 2.53 (s, 3H), 2.47-2.27 (dddd, $J_{\text{H-H}} = 1.6, 2.9, 7.9, 13.2$ Hz, 1H), 2.24-2.10 (m, 1H), 2.15-2.00 (s, 3H); ^{13}C NMR (CD_3OD , 125 MHz) δ : 175.35, 172.81, 153.75, 150.23, 138.38, 134.32, 132.30, 131.83, 130.80, 129.08 (q, $^2J_{\text{C-F}} = 30.9$ Hz), 127.43 (q, $^3J_{\text{C-F}} = 5.9$ Hz), 125.70 (q, $^1J_{\text{C-F}} = 273.6$ Hz), 70.98, 60.52, 57.53, 40.50 (q, $^4J_{\text{C-F}} = 3.2$ Hz), 39.61, 22.47, 15.90; ^{19}F NMR (CD_3OD , 470 MHz) δ : -60.45 (CF_3).

(2S,4R)-1-((S)-2-acetamido-3,3-dimethylbutanoyl)-N-(4-bromo-3-(trifluoromethyl)benzyl)-4-hydroxypyrrolidine-2-carboxamide (S6a - spy molecule 4):



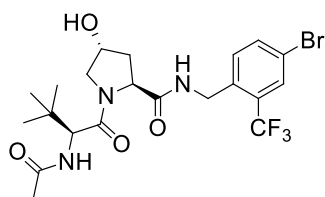
Overall yield of 93% (90 mg, 0.17 mmol), starting from amine **S3a** (77 mg, 0.18 mmol). Yield includes also step (vi). HRMS (ESI) $[M+H]^+$ (m/z): Calculated for $C_{19}H_{20}F_3N_3O_3S$: 428.1250; Observed: 428.1265; 1H NMR (CD_3OD , 500 MHz) δ : 7.82-7.58 (m, 3H), 4.63 (s, 1H), 4.60-4.45 (m, 3H), 4.34 (d, $J_{H-H} = 15.5$ Hz, 1H), 3.92 (d, $J_{H-H} = 11.0$ Hz, 1H), 3.81 (dd, $J_{H-H} = 3.8, 11.0$ Hz, 1H), 2.42-2.19 (m, 1H), 2.16-1.99 (m, 4H), 1.04 (s, 9H); ^{13}C NMR (CD_3OD , 125 MHz) δ : 174.75, 173.22, 172.39, 140.68, 136.32, 133.85, 131.02 (q, $^2J_{C-F} = 30.9$ Hz), 128.15 (q, $^3J_{C-F} = 5.4$ Hz), 124.56 (q, $^1J_{C-F} = 272.5$ Hz), 119.03, 71.17, 60.83, 59.25, 58.06, 43.21, 39.00, 36.53, 27.03, 22.39; ^{19}F NMR (CD_3OD , 470 MHz) δ : -62.30 (CF_3).

(2S,4R)-1-((S)-2-acetamido-3,3-dimethylbutanoyl)-4-hydroxy-N-(4-(4-methylthiazol-5-yl)-3-(trifluoromethyl)benzyl)pyrrolidine-2-carboxamide (S6b - spy molecule 8):



Overall yield of 84% (66 mg, 0.12 mmol), starting from amine **S3b** (63 mg, 0.15 mmol). Yield includes also step (vi). HRMS (ESI) $[M+H]^+$ (m/z): Calculated for $C_{25}H_{31}F_3N_4O_4S$: 541.2091; Observed: 541.2105; 1H NMR (CD_3OD , 500 MHz) δ : 9.03 (s, 1H), 7.87-7.84 (s, 1H), 7.78-7.69 (d, $J_{H-H} = 7.9$ Hz, 1H), 7.48-7.40 (d, $J_{H-H} = 7.9$ Hz, 1H), 4.82-4.43 (m, 5H), 3.96-3.92 (d, $J_{H-H} = 11.0$ Hz, 1H), 3.85-3.81 (dd, $J_{H-H} = 3.9, 11.0$ Hz, 1H), 2.42-2.23 (m, 1H), 2.23-2.19 (s, 3H), 2.18-2.06 (ddd, $J_{H-H} = 4.5, 9.2, 13.1$ Hz, 1H), 2.05-1.99 (s, 3H), 1.08-1.02 (s, 9H); ^{13}C NMR (CD_3OD , 125 MHz) δ : 174.83, 173.22, 172.39, 154.46, 152.09, 142.49, 135.13, 132.28, 131.57 (q, $^2J_{C-F} = 29.7$ Hz), 129.69 (q, $^3J_{C-F} = 1.9$ Hz), 128.81, 126.59 (q, $^3J_{C-F} = 5.3$ Hz), 125.24 (q, $^1J_{C-F} = 273.3$ Hz), 71.19, 60.87, 59.26, 58.08, 43.51, 39.05, 36.54, 27.06, 22.39, 15.35; ^{19}F NMR (CD_3OD , 470 MHz) δ : -60.65 (CF_3).

(2S,4R)-1-((S)-2-acetamido-3,3-dimethylbutanoyl)-N-(4-bromo-3-(trifluoromethyl)benzyl)-4-hydroxypyrrrolidine-2-carboxamide (S6c - spy molecule 12):

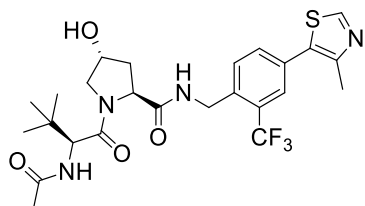


Prepared from 17 mg (0.042 mmol) of amine **S3c**, resulting in 18 mg (0.034 mmol, 82%) of the desired product as a white solid. Yield includes also step (vi). HRMS (ESI) $[M+H]^+$ (m/z): $C_{21}H_{27}BrF_3N_3O_4$: 522.1210; Observed: 522.1180; 1H NMR (CD_3OD , 500 MHz) δ : 7.87-7.83 (d, $J_{H-H} = 1.1$ Hz, 1H), 7.77-7.72 (m, 2H), 4.64 (s, 1H),

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4.71-4.41 (m, 4H), 3.96-3.93 (d, $J_{\text{H-H}} = 11.1$ Hz, 1H), 3.84-3.80 (dd, $J_{\text{H-H}} = 3.9, 11.1$ Hz, 1H), 2.39-2.22 (m, 1H), 2.14-2.07 (ddd, $J_{\text{H-H}} = 4.4, 9.4, 13.1$ Hz, 1H), 2.05-2.00 (s, 3H), 1.09-1.02 (s, 9H); ^{13}C NMR (CD_3OD , 125 MHz) δ : 175.02, 173.24, 172.47, 137.74, 136.54, 132.45, 130.32 (q, $^2J_{\text{C-F}} = 31.3$ Hz), 129.80 (q, $^3J_{\text{C-F}} = 6.1$ Hz), 125.05 (q, $^1J_{\text{C-F}} = 273.7$ Hz), 121.79, 71.23, 60.90, 59.30, 58.11, 40.40 (q, $^4J_{\text{C-F}} = 3.3$ Hz), 38.94, 36.58, 27.04, 22.39; ^{19}F NMR (CD_3OD , 470 MHz) δ : -62.12 (CF_3).

(2*S*,4*R*)-1-((*S*)-2-acetamido-3,3-dimethylbutanoyl)-4-hydroxy-*N*-(4-(4-methylthiazol-5-yl)-3-(trifluoromethyl)benzyl)pyrrolidine-2-carboxamide (S6d - spy molecule 16):

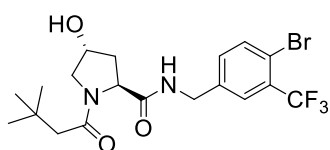


Prepared from 53 mg (0.123 mmol) of amine **S3d**, resulting in 51 mg (0.094 mmol, 77%) of the desired product as a white solid. Yield includes also step (vi). HRMS (ESI) $[\text{M}+\text{H}]^+$ (m/z): $\text{C}_{25}\text{H}_{31}\text{F}_3\text{N}_4\text{O}_4\text{S}$: 541.2091; Observed: 541.2091; ^1H NMR (CD_3OD , 500 MHz) δ : 8.98 (s, 1H), 7.97-7.93 (d, $J_{\text{H-H}} = 8.1$ Hz, 1H), 7.80-7.76 (d, $J_{\text{H-H}} = 1.3$ Hz, 1H), 7.76-7.69 (dd, $J_{\text{H-H}} = 1.3, 8.1$ Hz, 1H), 4.81-4.75 (d, $J_{\text{H-H}} = 16.2$ Hz, 1H), 4.67-4.51 (m, 4H), 3.97-3.93 (d, $J_{\text{H-H}} = 11.0$ Hz, 1H), 3.86-3.81 (dd, $J_{\text{H-H}} = 3.9, 11.0$ Hz, 1H), 2.54-2.51 (s, 3H), 2.31-2.24 (m, 1H), 2.17-2.10 (ddd, $J_{\text{H-H}} = 4.5, 9.2, 13.1$ Hz, 1H), 2.06-2.02 (s, 3H), 1.09-1.03 (s, 9H); ^{13}C NMR (CD_3OD , 125 MHz) δ : 175.06, 173.25, 172.49, 153.78, 150.21, 138.42, 134.11, 132.30, 131.83, 131.14, 129.17 (q, $^2J_{\text{C-F}} = 30.9$ Hz), 127.47 (q, $^3J_{\text{C-F}} = 5.9$ Hz), 125.70 (q, $^1J_{\text{C-F}} = 273.4$ Hz), 71.25, 60.93, 59.33, 58.13, 40.64 (q, $^4J_{\text{C-F}} = 3.3$ Hz), 38.97, 36.61, 27.06, 22.39, 15.89; ^{19}F NMR (CD_3OD , 470 MHz) δ : -61.96.

General procedure v. Amide coupling with 3,3-dimethylbutyric acid – Synthesis of S5a-d

To a solution of amine (1 equiv.) in DMF, 3,3-dimethylbutyric acid (1 equiv.) was added and the mixture was stirred at room temperature. DIPEA (2 equiv.) was added dropwise and the mixture was stirred for 5 minutes at room temperature. HATU (1.1 equiv.) was added and the mixture was stirred at room temperature for 60 minutes (LCMS monitoring). Water was added and the mixture was extracted with ethyl acetate. The combined organic phases were washed with brine, dried over MgSO_4 and evaporated under reduced pressure to give the corresponding crude, which was purified in the acidic Gilson preparative HPLC.

(2*S*,4*R*)-*N*-(4-bromo-3-(trifluoromethyl)benzyl)-1-(3,3-dimethylbutanoyl)-4-hydroxypyrrolidine-2-carboxamide (S5a - spy molecule 3):

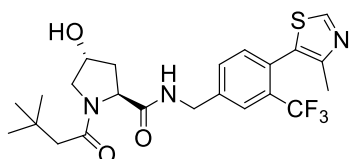


Prepared from 50.6 mg (0.125 mmol) of hydrochloride salt of the amine **S3a** resulting in 49.8 mg (0.107 mmol, 86%) of the desired product as a white solid. HRMS (ESI) $[\text{M}+\text{H}]^+$ (m/z): Calculated for $\text{C}_{19}\text{H}_{24}\text{BrF}_3\text{N}_2\text{O}_3$:

SUPPORTING INFORMATION

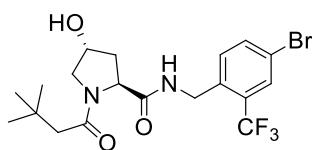
465.0995; Observed: 465.1013; ^1H NMR (CD_3OD , 400 MHz) δ : 7.82-7.75 (d, $J_{\text{H-H}} = 8.2$ Hz, 1H), 7.76-7.73 (m, 1H), 7.56-7.47 (dd, $J_{\text{H-H}} = 1.6, 8.2$ Hz, 1H), 4.58-4.36 (m, 4H), 3.80-3.75 (dd, $J_{\text{H-H}} = 4.1, 11.0$ Hz, 1H), 3.74-3.63 (dt, $J_{\text{H-H}} = 1.7, 11.0$ Hz, 1H), 2.37-1.81 (m, 4H), 1.10-0.97 (s, 9H); ^{13}C NMR (CD_3OD , 100 MHz) δ : 175.29, 173.80, 140.67, 136.36, 133.75, 131.03 (q, $^2J_{\text{C-F}} = 31.8$ Hz), 128.08 (q, $^3J_{\text{C-F}} = 5.5$ Hz), 124.56 (q, $^1J_{\text{C-F}} = 272.7$ Hz), 119.01, 71.10, 60.54, 57.82, 47.76, 43.17, 39.24, 32.74, 30.47; ^{19}F NMR (CD_3OD , 470 MHz) δ : -63.81 (CF_3).

(2*S*,4*R*)-1-(3,3-dimethylbutanoyl)-4-hydroxy-*N*-(4-(4-methylthiazol-5-yl)-3-(trifluoromethyl)benzyl)pyrrolidine-2-carboxamide (S5b - spy molecule 7):



Prepared with 0.023 g (0.053 mmol) of amine **S3b**, resulting in 0.020 mg (0.042 mmol, 80%) of product as a white solid. HRMS (ESI) $[\text{M}+\text{H}]^+$ (m/z): Calculated for $\text{C}_{23}\text{H}_{28}\text{F}_3\text{N}_3\text{O}_3\text{S}$: 484.1876; Observed: 484.1891; ^1H NMR (CD_3OD , 500 MHz) δ : 9.04-9.02 (s, 1H), 7.87-7.84 (m, 1H), 7.75-7.68 (m, 1H), 7.48-7.41 (d, $J_{\text{H-H}} = 7.8$ Hz, 1H), 4.65-4.43 (m, 4H), 3.82-3.78 (dd, $J_{\text{H-H}} = 4.2, 11.1$ Hz, 1H), 3.75-3.65 (dt, $J_{\text{H-H}} = 2.0, 11.1$ Hz, 1H), 2.38-2.24 (m, 3H), 2.22-2.20 (s, 3H), 2.19-2.06 (ddd, $J_{\text{H-H}} = 4.4, 8.4, 13.3$ Hz, 1H), 1.10-0.97 (s, 9H); ^{13}C NMR (CD_3OD , 125 MHz) δ : 175.37, 173.82, 154.44, 152.09, 142.49, 135.18, 132.21, 131.59 (q, $^2J_{\text{C-F}} = 30.5$ Hz), 129.69 (q, $^3J_{\text{C-F}} = 1.9$ Hz), 128.83, 126.55 (q, $^3J_{\text{C-F}} = 5.5$ Hz), 125.25 (q, $^1J_{\text{C-F}} = 273.1$ Hz), 71.13, 60.58, 57.85, 47.77, 43.65, 39.28, 32.77, 30.48, 15.89; ^{19}F NMR (CD_3OD , 470 MHz) δ : -60.68 (CF_3).

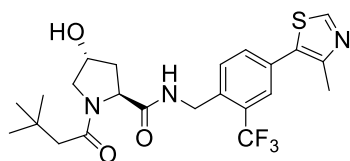
(2*S*,4*R*)-*N*-(4-bromo-2-(trifluoromethyl)benzyl)-1-(3,3-dimethylbutanoyl)-4-hydroxypyrrolidine-2-carboxamide (S5c - spy molecule 11):



Prepared from 17 mg (0.042 mmol) of amine **S3c**, resulting in 15 mg (0.032 mmol, 76%) of the desired product as a white solid. HRMS (ESI) $[\text{M}+\text{H}]^+$ (m/z): Calculated for $\text{C}_{19}\text{H}_{24}\text{BrF}_3\text{N}_2\text{O}_3$: 465.0995; Observed: 465.0997; ^1H NMR (CD_3OD , 500 MHz) δ : 7.90-7.83 (d, $J_{\text{H-H}} = 1.8$ Hz, 1H), 7.83-7.60 (dd, $J_{\text{H-H}} = 1.8, 8.3$ Hz, 1H), 7.72-7.50 (d, $J_{\text{H-H}} = 8.3$ Hz, 1H), 4.66-4.42 (m, 4H), 3.81-3.77 (dd, $J_{\text{H-H}} = 4.1, 11.1$ Hz, 1H), 3.74-3.66 (dd, $J_{\text{H-H}} = 1.9, 11.1$ Hz, 1H), 2.40-2.16 (m, 3H), 2.14-2.07 (ddd, $J_{\text{H-H}} = 4.6, 8.5, 13.2$ Hz, 1H), 1.12-1.02 (s, 9H); ^{13}C NMR (CD_3OD , 125 MHz) δ : 175.51, 173.92, 137.74 (q, $^3J_{\text{C-F}} = 1.2$ Hz), 136.65, 132.32, 130.32 (q, $^2J_{\text{C-F}} = 31.3$ Hz), 129.80 (q, $^3J_{\text{C-F}} = 6.1$ Hz), 125.05 (q, $^1J_{\text{C-F}} = 273.8$ Hz), 121.79, 71.14, 60.58, 57.91, 47.82, 40.36 (q, $^4J_{\text{C-F}} = 3.3$ Hz), 39.21, 32.87, 30.50; ^{19}F NMR (CD_3OD , 470 MHz) δ : -62.11 (CF_3).

SUPPORTING INFORMATION

(2S,4R)-1-(3,3-dimethylbutanoyl)-4-hydroxy-N-(4-(4-methylthiazol-5-yl)-2-(trifluoromethyl)benzyl)pyrrolidine-2-carboxamide (S5d - spy molecule 15):



Prepared from 24 mg (0.056 mmol) of amine **S3d**, resulting in 22 mg (0.045 mmol, 82%) of the desired product as a white solid. HRMS (ESI) $[M+H]^+$ (m/z): Calculated for $C_{23}H_{28}F_3N_3O_3S$: 484.1876; Observed: 484.1894; 1H NMR (CD_3OD , 500 MHz) δ : 9.00-8.98 (s, 1H), 7.91-7.87 (d, $J_{H-H} = 7.9$ Hz, 1H), 4.76-4.51 (m, 4H), 3.83-3.78 (dd, $J_{H-H} = 4.1, 11.1$ Hz, 1H), 3.74-3.67 (dt, $J_{H-H} = 1.5, 11.1$ Hz, 1H), 2.53 (s, 3H), 2.41-2.11 (m, 4H), 1.12-1.02 (s, 9H); ^{13}C NMR (CD_3OD , 125 MHz) δ : 175.55, 173.95, 154.03, 149.74, 138.59, 134.26, 132.13, 132.03, 131.06, 129.16 (q, $^2J_{C-F} = 30.9$ Hz), 127.48 (q, $^3J_{C-F} = 5.8$ Hz), 125.69 (q, $^1J_{C-F} = 273.5$ Hz), 71.16, 60.61, 57.94, 47.84, 40.59 (q, $^4J_{C-F} = 3.1$ Hz), 39.25, 32.89, 30.51, 15.68; ^{19}F NMR (CD_3OD , 470 MHz) δ : -61.95.

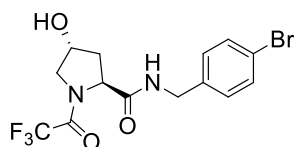
General procedure vi. Amide coupling with Boc-L-tert-leucine and deprotection - Synthesis of intermediates of compounds S6a-d and S12a-b

Same as “general procedure iii”, just replacing the Boc-L-hydroxyproline with Boc-L-tert-leucine. All the crude intermediates prepared at this step were directly used in the next steps without further purification and characterization after deprotection of the Boc group.

General procedure vii. Amine trifluoroacetylation – Synthesis of S10a-b and S12a-b

To a solution of the amine (1 equiv.) in dry MeOH (1 ml per mmol of amine) was added triethylamine (2 equiv.). Ethyltrifluoroacetate (1.25 equiv.) was added and the reaction was stirred at room temperature for approximately 24 hours (LCMS monitoring). The solvent was evaporated under reduced pressure and the crude mixture was extracted with ethyl acetate and 1.0 M HCl solution. Combined organic phases were dried over $MgSO_4$, concentrated and purified in the acidic Gilson preparative HPLC system, yielding the desired product.

(2S,4R)-N-(4-bromobenzyl)-4-hydroxy-1-(2,2,2-trifluoroacetyl)pyrrolidine-2-carboxamide (S10a - spy molecule 17):

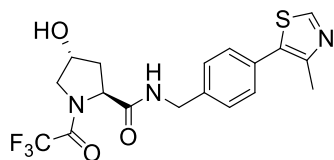


Prepared from 32 mg (0.093 mmol) of amine **S9a**, resulting in 25 mg (0.063 mmol, 68%) of the desired product as a white solid. HRMS (ESI) $[M+H]^+$ (m/z): Calculated for $C_{14}H_{14}BrF_3N_2O_3$: 395.0213; Observed: 395.0230; 1H NMR (CD_3OD , 500 MHz) δ : 7.52-7.48 (m, 2H), 7.29-7.22 (m, 2H), 4.66-4.61 (t, $J_{H-H} = 8.6$ Hz, 1H), 4.57-4.54 (m, 1H), 4.50-4.30 (m, 2H), 3.88-3.84 (dd, $J_{H-H} = 3.5, 11.5$ Hz, 1H), 3.82-3.78 (dd, $J_{H-H} = 1.5, 11.5$ Hz, 1H), 2.48-2.29 (ddt, $J_{H-H} = 1.8, 7.8, 13.2$ Hz, 1H), 2.24-2.04 (ddd, $J_{H-H} = 4.3, 9.3, 13.2$ Hz, 1H); ^{13}C NMR (CD_3OD , 125

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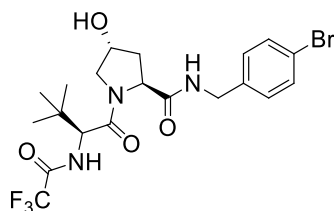
MHz) δ : 173.19, 157.56 (q, $^2J_{\text{C-F}} = 37.11$ Hz), 139.11, 132.69, 130.44, 121.98, 117.73 (q, $^1J_{\text{C-F}} = 286.8$ Hz), 71.08, 61.89, 57.21 (q, $^4J_{\text{C-F}} = 3.0$ Hz), 43.53, 38.57; ^{19}F NMR (CD_3OD , 470 MHz) δ : -72.35 (CF_3).

(2*S*,4*R*)-4-hydroxy-*N*-(4-(4-methylthiazol-5-yl)benzyl)-1-(2,2,2-trifluoroacetyl)pyrrolidine-2-carboxamide (S10b - spy molecule 18):



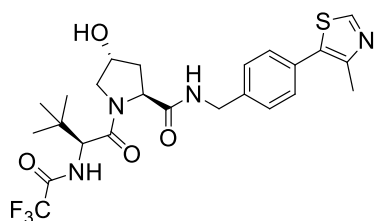
Prepared from 28 mg (0.079 mmol) of amine **S9b**, resulting in 19 mg (0.046 mmol, 58%) of the desired product as a white solid. HRMS (ESI) $[\text{M}+\text{H}]^+$ (m/z): Calculated for $\text{C}_{18}\text{H}_{19}\text{F}_3\text{N}_3\text{O}_3\text{S}$: 414.1094; Observed: 414.1117; ^1H NMR (CD_3OD , 500 MHz) δ : 8.91 (s, 1H), 7.50-7.41 (m, 4H), 4.69-4.64 (d, $J_{\text{H-H}} = 8.5$ Hz, 1H), 4.59-4.40 (m, 3H), 3.89-3.85 (dd, $J_{\text{H-H}} = 3.6, 11.5$ Hz, 1H), 3.83-3.79 (dd, $J_{\text{H-H}} = 1.4, 11.5$ Hz, 1H), 2.54-2.50 (s, 3H), 2.37-2.31 (ddt, $J_{\text{H-H}} = 1.8, 7.8, 13.3$ Hz, 1H), 2.14-2.07 (ddd, $J_{\text{H-H}} = 4.3, 9.3, 13.3$ Hz, 1H); ^{13}C NMR (CD_3OD , MHz) δ : 173.23, 157.58 (q, $^2J_{\text{C-F}} = 36.8$ Hz), 153.00, 149.20, 140.12, 133.48, 131.79, 130.59, 129.03, 117.75 (q, $^1J_{\text{C-F}} = 286.8$ Hz), 71.10, 61.93, 57.23 (q, $^4J_{\text{C-F}} = 3.0$ Hz), 43.82, 38.61, 15.89; ^{19}F NMR (CD_3OD , 470 MHz) δ : -73.74 (CF_3).

(2*S*,4*R*)-*N*-(4-bromobenzyl)-1-((*S*)-3,3-dimethyl-2-(2,2,2-trifluoroacetamido)butanoyl)-4-hydroxypyrrolidine-2-carboxamide (S12a - spy molecule 21):



Prepared from 24 mg (0.070 mmol) of amine **S9a**, resulting in 23 mg (0.045 mmol, 65%) of the desired product as a white solid. HRMS (ESI) $[\text{M}+\text{H}]^+$ (m/z): Calculated for $\text{C}_{20}\text{H}_{25}\text{BrF}_3\text{N}_3\text{O}_4$: 508.1053; Observed: 508.1046; ^1H NMR (CD_3OD , 500 MHz) δ : 8.58 (s, NH), 7.53-7.29 (m, 4H), 4.77-4.75 (s, 1H), 4.61-4.26 (m, 4H), 3.88-3.81 (m, 2H), 2.42-2.22 (m, 1H), 2.12-2.06 (ddd, $J_{\text{H-H}} = 4.4, 9.4, 13.1$ Hz, 1H), 1.10-1.04 (s, 9H); ^{13}C NMR (CD_3OD , 125 MHz) δ : 174.38, 170.74, 158.81 (q, $^2J_{\text{C-F}} = 37.6$ Hz), 139.34, 132.57, 130.46, 121.82, 117.58 (q, $^1J_{\text{C-F}} = 286.3$ Hz), 71.17, 60.95, 59.62, 58.30, 43.50, 39.03, 37.38, 26.93; ^{19}F NMR (CD_3OD , 470 MHz) δ : -76.65 (CF_3).

(2*S*,4*R*)-1-((*S*)-3,3-dimethyl-2-(2,2,2-trifluoroacetamido)butanoyl)-4-hydroxy-*N*-(4-(4-methylthiazol-5-yl)benzyl)pyrrolidine-2-carboxamide (S12b - spy molecule 22):



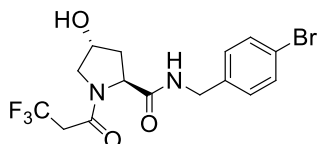
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Prepared from 60 mg (0.17 mmol) of amine **S9b**, resulting in 63 mg (0.12 mmol, 71%) of the desired product as a white solid. HRMS (ESI) $[M+H]^+$ (m/z): Calculated for $C_{24}H_{29}F_3N_4O_4S$: 527.1934; Observed: 527.1901; 1H NMR (CD_3OD , 500 MHz) δ : 8.91-8.90 (s, 1H), 7.51-7.43 (m, 4H), 4.77 (s, 1H), 4.81-4.36 (m, 4H), 3.89-3.62 (m, 2H), 2.52-2.49 (s, 3H), 2.45-2.24 (m, 1H), 2.18-2.08 (ddd, $J_{H-H} = 4.4, 9.3, 13.1$ Hz, 1H), 1.11-1.05 (s, 9H); ^{13}C NMR (CD_3OD , 100 MHz) δ : 174.38, 170.77, 158.80 (q, $^2J_{C-F} = 38.4$ Hz), 152.94, 149.15, 140.34, 133.50, 131.64, 130.47, 129.04, 117.58 (q, $^1J_{C-F} = 285.9$ Hz), 71.18, 60.98, 59.64, 58.31, 43.81, 39.05, 37.40, 26.96, 15.89; ^{19}F NMR (CD_3OD , 470 MHz) δ : -75.18.

General procedure viii. Amide coupling with 3,3,3-trifluoropropanoic acid – Synthesis of S11a-b

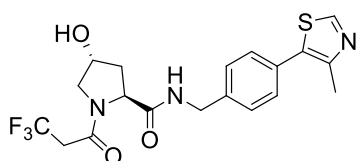
To a solution of amine (1 equiv.) in DMF, 3,3,3-Trifluoropropanoic acid (1 equiv.) was added and the mixture was stirred at room temperature. DIPEA (2 equiv.) was added dropwise and the mixture was stirred for 5 minutes at room temperature. HATU (1.1 equiv.) was added and the mixture was stirred at room temperature for 30-90 minutes (TLC monitoring). Water was added and the mixture was extracted with ethyl acetate. The combined organic phases were washed with brine, dried over $MgSO_4$ and evaporated under reduced pressure to give the corresponding crude, which was purified by flash column chromatography with an increasing gradient of DCM and 20% MeOH in DCM to yield the desired product.

(2S,4R)-N-(4-bromobenzyl)-4-hydroxy-1-(3,3,3-trifluoropropanoyl)pyrrolidine-2-carboxamide (S11a - spy molecule 19):



Prepared from 37 mg (0.107 mmol) of amine **S9a**, resulting in 33 mg (0.081 mmol, 75%) of the desired product as a white solid. HRMS (ESI) $[M+H]^+$ (m/z): Calculated for $C_{15}H_{17}BrF_3N_2O_3$: 409.0369; Observed: 409.0389; 1H NMR (CD_3OD , 500 MHz) δ : 7.52-7.47 (m, 2H), 7.29-7.25 (m, 2H), 4.59-4.33 (m, 4H), 3.81-3.77 (dd, $J_{H-H} = 4.3, 11.0$ Hz, 1H), 3.61-3.58 (m, 1H), 3.58-2.93 (m, 2H), 2.45-2.24 (m, 1H), 2.20-2.05 (ddd, $J_{H-H} = 4.7, 8.2, 13.2$ Hz, 1H); ^{13}C NMR (CD_3OD , 125 MHz) δ : 174.37, 165.27 (q, $^3J_{C-F} = 3.5$ Hz), 139.20, 132.66, 130.43, 125.90 (q, $^1J_{C-F} = 275.3$ Hz), 121.89, 70.83, 60.70, 56.98, 43.48, 39.75 (q, $^1J_{C-F} = 28.9$ Hz), 39.39; ^{19}F NMR (CD_3OD , 470 MHz) δ : -64.04 (CF_3).

(2S,4R)-4-hydroxy-N-(4-(4-methylthiazol-5-yl)benzyl)-1-(3,3,3-trifluoropropanoyl)pyrrolidine-2-carboxamide (S11b - spy molecule 20):



Prepared from 22 mg (0.062 mmol) of amine **S9b**, resulting in 22 mg (0.051 mmol, 83%) of the desired product as a white solid. HRMS (ESI) $[M+H]^+$ (m/z): Calculated for $C_{19}H_{20}F_3N_3O_3S$: 428.1250; Observed: 428.1262; 1H

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NMR (CD₃OD, 500 MHz) δ : 8.93-8.89 (s, 1H), 7.50-7.44 (m, 4H), 4.65-4.44 (m, 4H), 3.83-3.79 (dd, $J_{\text{H-H}} = 4.3, 10.9$ Hz, 1H), 3.78-3.58 (dd, $J_{\text{H-H}} = 2.0, 10.9$ Hz, 1H), 3.58-2.93 (m, 2H), 2.52-2.50 (s, 3H), 2.47-2.27 (m, 1H), 2.24-2.09 (ddd, $J_{\text{H-H}} = 4.7, 8.2, 13.2$ Hz, 1H); ¹³C NMR (CD₃OD, 125 MHz) δ : 174.41, 165.29 (q, $^3J_{\text{C-F}} = 3.3$ Hz), 152.98, 149.17, 140.23, 133.51, 131.70, 130.56, 129.03, 125.92 (q, $^1J_{\text{C-F}} = 275.3$ Hz), 70.85, 60.74, 57.00, 43.78, 39.77 (q, $^2J_{\text{C-F}} = 29.0$ Hz), 39.43, 15.88; ¹⁹F NMR (CD₃OD, 470 MHz) δ : -64.04 (CF₃).

3. Protein expression, purification and biotinylation

The VHL E3 ligase is a multi-protein complex composed of five proteins: VHL protein (pVHL), elongin B (eloB), elongin C (eloC), Cullin-2 (Cul2) and Ring-box protein 1 (Rbx1).^[3] Since the compounds developed in this work bind solely to the VHL protein, the VBC complex (equimolar complex of pVHL₅₄₋₂₁₃, eloB₁₋₁₀₄ and eloC₁₇₋₁₁₂) was used in all experiments, as it can be readily expressed in *E. coli* with high yields,^[4] while the full E3 ligase would require baculovirus-insect cells expression system.^[5] The expression and purification of VBC was performed as described previously by our group^[2] and employed directly in all NMR experiments.

For the surface plasmon resonance (SPR) experiments, a VBC complex containing an AviTagTM in the N-terminus of eloB (AviVBC) was purified using the same procedure described for VBC. The modified eloB/eloC expression plasmid was previously developed in-house by Dr. Michael Roy and kindly shared. The AviVBC complex was site-specifically biotinylated in the AviTag using the GST-BirA method previously described by Fairhead and Howarth.^[6]

4. Surface plasmon resonance experiments

The SPR experiments were performed with a Biacore T200 instrument (GE Healthcare). All measurements were performed at 20 °C with buffer containing 10 mM HEPES, pH 7.5, 150 mM NaCl, 1 mM TCEP, 0.005% (v/v) Tween® 20 and 2% (v/v) dimethyl sulfoxide (DMSO). Biotinylated AviVBC (~0.5 µM) was immobilised at 22 °C onto a Series S sensor chip SA (GE Healthcare) to levels of approximately 3500-4000 response units (RU).

Solutions of each spy molecule were prepared in buffer at concentrations based on previous structure-activity relationship studies of VHL ligands (K_D expected in the nanomolar range for structures like spy molecules **16** and **22**, or in the millimolar range for spy molecules similar to **1** and **9**). From this first screen, the binding affinities were roughly estimated using the Biacore T200 evaluation software (GE Healthcare), then measurements were repeated using concentrations above and below the K_D obtained in the first round to generate better curves for fitting the data accurately. Contact and dissociation times varied across the different compounds tested, but in general fast binding kinetics were observed for all compounds, fully reaching steady-state or being completely dissociated from the surface in less than sixty seconds.

Data analysis was performed using the steady state responses of the double-referenced sensorgrams (raw data subtracted from blank and reference surface injections) obtained for each concentration tested. These responses were plotted against the respective concentrations and the data fitted to a 1:1 binding model using the Biacore T200 evaluation software and the following equation:

$$R_{eq} = \frac{C \times R_{MAX}}{K_D + C} + offset$$

Where R_{eq} is the steady-state response at a given concentration C . Deviations in R_{eq} were corrected by adding an 'offset' term to the equation. K_D is the dissociation constant to be determined and R_{MAX} is the maximum response expected for a given compound according to the equation below:

$$R_{MAX} = n \times R_{Protein} \frac{MW_{Compound}}{MW_{Protein}}$$

Where n is the stoichiometry of the interaction (in this case, $n = 1$) and $R_{Protein}$ is the immobilization level of protein. $MW_{Compound}$ and $MW_{Protein}$ are the molecular weights of compound and protein, respectively. Sensorgrams and fitting parameters for all spy molecules can be found in section 7.

5. Measurement of the transverse relaxation rates by ^{19}F CPMG NMR

All the NMR experiments were performed in a 500 MHz Bruker AVANCE NMR spectrometer equipped with a CPQCI-F cryoprobe. To measure the transverse relaxation rates (R_2), a solution of each spy molecule at 100 μM was prepared in 50 mM potassium phosphate monobasic (KH_2PO_4), pH 7.5, 100 mM NaCl, 1 mM TCEP, 2% (v/v) DMSO, 20% D_2O and 10 μM trifluoroacetic acid (TFA). For each solution, ^{19}F CPMG experiments (decoupled) were performed varying the total CPMG filter (50, 100, 200, 400, 800, 1200, 1600 and 3200 ms). Due to the fast relaxation of molecules **21** and **22**, these experiments were repeated with shorter CPMG filters (50, 100, 150, 200, 300, 400, 700 and 1000 ms). The fluorine peaks were integrated and plotted against the respective CPMG filters. The R_2 relaxation rates were obtained from fitting the data as an exponential decay (GraphPad Prism 6) according to the equation below: ^[7]

$$I(t) = I(0) \times e^{-R_2 t}$$

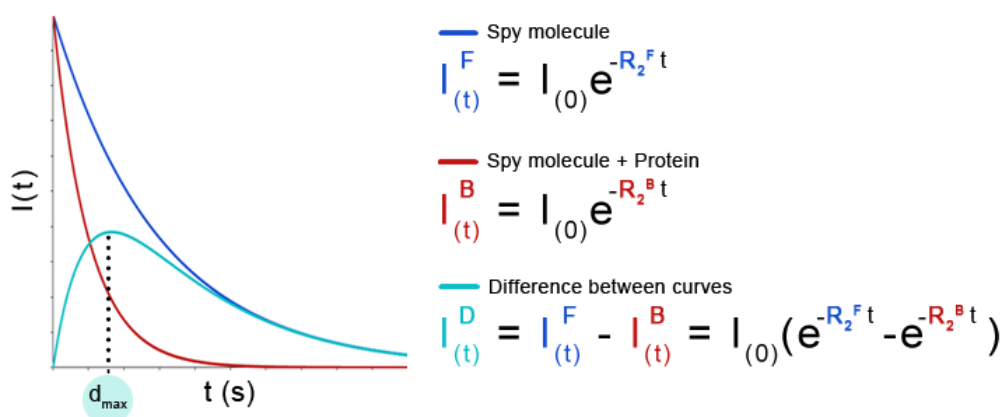
Where $I(t)$ is the ^{19}F signal intensity or integral, t is the total CPMG filter in seconds, $I(0)$ is the signal intensity when $t = 0$. To obtain the R_2 upon addition of protein, all the experiments above were repeated in presence of VBC at 1 μM . Experiments were performed as triplicates and the R_2 contrasts (C_2) were obtained according to the equation below: ^[8]

$$C_2 = \frac{R_2^{\text{Observed}} - R_2^{\text{Free}}}{R_2^{\text{Observed}}}$$

Where R_2^{Free} is the R_2 obtained for the spy molecule free in solution and R_2^{Observed} is the R_2 obtained with the spy molecule in presence of VBC. The raw data, fitting, R_2 and C_2 values for each spy molecule can be found in section 8.1. For the R_2 measurements using molecule **19** at varied concentrations of spy molecule and protein, see section 8.2. For the R_2 measurements using spy molecules **6** and **11** (used to generate figures S2, S3 and S4) see section 8.3.

6. Competition experiments by ^{19}F NMR

As full measurements of R_2 would be very time consuming, the competition experiments were performed using a single ^{19}F CPMG experiment (decoupled) per sample with a fixed CPMG delay. To determine the best CPMG delay for a given spy molecule and assay condition, the procedure described in Figure S5 was developed based on the transverse relaxation rate equations.^[7] By knowing the relaxation rates of the spy molecule free in solution and in presence of protein, the CPMG delay where the difference between the NMR peaks is maximum is hereon referred as d_{\max} . The d_{\max} for all the conditions tested for each spy molecule can be found in section 8, together with the respective values of R_2 and C_2 .



When $t = d_{\max}$, the maximum of the difference curve is described as:

$$\frac{d}{dt} I_{(t)}^D = 0$$

$$\frac{d}{dt} I_{(0)} (e^{-R_2^F t} - e^{-R_2^B t}) = 0$$

$$R_2^B e^{-R_2^B t} - R_2^F e^{-R_2^F t} = 0 \quad \leftrightarrow$$

Best CPMG filter

$$t = d_{\max} = \frac{\ln\left(\frac{R_2^F}{R_2^B}\right)}{R_2^F - R_2^B}$$

Method for selecting the best CPMG delay for competition experiments. With the transverse relaxation rates of the spy molecule free in solution (R_2^F , resulting in the blue plot) and bound to protein (R_2^B , resulting in the red plot), the difference between the two curves is described by $I(t)^D$ (cyan curve). To obtain d_{\max} , the delay where the difference curve reaches its maximum, the first derivative of $I(t)^D$ was obtained and equalled to 0, subsequently isolating t (CPMG filter).

After the d_{\max} for each condition was established, the competition experiments with spy molecules **6** (Figure S3), **11** (Figure S4) and **19** (Figure 5 and S5) were performed with solutions containing 50 mM potassium phosphate monobasic (KH_2PO_4), pH 7.5, 100 mM NaCl, 1 mM TCEP, 2% (v/v) DMSO, 20% D_2O and 10 μM trifluoroacetic acid (TFA). Each assay consisted of two controls:

SUPPORTING INFORMATION

- 1) Spy molecule free in solution
- 2) Spy molecule in presence of protein

Samples containing different concentrations of competitors were prepared in presence of spy molecule and protein and ^{19}F CPMG spectra at the respective d_{max} were collected. The displacement of the spy molecule was obtained from the equation below:

$$\text{Displacement} = \frac{I_C - I_P}{I_F - I_P} \times 100\%$$

Where I_F is the integral of the fluorine peak of the spy molecule free in solution, I_P is the integral of the fluorine peak of the spy molecule in presence of protein and I_C is the integral of the fluorine peak of the spy molecule in presence of protein and a competitor at a given concentration.

To determine the dissociation constant of a competitor, the displacement of the spy molecule was plotted against the concentration of the competitors, and then fitted to a “ \log_{10} [Inhibitor] versus Normalised response” model using GraphPad Prism 6.0, resulting in the plots observed in Figures 5, S3 and S4. By knowing the concentrations of spy molecule, protein and K_D of the spy molecule (K_S), the IC_{50} values obtained from the fitting were converted into the K_D of competitor (K_i) using the Nikolovska-Coleska relationship.^[9]

$$K_i = \frac{[C]_{50}}{\frac{[S]_{50}}{K_S} + \frac{[P]_0}{K_S} + 1}$$

Where $[C]_{50}$ and $[S]_{50}$ are, respectively, the free concentrations of competitor and spy molecule at 50% inhibition (concentration of competitor equals the IC_{50}), and $[P]_0$ is the free concentration of protein in presence of just the spy molecule. As the total concentrations of protein (P_t) and spy molecule (S_t) are known, these values can be obtained from the equations below:^[9a]

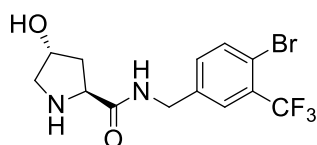
$$P_0 = \frac{[P]_t - K_S - [S]_t - \sqrt{(K_S + [S]_t - [P]_t)^2 + 4[P]_t K_S}}{2}$$

$$[S]_{50} = [S]_t - \frac{([P]_t - [P]_0)}{2}$$

$$[C]_{50} = IC_{50} - [P]_0 + \frac{([P]_t - [P]_0)}{2} \times \left(1 + \frac{K_S}{[S]_{50}}\right)$$

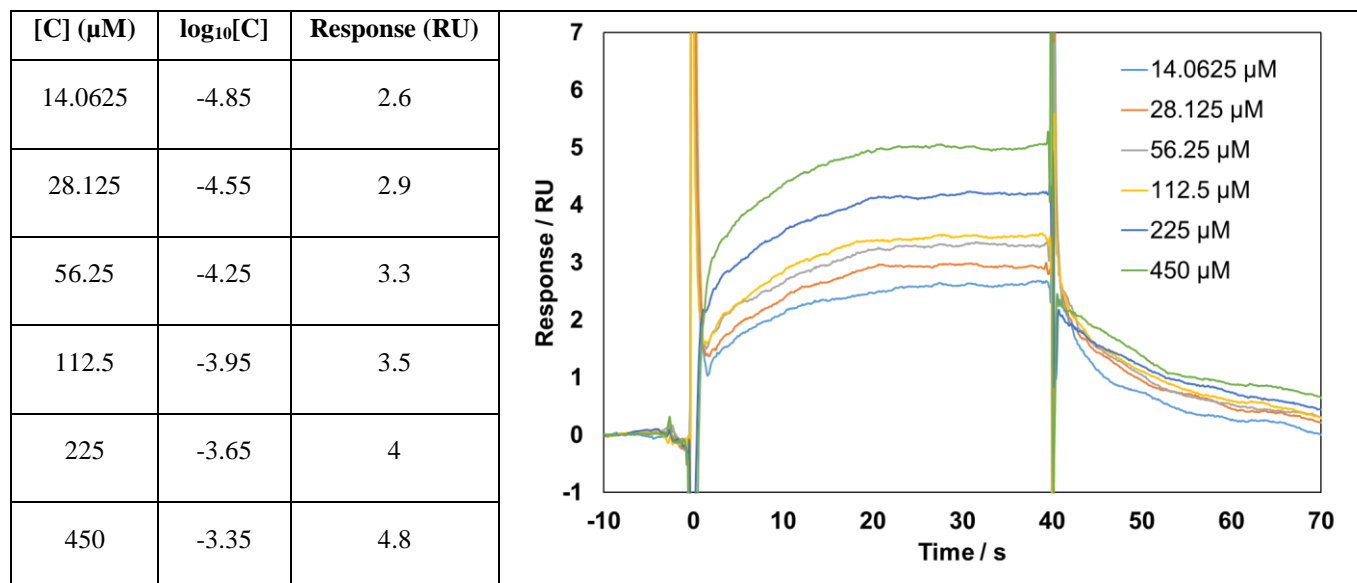
SUPPORTING INFORMATION

7. Surface plasmon resonance – Sensorgrams and data fitting



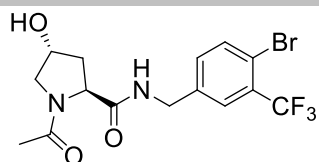
Spy molecule 1 (Compound S3a)

Blank and reference surface subtracted responses according to the concentration of spy molecule. **Theoretical $R_{MAX} = 25.4$**



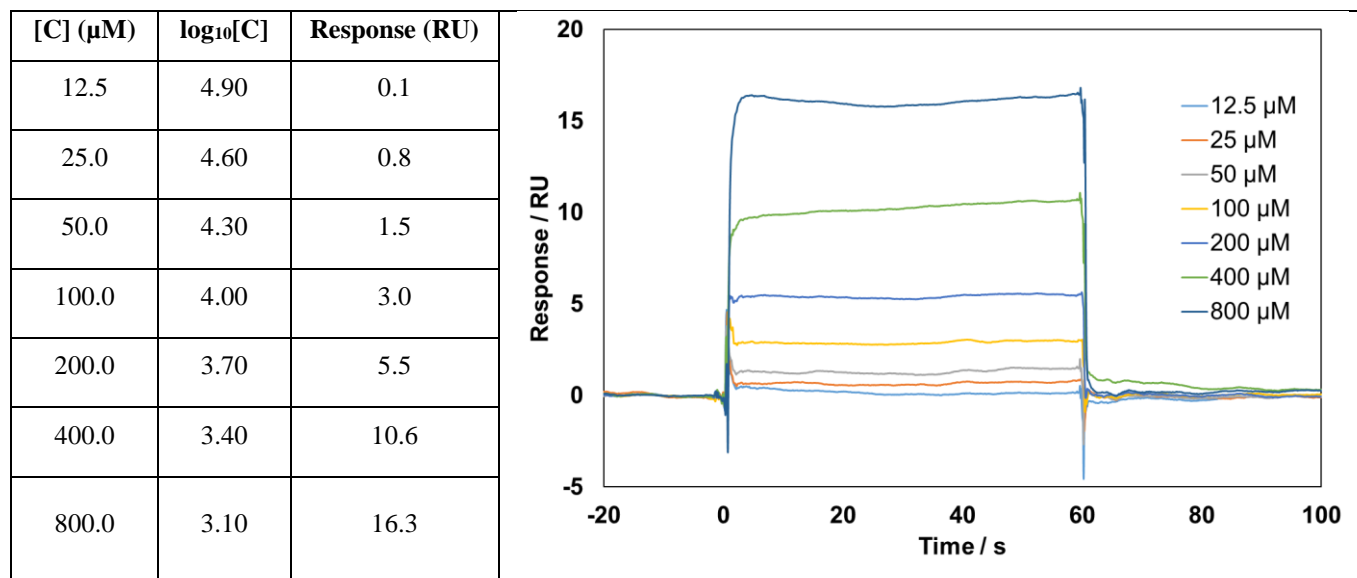
Data could not be fitted. Responses increase with concentration, but too far from the theoretical R_{MAX} . $K_D \gg \gg 1.0$ mM.

SUPPORTING INFORMATION

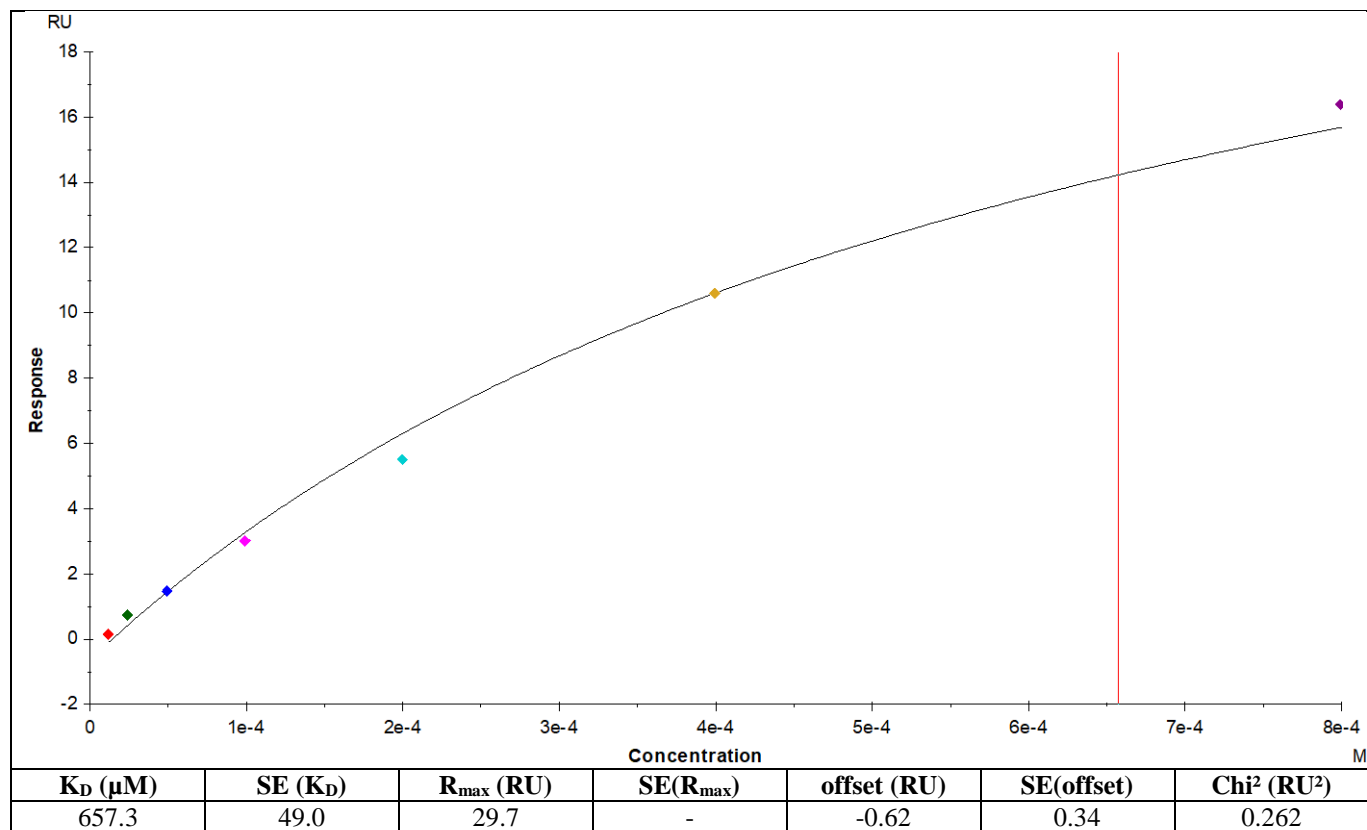


Spy molecule 2 (Compound S4a)

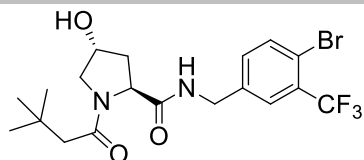
Blank and reference surface subtracted responses according to the concentration of spy molecule. **Theoretical $R_{MAX} = 29.7$**



Data fitting using the Biacore T200 evaluation software. As responses were lower than the theoretical R_{MAX} , fitting was performed with a fixed R_{MAX} .

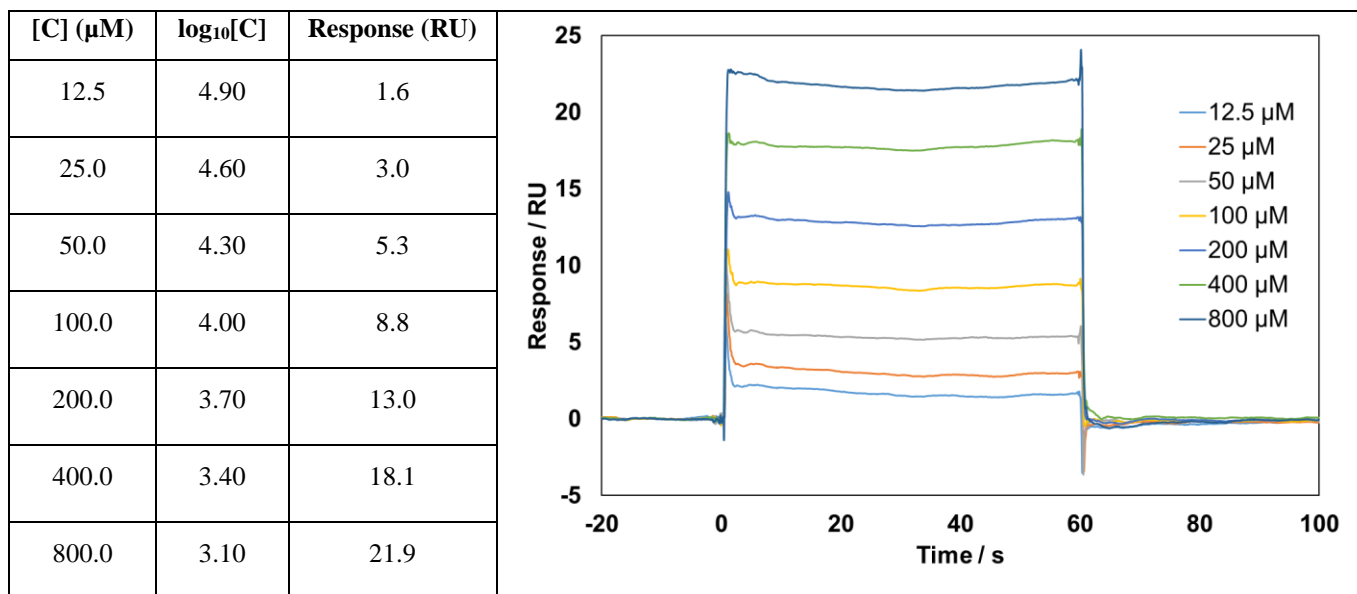


SUPPORTING INFORMATION

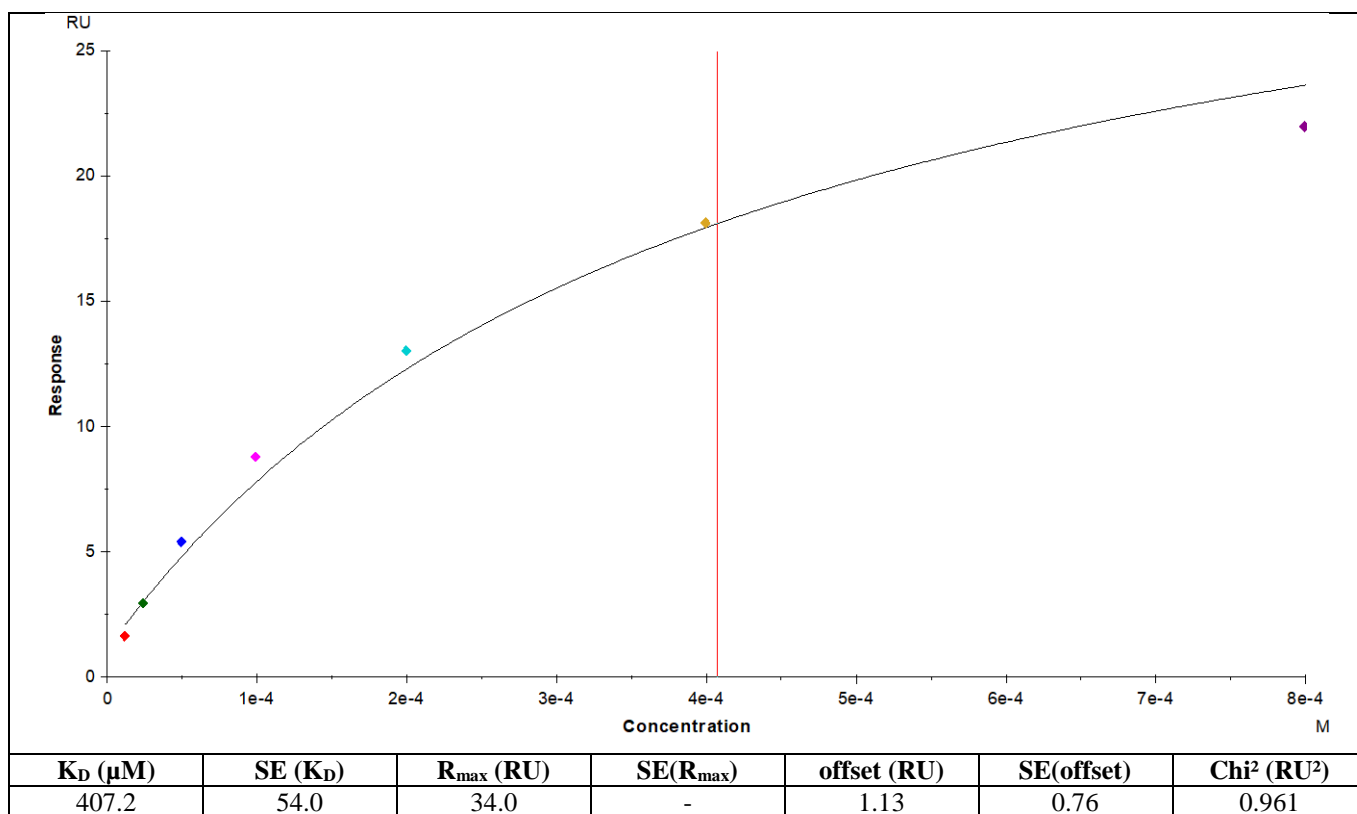


Spy molecule 3 (Compound S5a)

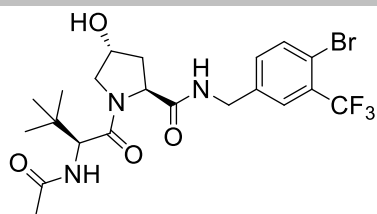
Blank and reference surface subtracted responses according to the concentration of spy molecule. **Theoretical R_{MAX} = 34.0**



Data fitting using the Biacore T200 evaluation software. As responses were lower than the theoretical R_{MAX} , fitting was performed with a fixed R_{MAX} .



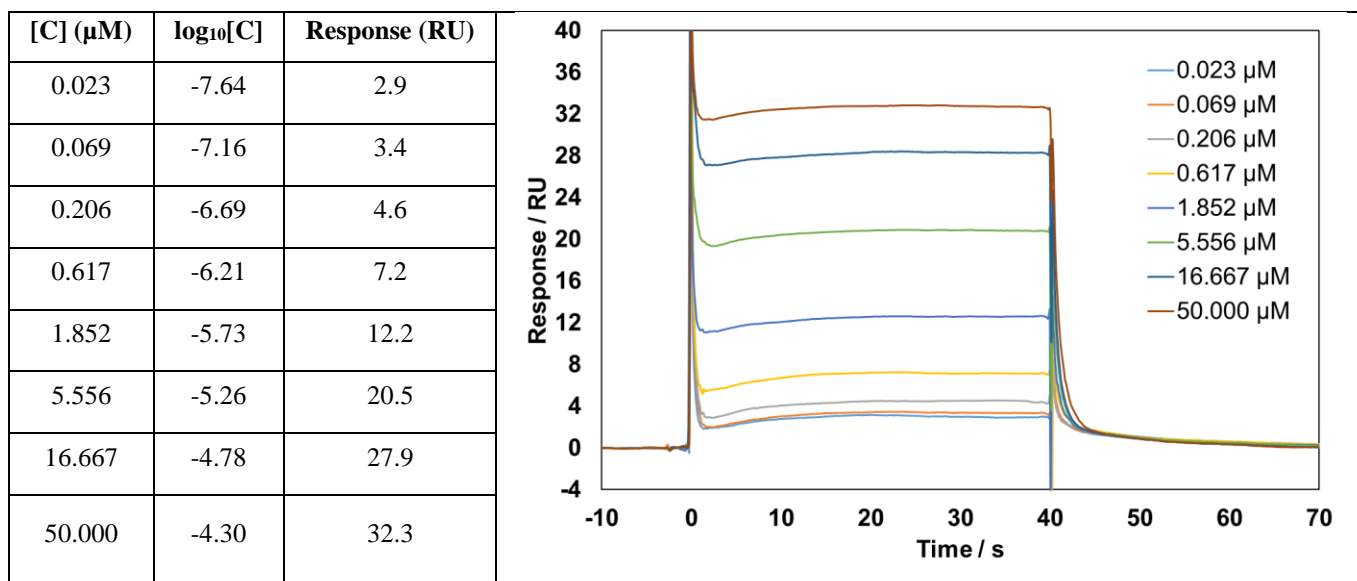
SUPPORTING INFORMATION



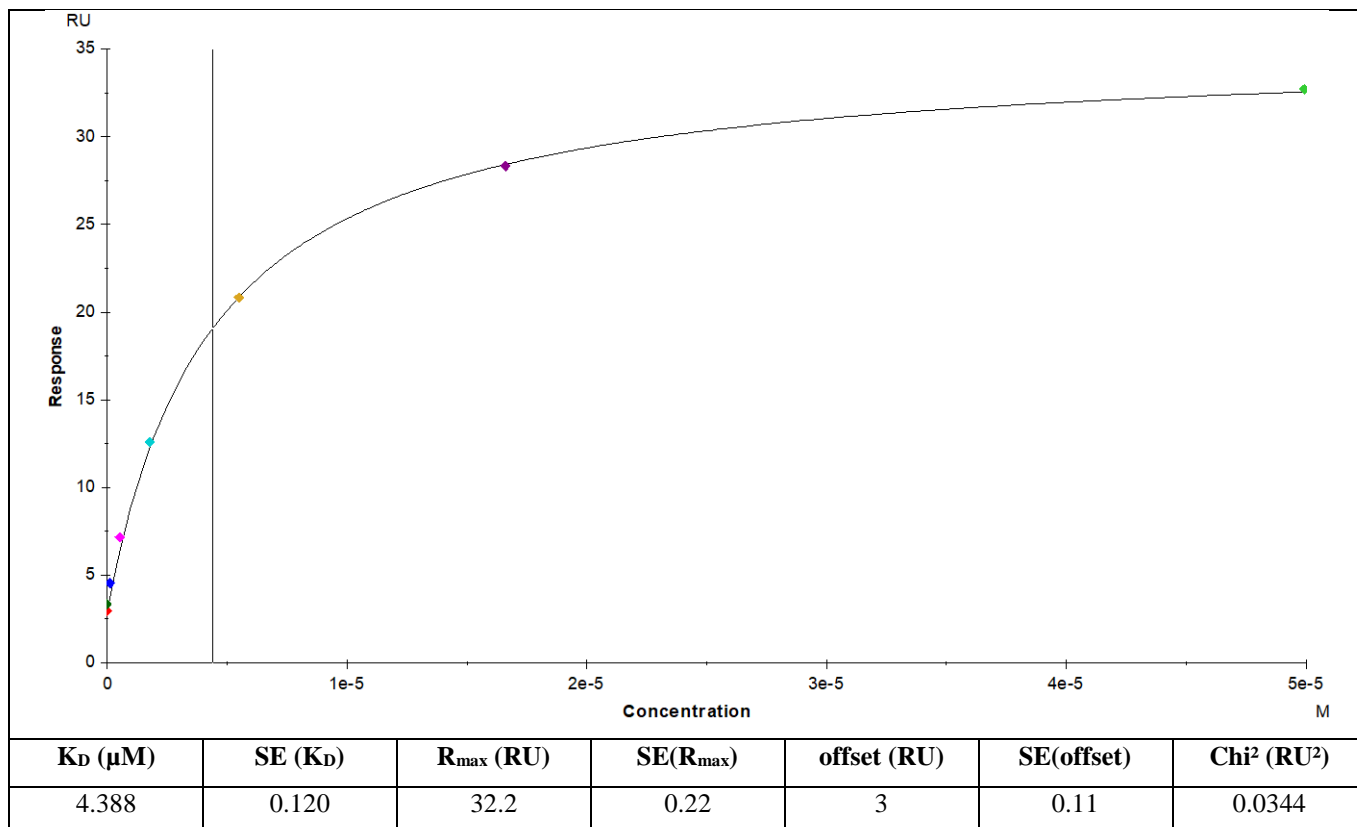
Spy molecule 4 (Compound S6a)

Blank and reference surface subtracted responses according to the concentration of spy molecule.

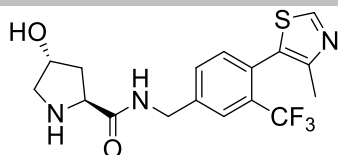
Theoretical $R_{MAX} = 35.3$



Data fitting using the Biacore T200 evaluation software:



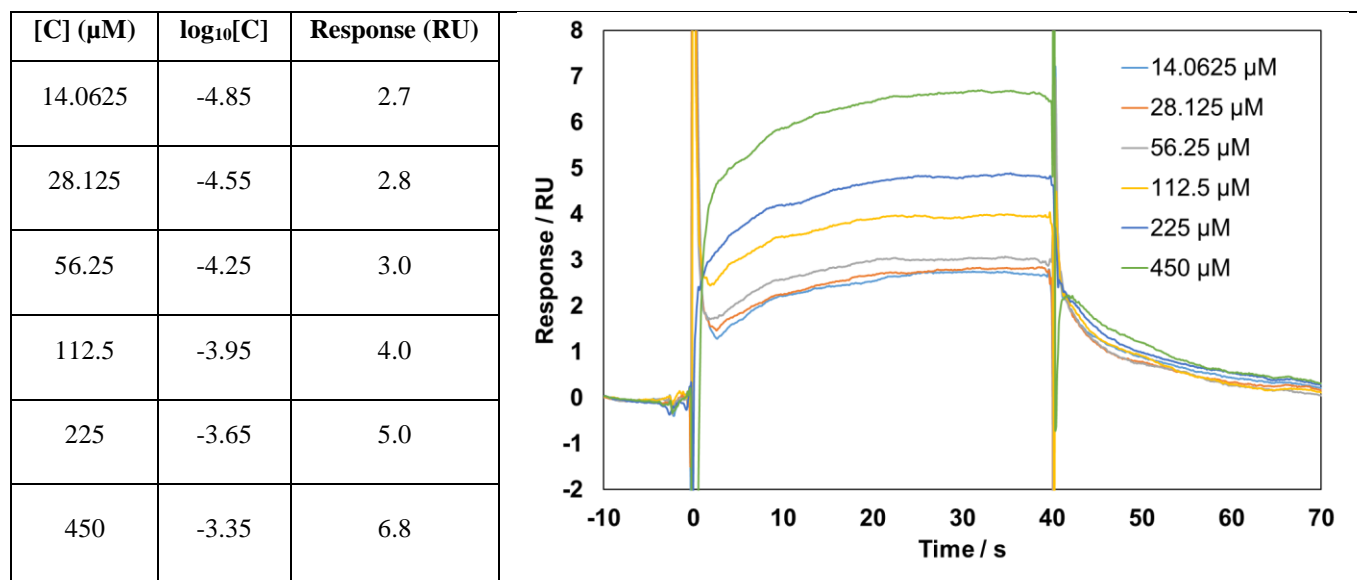
SUPPORTING INFORMATION



Spy molecule 5 (Compound S3b)

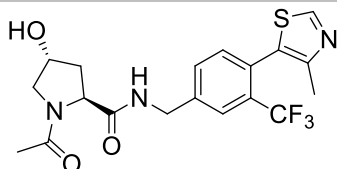
Blank and reference surface subtracted responses according to the concentration of spy molecule:

Theoretical $R_{MAX} = 26.0$



Data could not be fitted. Responses increase with concentration, but too far from the theoretical R_{MAX} . $K_D \gg \gg 1.0 \text{ mM}$.

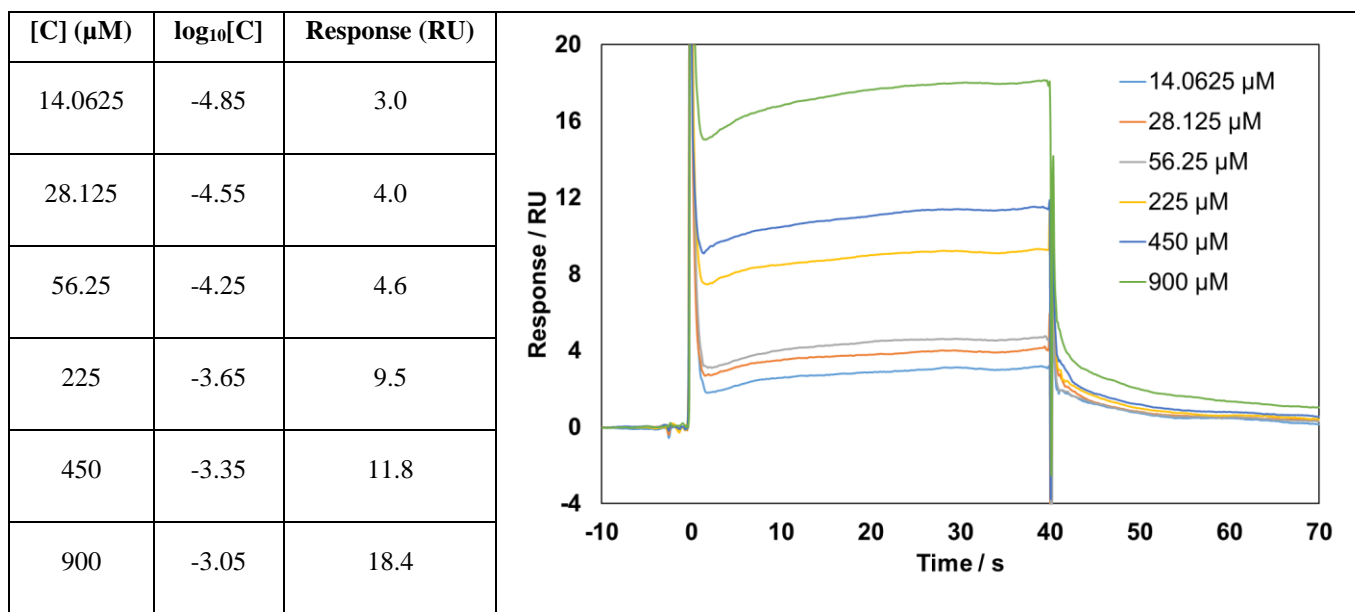
SUPPORTING INFORMATION



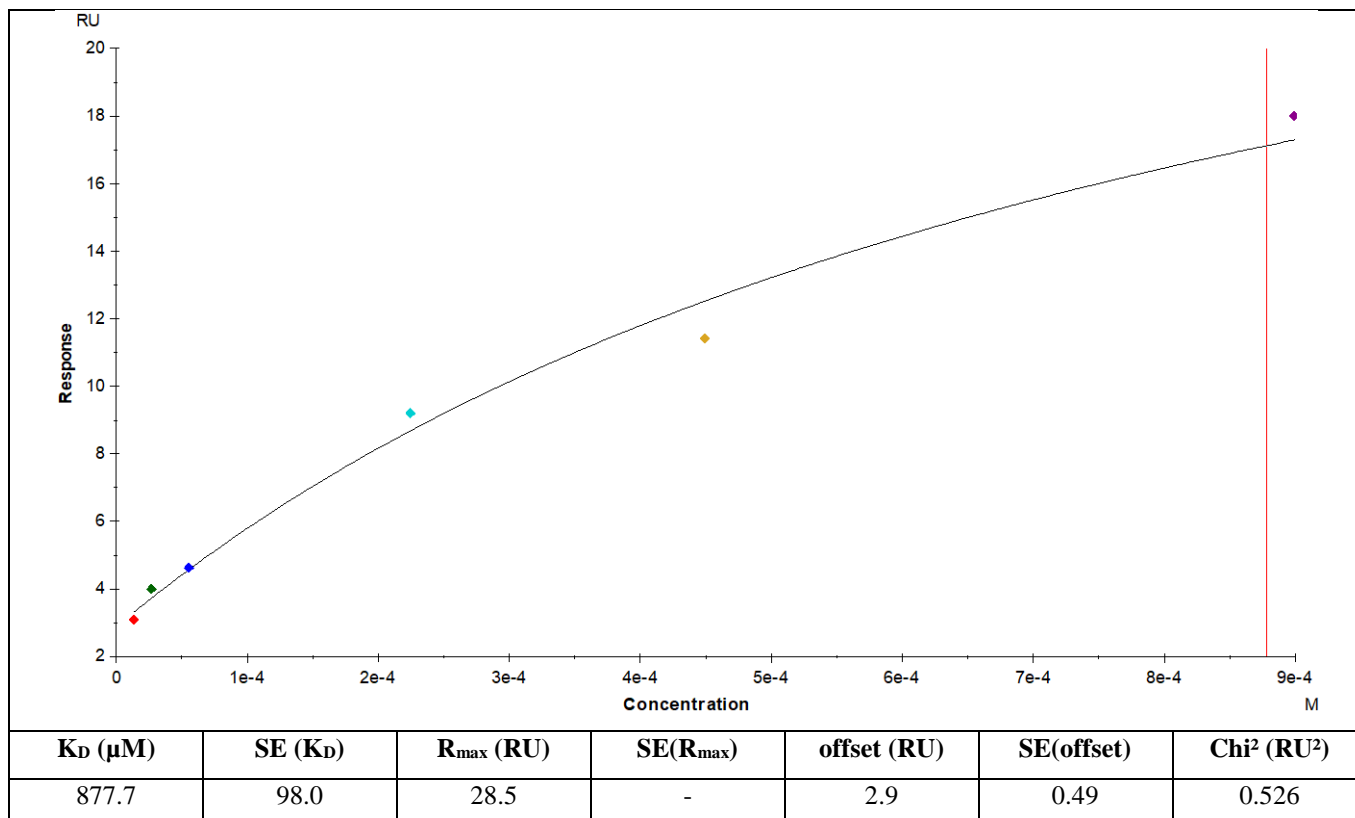
Spy molecule 6 (Compound S4b)

Blank and reference surface subtracted responses according to the concentration of spy molecule:

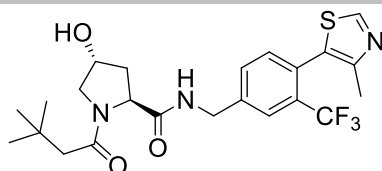
Theoretical $R_{MAX} = 28.5$



Data fitting using the Biacore T200 evaluation software. As responses were much lower than the theoretical R_{MAX} , fitting was performed with a fixed R_{MAX} :



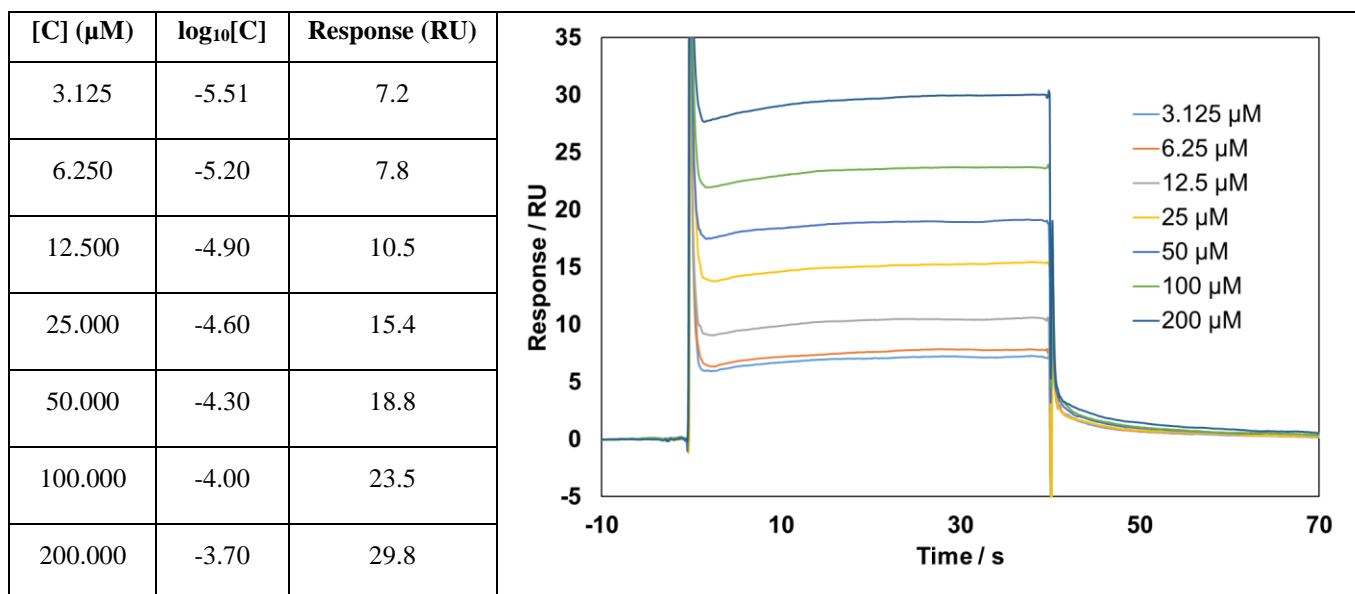
SUPPORTING INFORMATION



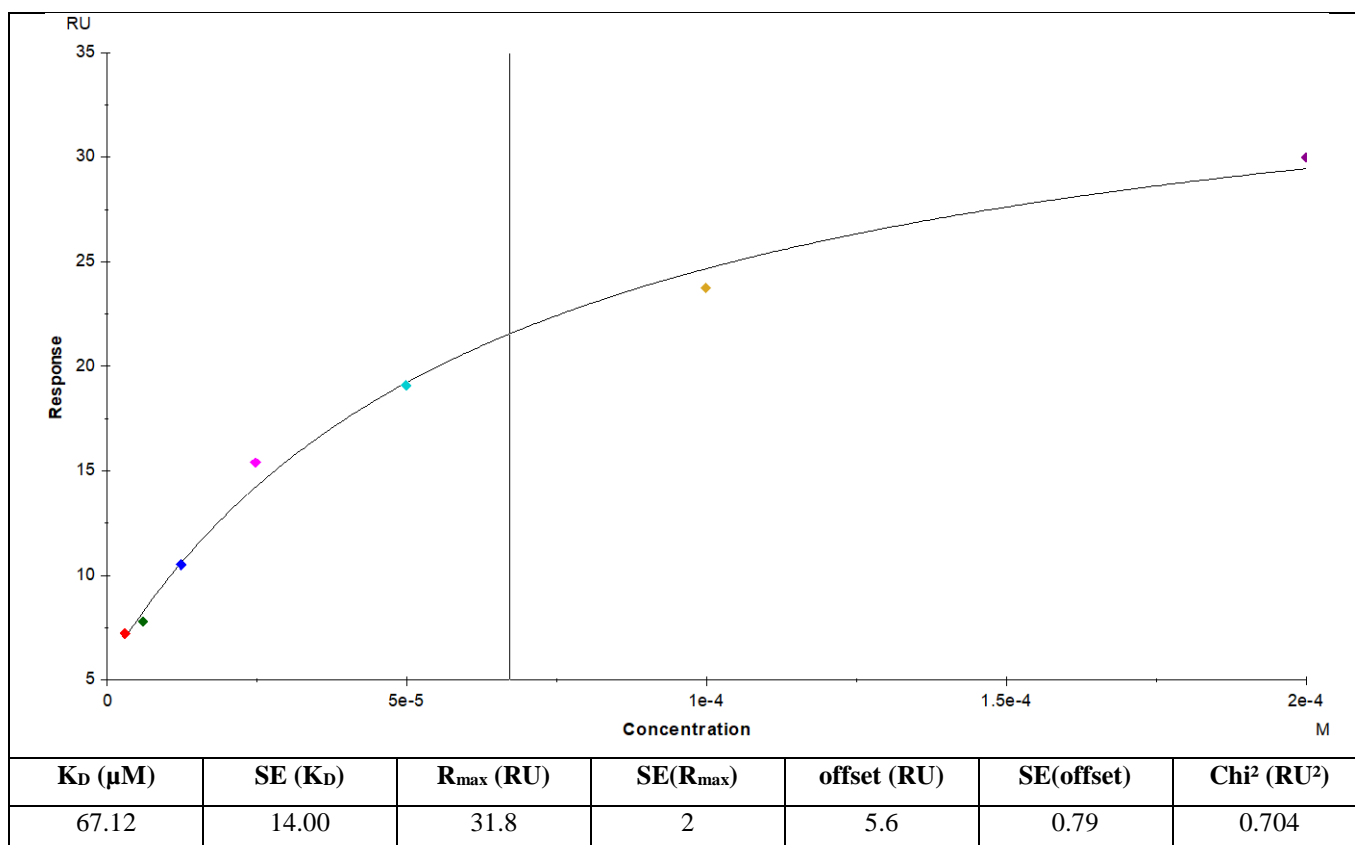
Spy molecule 7 (Compound S5b)

Blank and reference surface subtracted responses according to the concentration of spy molecule.

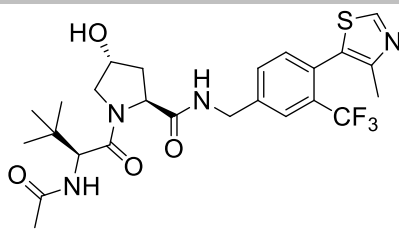
Theoretical $R_{MAX} = 32.0$



Data fitting using the Biacore T200 evaluation software:



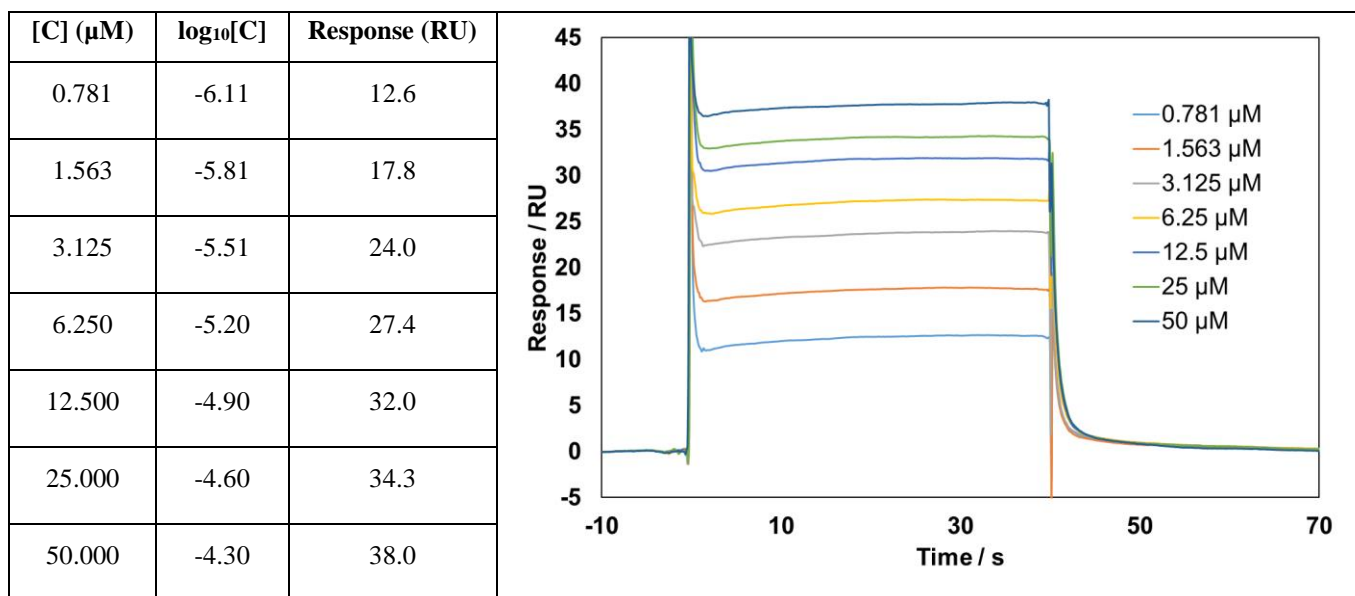
SUPPORTING INFORMATION



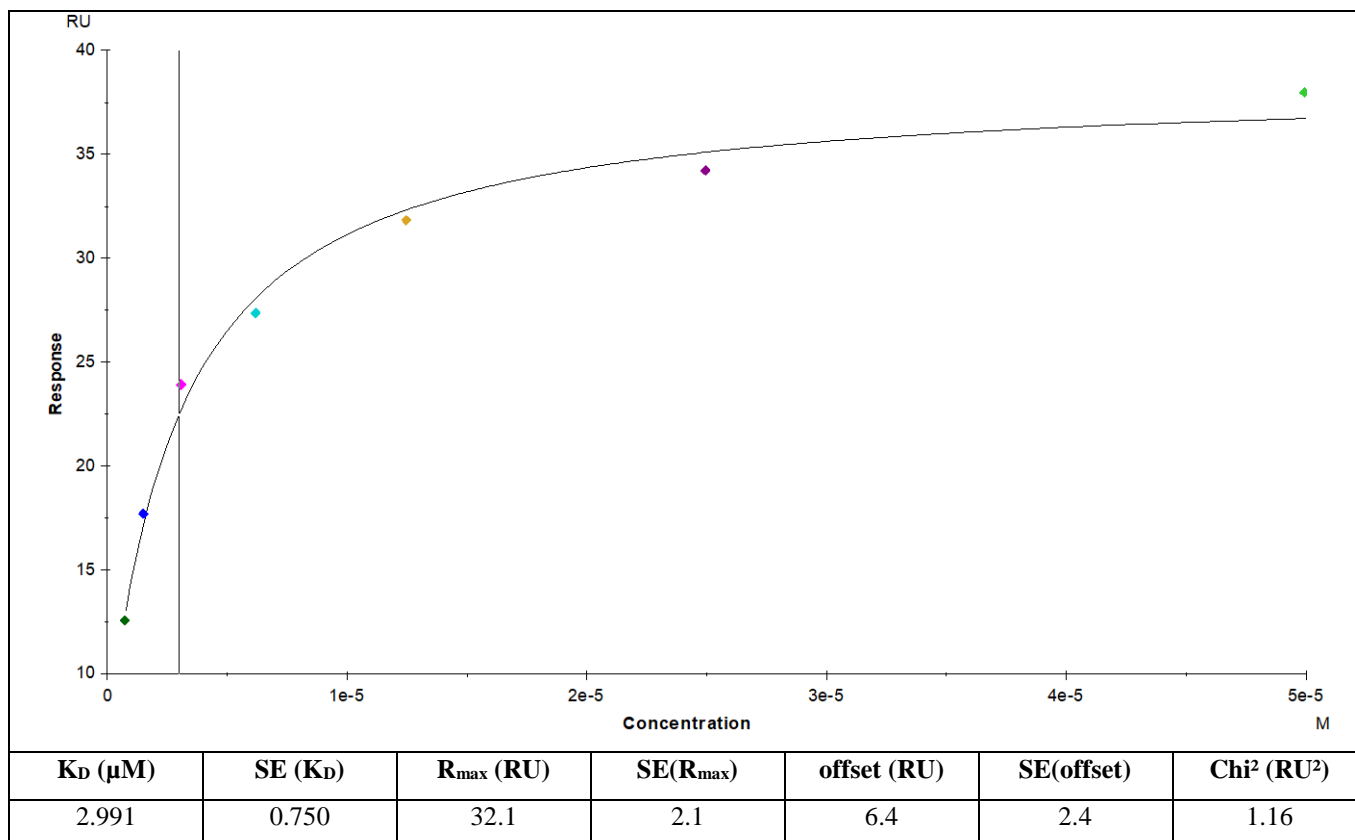
Spy molecule 8 (Compound S6b)

Blank and reference surface subtracted responses according to the concentration of spy molecule.

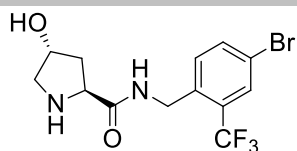
Theoretical $R_{MAX} = 35.5$



Data fitting using the Biacore T200 evaluation software:



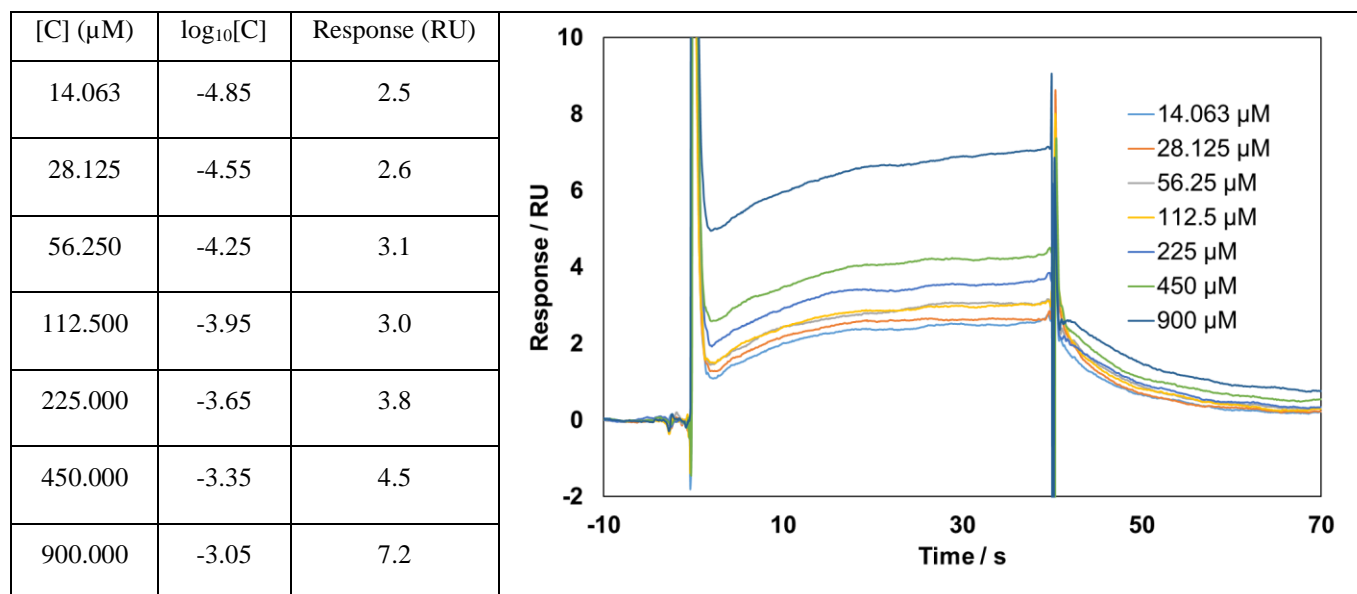
SUPPORTING INFORMATION



Spy molecule 9 (Compound S3c)

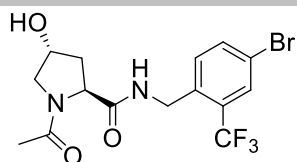
Blank and reference surface subtracted responses according to the concentration of spy molecule.

Theoretical $R_{MAX} = 24.0$



Data could not be fitted. Responses increase with concentration, but too far from the theoretical R_{MAX} . $K_D \gg \gg 1.0 \text{ mM}$.

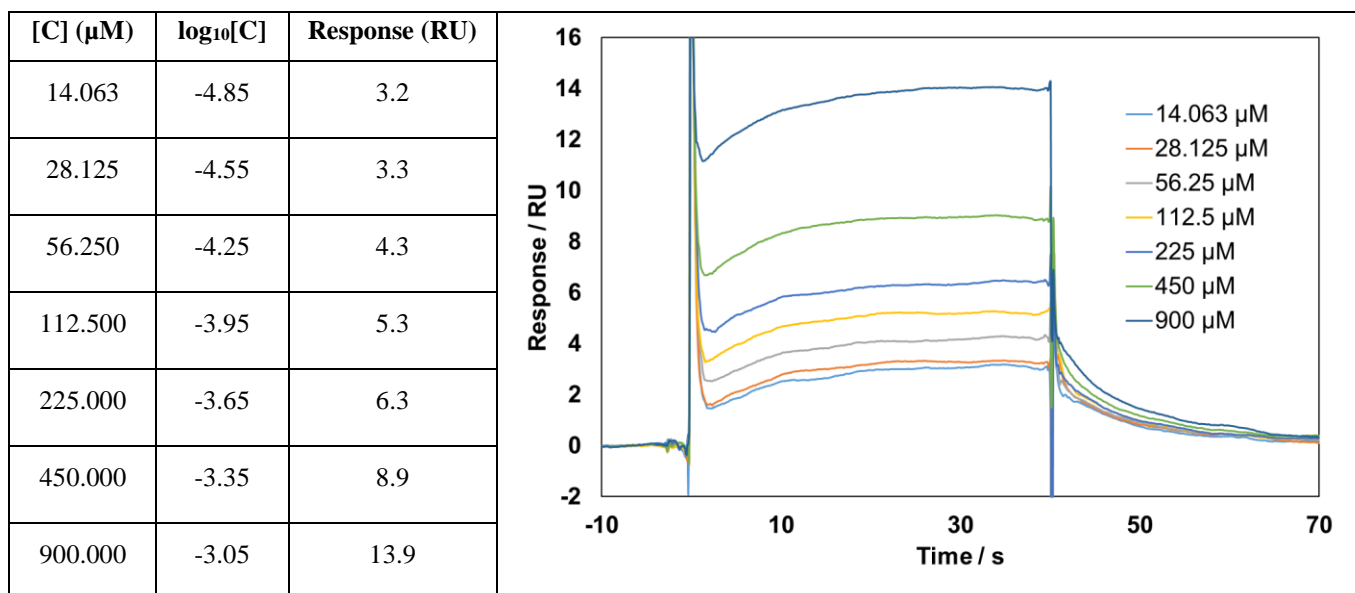
SUPPORTING INFORMATION



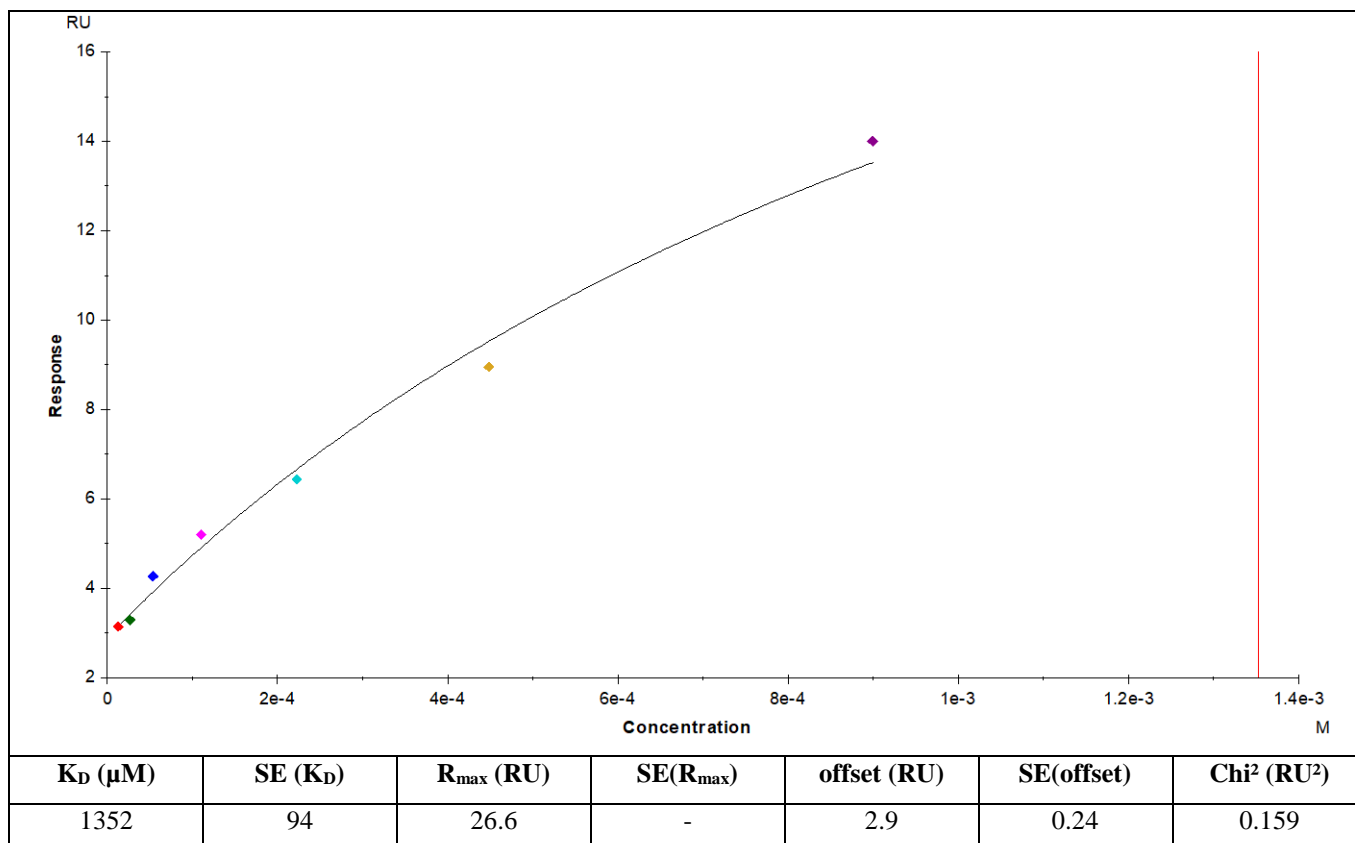
Spy molecule 10 (Compound S4c)

Blank and reference surface subtracted responses according to the concentration of spy molecule.

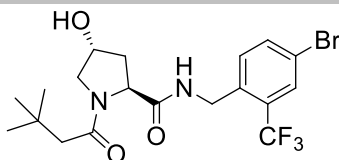
Theoretical $R_{MAX} = 26.6$



Data fitting using the Biacore T200 evaluation software. As responses were lower than the theoretical R_{MAX} , fitting was performed with a fixed R_{MAX} .



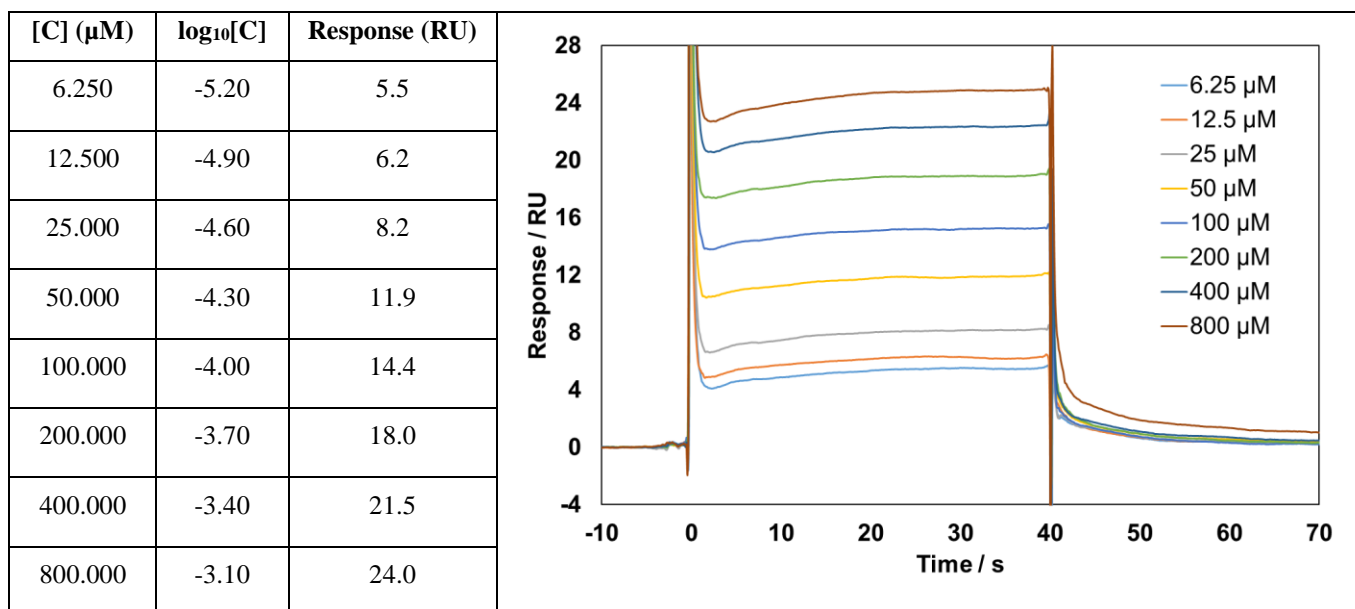
SUPPORTING INFORMATION



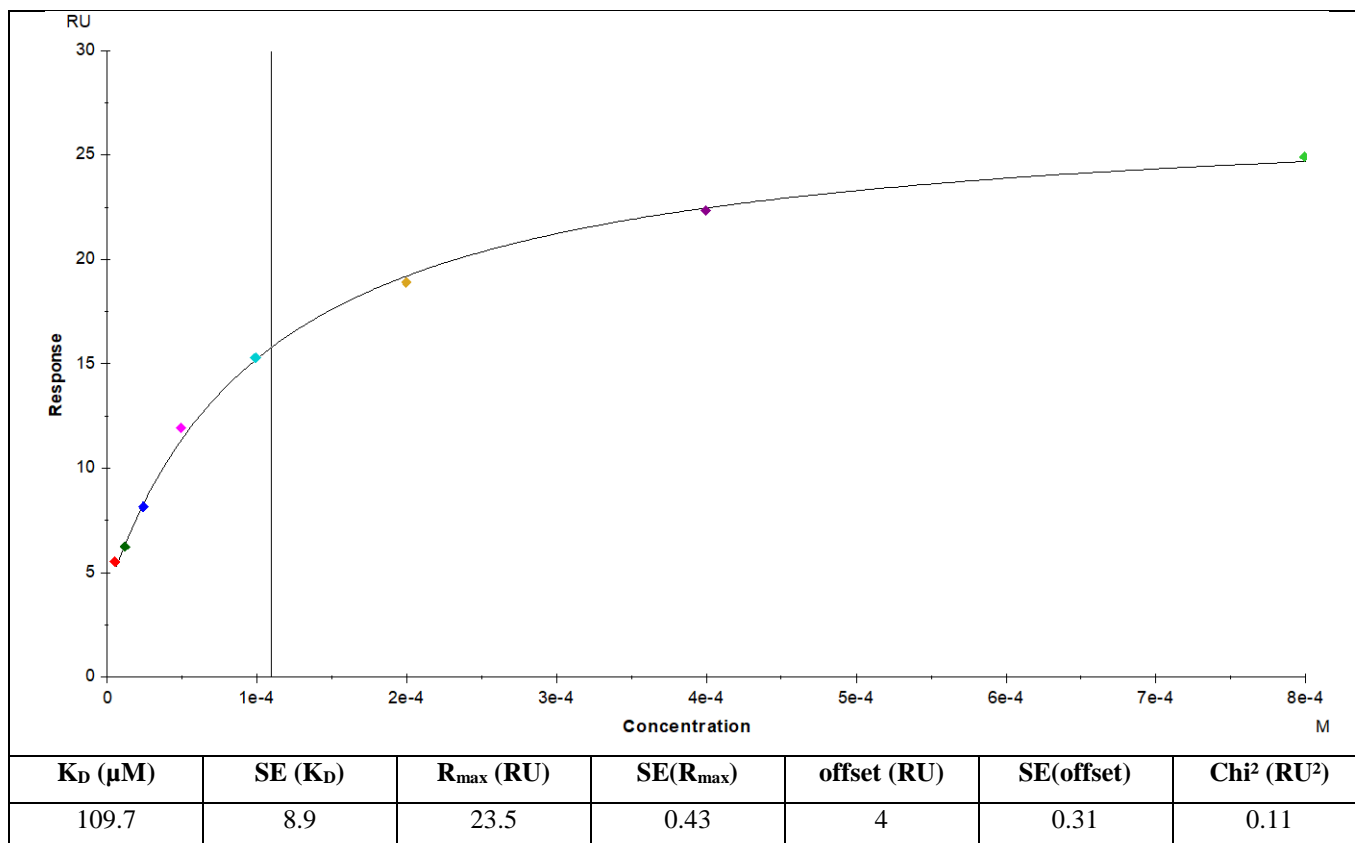
Spy molecule 11 (Compound S5c)

Blank and reference surface subtracted responses according to the concentration of spy molecule.

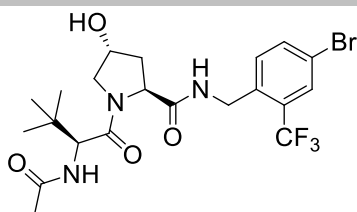
Theoretical $R_{MAX} = 29.8$



Data fitting using the Biacore T200 evaluation software:



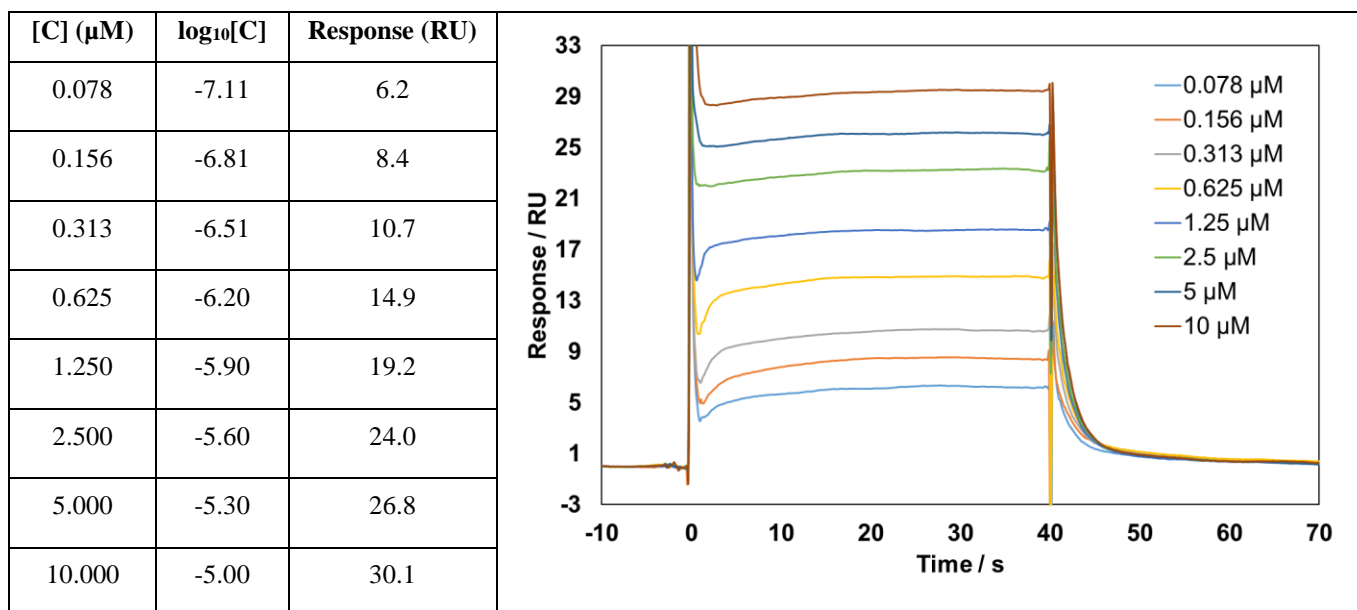
SUPPORTING INFORMATION



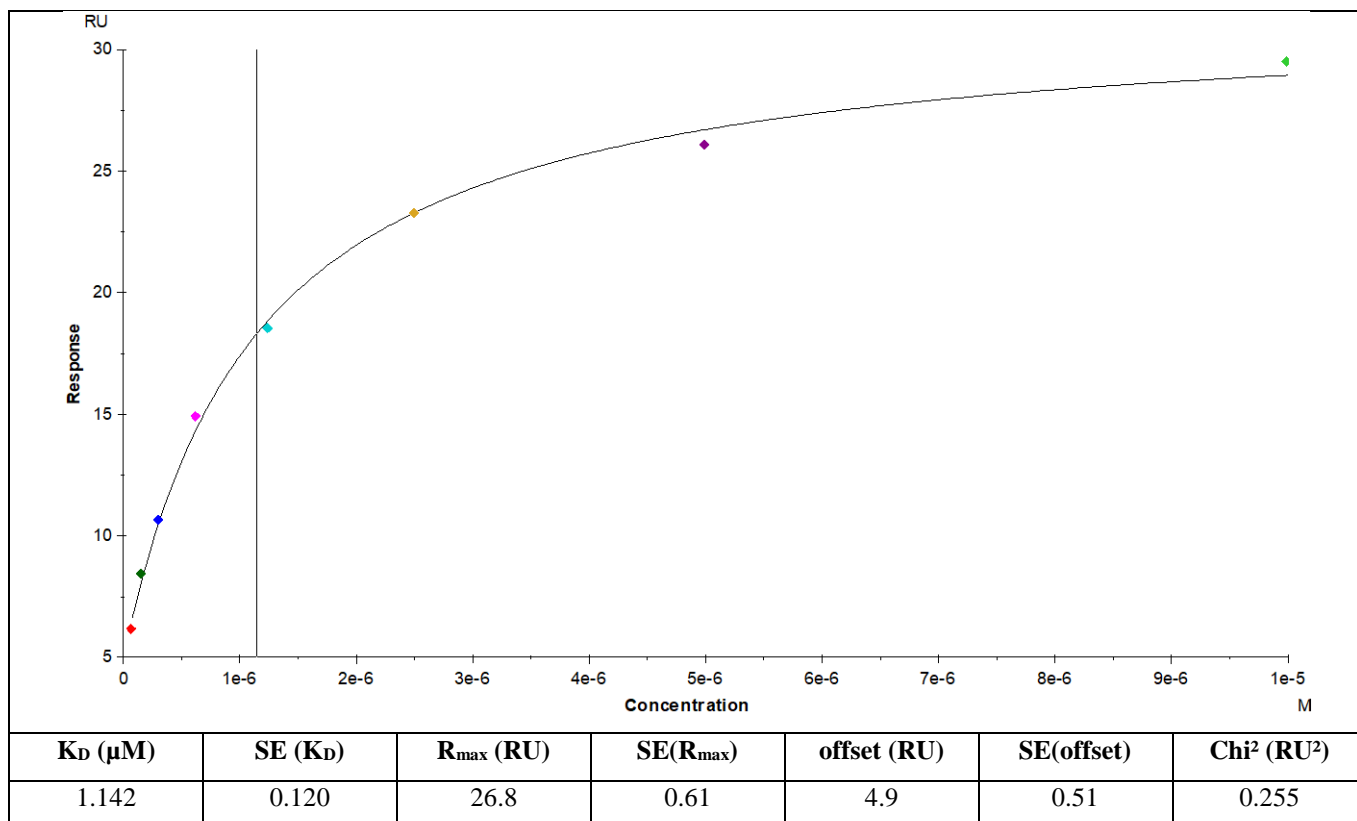
Spy molecule 12 (Compound S6c)

Blank and reference surface subtracted responses according to the concentration of spy molecule.

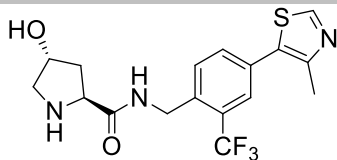
Theoretical $R_{MAX} = 33.2$



Data fitting using the Biacore T200 evaluation software:



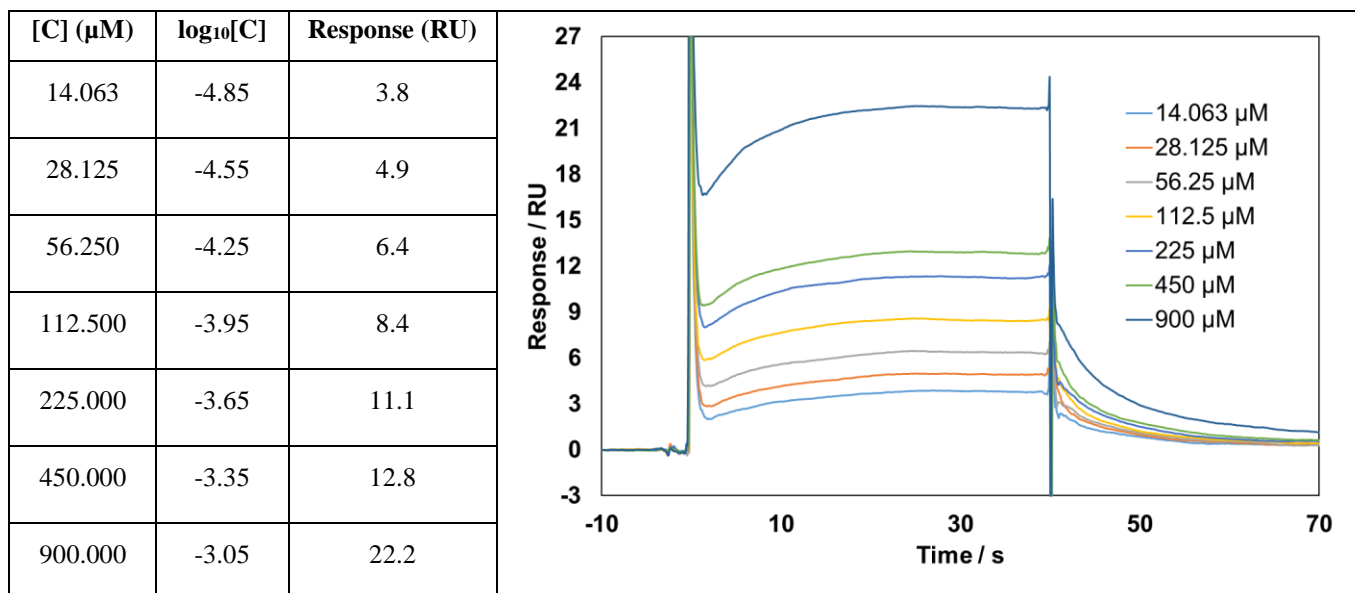
SUPPORTING INFORMATION



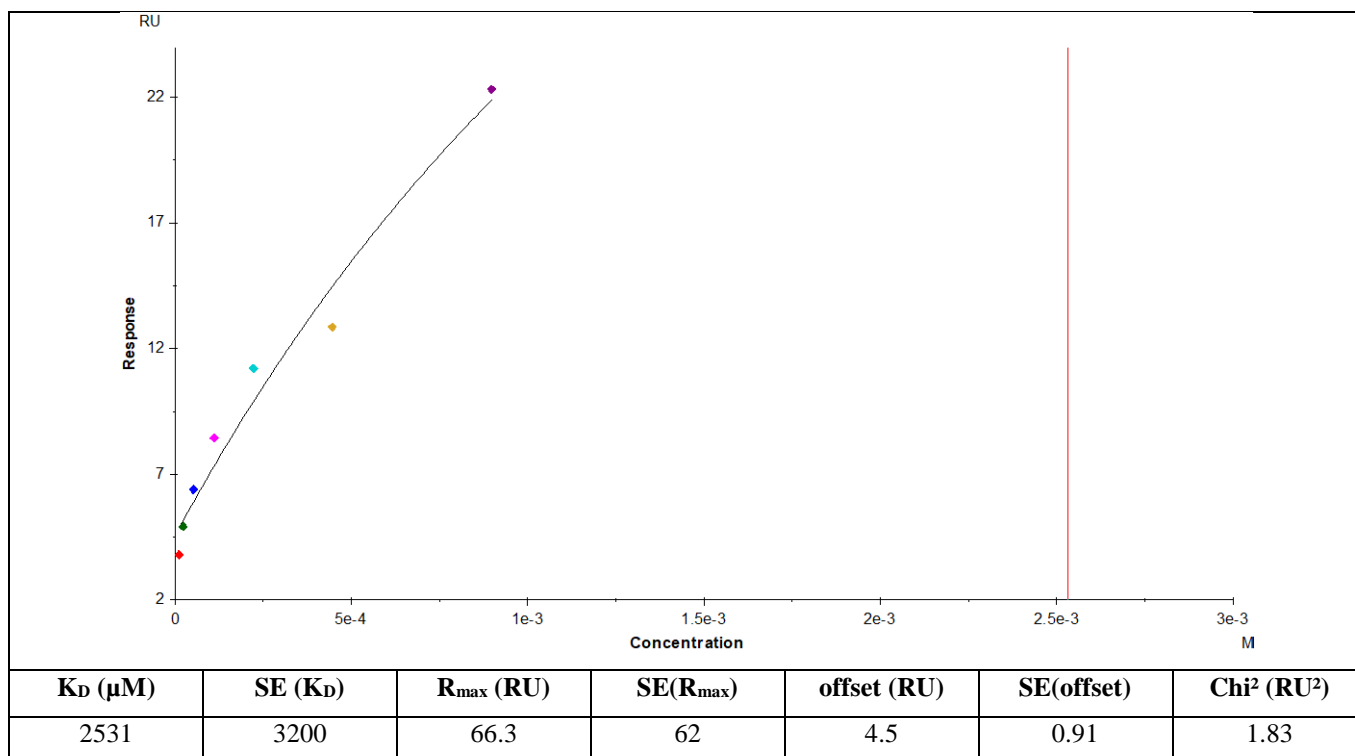
Spy molecule 13 (Compound S3d)

Blank and reference surface subtracted responses according to the concentration of spy molecule.

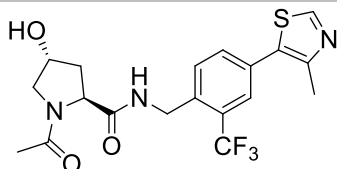
Theoretical $R_{MAX} = 24.3$



Data fitting using the Biacore T200 evaluation software: The fitted K_D presented a very large error and high R_{MAX} . The results not significant even using different fitting methods (fixing R_{MAX} or the offset). Large difference in response between concentrations of 450 and 900 μM might indicate unspecific / promiscuous binding to the protein surface. Similar result obtained from different repeats of different stocks of the compound.



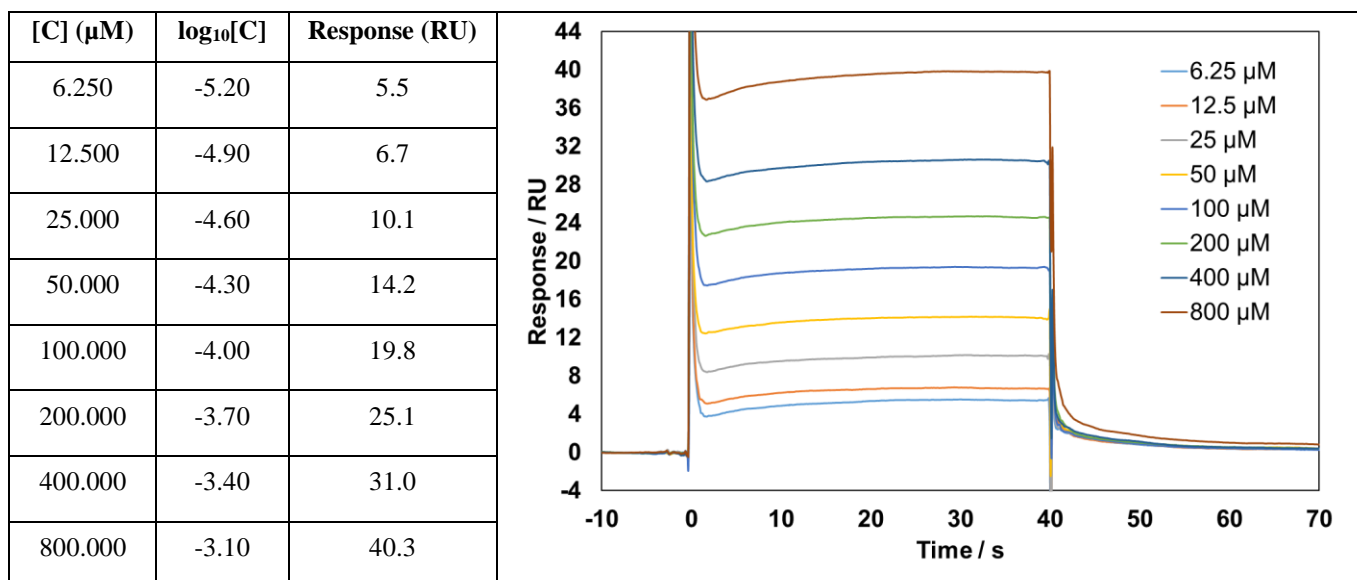
SUPPORTING INFORMATION



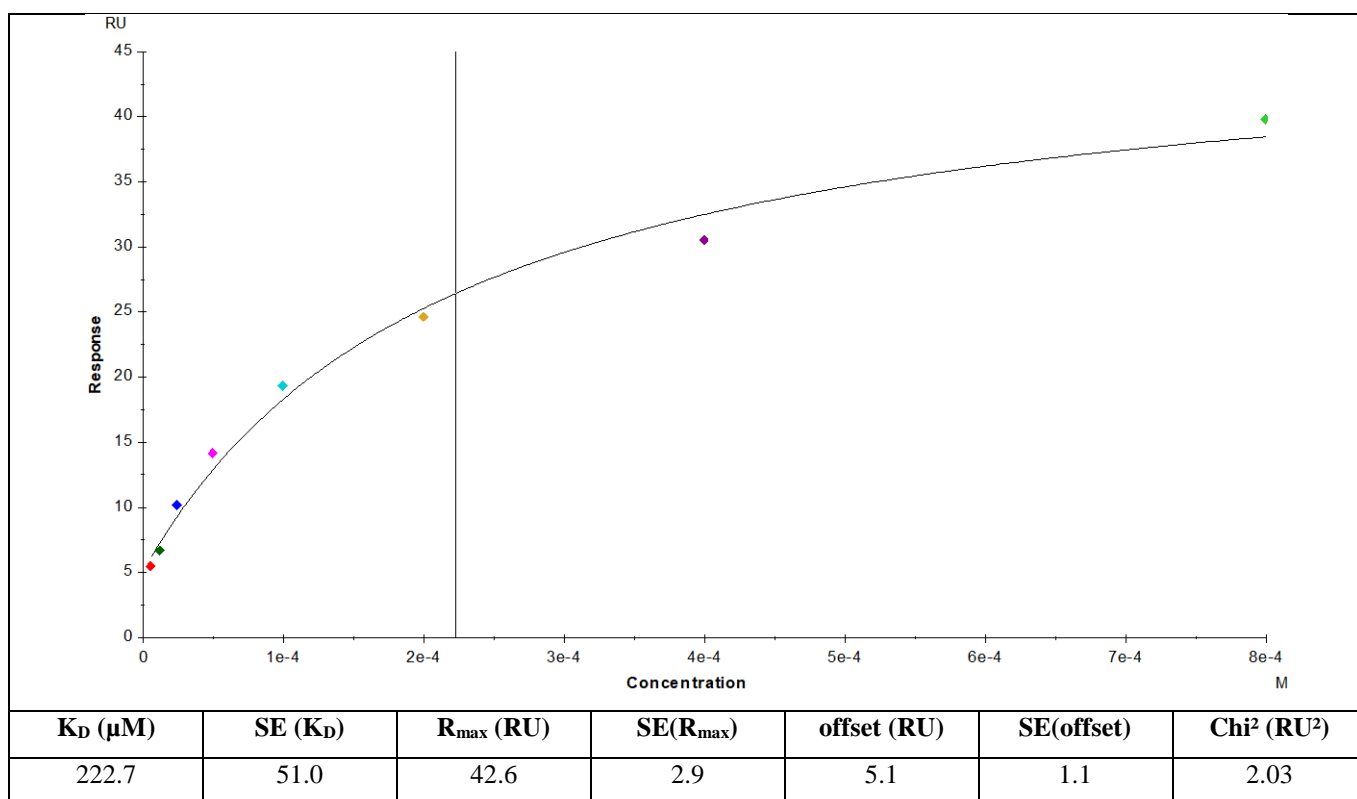
Spy molecule 14 (Compound S4d)

Blank and reference surface subtracted responses according to the concentration of spy molecule.

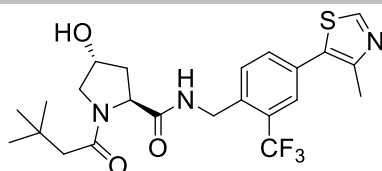
Theoretical $R_{MAX} = 26.8$



Data fitting using the Biacore T200 evaluation software: The only acceptable fitting resulted in a very large R_{MAX} . (1.6 times larger than the theoretical). Fitting with fixed theoretical R_{MAX} presented a large error in the K_D . Compound might bind to the VHL-HIF site, but also bind unspecifically / promiscuously to the protein surface.



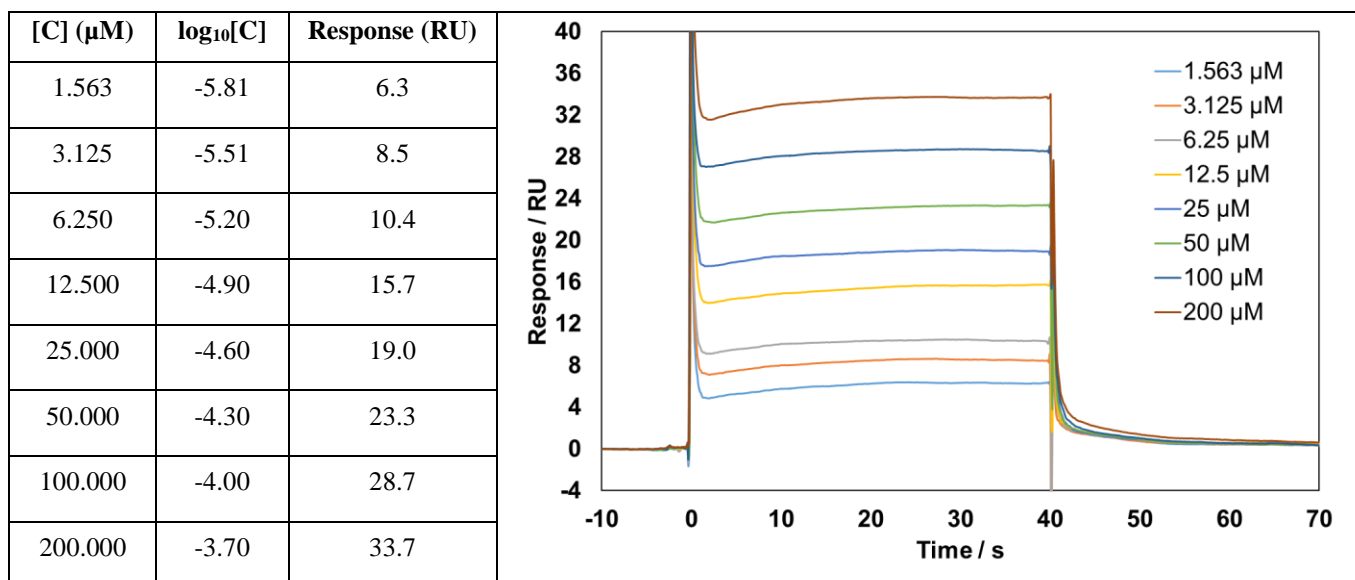
SUPPORTING INFORMATION



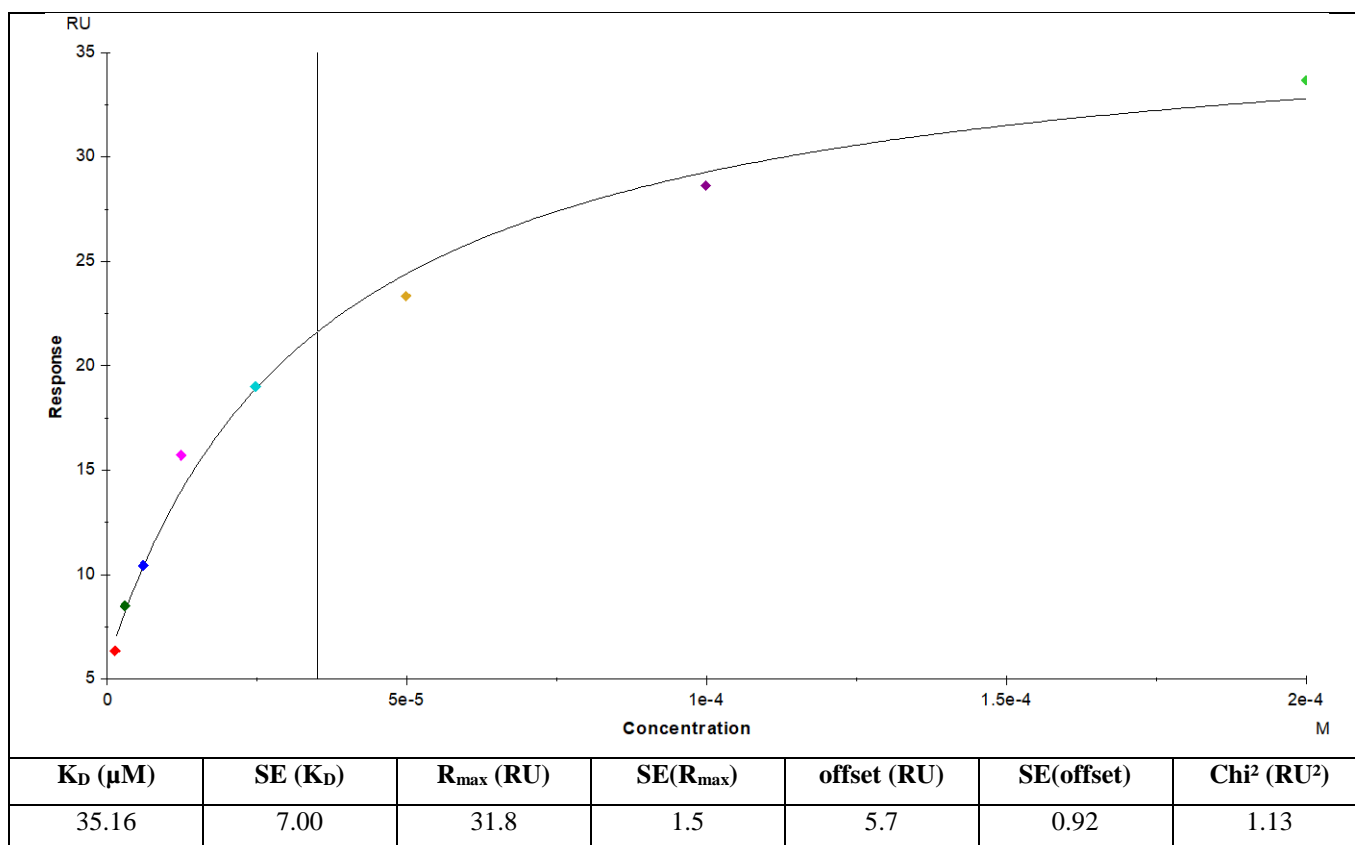
Spy molecule 15 (Compound S5d)

Blank and reference surface subtracted responses according to the concentration of spy molecule.

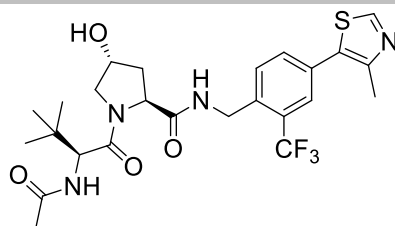
Theoretical $R_{MAX} = 30.0$



Data fitting using the Biacore T200 evaluation software:



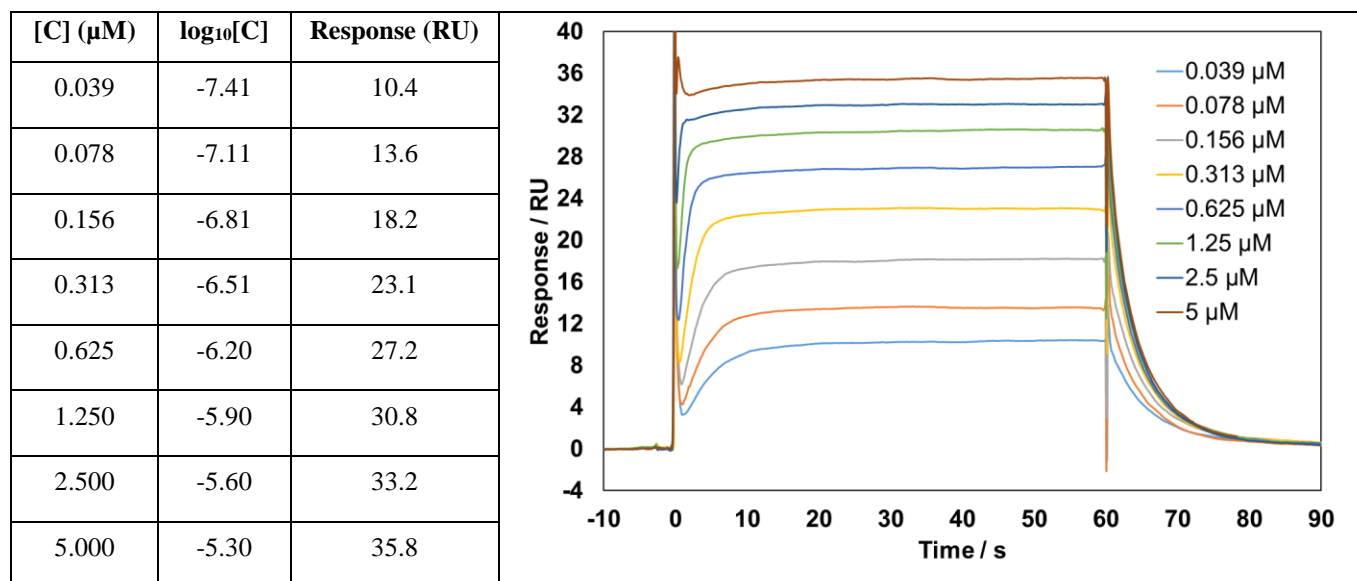
SUPPORTING INFORMATION



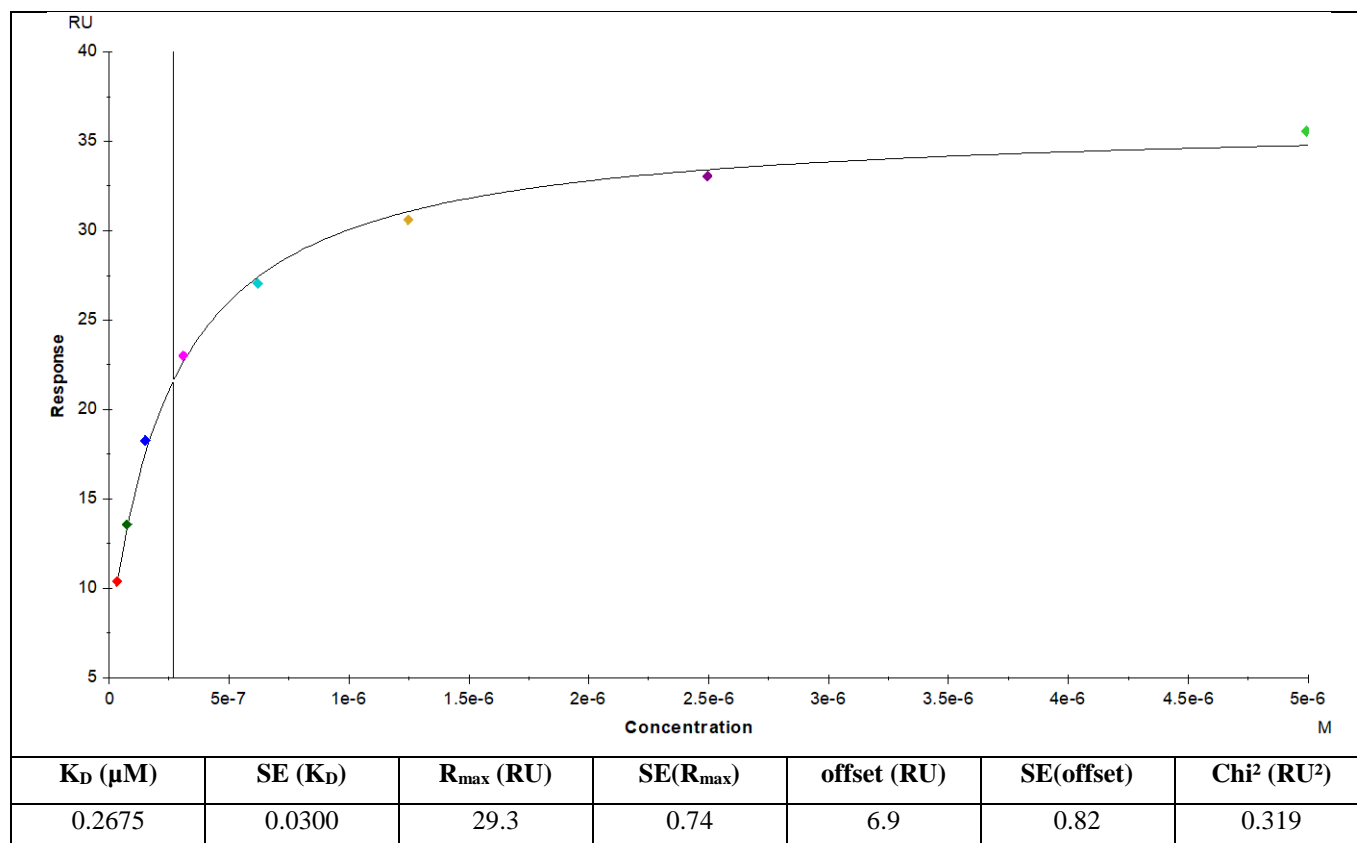
Spy molecule 16 (Compound S6d)

Blank and reference surface subtracted responses according to the concentration of spy molecule.

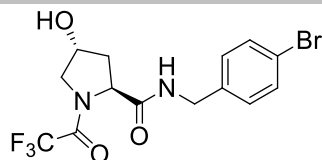
Theoretical $R_{MAX} = 33.3$



Data fitting using the Biacore T200 evaluation software:



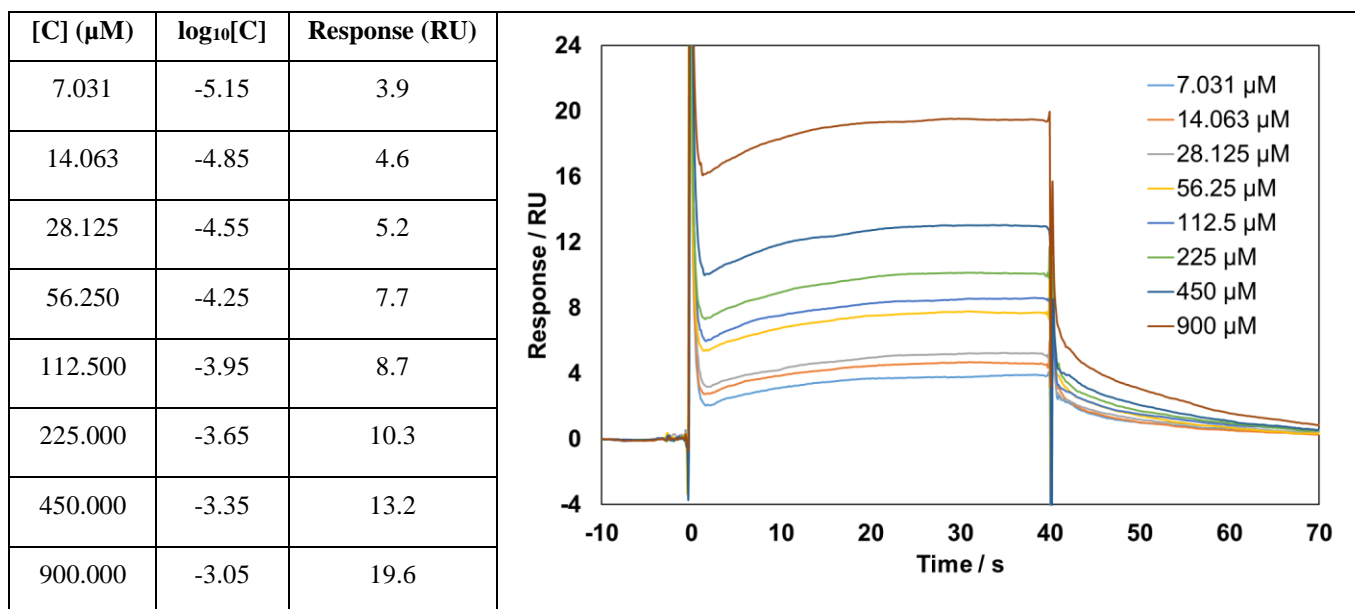
SUPPORTING INFORMATION



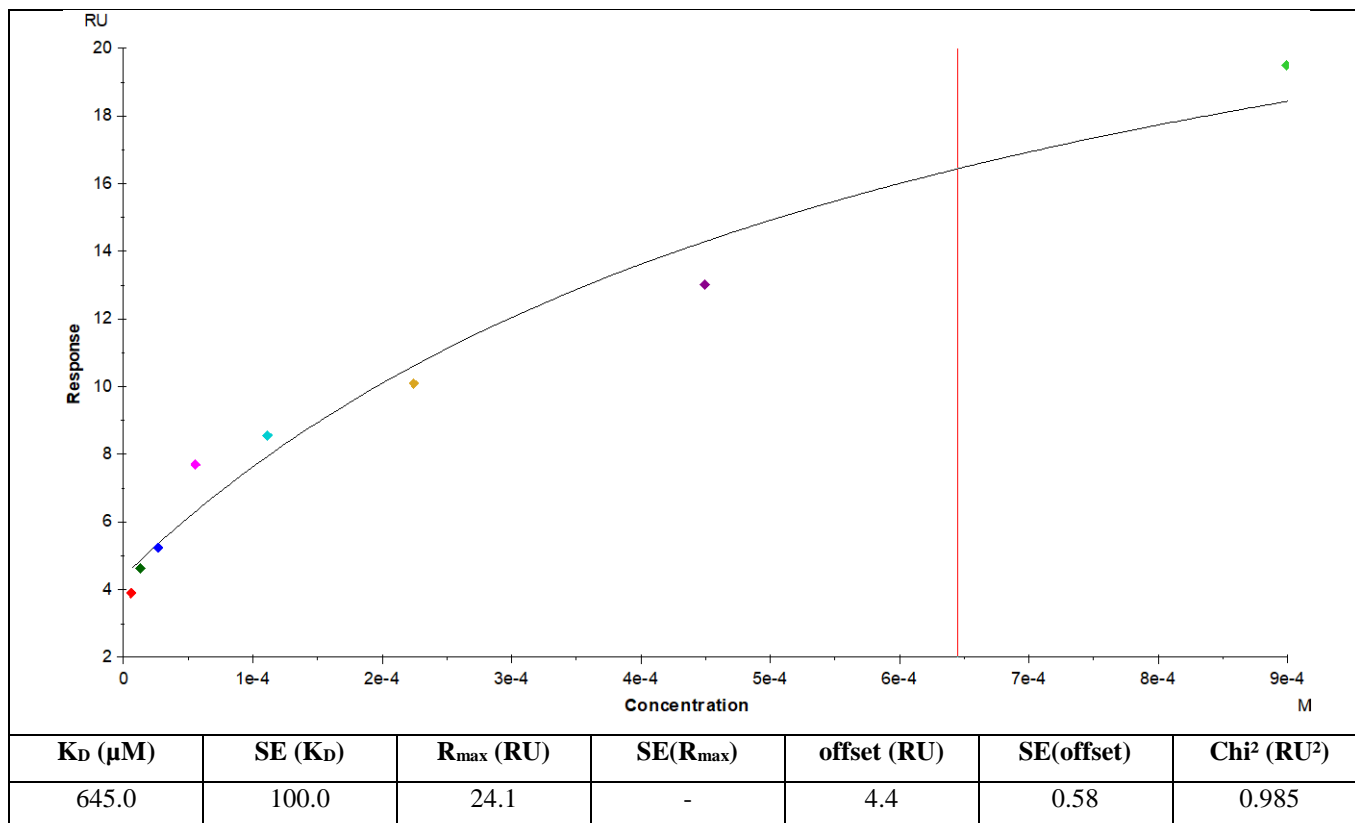
Spy molecule 17 (Compound S10a)

Blank and reference surface subtracted responses according to the concentration of spy molecule.

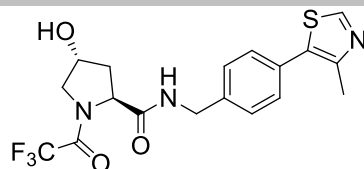
Theoretical $R_{MAX} = 24.1$



Data fitting using the Biacore T200 evaluation software. As responses were lower than the theoretical R_{MAX} , fitting was performed with a fixed R_{MAX} .



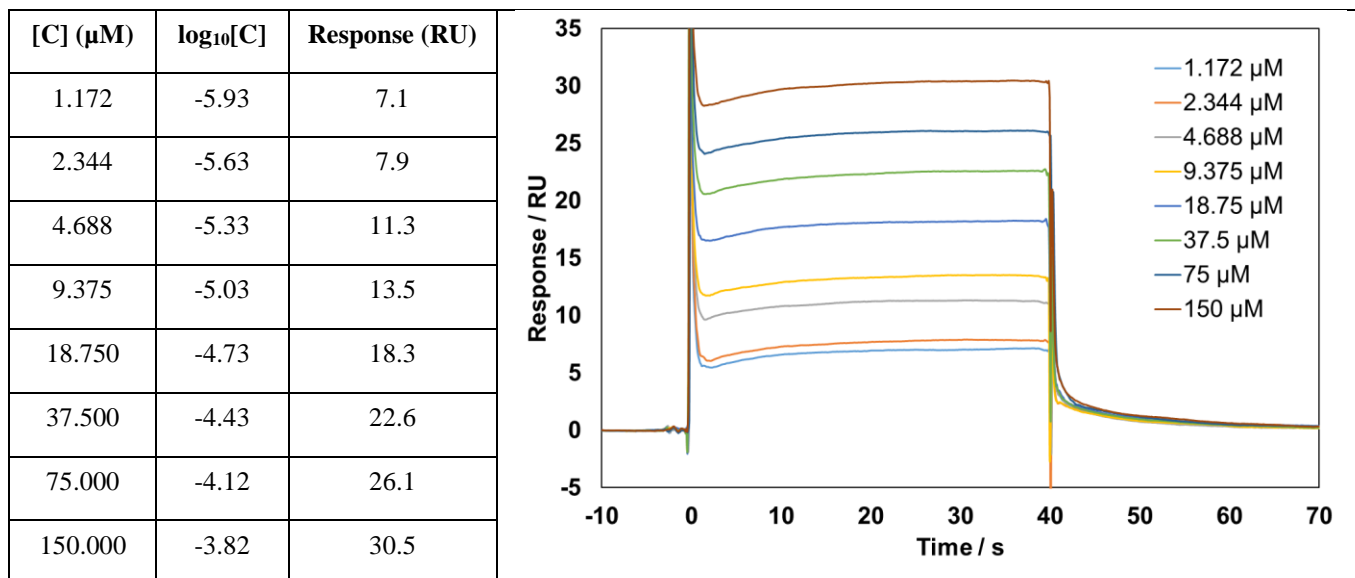
SUPPORTING INFORMATION



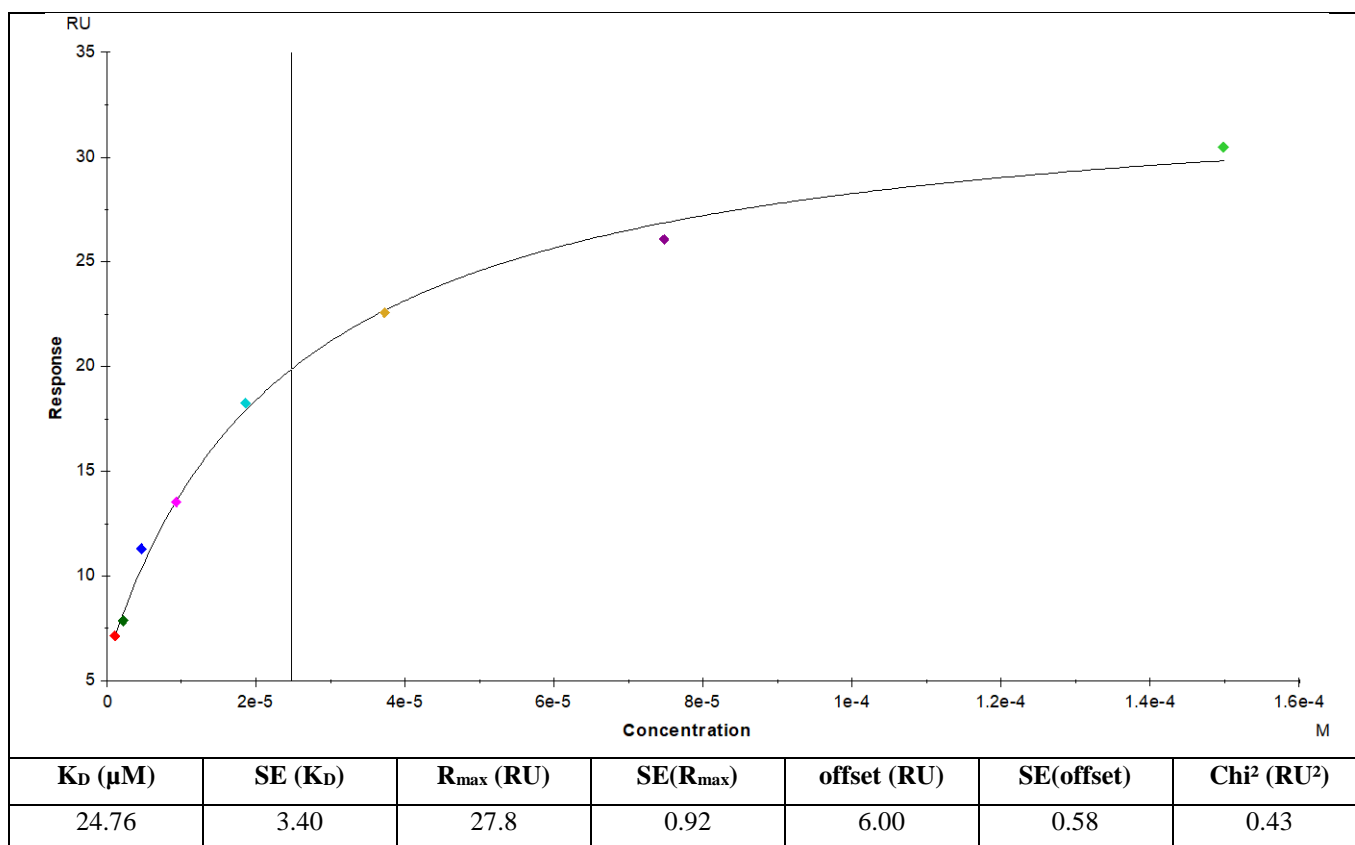
Spy molecule 18 (Compound S10b)

Blank and reference surface subtracted responses according to the concentration of spy molecule.

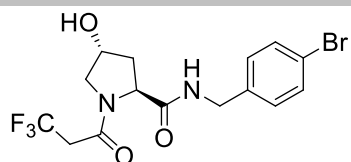
Theoretical $R_{MAX} = 24.7$



Data fitting using the Biacore T200 evaluation software:



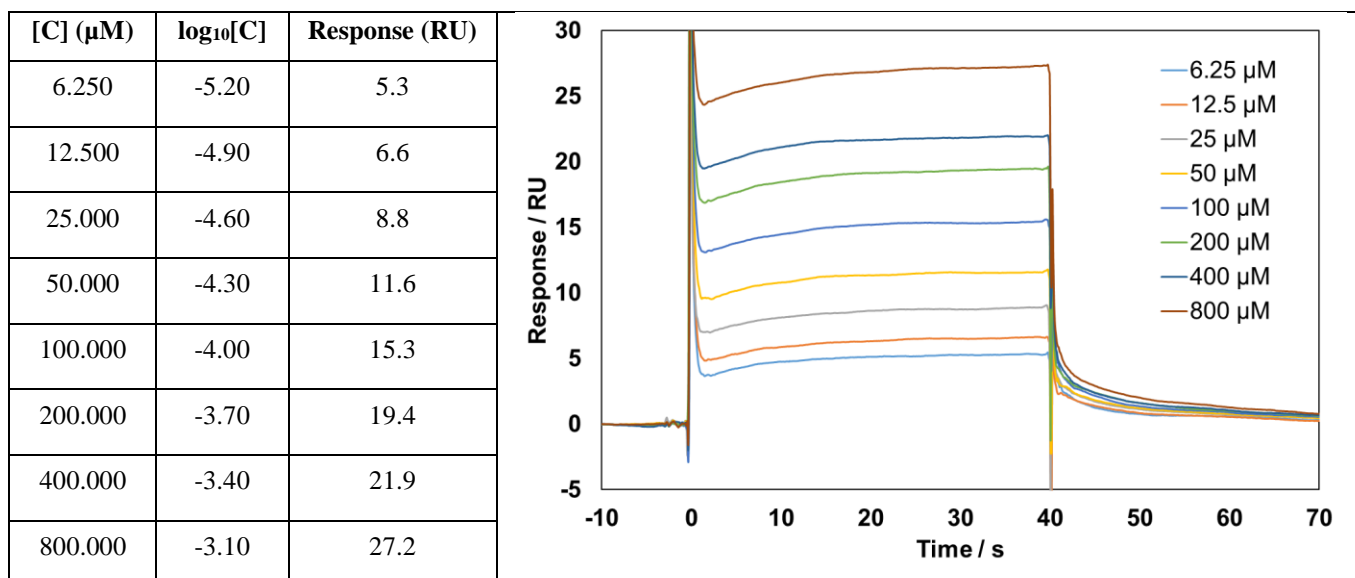
SUPPORTING INFORMATION



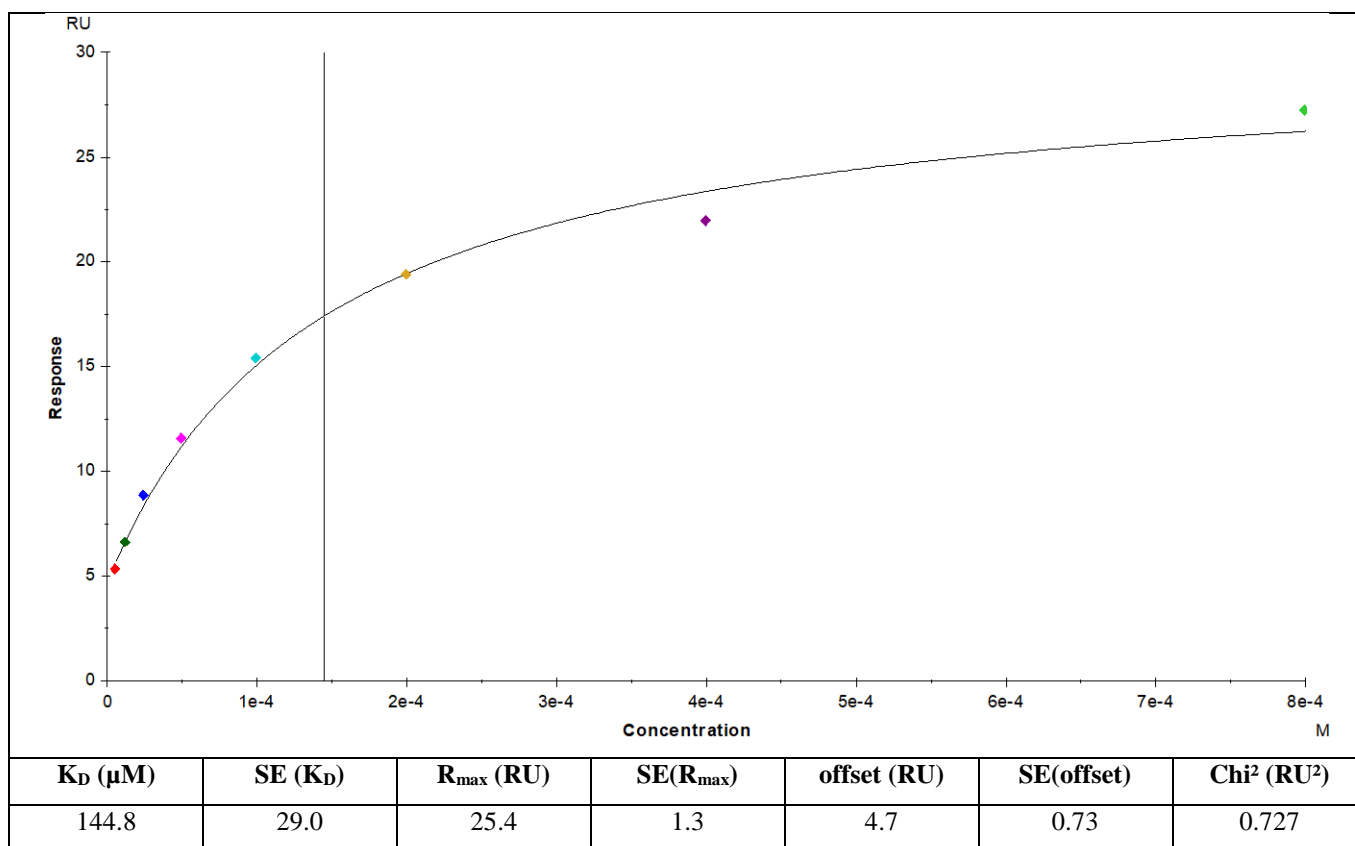
Spy molecule 19 (Compound S11a)

Blank and reference surface subtracted responses according to the concentration of spy molecule.

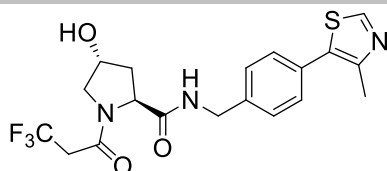
Theoretical $R_{MAX} = 24.7$



Data fitting using the Biacore T200 evaluation software:



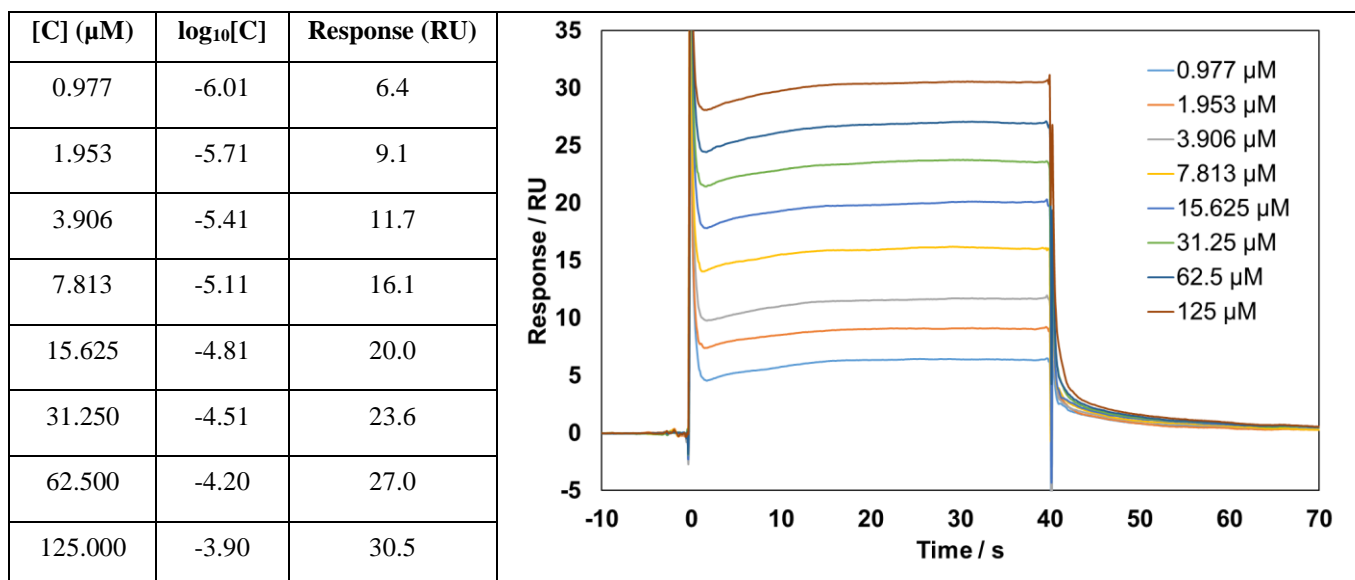
SUPPORTING INFORMATION



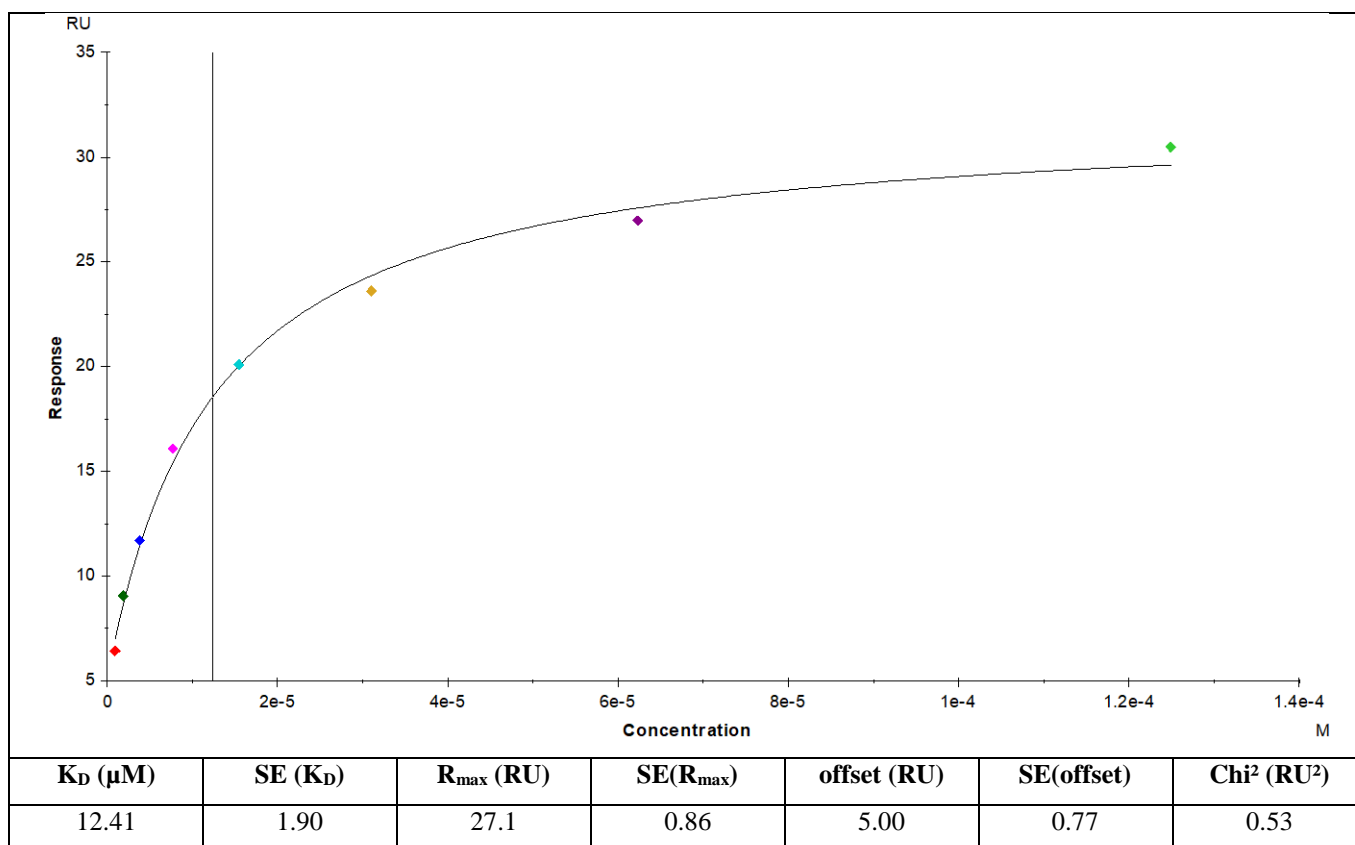
Spy molecule 20 (Compound S11b)

Blank and reference surface subtracted responses according to the concentration of spy molecule.

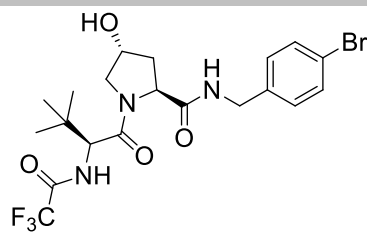
Theoretical $R_{MAX} = 25.4$



Data fitting using the Biacore T200 evaluation software:



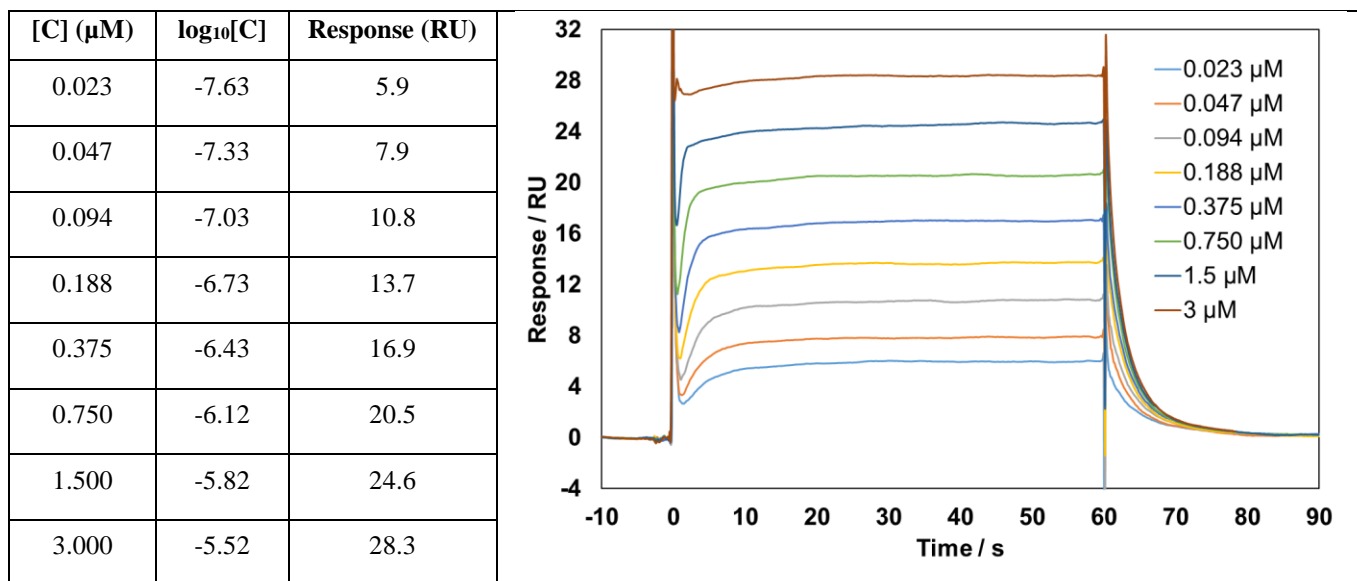
SUPPORTING INFORMATION



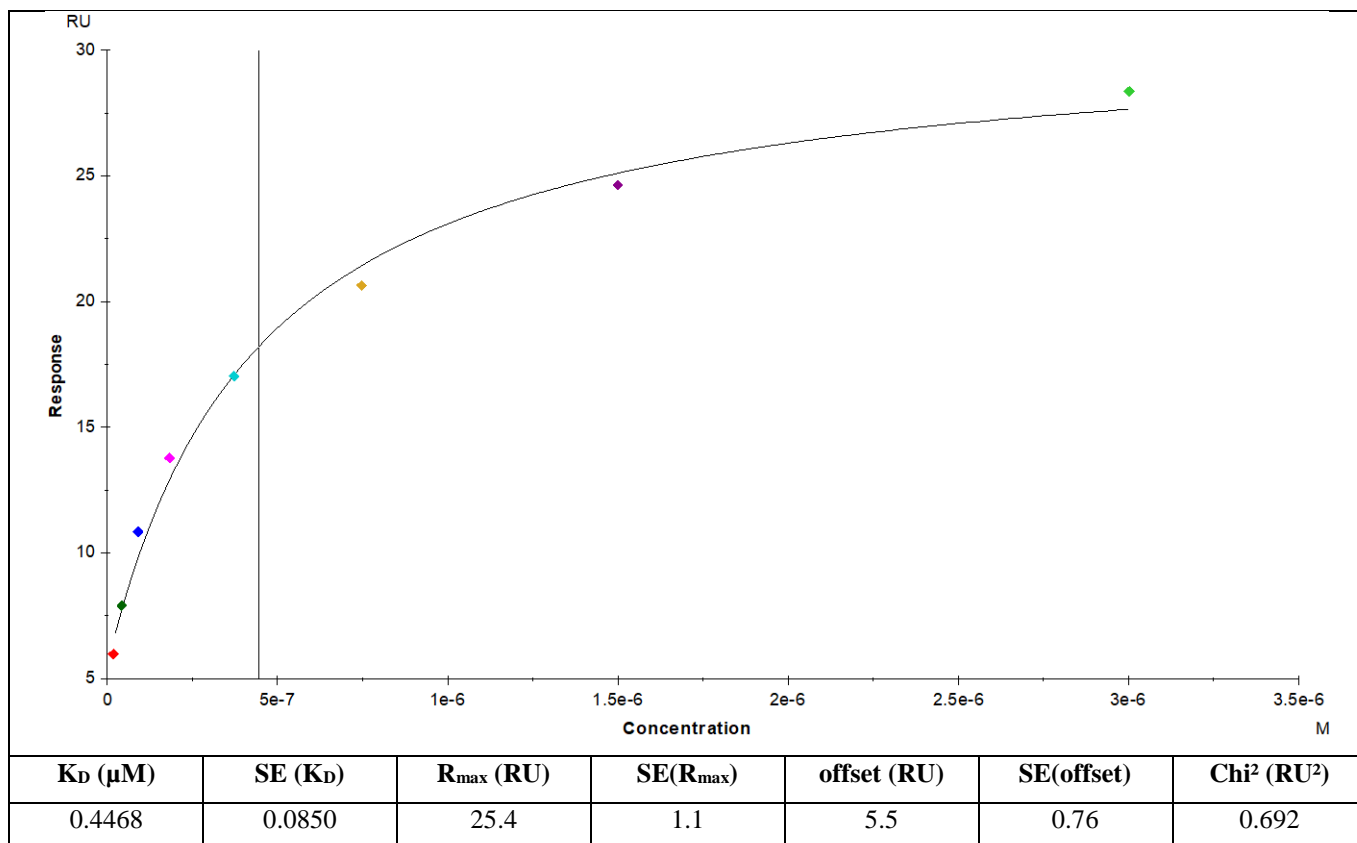
Spy molecule 21 (Compound S12a)

Blank and reference surface subtracted responses according to the concentration of spy molecule.

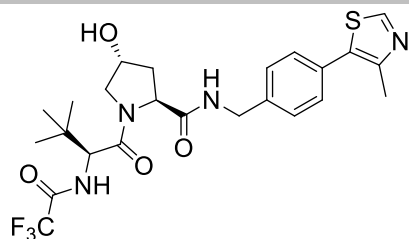
Theoretical $R_{MAX} = 30.5$



Data fitting using the Biacore T200 evaluation software:



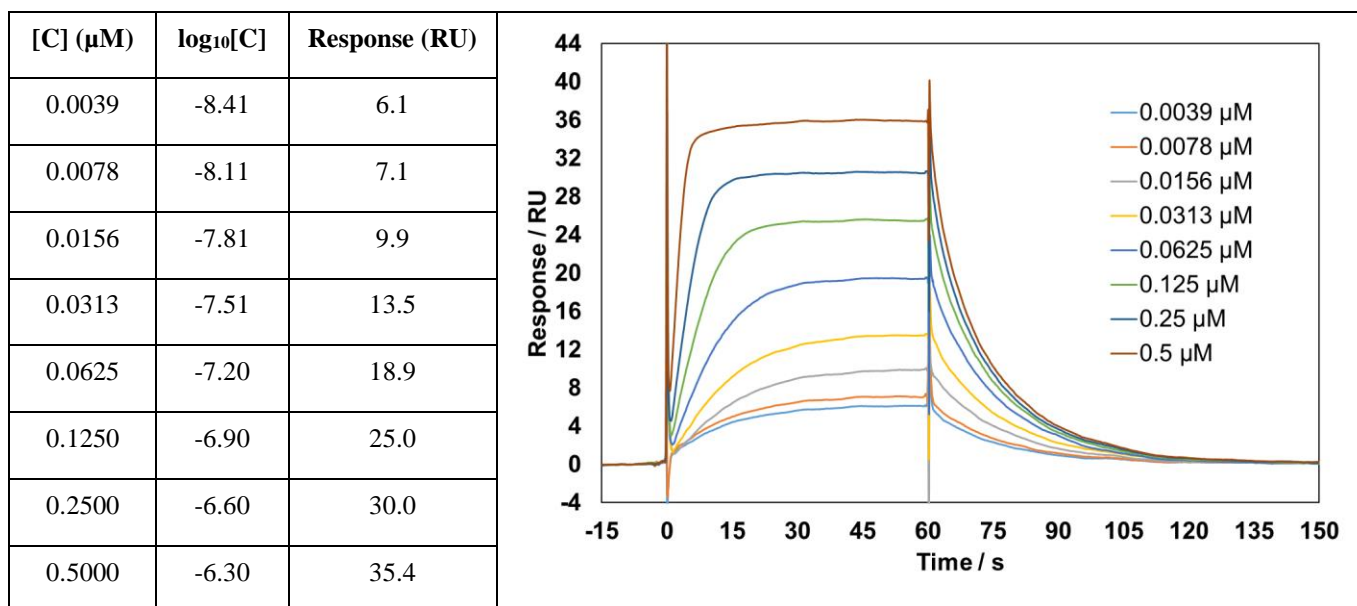
SUPPORTING INFORMATION



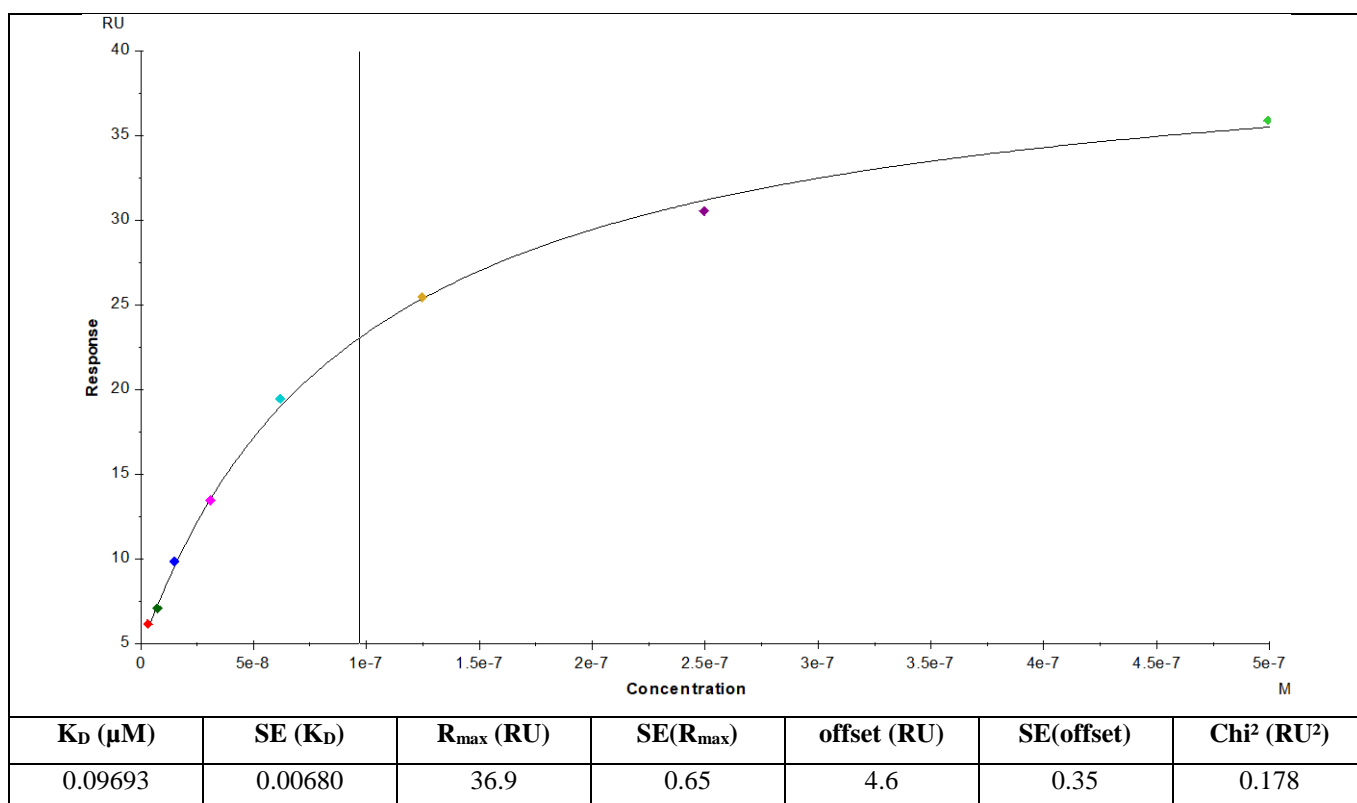
Spy molecule 22 (Compound S12b)

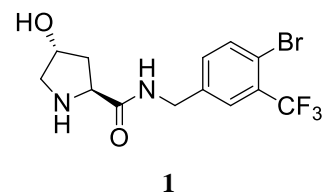
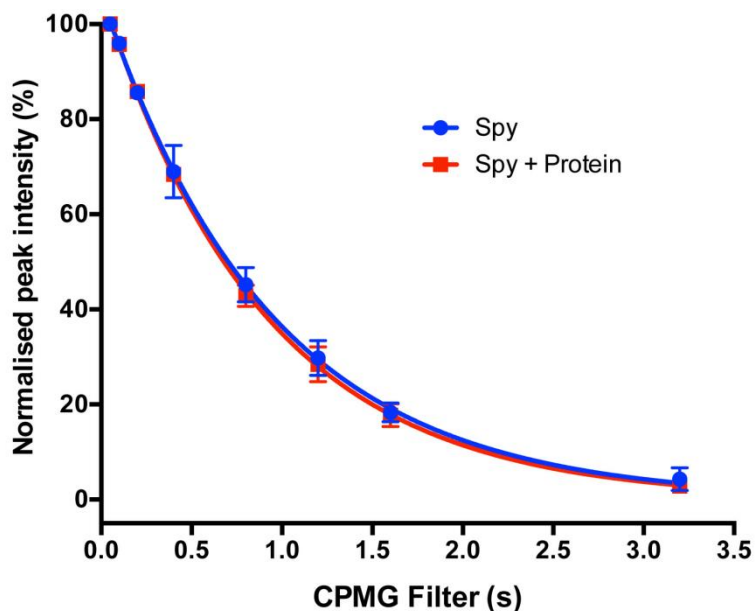
Blank and reference surface subtracted responses according to the concentration of spy molecule.

Theoretical $R_{MAX} = 36.6$

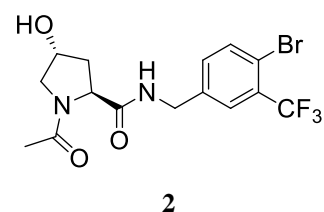
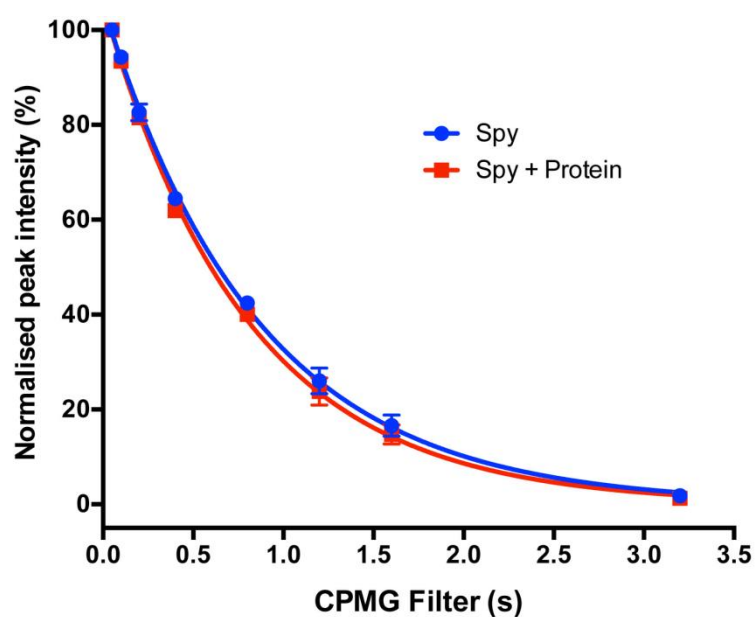


Data fitting using the Biacore T200 evaluation software:



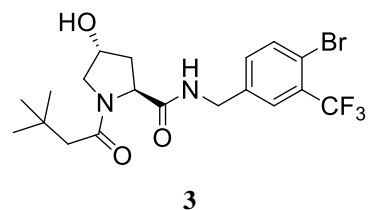
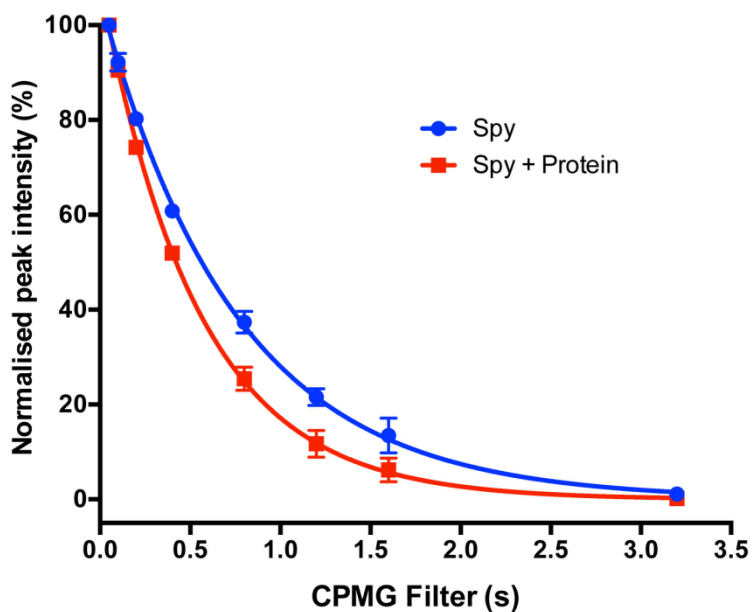
8. ^{19}F CPMG signal intensity versus CPMG filter8.1 Spy molecules at 100 μM in absence or in presence of VBC 1 μM 

Spy molecule 100 μM	
R_2 (s^{-1})	1.069 ± 0.028
R-square	0.995
Spy molecule 100 μM + VBC 1 μM	
R_2 (s^{-1})	1.115 ± 0.020
R-square	0.998
Contrast	
R_2 contrast – C_2 (%)	4.1 ± 3.1
d_{max} (s)	0.916

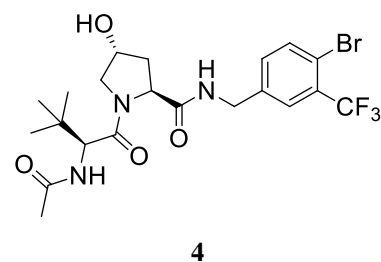
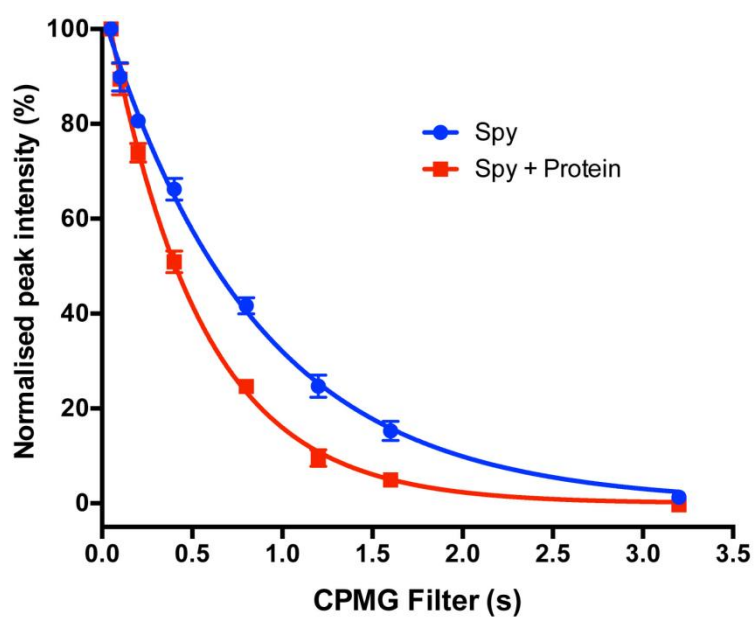


Spy molecule 100 μM	
R_2 (s^{-1})	1.170 ± 0.019
R-square	0.998
Spy molecule 100 μM + VBC 1 μM	
R_2 (s^{-1})	1.251 ± 0.021
R-square	0.998
Contrast	
R_2 contrast – C_2 (%)	6.5 ± 2.2
d_{max} (s)	0.826

SUPPORTING INFORMATION

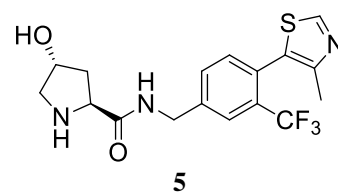
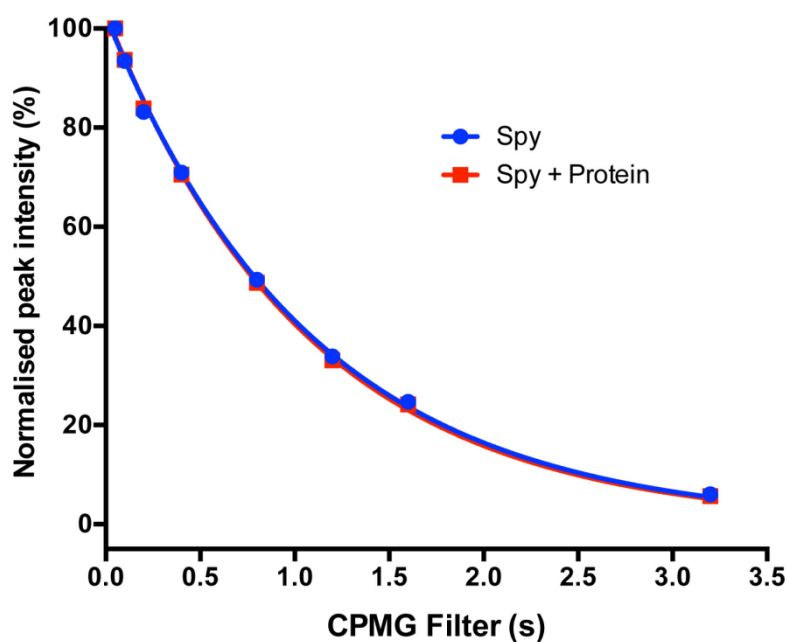


Spy molecule 100 μM	
R_2 (s^{-1})	1.326 ± 0.025
R-square	0.998
Spy molecule 100 μM + VBC 1 μM	
R_2 (s^{-1})	1.847 ± 0.033
R-square	0.998
Contrast	
R_2 contrast – C_2 (%)	28.2 ± 2.3
d_{max} (s)	0.636

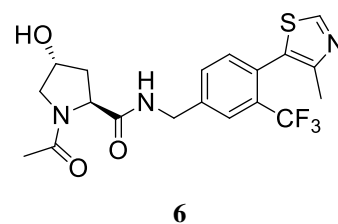
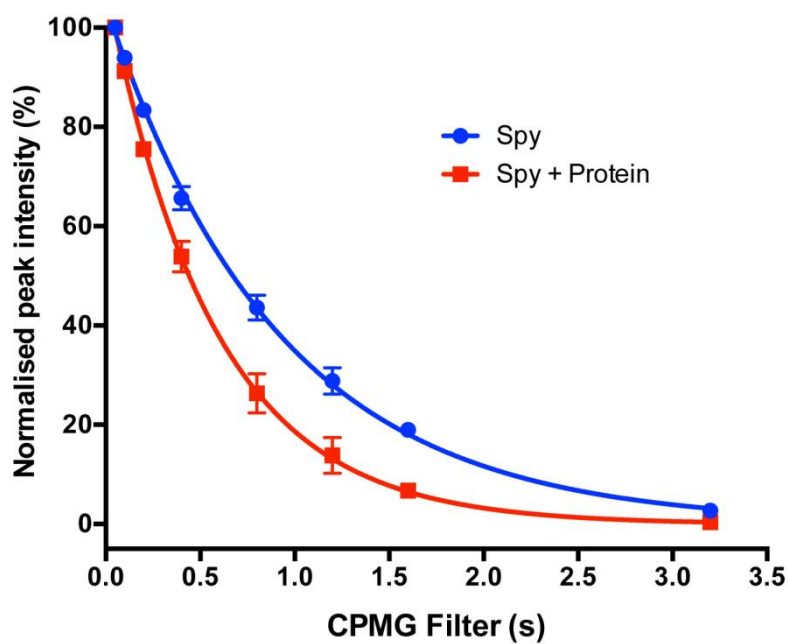


Spy molecule 100 μM	
R_2 (s^{-1})	1.175 ± 0.027
R-square	0.996
Spy molecule 100 μM + VBC 1 μM	
R_2 (s^{-1})	1.924 ± 0.037
R-square	0.998
Contrast	
R_2 contrast – C_2 (%)	38.9 ± 2.5
d_{max} (s)	0.658

SUPPORTING INFORMATION

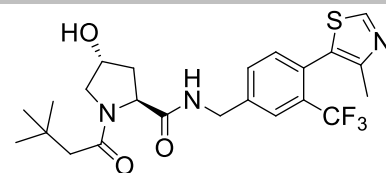
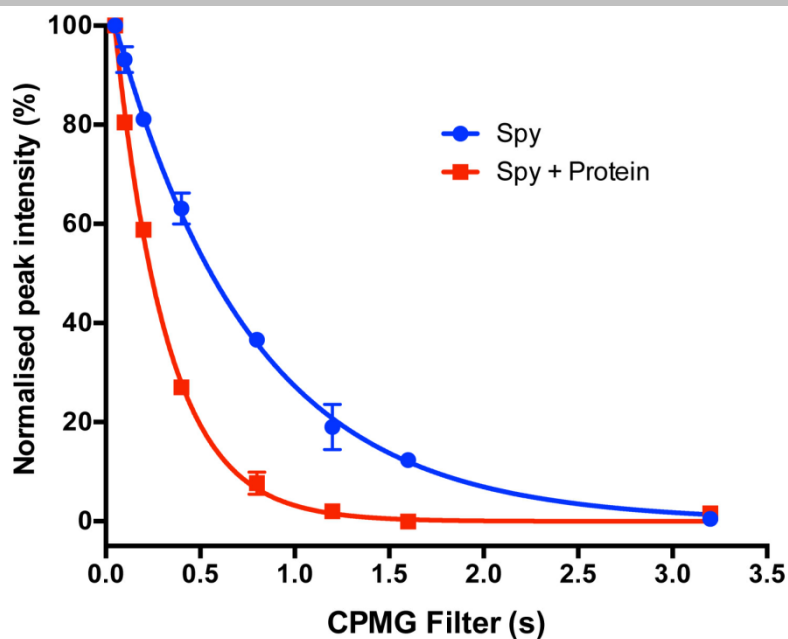


Spy molecule 100 μM	
R_2 (s^{-1})	0.915 ± 0.012
R-square	0.999
Spy molecule 100 μM + VBC 1 μM	
R_2 (s^{-1})	0.936 ± 0.011
R-square	0.999
Contrast	
R_2 contrast - C_2 (%)	2.2 ± 1.8
d_{max} (s)	1.080



Spy molecule 100 μM	
R_2 (s^{-1})	1.096 ± 0.020
R-square	0.998
Spy molecule 100 μM + VBC 1 μM	
R_2 (s^{-1})	1.760 ± 0.039
R-square	0.997
Contrast	
R_2 contrast - C_2 (%)	37.7 ± 2.6
d_{max} (s)	0.713

SUPPORTING INFORMATION



7

Spy molecule 100 μM

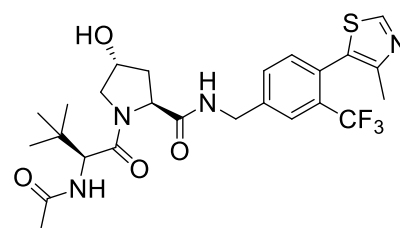
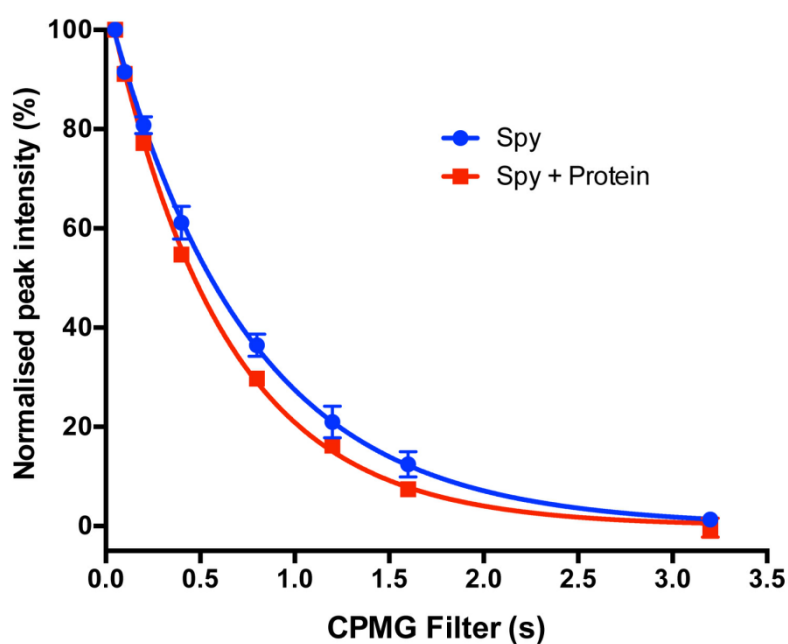
R_2 (s^{-1})	1.368 ± 0.033
R-square	0.998

Spy molecule 100 μM + VBC 1 μM

R_2 (s^{-1})	3.614 ± 0.088
R-square	0.999

Contrast

R_2 contrast - C_2 (%)	62.1 ± 3.0
d_{max} (s)	0.433



8

Spy molecule 100 μM

R_2 (s^{-1})	1.349 ± 0.027
R-square	0.997

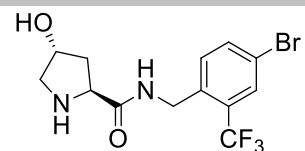
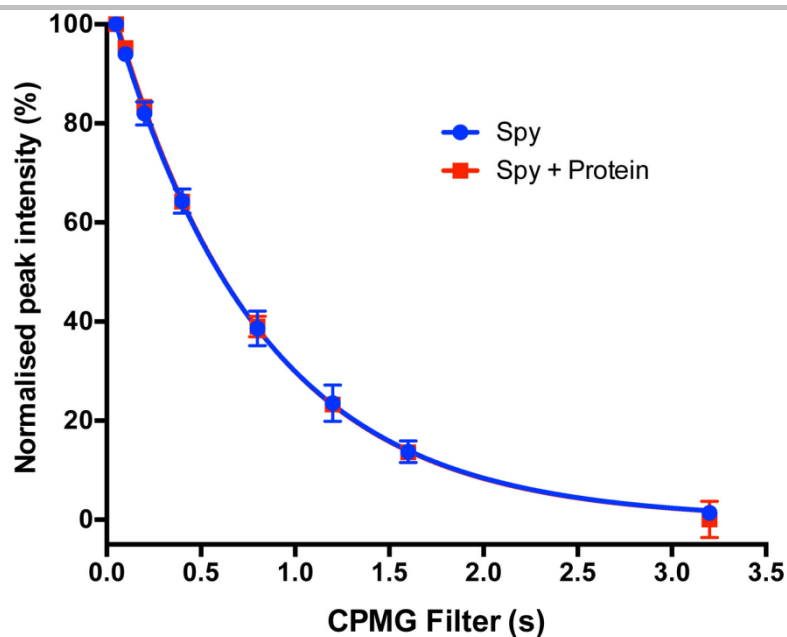
Spy molecule 100 μM + VBC 1 μM

R_2 (s^{-1})	1.639 ± 0.022
R-square	0.999

Contrast

R_2 contrast - C_2 (%)	17.7 ± 2.2
d_{max} (s)	0.671

SUPPORTING INFORMATION



9

Spy molecule 100 μM

R_2 (s^{-1}) 1.268 ± 0.026

R-square 0.997

Spy molecule 100 μM + VBC 1 μM

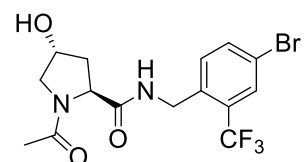
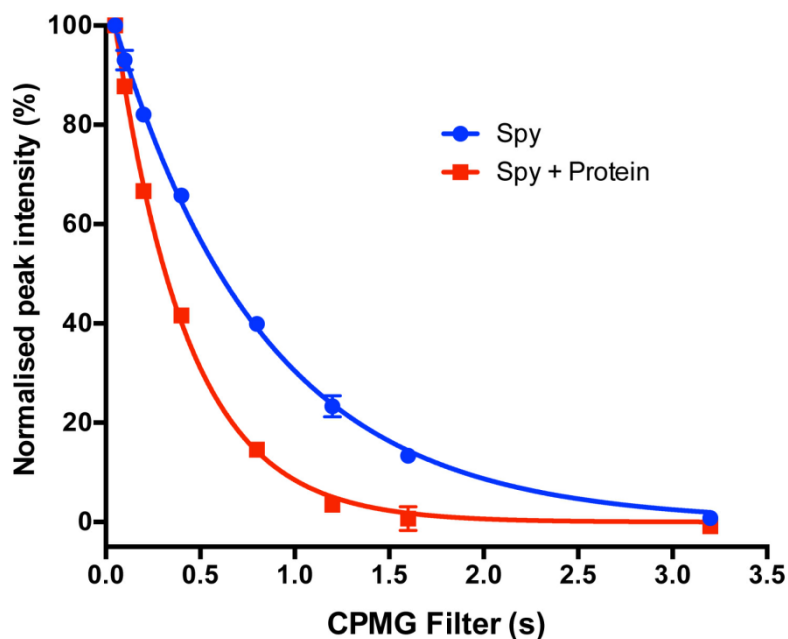
R_2 (s^{-1}) 1.279 ± 0.022

R-square 0.998

Contrast

R_2 contrast - C_2 (%) 0.9 ± 2.7

d_{max} (s) 0.785



10

Spy molecule 100 μM

R_2 (s^{-1}) 1.249 ± 0.018

R-square 0.999

Spy molecule 100 μM + VBC 1 μM

R_2 (s^{-1}) 2.596 ± 0.046

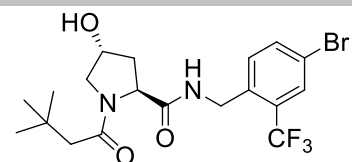
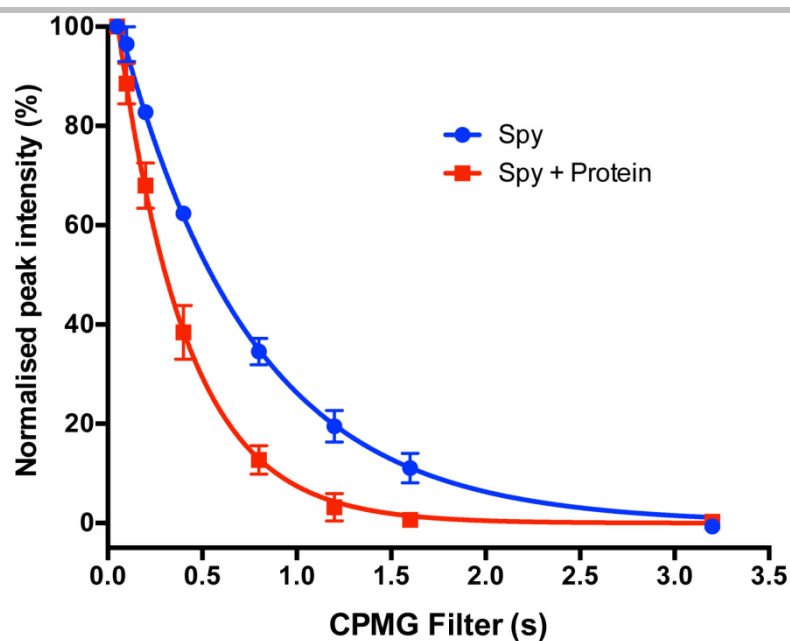
R-square 0.999

Contrast

R_2 contrast - C_2 (%) 51.9 ± 2.1

d_{max} (s) 0.543

SUPPORTING INFORMATION



11

Spy molecule 100 μ M

R_2 (s^{-1}) 1.427 ± 0.033

R-square 0.997

Spy molecule 100 μ M + VBC 1 μ M

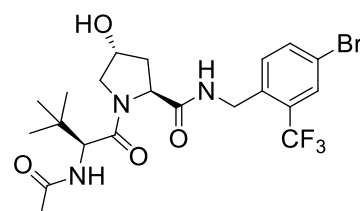
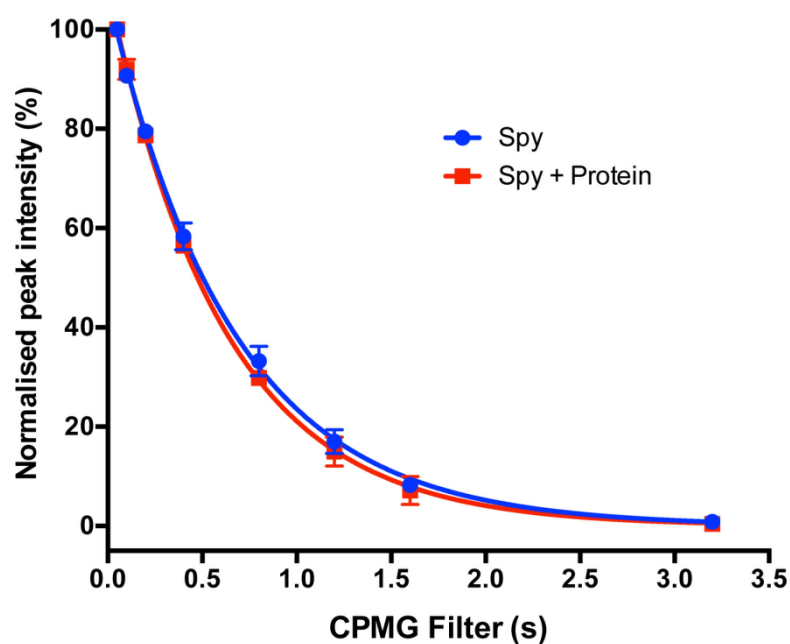
R_2 (s^{-1}) 2.740 ± 0.096

R-square 0.995

Contrast

R_2 contrast - C_2 (%) 47.9 ± 4.1

d_{\max} (s) 0.497



12

Spy molecule 100 μ M

R_2 (s^{-1}) 1.512 ± 0.029

R-square 0.998

Spy molecule 100 μ M + VBC 1 μ M

R_2 (s^{-1}) 1.637 ± 0.028

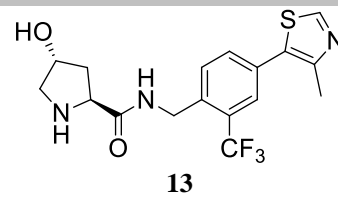
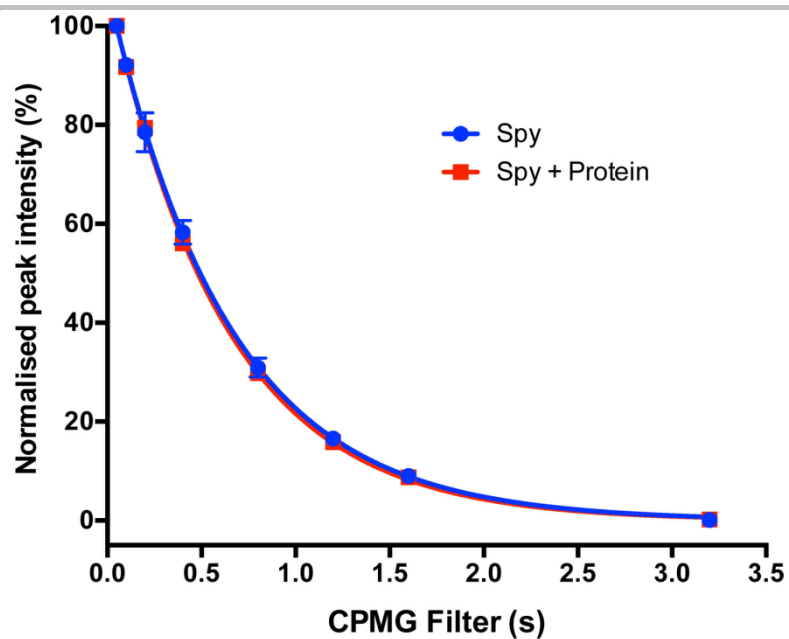
R-square 0.998

Contrast

R_2 contrast - C_2 (%) 7.6 ± 2.4

d_{\max} (s) 0.635

SUPPORTING INFORMATION



Spy molecule 100 μM

$$R_2 \text{ (s}^{-1}\text{)} = 1.558 \pm 0.028$$

$$\text{R-square} = 0.998$$

Spy molecule 100 μM + VBC 1 μM

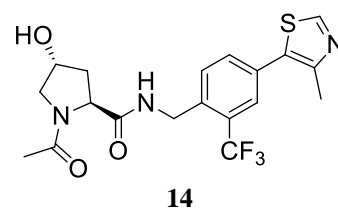
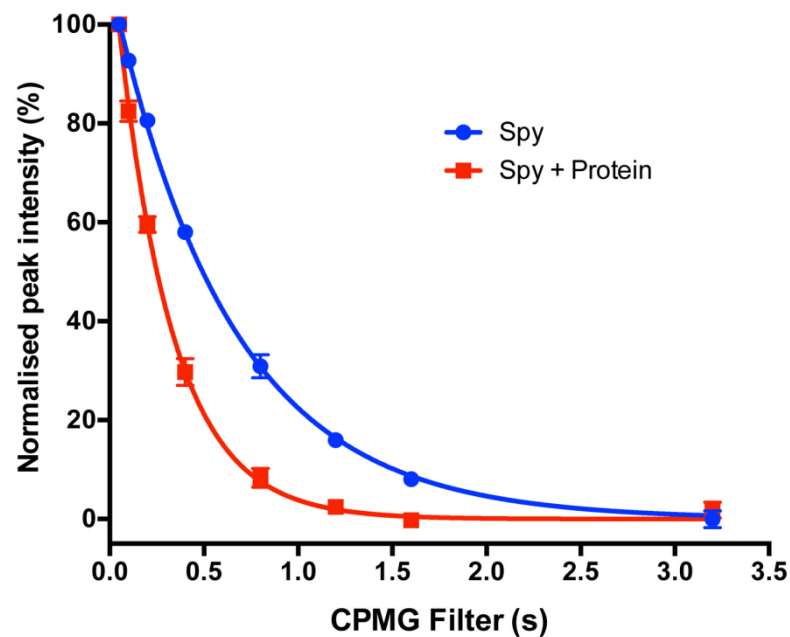
$$R_2 \text{ (s}^{-1}\text{)} = 1.609 \pm 0.017$$

$$\text{R-square} = 0.999$$

Contrast

$$R_2 \text{ contrast} - C_2 \text{ (\%)} = 3.2 \pm 2.0$$

$$d_{\text{max}} \text{ (s)} = 0.632$$



Spy molecule 100 μM

$$R_2 \text{ (s}^{-1}\text{)} = 1.579 \pm 0.022$$

$$\text{R-square} = 0.999$$

Spy molecule 100 μM + VBC 1 μM

$$R_2 \text{ (s}^{-1}\text{)} = 3.425 \pm 0.073$$

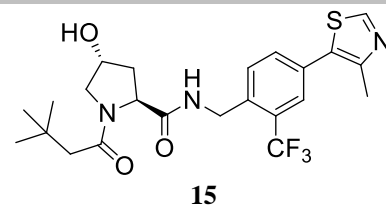
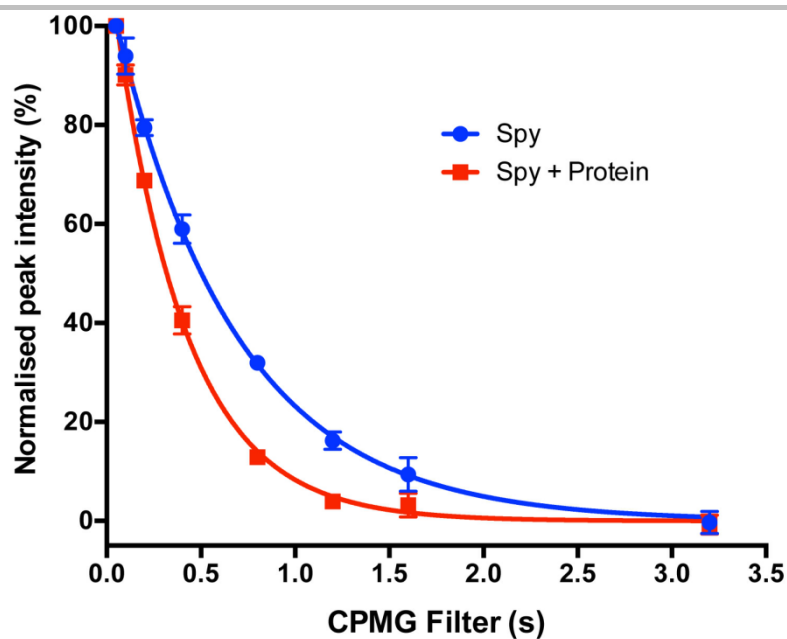
$$\text{R-square} = 0.998$$

Contrast

$$R_2 \text{ contrast} - C_2 \text{ (\%)} = 53.9 \pm 2.5$$

$$d_{\text{max}} \text{ (s)} = 0.419$$

SUPPORTING INFORMATION



Spy molecule 100 μ M

$$R_2 \text{ (s}^{-1}\text{)} = 1.545 \pm 0.035$$

$$\text{R-square} = 0.997$$

Spy molecule 100 μ M + VBC 1 μ M

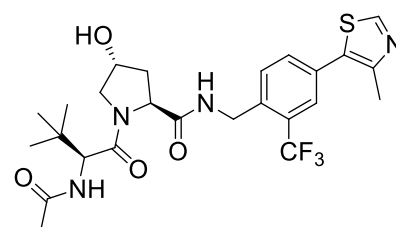
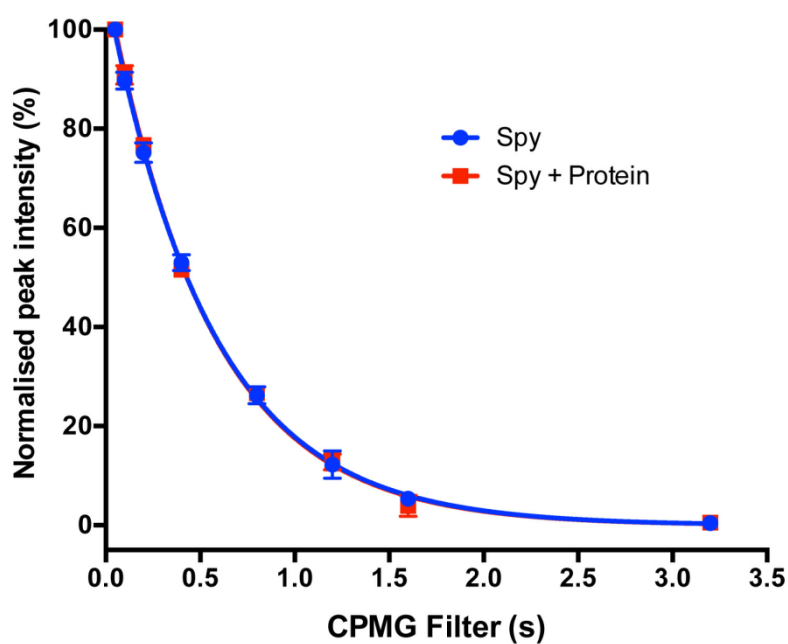
$$R_2 \text{ (s}^{-1}\text{)} = 2.634 \pm 0.058$$

$$\text{R-square} = 0.998$$

Contrast

$$R_2 \text{ contrast} - C_2 \text{ (\%)} = 41.3 \pm 2.7$$

$$d_{\text{max}} \text{ (s)} = 0.490$$



Spy molecule 100 μ M

$$R_2 \text{ (s}^{-1}\text{)} = 1.805 \pm 0.032$$

$$\text{R-square} = 0.998$$

Spy molecule 100 μ M + VBC 1 μ M

$$R_2 \text{ (s}^{-1}\text{)} = 1.828 \pm 0.031$$

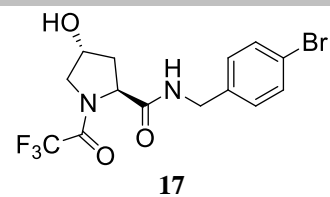
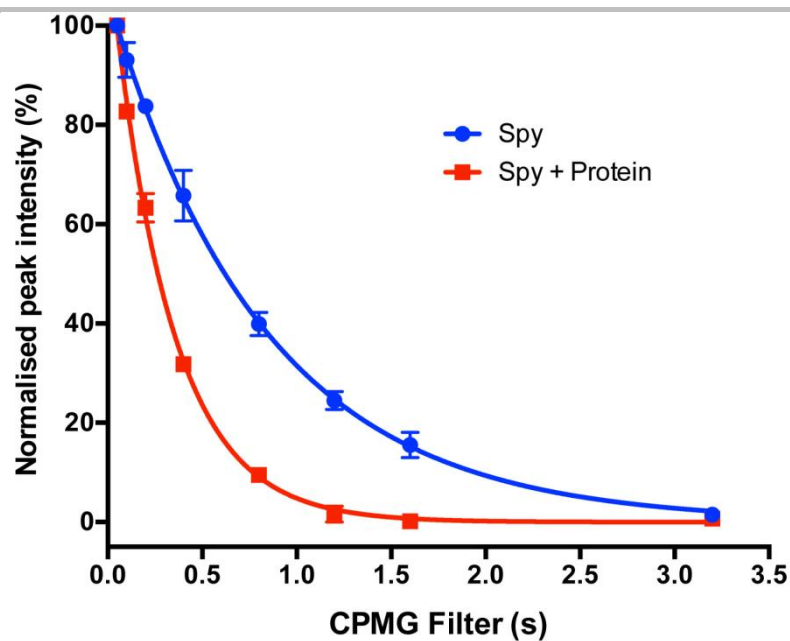
$$\text{R-square} = 0.999$$

Contrast

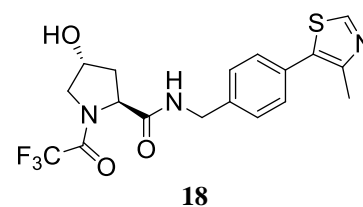
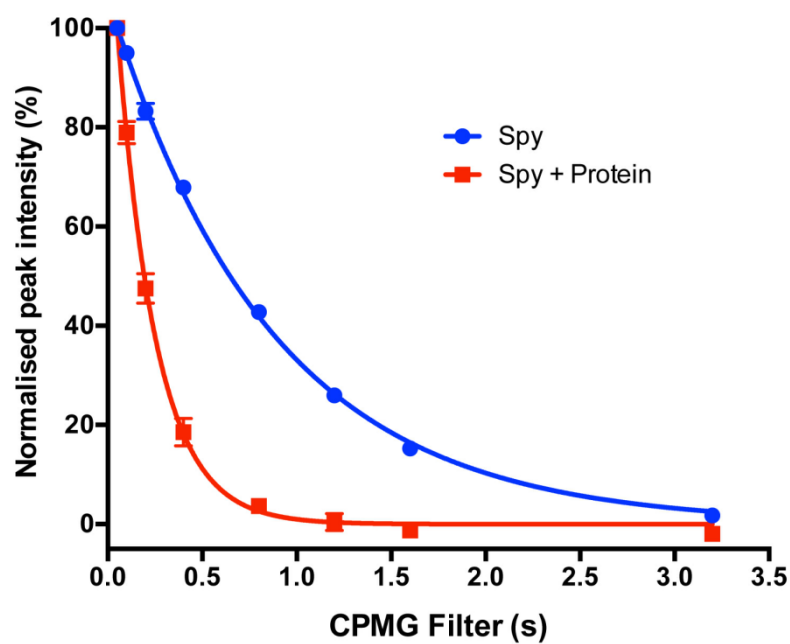
$$R_2 \text{ contrast} - C_2 \text{ (\%)} = 1.3 \pm 2.4$$

$$d_{\text{max}} \text{ (s)} = 0.551$$

SUPPORTING INFORMATION

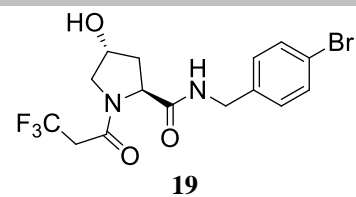
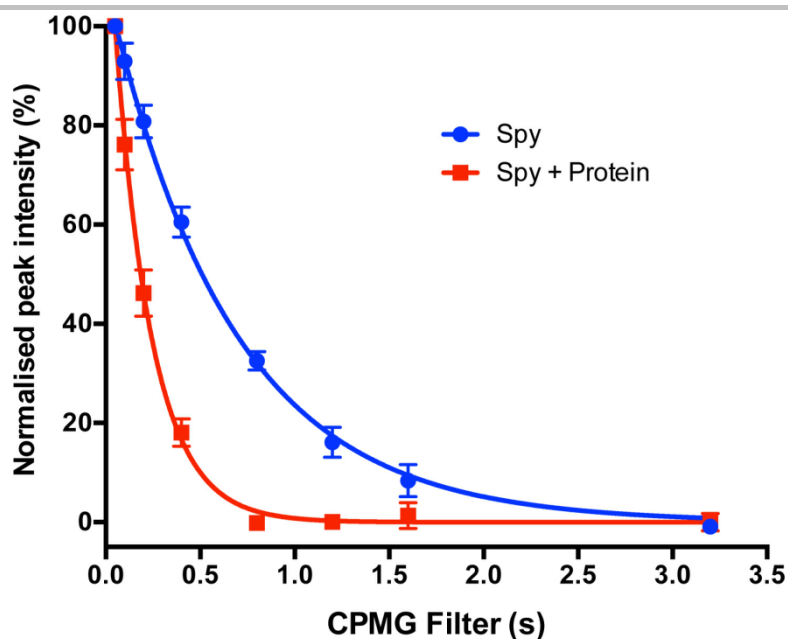


Spy molecule 100 μ M	
R_2 (s^{-1})	1.213 ± 0.028
R-square	0.996
Spy molecule 100 μ M + VBC 1 μ M	
R_2 (s^{-1})	3.189 ± 0.066
R-square	0.998
Contrast	
R_2 contrast - C_2 (%)	62.0 ± 2.6
d_{\max} (s)	0.489

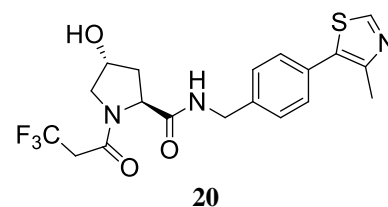
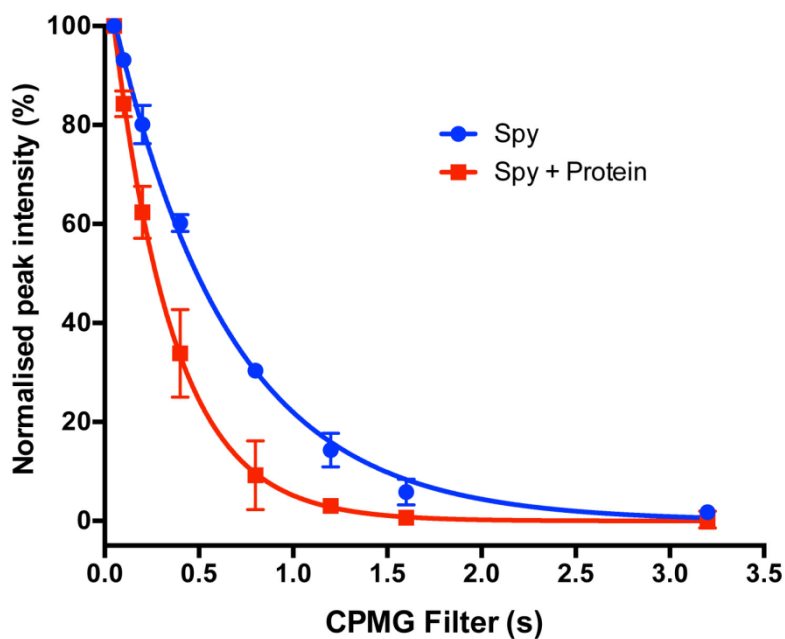


Spy molecule 100 μ M	
R_2 (s^{-1})	1.167 ± 0.014
R-square	0.999
Spy molecule 100 μ M + VBC 1 μ M	
R_2 (s^{-1})	4.858 ± 0.123
R-square	0.998
Contrast	
R_2 contrast - C_2 (%)	76.0 ± 3.2
d_{\max} (s)	0.386

SUPPORTING INFORMATION

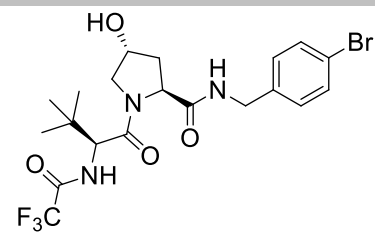
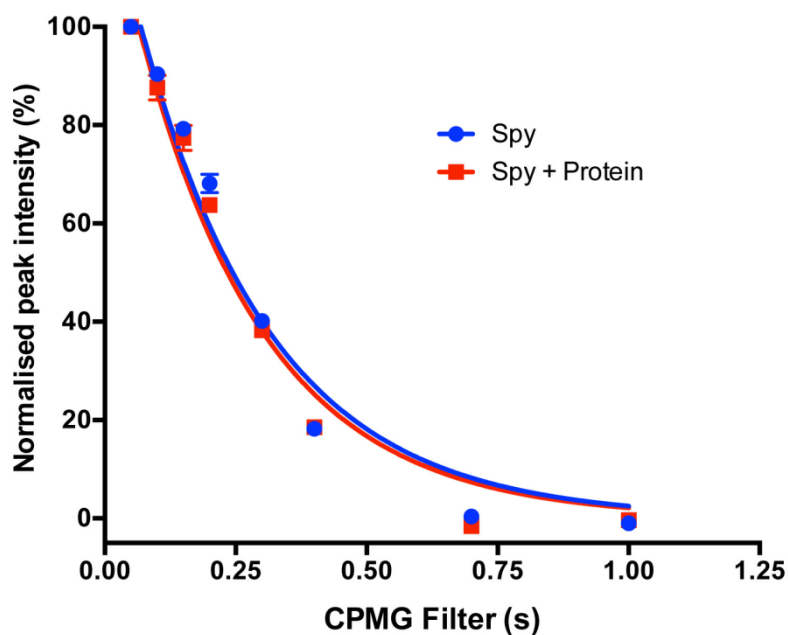


Spy molecule 100 μM	
R_2 (s^{-1})	1.526 ± 0.041
R-square	0.996
Spy molecule 100 μM + VBC 1 μM	
R_2 (s^{-1})	5.096 ± 0.186
R-square	0.995
Contrast	
R_2 contrast - C_2 (%)	70.1 ± 4.5
d_{max} (s)	0.338



Spy molecule 100 μM	
R_2 (s^{-1})	1.609 ± 0.042
R-square	0.996
Spy molecule 100 μM + VBC 1 μM	
R_2 (s^{-1})	3.116 ± 0.153
R-square	0.990
Contrast	
R_2 contrast - C_2 (%)	48.4 ± 5.6
d_{max} (s)	0.439

SUPPORTING INFORMATION

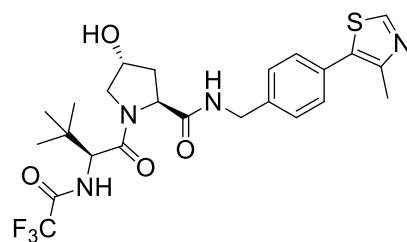
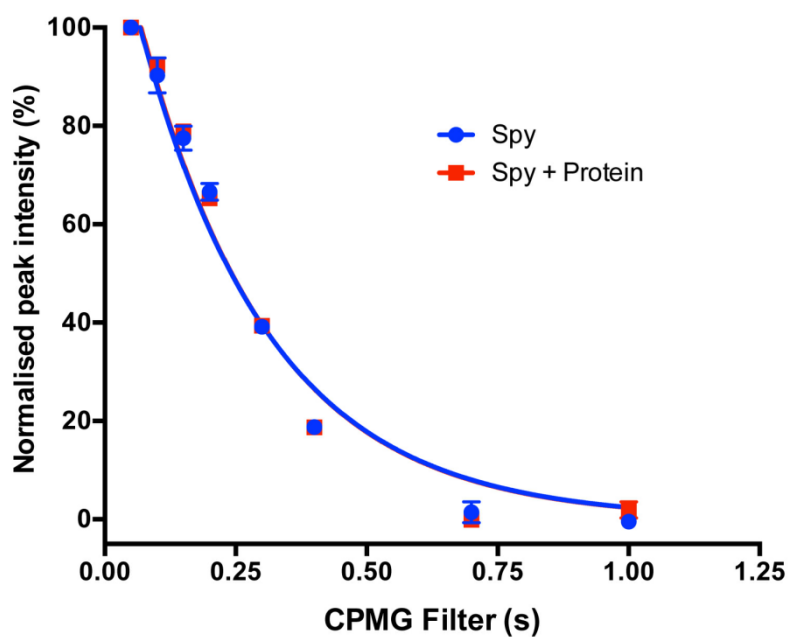


21

Spy molecule 100 μM	
R_2 (s^{-1})	3.959 ± 0.262
R-square	0.970

Spy molecule 100 μM + VBC 1 μM	
R_2 (s^{-1})	4.109 ± 0.247
R-square	0.976

Contrast	
R_2 contrast - C_2 (%)	3.7 ± 8.8
d_{max} (s)	0.248



22

Spy molecule 100 μM	
R_2 (s^{-1})	3.987 ± 0.241
R-square	0.975

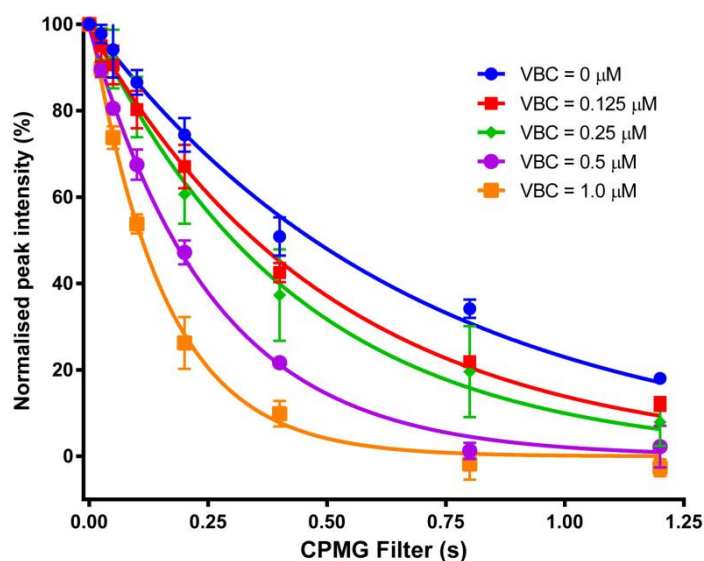
Spy molecule 100 μM + VBC 1 μM	
R_2 (s^{-1})	4.018 ± 0.245
R-square	0.974

Contrast	
R_2 contrast - C_2 (%)	0.8 ± 8.6
d_{max} (s)	0.250

SUPPORTING INFORMATION

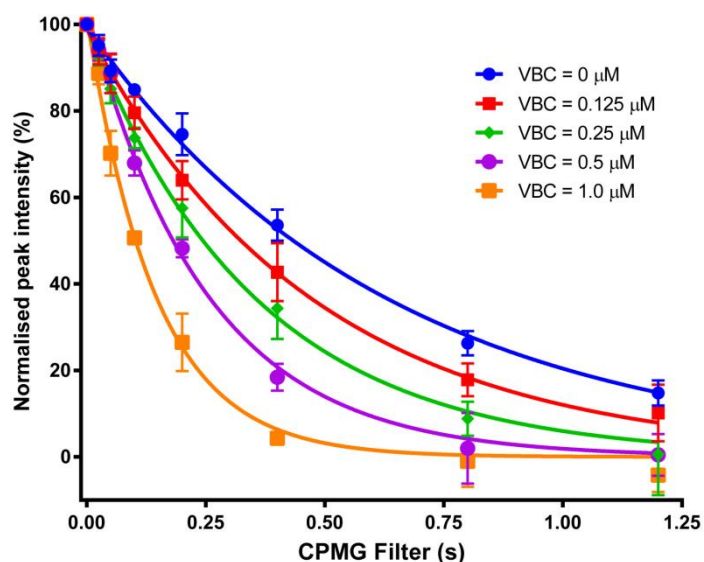
8.2 Spy molecule **19** and VBC at different concentrations

Spy molecule 19 at 50 μM

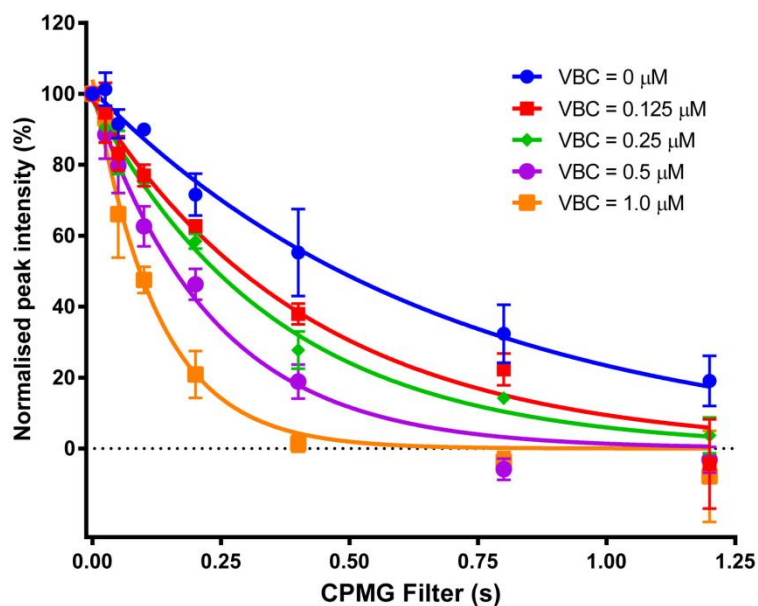


Parameter	[VBC] μM				
	0.000	0.125	0.250	0.500	1.000
R_2 (s^{-1})	1.475 ± 0.059	1.969 ± 0.071	2.316 ± 0.168	3.857 ± 0.121	6.424 ± 0.262
R-square	0.986	0.991	0.967	0.995	0.993
R_2 contrast – C_2 (%)	-	22.5 ± 4.2	34.1 ± 7.9	60.4 ± 3.8	76.2 ± 5.2
d_{max} (s)	-	0.575	0.528	0.398	0.293

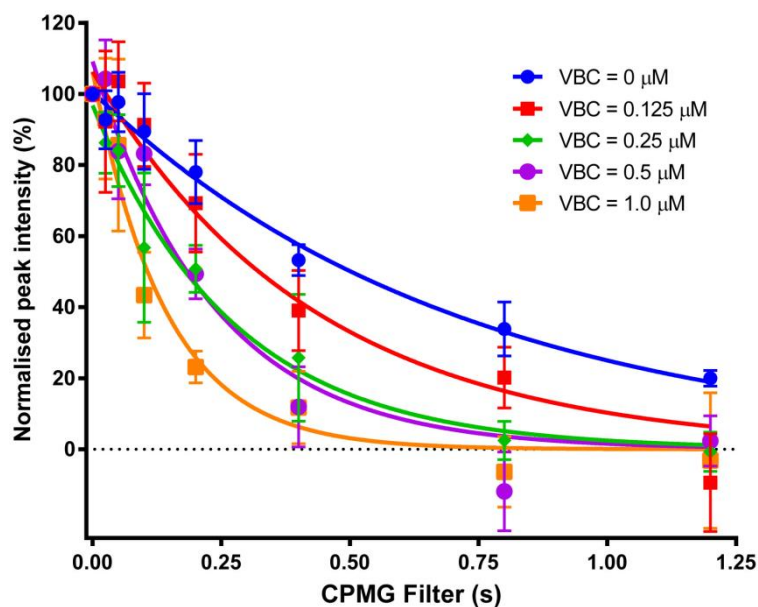
Spy molecule 19 at 25 μM



Parameter	[VBC] μM				
	0.000	0.125	0.250	0.500	1.000
R_2 (s^{-1})	1.582 ± 0.049	2.107 ± 0.095	2.811 ± 0.156	4.008 ± 0.200	6.934 ± 0.348
R-square	0.992	0.986	0.984	0.989	0.989
R_2 contrast – C_2 (%)	-	27.6 ± 5.1	45.7 ± 6.3	61.9 ± 5.9	78.0 ± 6.4
d_{max} (s)	-	0.555	0.475	0.389	0.280

Spy molecule 19 at 10 μM 

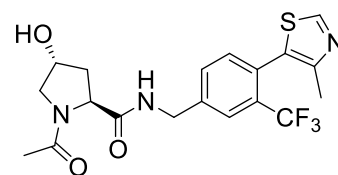
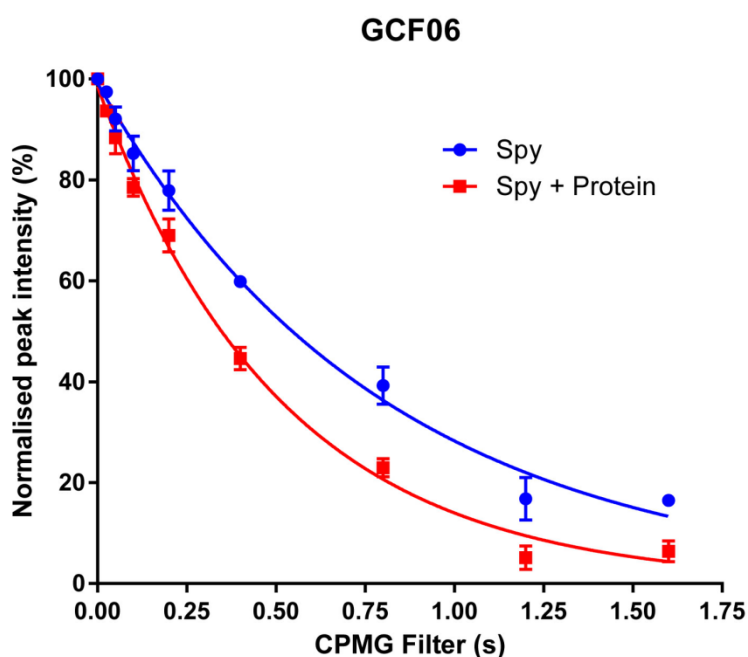
Parameter	[VBC] μM				
	0.000	0.125	0.250	0.500	1.000
R_2 (s^{-1})	1.464 ± 0.096	2.346 ± 0.193	2.807 ± 0.131	4.288 ± 0.304	7.974 ± 0.727
R-square	0.962	0.961	0.988	0.979	0.967
R_2 contrast – C_2 (%)	-	35.0 ± 8.9	45.6 ± 5.3	64.4 ± 8.5	80.9 ± 11.7
d_{max} (s)	-	0.524	0.476	0.374	0.256



Parameter	[VBC] μM				
	0.000	0.125	0.250	0.500	1.000
R_2 (s^{-1})	1.392 ± 0.101	2.337 ± 0.335	3.678 ± 0.489	4.255 ± 0.566	7.063 ± 1.128
R-square	0.952	0.898	0.922	0.931	0.903
R_2 contrast – C_2 (%)	-	34.7 ± 15.3	58.5 ± 15.5	64.1 ± 15.8	78.4 ± 20.3
d_{max} (s)	-	0.526	0.409	0.376	0.277

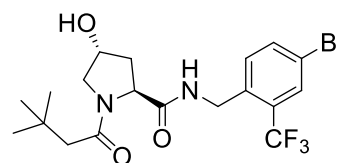
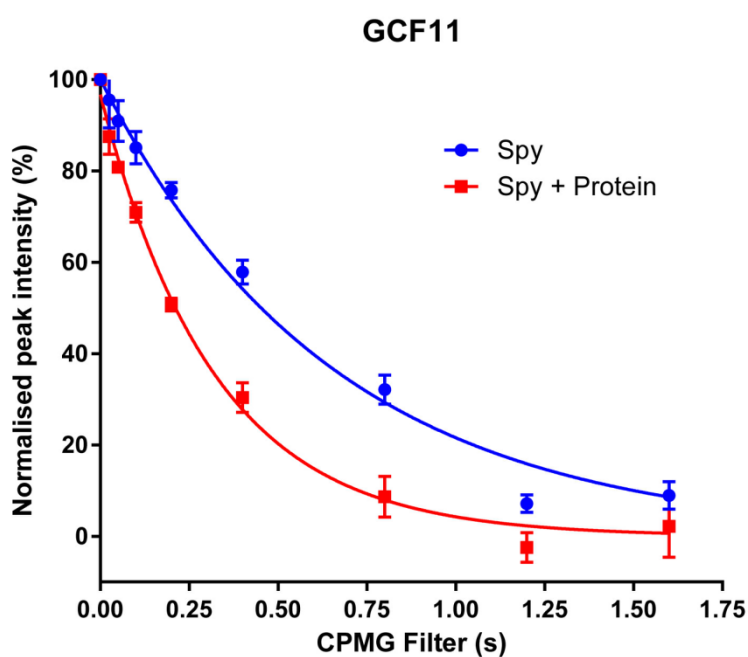
SUPPORTING INFORMATION

8.3 Spy molecules **6** and **11** at 50 μM in absence or in presence of VBC 1 μM



6

Spy molecule 100 μM	
R_2 (s^{-1})	1.255 ± 0.043
R-square	0.989
Spy molecule 100 μM + VBC 1 μM	
R_2 (s^{-1})	1.951 ± 0.062
R-square	0.994
Contrast	
R_2 contrast – C_2 (%)	35.7 ± 4.0
d_{max} (s)	0.634

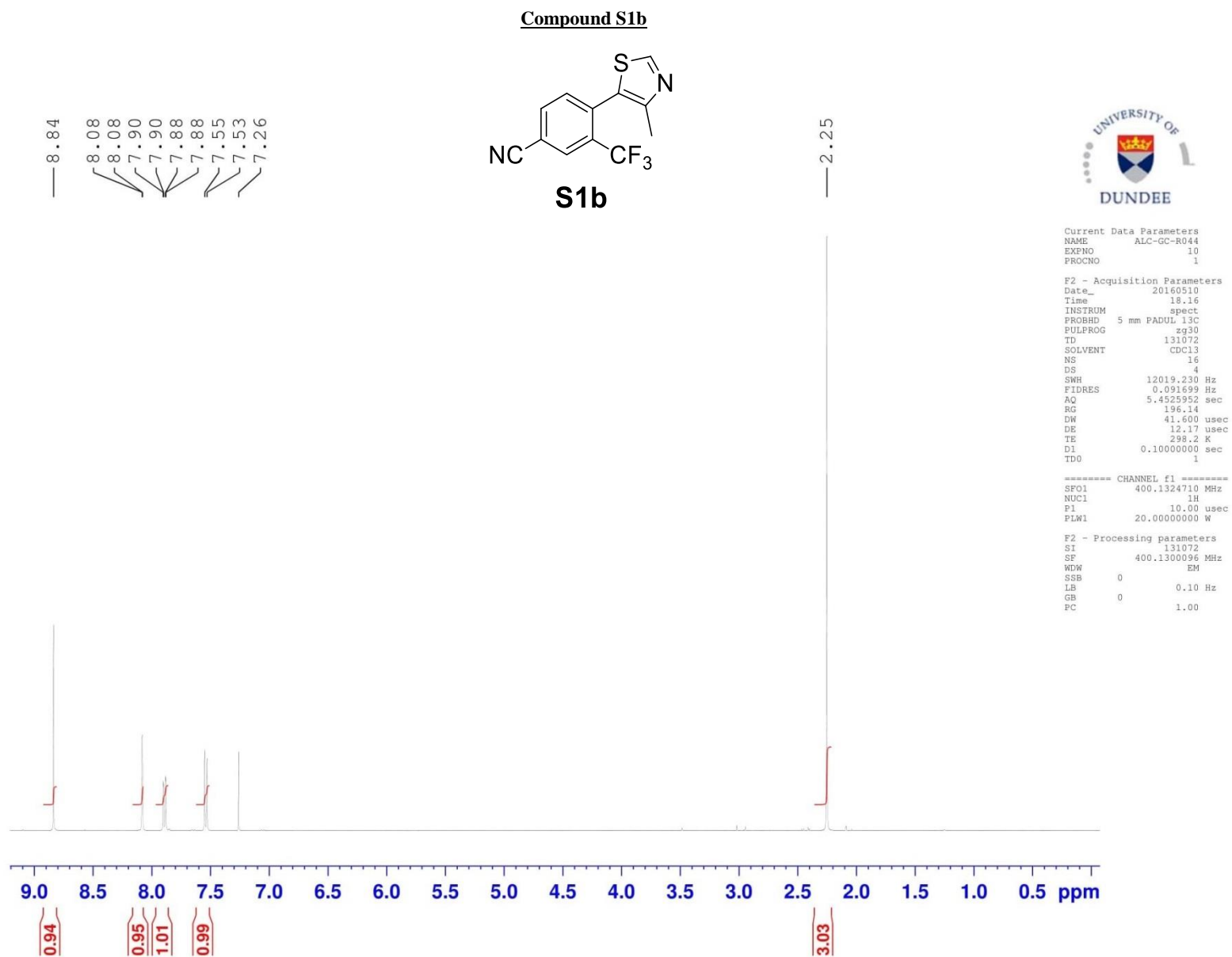


11

Spy molecule 100 μM	
R_2 (s^{-1})	1.534 ± 0.071
R-square	0.984
Spy molecule 100 μM + VBC 1 μM	
R_2 (s^{-1})	3.122 ± 0.145
R-square	0.990
Contrast	
R_2 contrast – C_2 (%)	50.9 ± 5.7
d_{max} (s)	0.447

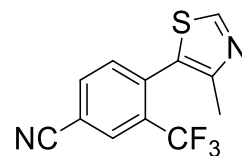
SUPPORTING INFORMATION

9. NMR spectra of synthesized compounds

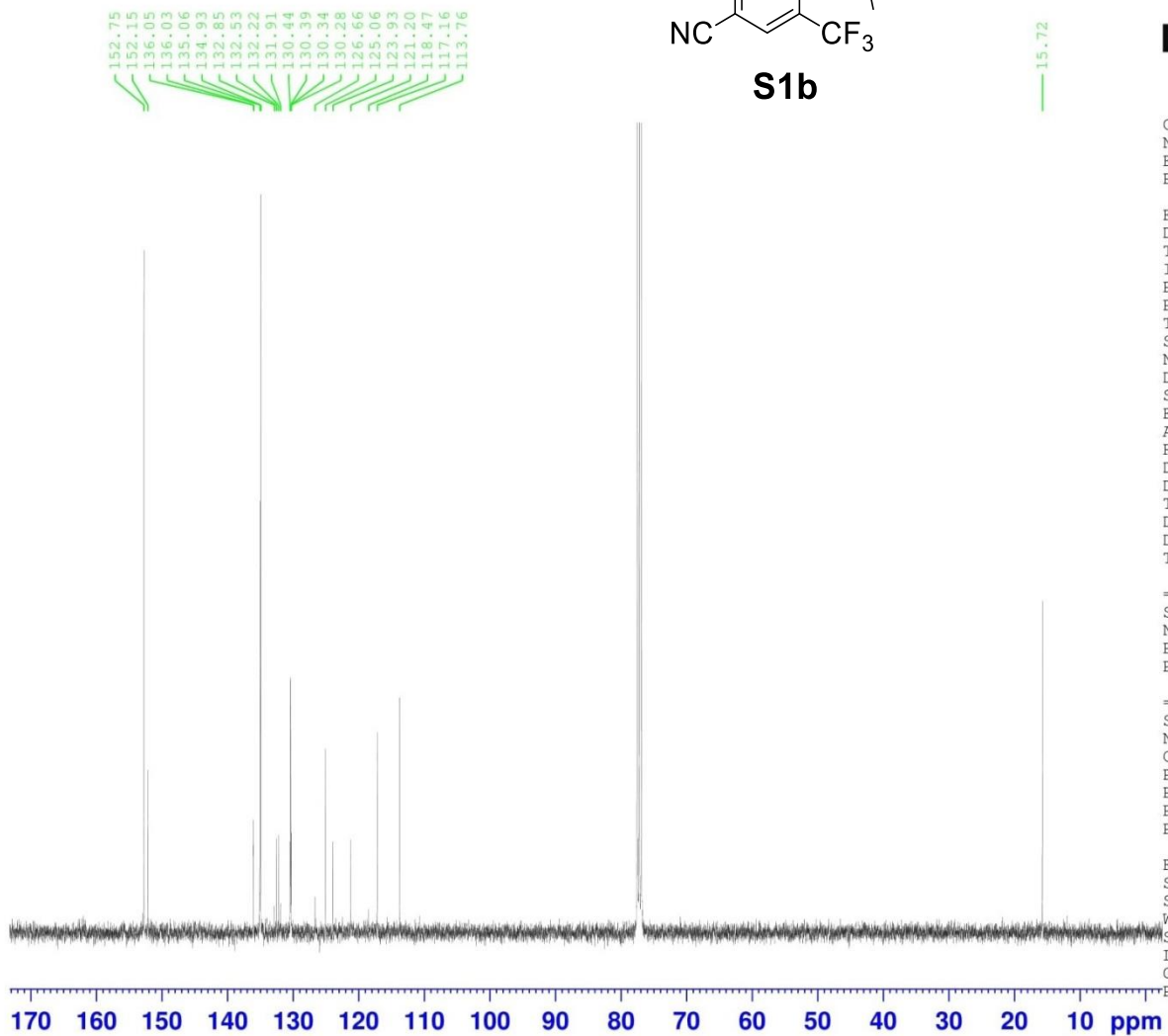


SUPPORTING INFORMATION

Compound S1b



S1b



Current Data Parameters
 NAME ALC-GC-R044
 EXPNO 14
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20160511
 Time 19.04
 INSTRUM spect
 PROBHD 5 mm PADUL 13C
 PULPROG zgpg30
 TD 119044
 SOLVENT CDCl3
 NS 512
 DS 4
 SWH 25000.000 Hz
 FIDRES 0.210006 Hz
 AQ 2.3808801 sec
 RG 196.14
 DW 20.000 usec
 DE 7.92 usec
 TE 298.2 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TD0 1

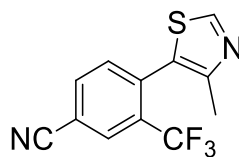
==== CHANNEL f1 =====
 SFO1 100.6238351 MHz
 NUC1 13C
 P1 10.00 usec
 PLW1 36.00000000 W

==== CHANNEL f2 =====
 SFO2 400.1316005 MHz
 NUC2 1H
 CPDPRG[2] waltz64
 PCPD2 90.00 usec
 PLW2 20.00000000 W
 PLW12 0.24691001 W
 PLW13 0.20000000 W

F2 - Processing parameters
 SI 131072
 SF 100.6127525 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

SUPPORTING INFORMATION

Compound S1b



S1b



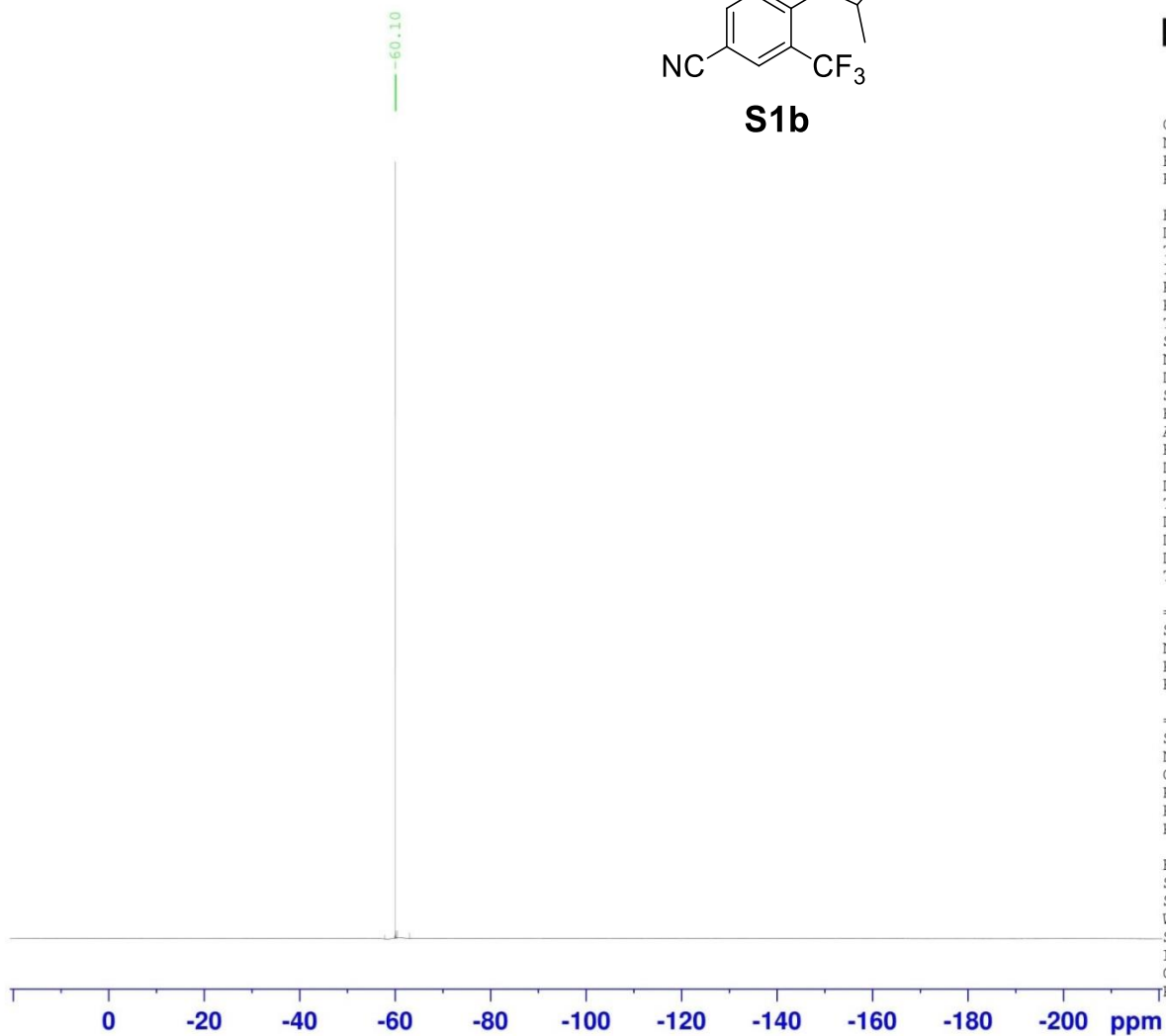
Current Data Parameters
 NAME GC-R044
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20160517
 Time 10.37
 INSTRUM spect
 PROBHD 5 mm QNP 1H/13
 PULPROG zgfhigqn.2
 TD 131072
 SOLVENT CDCl3
 NS 16
 DS 4
 SWH 113636.367 Hz
 FIDRES 0.866977 Hz
 AQ 0.5767168 sec
 RG 724
 DW 4.400 usec
 DE 6.50 usec
 TE 298.2 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 D12 0.00002000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 470.5453180 MHz
 NUC1 19F
 P1 13.50 usec
 PLW1 25.00000000 W

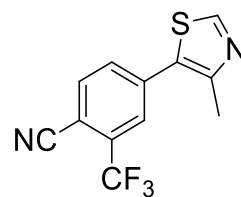
===== CHANNEL f2 =====
 SFO2 500.1320005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 80.00 usec
 PLW2 25.00000000 W
 PLW12 0.39063001 W

F2 - Processing parameters
 SI 65536
 SF 470.5923772 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

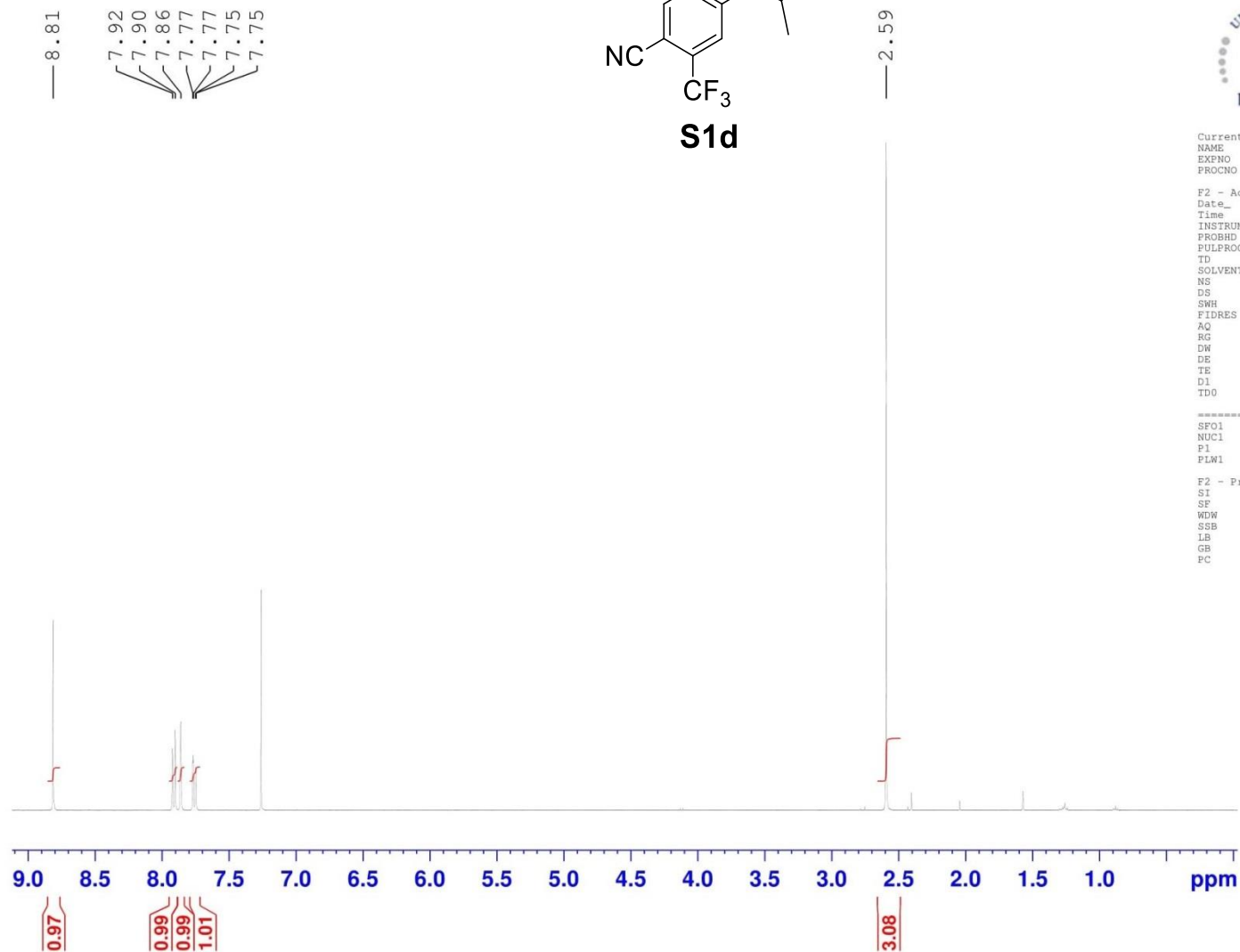


SUPPORTING INFORMATION

Compound S1d



S1d



```

Current Data Parameters
NAME      ALC-GC-R045
EXPNO     10
PROCNO    1

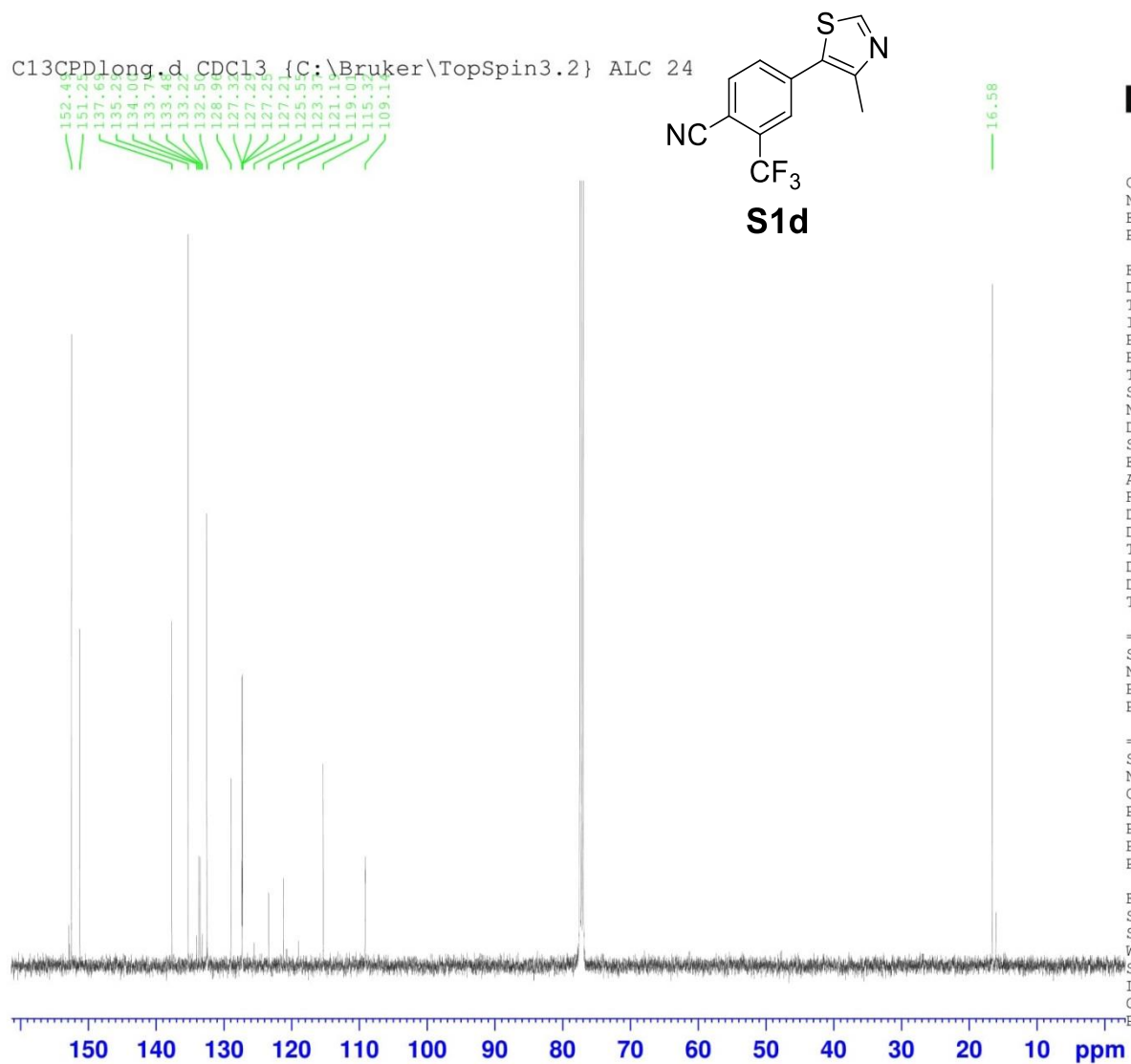
F2 - Acquisition Parameters
Date_     20160511
Time      17.29
INSTRUM   spect
PROBHD    5 mm PADUL 13C
PULPROG   zg30
TD         131072
SOLVENT   CDCl3
NS         16
DS         4
SWH        12019.230 Hz
FIDRES     0.091699 Hz
AQ          5.4525952 sec
RG          196.14
DW          41.600 usec
DE          12.17 usec
TE          298.2 K
D1          0.10000000 sec
TD0         1

===== CHANNEL f1 =====
SFO1      400.1324710 MHz
NUC1       1H
P1         10.00 usec
PLW1       20.00000000 W

F2 - Processing parameters
SI         131072
SF         400.1300097 MHz
WDW        EM
SSB        0
LB         0.10 Hz
GB         0
PC         1.00
    
```

SUPPORTING INFORMATION

Compound S1d



Current Data Parameters
 NAME GC-R045
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20160518
 Time 0.18
 INSTRUM spect
 PROBHD 5 mm QNP 1H/13
 PULPROG zgpg30
 TD 65536
 SOLVENT CDC13
 NS 2048
 DS 4
 SWH 29761.904 Hz
 FIDRES 0.454131 Hz
 AQ 1.1010048 sec
 RG 2050
 DW 16.800 usec
 DE 6.50 usec
 TE 298.2 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TD0 2

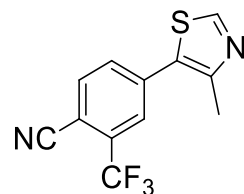
===== CHANNEL f1 =====
 SFO1 125.7703637 MHz
 NUC1 13C
 P1 7.50 usec
 PLW1 92.00000000 W

===== CHANNEL f2 =====
 SFO2 500.1320005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 80.00 usec
 PLW2 25.00000000 W
 PLW12 0.39063001 W
 PLW13 0.25000000 W

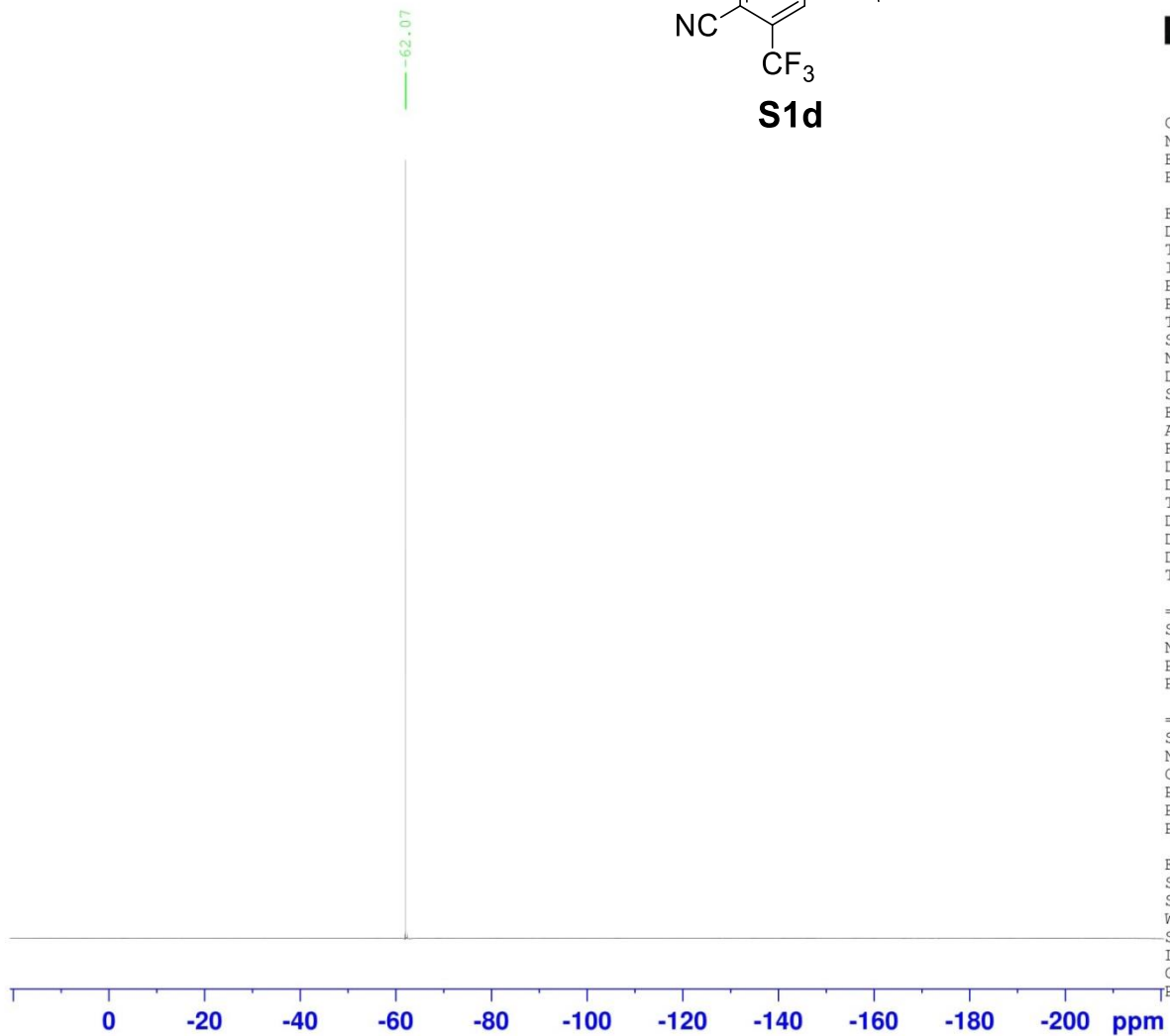
F2 - Processing parameters
 SI 32768
 SF 125.7577673 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

SUPPORTING INFORMATION

Compound S1d



S1d



Current Data Parameters
NAME GC-R045
EXPNO 9
PROCNO 1

F2 - Acquisition Parameters
Date_ 20160517
Time 10.50
INSTRUM spect
PROBHD 5 mm QNP 1H/13
PULPROG zgfhigqn.2
TD 131072
SOLVENT CDC13
NS 16
DS 4
SWH 113636.367 Hz
FIDRES 0.866977 Hz
AQ 0.5767168 sec
RG 724
DW 4.400 usec
DE 6.50 usec
TE 298.2 K
D1 1.00000000 sec
D11 0.03000000 sec
D12 0.00002000 sec
TD0 1

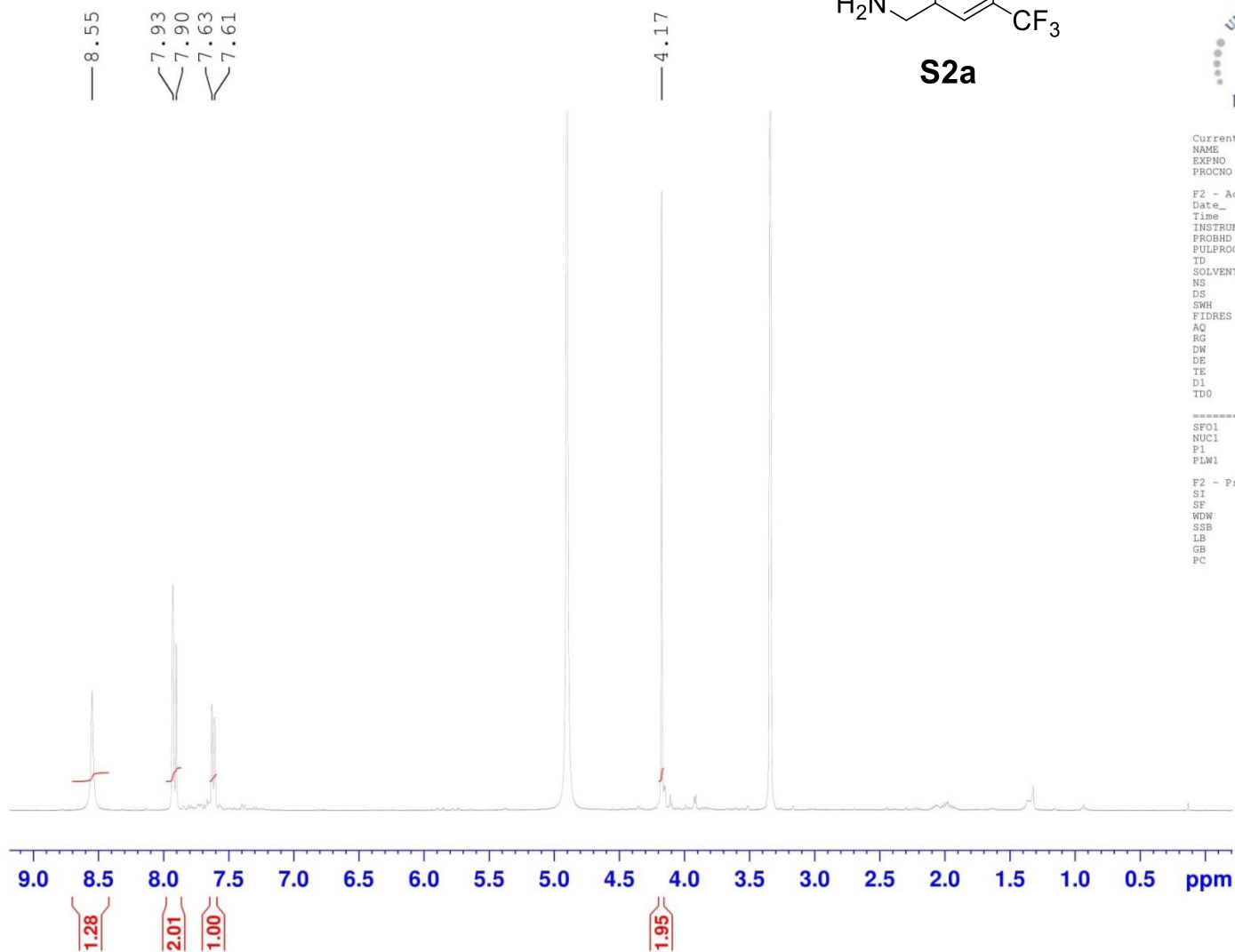
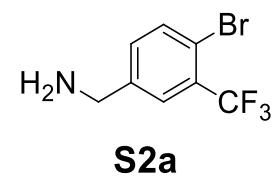
==== CHANNEL f1 =====
SFO1 470.5453180 MHz
NUC1 19F
P1 13.50 usec
PLW1 25.00000000 W

==== CHANNEL f2 =====
SFO2 500.1320005 MHz
NUC2 1H
CPDPRG[2] waltz16
PCPD2 80.00 usec
PLW2 25.00000000 W
PLW12 0.39063001 W

F2 - Processing parameters
SI 65536
SF 470.5923772 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

SUPPORTING INFORMATION

Compound S2a



```

Current Data Parameters
NAME      ALC-GC-S3A-3
EXPNO    20
PROCNO    1

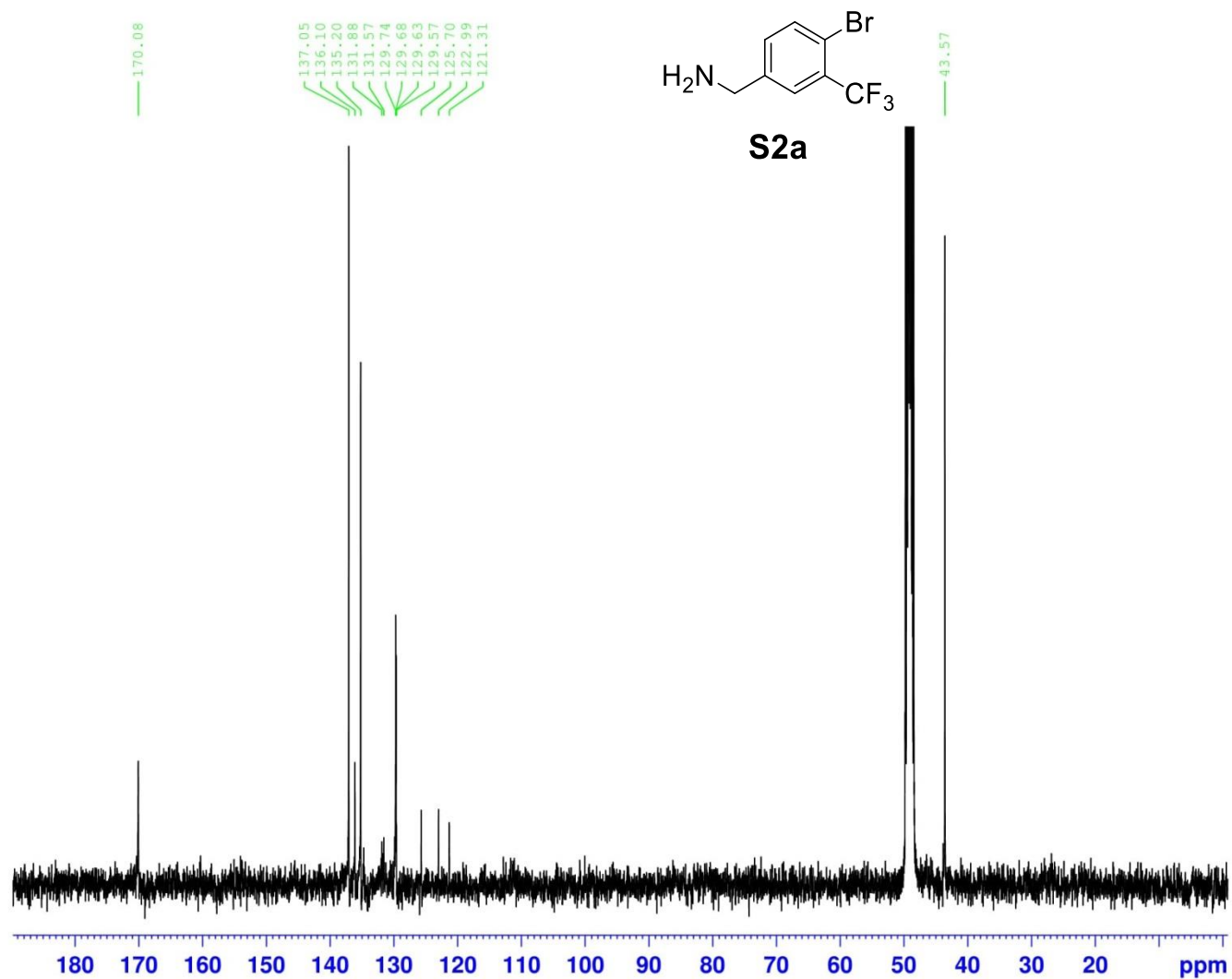
F2 - Acquisition Parameters
Date_    20181123
Time     21.31
INSTRUM  spect
PROBHD   5 mm PADUL 13C
PULPROG  zg30
ID       65536
SOLVENT  MeOD
NS       64
DS       2
SWH      8000.000 Hz
FIDRES   0.122070 Hz
AQ       4.0960002 sec
RG       196.14
DW       62.500 usec
DE       11.07 usec
TE       294.6 K
D1       1.0000000 sec
TD0      1

===== CHANNEL f1 =====
SF01    400.1324710 MHz
NUC1     1H
P1       10.00 usec
PLW1    20.00000000 W

F2 - Processing parameters
SI       65536
SF       400.1299954 MHz
WDW      EM
SSB      0
LB       0.30 Hz
GB       0
PC       1.00
    
```


SUPPORTING INFORMATION

Compound S2a



Current Data Parameters
 NAME ALC-GC-S3A-3
 EXPNO 21
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20181124
 Time 2.51
 INSTRUM spect
 PROBHD 5 mm PADUL 13C
 PULPROG udeflt
 TD 17996
 SOLVENT MeOD
 NS 5000
 DS 0
 SWH 25000.000 Hz
 FIDRES 1.389198 Hz
 AQ 0.3599200 sec
 RG 196.14
 DW 20.000 usec
 DE 8.66 usec
 TE 294.6 K
 D1 3.00000000 sec
 D11 0.03000000 sec
 D12 0.00002000 sec
 D20 200.00000000 sec
 TDO 1

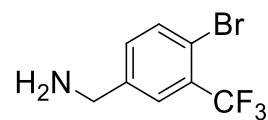
===== CHANNEL f1 =====
 SF01 100.6238346 MHz
 NUC1 13C
 P1 10.00 usec
 P13 2000.00 usec
 P26 500.00 usec
 PLW1 36.00000000 W
 SPNAM[5] Crp60comp.4
 SPOAL5 0.500
 SPOFF5 0 Hz
 SPW5 5.50040007 W
 SPNAM[8] Crp60,0.5,20.1
 SPOAL8 0.500
 SPOFF8 0 Hz
 SPW8 5.50040007 W

===== CHANNEL f2 =====
 SF02 400.1316005 MHz
 NUC2 1H
 CPDPRG[2] waltz64
 PCPD2 90.00 usec
 PLW2 20.00000000 W
 PLW12 0.24691001 W

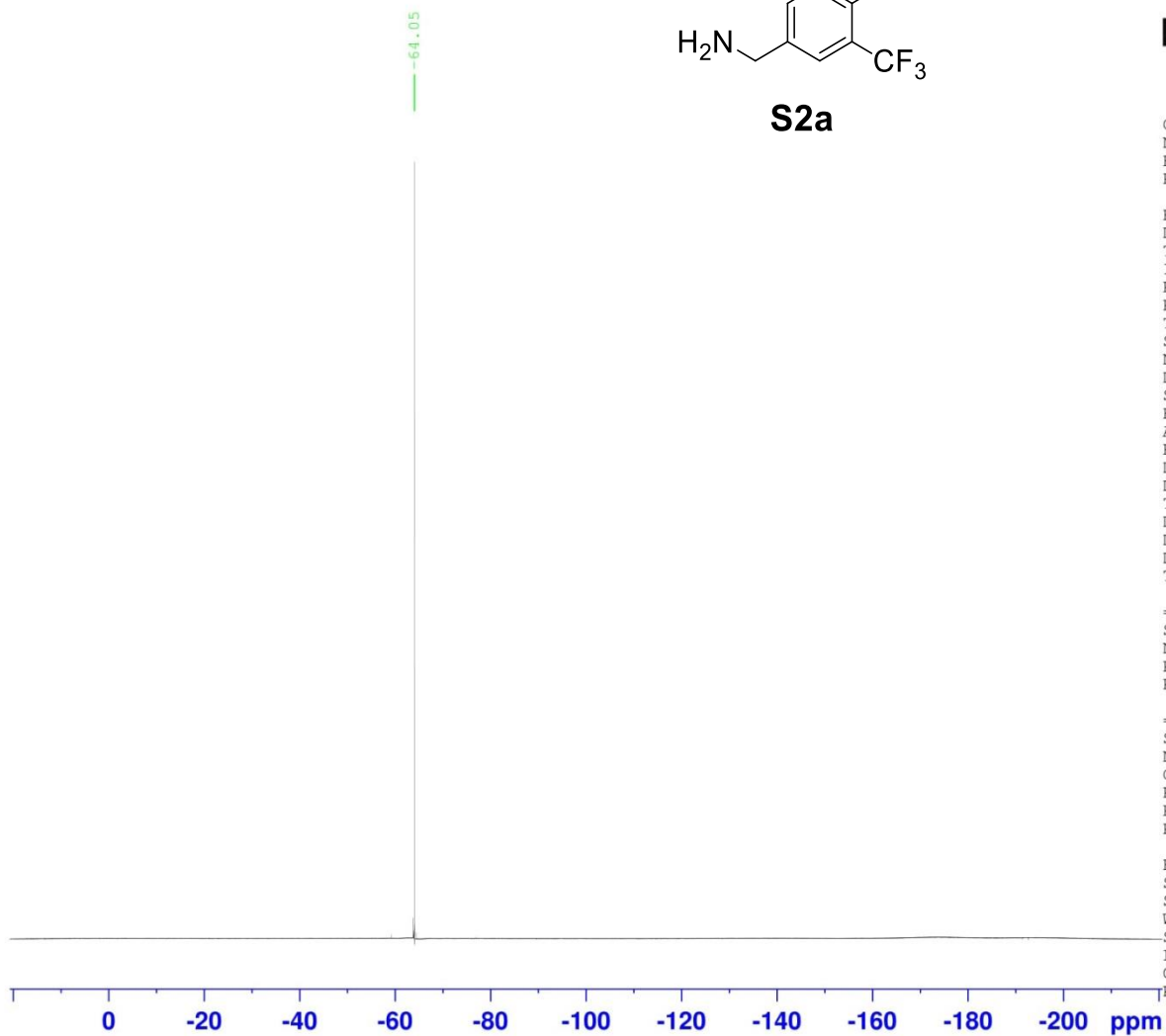
F2 - Processing parameters
 SI 262144
 SF 100.6126171 MHz
 WDW EM
 SSB 0
 LB 2.00 Hz
 GB 0
 PC 1.40

SUPPORTING INFORMATION

Compound S2a



S2a



Current Data Parameters
 NAME GC-S2A-F3
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20181126
 Time 9.55
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zgfhigqn.2
 TD 131072
 SOLVENT MeOD
 NS 16
 DS 4
 SWH 113636.367 Hz
 FIDRES 0.866977 Hz
 AQ 0.5767168 sec
 RG 322
 DW 4.400 usec
 DE 6.50 usec
 TE 303.4 K
 D1 1.0000000 sec
 D11 0.0300000 sec
 D12 0.00002000 sec
 TD0 1

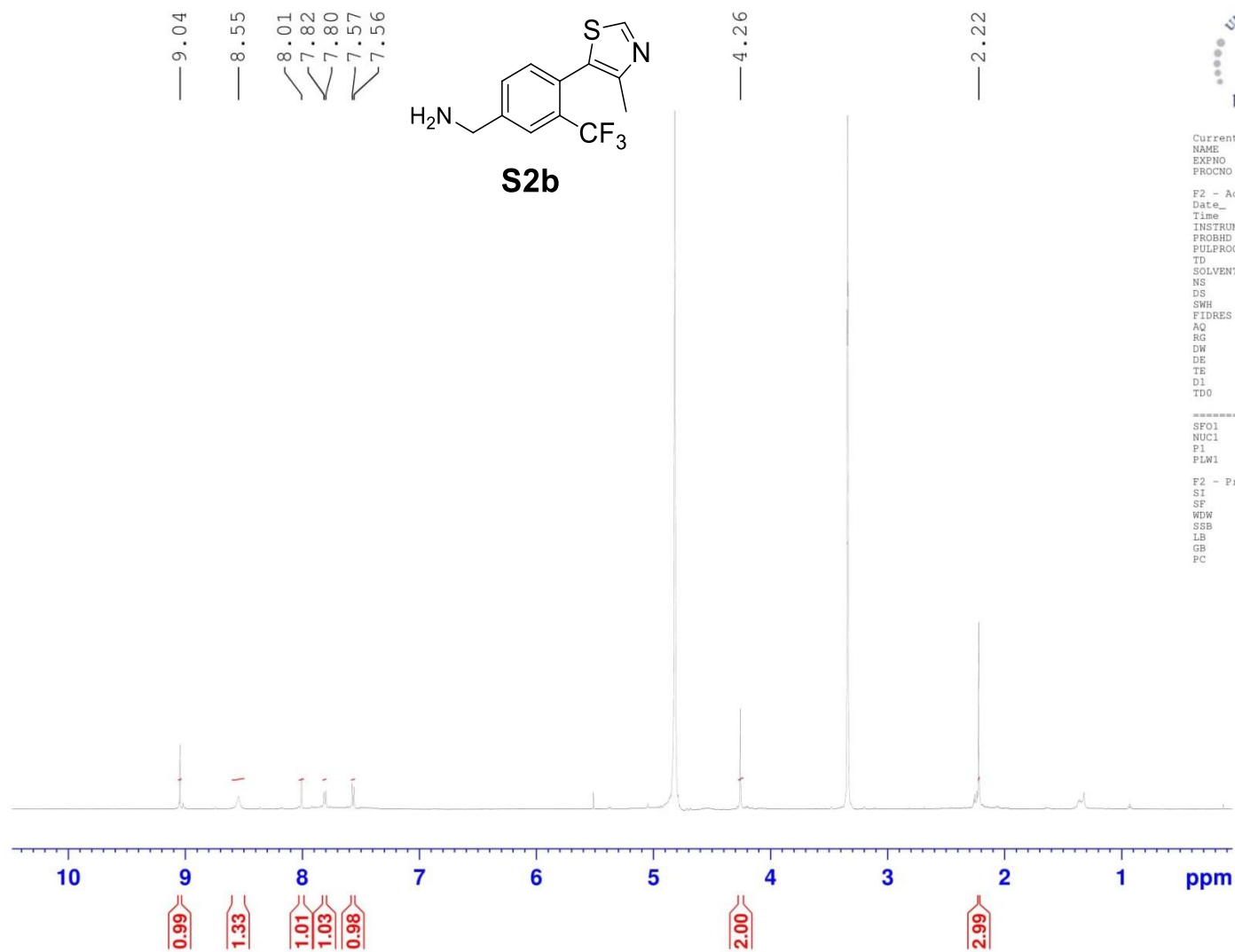
===== CHANNEL f1 =====
 SFO1 470.5453180 MHz
 NUC1 19F
 P1 14.50 usec
 PLW1 45.00000000 W

===== CHANNEL f2 =====
 SFO2 500.1320005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 80.00 usec
 PLW2 20.85000038 W
 PLW12 0.35916999 W

F2 - Processing parameters
 SI 65536
 SF 470.5923772 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

SUPPORTING INFORMATION

Compound S2b



Current Data Parameters
 NAME GC-S2B-2
 EXPNO 10
 PROCNO 1

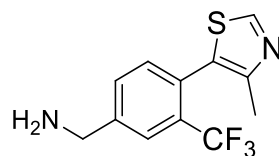
F2 - Acquisition Parameters
 Date_ 20181126
 Time 10.27
 INSTRUM spect
 PROBHD 5 mm FABBO BB/
 PULPROG zg30
 ID 65536
 SOLVENT MeOD
 NS 64
 DS 2
 SWH 10000.000 Hz
 FIDRES 0.152588 Hz
 AQ 3.2767999 sec
 RG 181
 DW 50.000 usec
 DE 6.50 usec
 TE 303.1 K
 D1 1.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 SF01 500.1330885 MHz
 NUC1 1H
 P1 10.00 usec
 PLW1 20.85000038 W

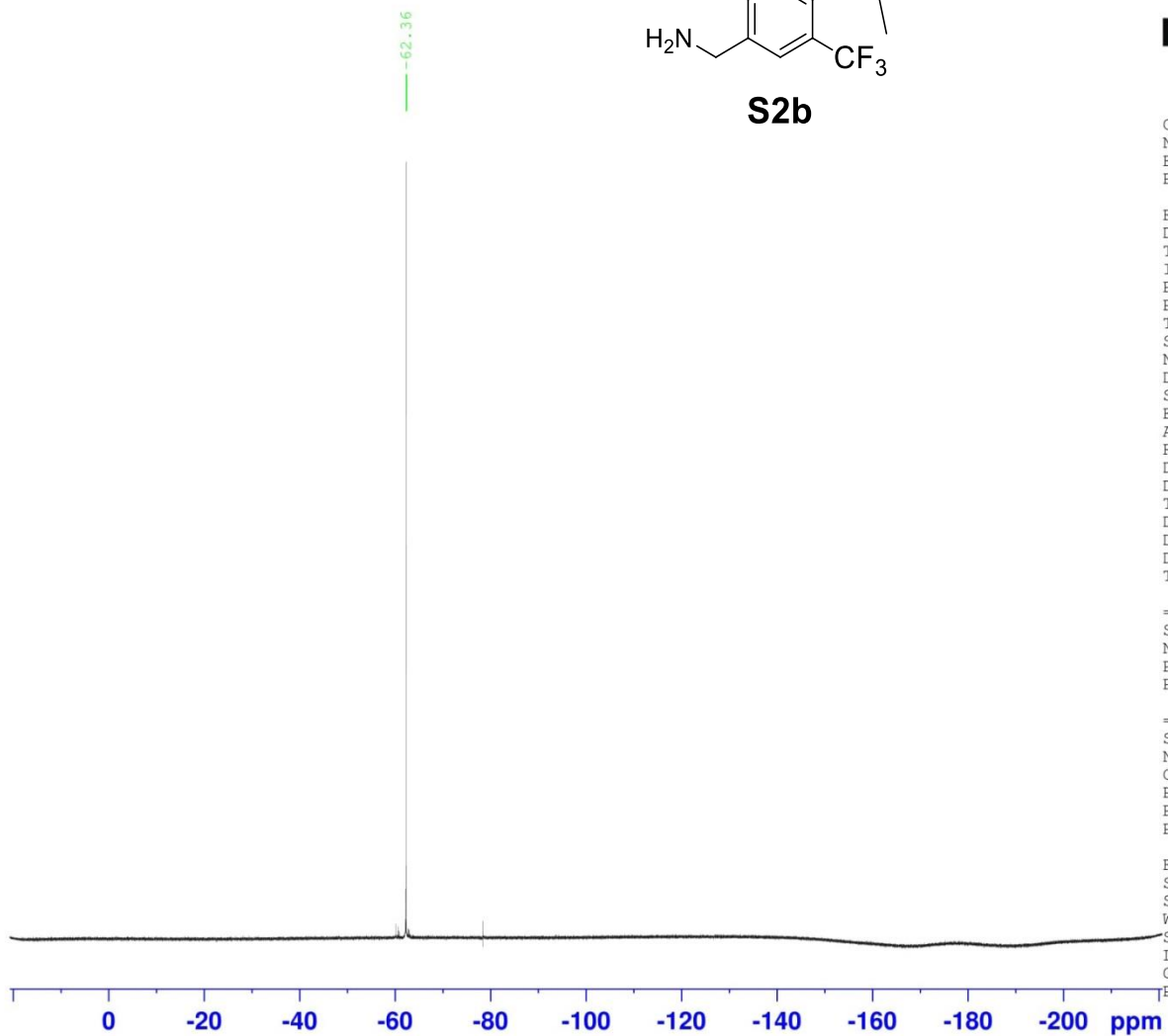
F2 - Processing parameters
 SI 65536
 SF 500.1292670 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

SUPPORTING INFORMATION

Compound S2b



S2b



Current Data Parameters
 NAME GC-S2B-2
 EXPNO 11
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20181126
 Time 10.29
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zgfhigqn.2
 TD 131072
 SOLVENT MeOD
 NS 16
 DS 4
 SWH 113636.367 Hz
 FIDRES 0.866977 Hz
 AQ 0.5767168 sec
 RG 322
 DW 4.400 usec
 DE 6.50 usec
 TE 303.4 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 D12 0.00002000 sec
 TD0 1

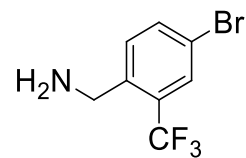
==== CHANNEL f1 =====
 SFO1 470.5453180 MHz
 NUC1 19F
 P1 14.50 usec
 PLW1 45.00000000 W

==== CHANNEL f2 =====
 SFO2 500.1320005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 80.00 usec
 PLW2 20.85000038 W
 PLW12 0.35916999 W

F2 - Processing parameters
 SI 65536
 SF 470.5923772 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

SUPPORTING INFORMATION

Compound S2c



S2c

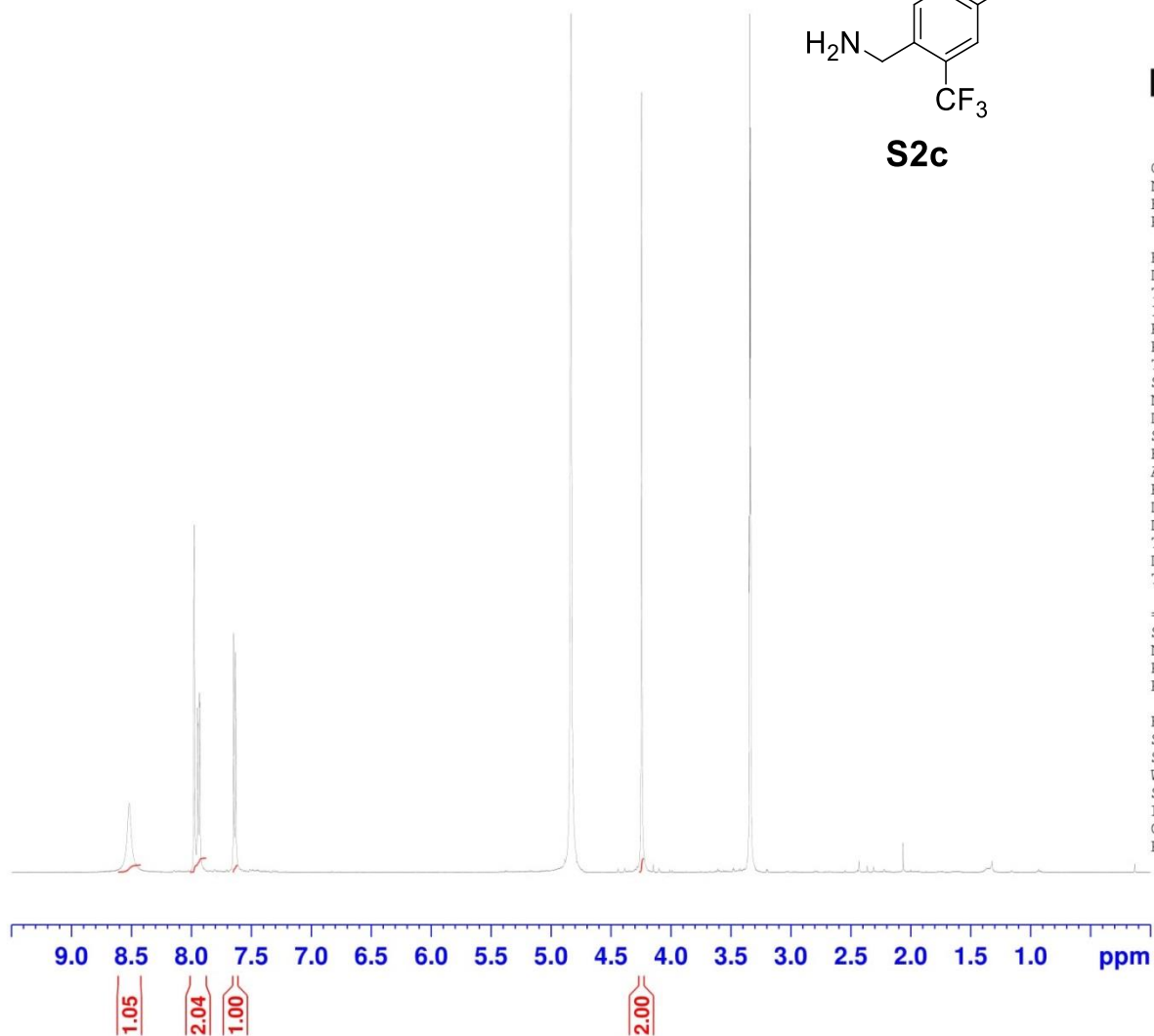


Current Data Parameters
 NAME GC-S2C-GILSON
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20181126
 Time 9.46
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg30
 TD 65536
 SOLVENT MeOD
 NS 64
 DS 2
 SWH 10000.000 Hz
 FIDRES 0.152588 Hz
 AQ 3.2767999 sec
 RG 181
 DW 50.000 usec
 DE 6.50 usec
 TE 303.1 K
 D1 1.00000000 sec
 TD0 1

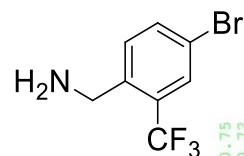
==== CHANNEL f1 =====
 SFO1 500.1330885 MHz
 NUC1 1H
 P1 10.00 usec
 PLW1 20.85000038 W

F2 - Processing parameters
 SI 65536
 SF 500.1299958 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

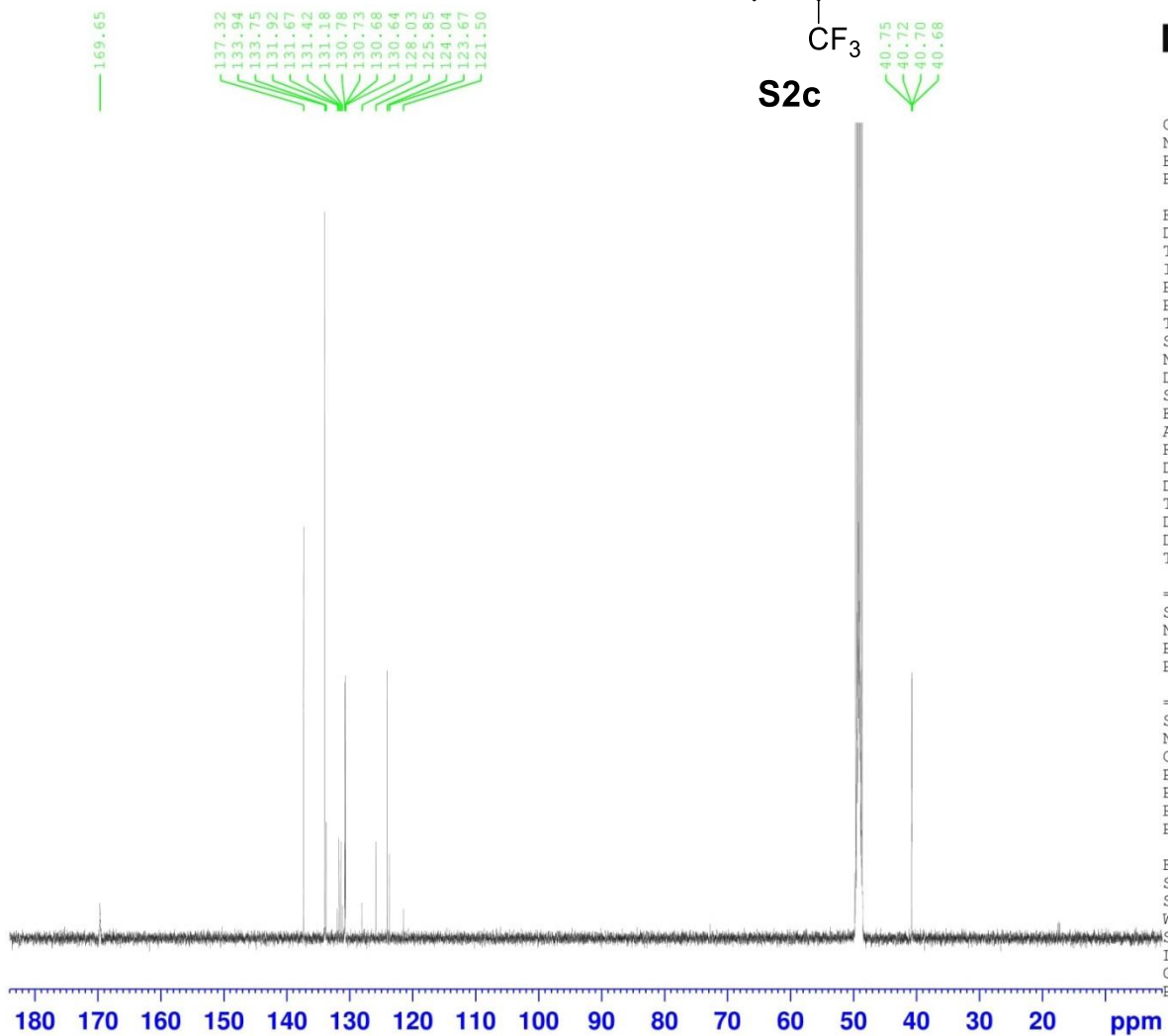


SUPPORTING INFORMATION

Compound S2c



S2c



Current Data Parameters
 NAME GC-S2C+
 EXPNO 11
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20181126
 Time 11.50
 INSTRUM spect
 PROBHD 5 mm CPQCI 1H/
 PULPROG zgpg30
 TD 65400
 SOLVENT MeOD
 NS 2048
 DS 4
 SWH 29761.904 Hz
 FIDRES 0.455075 Hz
 AQ 1.0987200 sec
 RG 190.78
 DW 16.800 usec
 DE 18.00 usec
 TE 303.1 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TD0 1

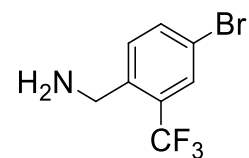
===== CHANNEL f1 =====
 SFO1 125.7703637 MHz
 NUC1 13C
 P1 12.00 usec
 PLW1 133.00000000 W

===== CHANNEL f2 =====
 SFO2 500.1320005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 100.00 usec
 PLW2 5.19999981 W
 PLW12 0.06406900 W
 PLW13 0.06406900 W

F2 - Processing parameters
 SI 131072
 SF 125.7575991 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

SUPPORTING INFORMATION

Compound S2c



S2c



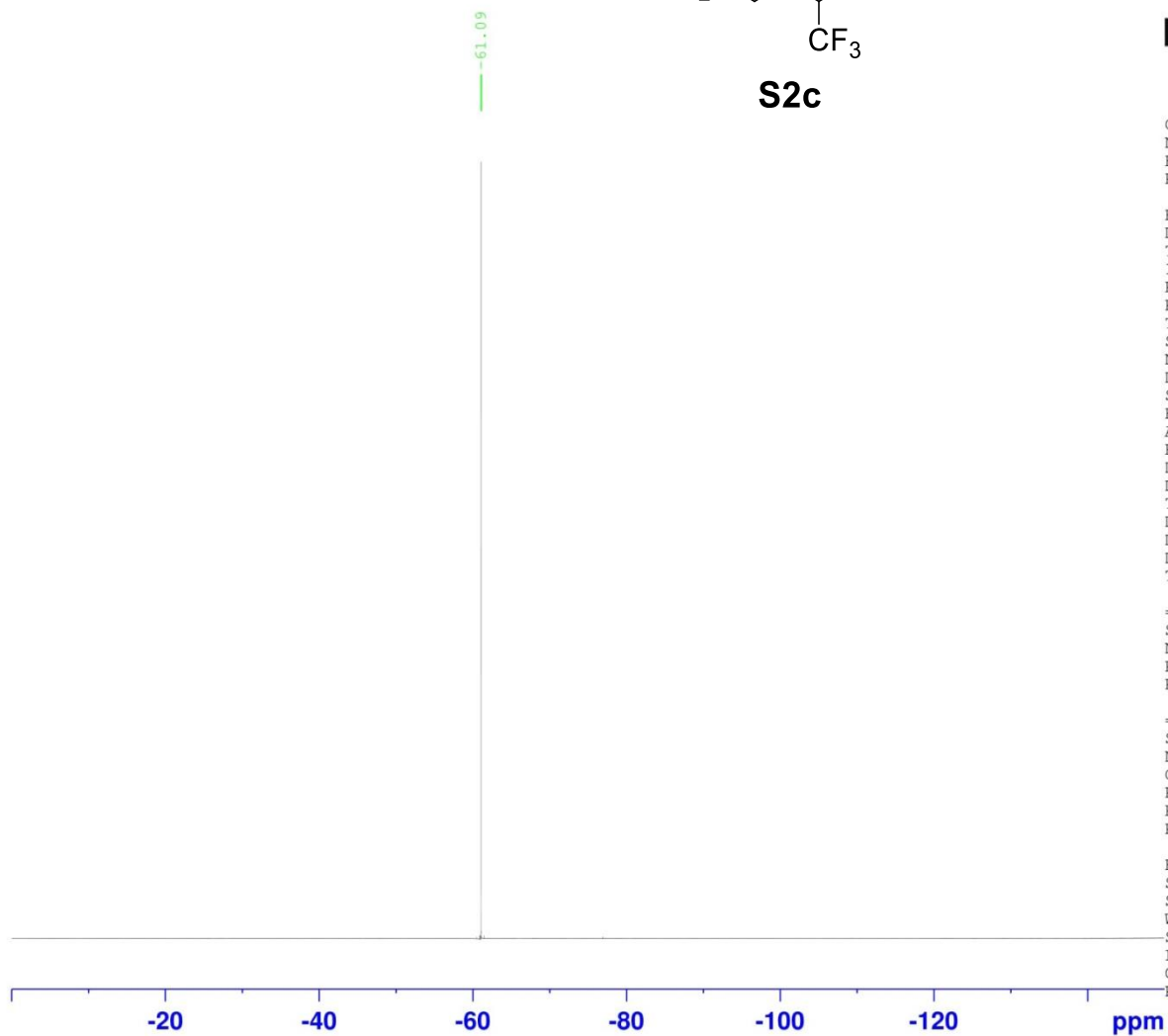
Current Data Parameters
NAME GC-S2C-GILSON
EXPNO 11
PROCNO 1

F2 - Acquisition Parameters
Date_ 20181126
Time 9.49
INSTRUM spect
PROBHD 5 mm PABBO BB/
PULPROG zgfhigqn.2
TD 131072
SOLVENT MeOD
NS 16
DS 4
SWH 113636.367 Hz
FIDRES 0.866977 Hz
AQ 0.5767168 sec
RG 322
DW 4.400 usec
DE 6.50 usec
TE 303.4 K
D1 1.00000000 sec
D11 0.03000000 sec
D12 0.00002000 sec
TD0 1

==== CHANNEL f1 =====
SFO1 470.5453180 MHz
NUC1 19F
P1 14.50 usec
PLW1 45.00000000 W

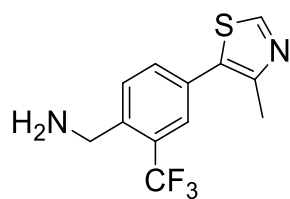
==== CHANNEL f2 =====
SFO2 500.1320005 MHz
NUC2 1H
CPDPRG[2] waltz16
PCPD2 80.00 usec
PLW2 20.85000038 W
PLW12 0.35916999 W

F2 - Processing parameters
SI 65536
SF 470.5923772 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

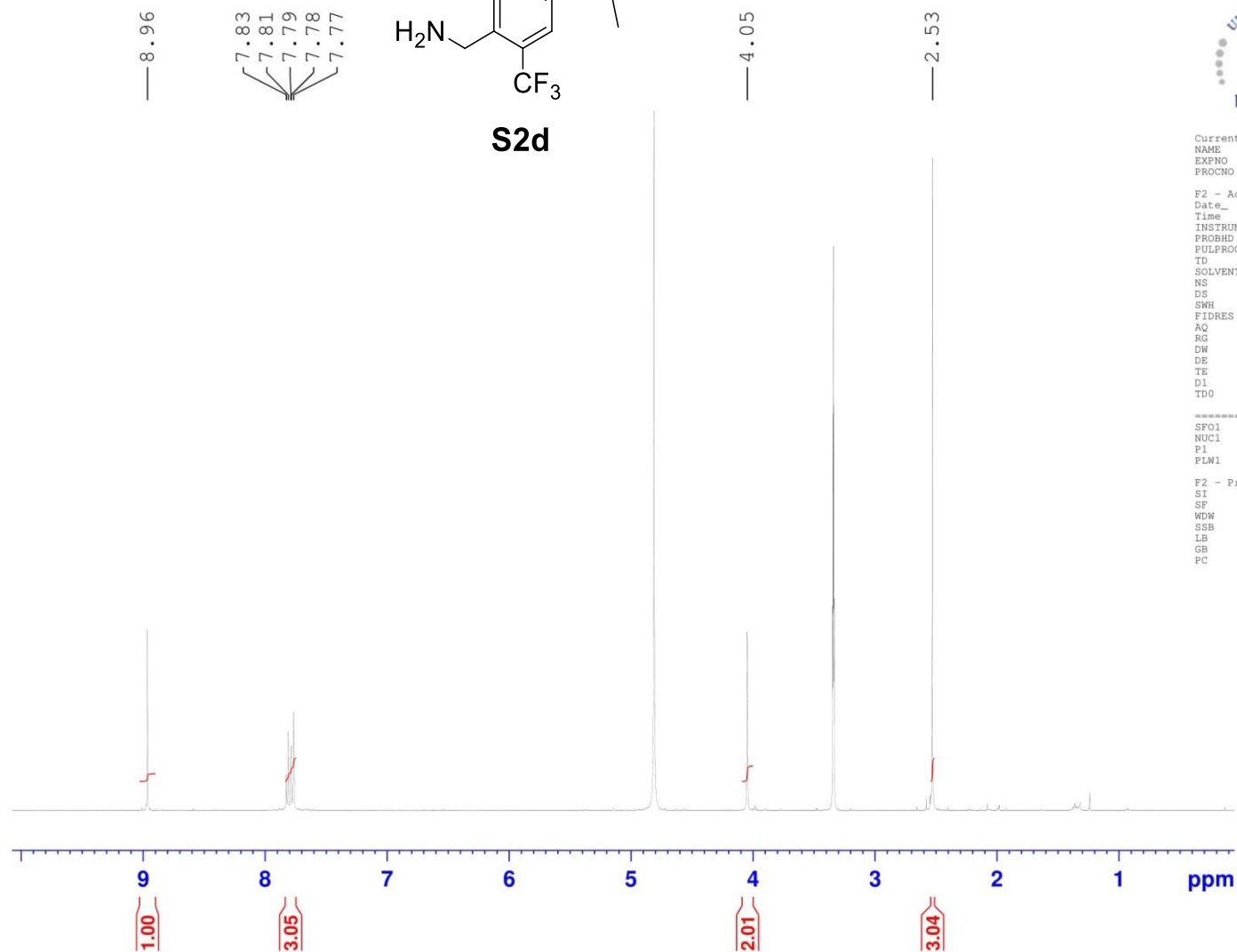


SUPPORTING INFORMATION

Compound S2d



S2d



```

Current Data Parameters
NAME      ALC-GC-S2D-E2
EXPNO     10
PROCNO    1

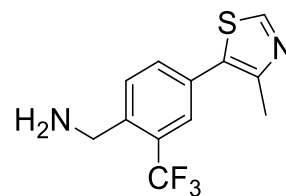
F2 - Acquisition Parameters
Date_     20181120
Time      14.26
INSTRUM   spect
PROBHD    5 mm FABBO BB/
PULPROG   zg30
ID         65536
SOLVENT   MeOD
NS         64
DS         2
SWH        10000.000 Hz
FIDRES     0.152588 Hz
AQ         3.2767999 sec
RG         181
DW         50.000 usec
DE         6.50 usec
TE         303.1 K
D1         1.00000000 sec
TD0        1

===== CHANNEL f1 =====
SF01      500.1330885 MHz
NUC1       1H
P1         10.00 usec
PLW1       20.85000038 W

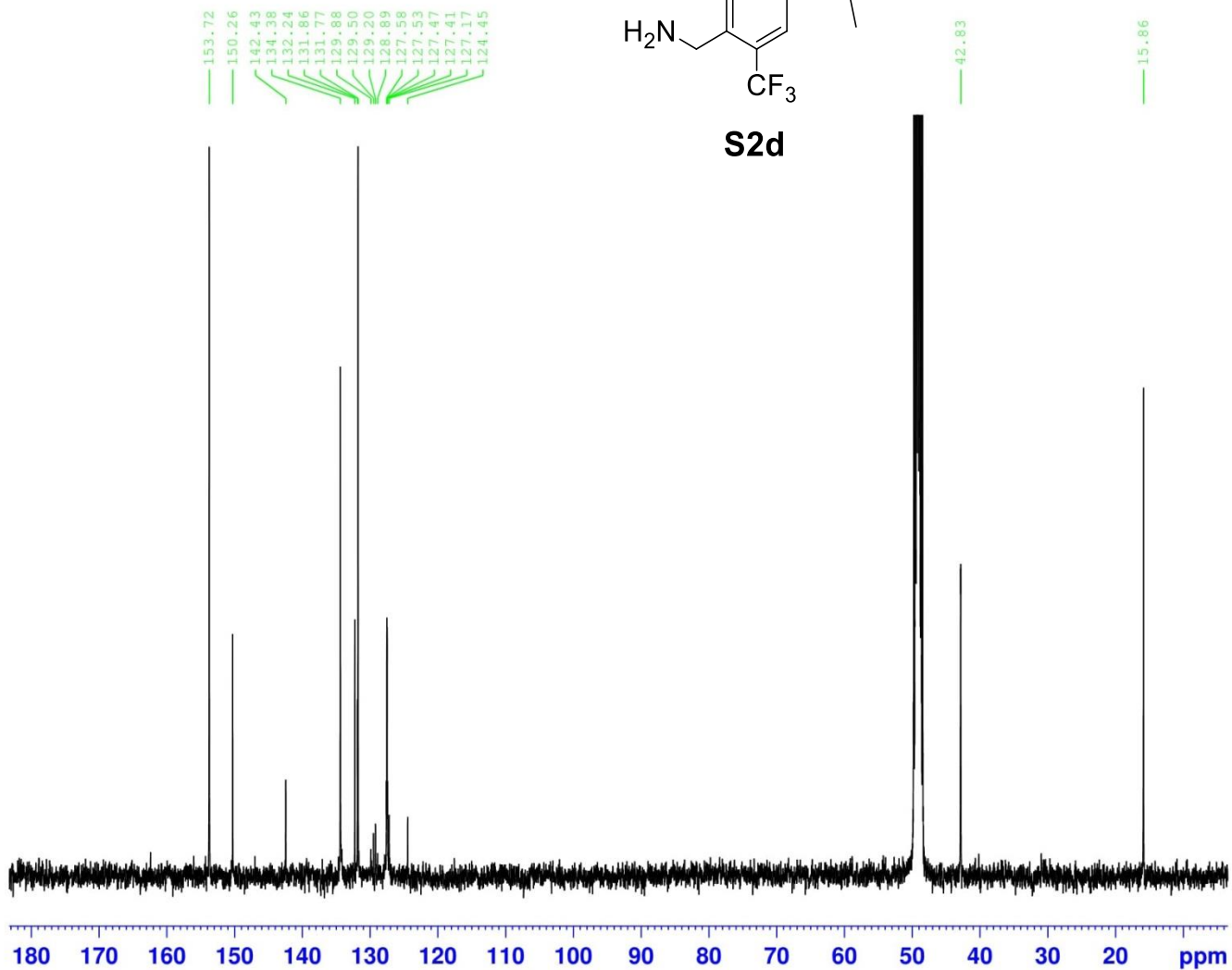
F2 - Processing parameters
SI         65536
SF         500.1299958 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00
    
```


SUPPORTING INFORMATION

Compound S2d



S2d



```

Current Data Parameters
NAME      ALC-GC-S2D
EXPNO    30
PROCNO   1

F2 - Acquisition Parameters
Date_    20181121
Time     5.05
INSTRUM  spect
PROBHD   5 mm PADUL 13C
PULPROG  udef
TD        17996
SOLVENT  MeOD
NS        5000
DS        0
SWH       25000.000 Hz
FIDRES    1.389198 Hz
AQ         0.3599200 sec
RG         196.14
DW         20.000 usec
DE         8.66 usec
TE        294.6 K
D1         3.00000000 sec
D11        0.03000000 sec
D12        0.00002000 sec
D20        200.00000000 sec
TD0        1

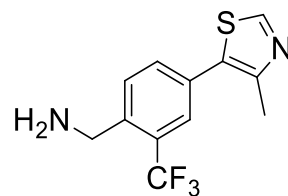
===== CHANNEL f1 =====
SF01      100.6238346 MHz
NUC1       13C
P1         10.00 usec
P13        2000.00 usec
P26         500.00 usec
PLW1       36.00000000 W
SPNAM[5]   Crp60comp.4
SPOAL5     0.500
SPOFFS5    0 Hz
SPW5       5.50040007 W
SPNAM[8]   Crp60,0.5,20.1
SPOAL8     0.500
SPOFFS8    0 Hz
SPW8       5.50040007 W

===== CHANNEL f2 =====
SF02      400.1316005 MHz
NUC2       1H
CPDPRG[2]  waltz64
PCPD2      90.00 usec
PLW2       20.00000000 W
PLW12      0.24691001 W

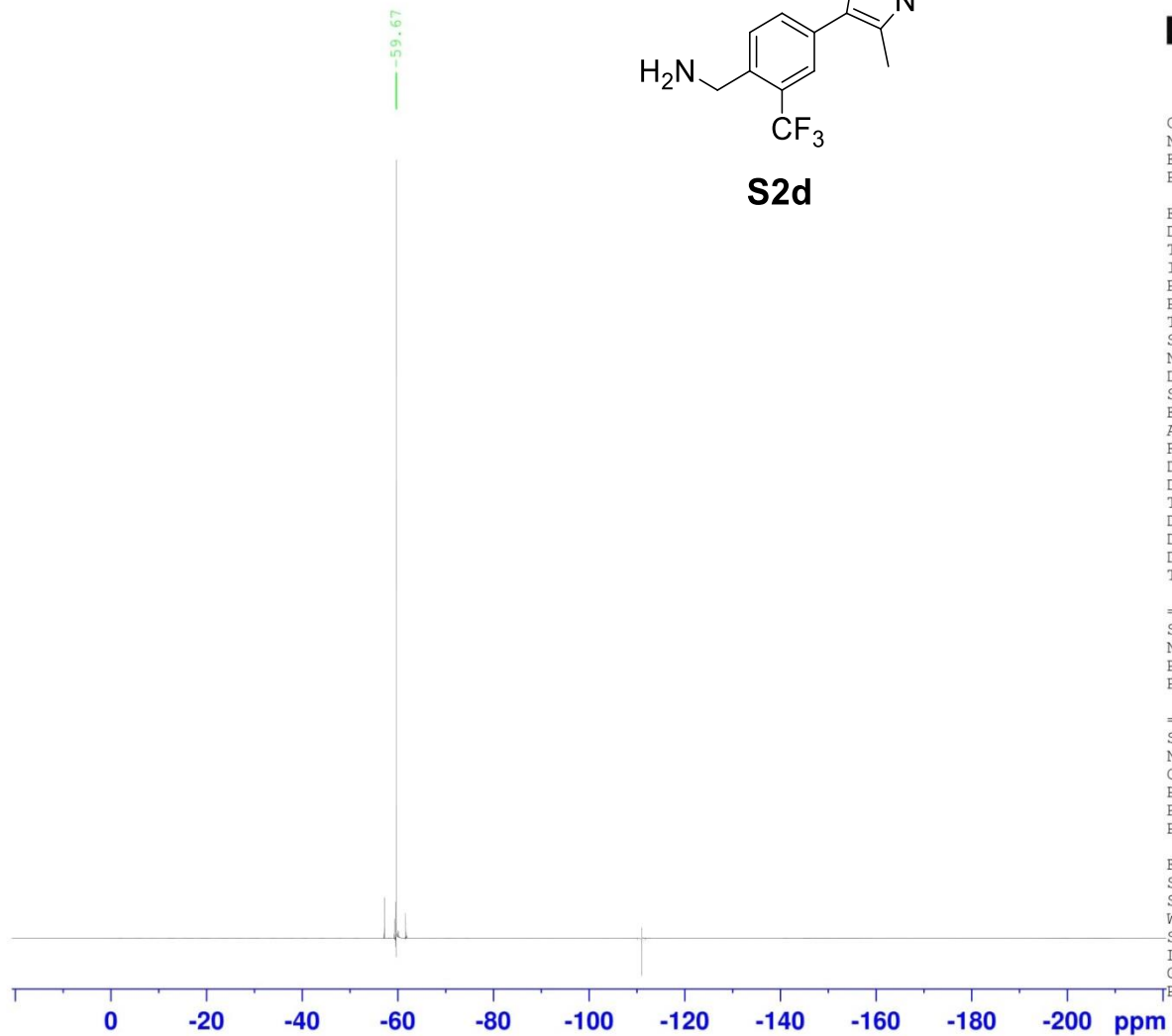
F2 - Processing parameters
SI         262144
SF         100.6126170 MHz
WDW        EM
SSB        0
LB         2.00 Hz
GB         0
PC         1.40
    
```

SUPPORTING INFORMATION

Compound S2d



S2d



Current Data Parameters
 NAME GC-R049
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20160519
 Time 13.52
 INSTRUM spect
 PROBHD 5 mm QNP 1H/13
 PULPROG zgfhigqn.2
 TD 131072
 SOLVENT MeOD
 NS 16
 DS 4
 SWH 113636.367 Hz
 FIDRES 0.866977 Hz
 AQ 0.5767168 sec
 RG 203
 DW 4.400 usec
 DE 6.50 usec
 TE 298.2 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 D12 0.00002000 sec
 TD0 1

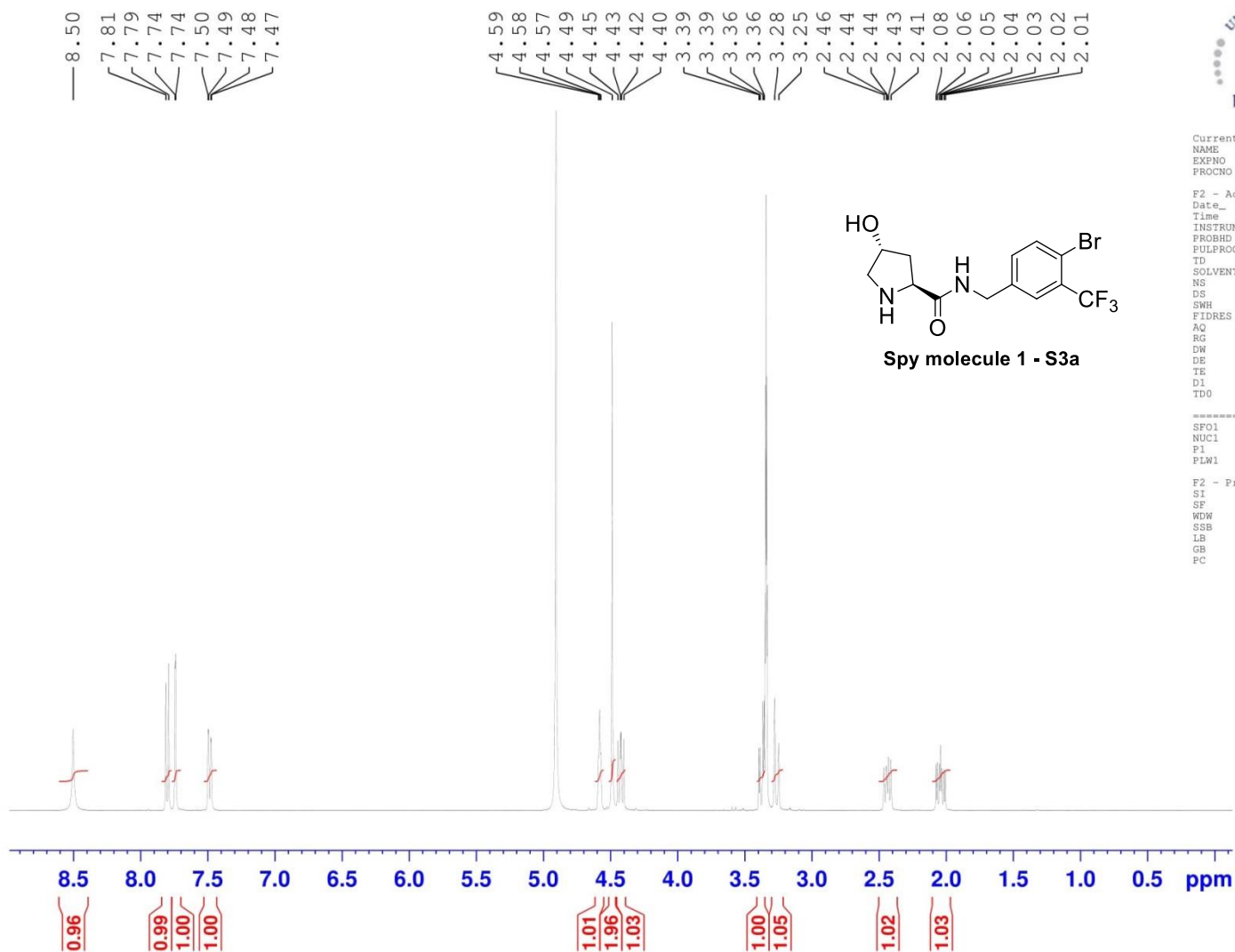
===== CHANNEL f1 =====
 SFO1 470.5453180 MHz
 NUC1 19F
 P1 13.50 usec
 PLW1 25.00000000 W

===== CHANNEL f2 =====
 SFO2 500.1320005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 80.00 usec
 PLW2 25.00000000 W
 PLW12 0.39063001 W

F2 - Processing parameters
 SI 65536
 SF 470.5923772 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

SUPPORTING INFORMATION

Spy molecule 1 (Compound S3a)



Current Data Parameters
 NAME ALC-GC-R028-HPLC
 EXPNO 10
 PROCNO 1

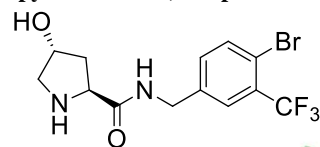
F2 - Acquisition Parameters
 Date_ 20160202
 Time 19.28
 INSTRUM spect
 PROBHD 5 mm PADUL13C
 PULPROG zg30
 TD 131072
 SOLVENT MeOD
 NS 320
 DS 4
 SWH 12019.230 Hz
 FIDRES 0.091699 Hz
 AQ 5.4525952 sec
 RG 196.14
 DW 41.600 usec
 DE 12.17 usec
 TE 298.2 K
 D1 0.10000000 sec
 TDO 1

===== CHANNEL f1 =====
 SF01 400.1324710 MHz
 NUC1 1H
 P1 10.00 usec
 PLW1 20.00000000 W

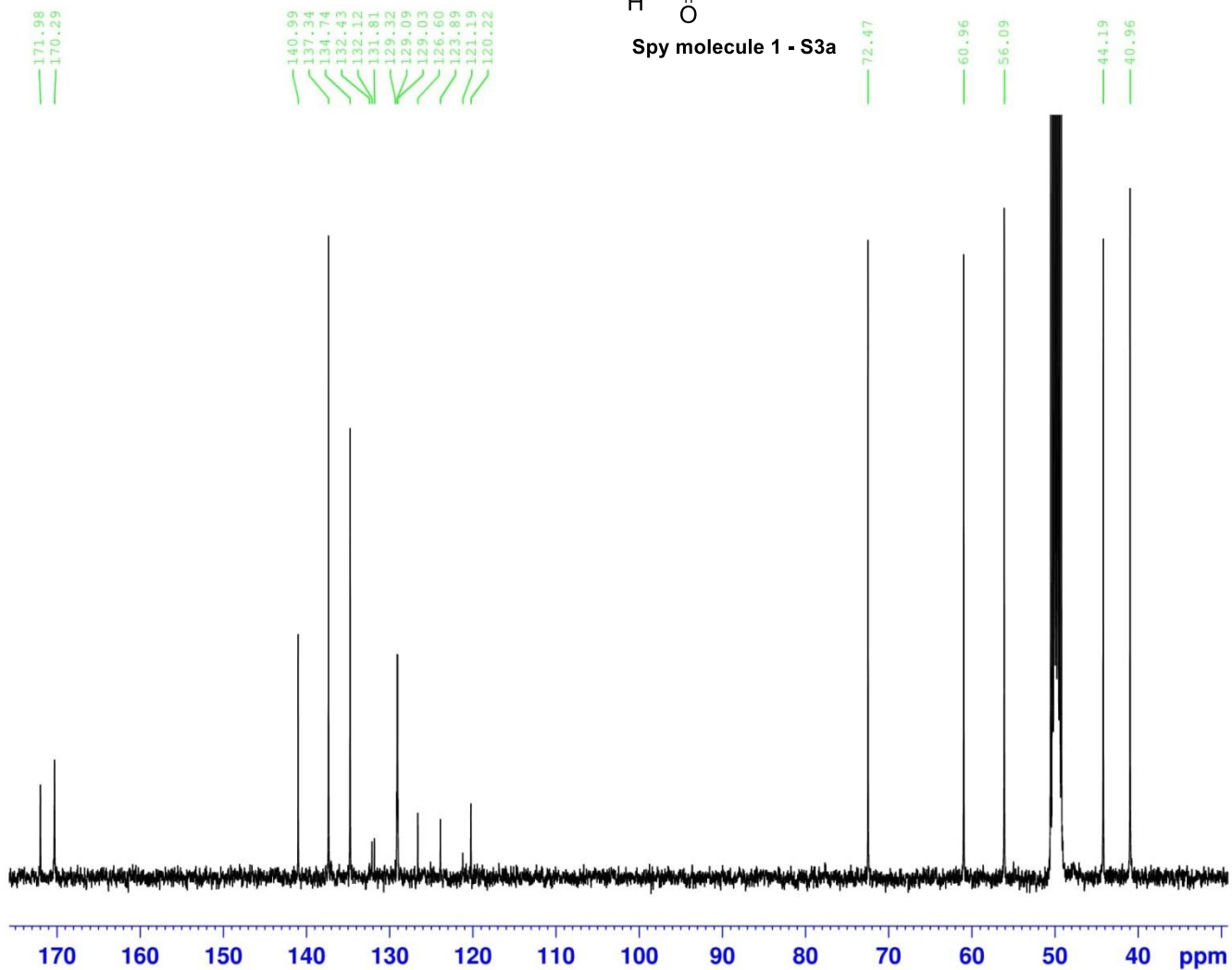
F2 - Processing parameters
 SI 131072
 SF 400.1299957 MHz
 WDW EM
 SSB 0
 LB 0.10 Hz
 GB 0
 PC 1.00

SUPPORTING INFORMATION

Spy molecule 1 (Compound S3a)



Spy molecule 1 - S3a



```

Current Data Parameters
NAME      ALC-GC-R028-HPLC
EXPNO     11
PROCNO    1

F2 - Acquisition Parameters
Date_     20160203
Time      0.07
INSTRUM   spect
PROBHD    5 mm PADUL 13C
PULPROG   udef1
TD         17996
SOLVENT   MeOD
NS         2000
DS         0
SWH        25000.000 Hz
FIDRES     1.389198 Hz
AQ         0.3599200 sec
RG         196.14
DW         20.000 usec
DE         8.66 usec
TE         298.2 K
D1         3.00000000 sec
D11        0.03000000 sec
D12        0.00002000 sec
D20        200.00000000 sec
TD0        1

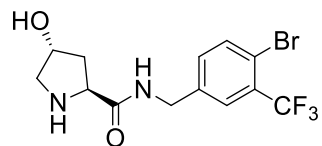
===== CHANNEL f1 =====
SF01      100.6238346 MHz
NUC1       13C
P1         10.00 usec
P13        2000.00 usec
P26        500.00 usec
PLW1       36.00000000 W
SPNAM[5]   Crp60comp.4
SPOAL5     0.500
SPOFFS5    0 Hz
SPW5       5.50040007 W
SPNAM[8]   Crp60,0.5,20.1
SPOAL8     0.500
SPOFFS8    0 Hz
SPW8       5.50040007 W

===== CHANNEL f2 =====
SF02      400.1316005 MHz
NUC2       1H
CPDPRG[2]  waltz64
PCPD2      90.00 usec
PLW2       20.00000000 W
PLW12      0.24691001 W

F2 - Processing parameters
SI         262144
SF         100.6125411 MHz
WDW        EM
SSB        0
LB         2.00 Hz
GB         0
PC         1.40
    
```

SUPPORTING INFORMATION

Spy molecule 1 (Compound S3a)



Spy molecule 1 - S3a



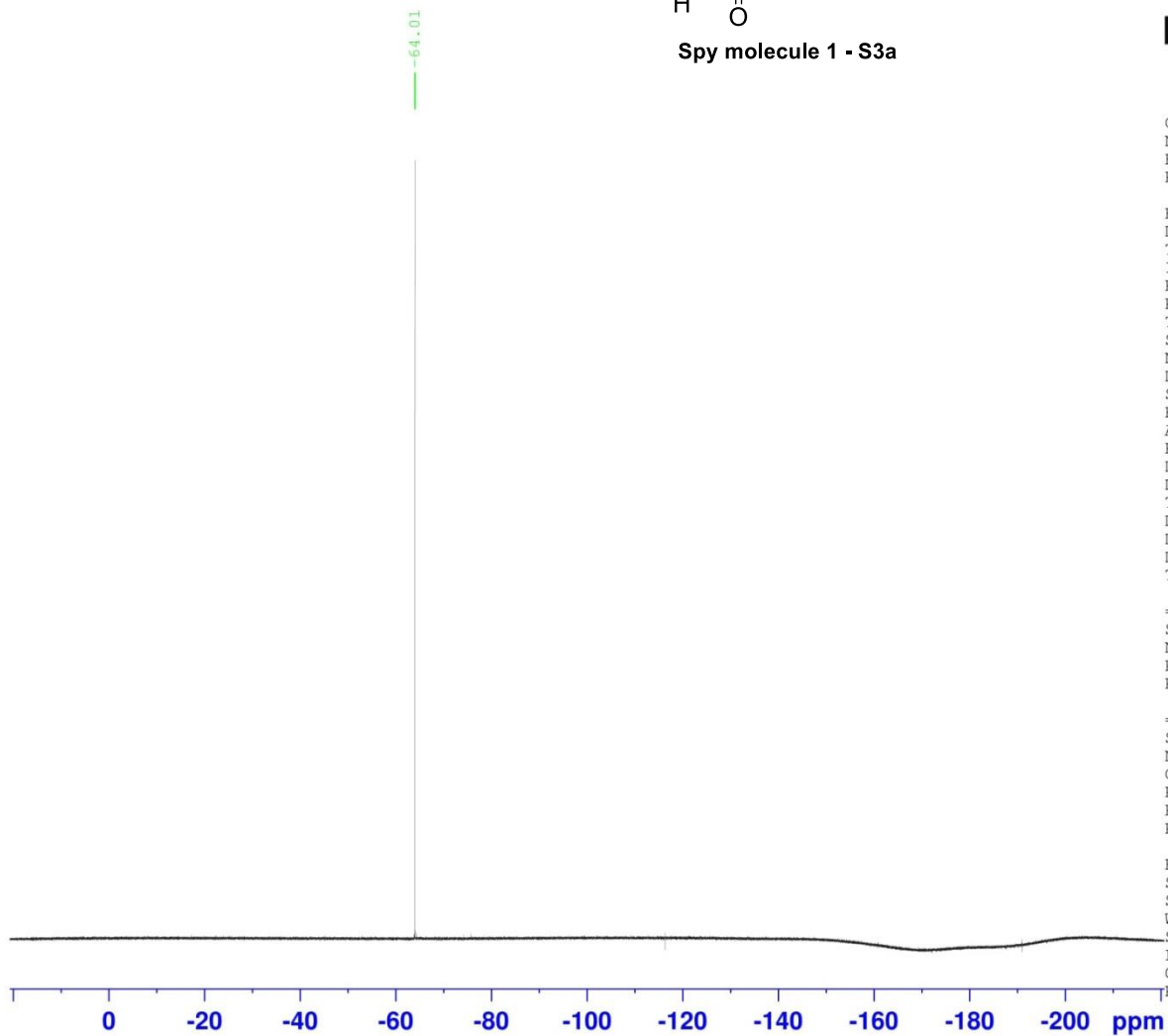
Current Data Parameters
 NAME AC-GC-R028
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20160125
 Time 9.49
 INSTRUM spect
 PROBHD 5 mm QNP 1H/13
 PULPROG zgfhigqn.2
 TD 131072
 SOLVENT MeOD
 NS 64
 DS 4
 SWH 113636.367 Hz
 FIDRES 0.866977 Hz
 AQ 0.5767168 sec
 RG 456
 DW 4.400 usec
 DE 6.50 usec
 TE 298.2 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 D12 0.00002000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 470.5453180 MHz
 NUC1 19F
 P1 11.50 usec
 PLW1 25.00000000 W

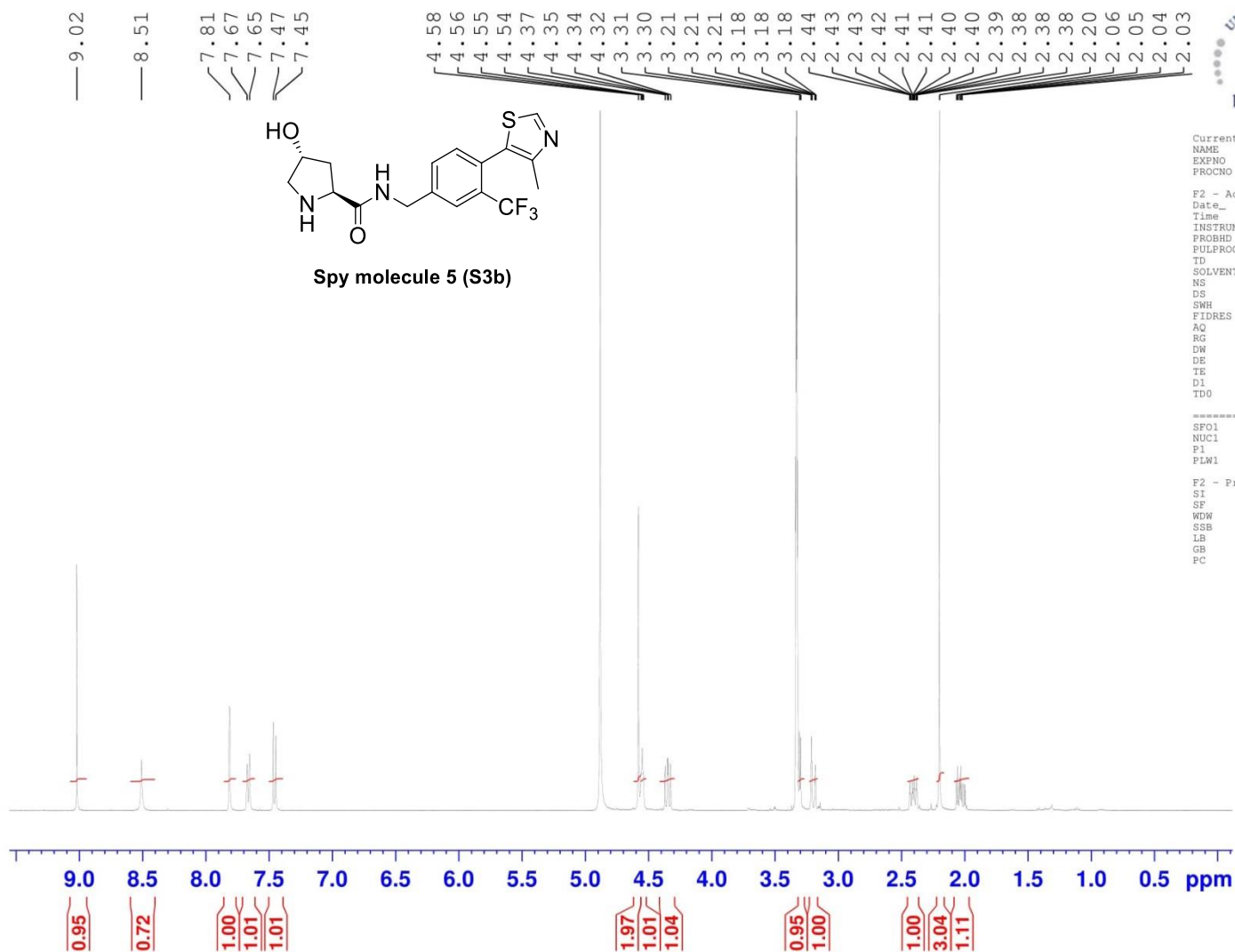
===== CHANNEL f2 =====
 SFO2 500.1320005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 80.00 usec
 PLW2 25.00000000 W
 PLW12 0.39063001 W

F2 - Processing parameters
 SI 65536
 SF 470.5923772 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



SUPPORTING INFORMATION

Spy molecule 5 (Compound S3b)



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Current Data Parameters
 NAME ALC-GC-R038
 EXPNO 1
 PROCNO 1

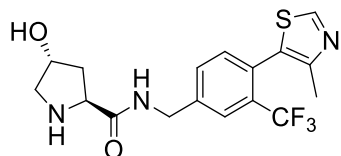
F2 - Acquisition Parameters
 Date_ 20160315
 Time 23.01
 INSTRUM spect
 PROBHD 5 mm PADUL13C
 PULPROG zg30
 ID 131072
 SOLVENT MeOD
 NS 600
 DS 4
 SWH 12019.230 Hz
 FIDRES 0.091699 Hz
 AQ 5.4525952 sec
 RG 196.14
 DW 41.600 usec
 DE 12.17 usec
 TE 298.2 K
 D1 0.1000000 sec
 TDO 1

===== CHANNEL f1 =====
 SF01 400.1324710 MHz
 NUC1 1H
 P1 10.00 usec
 PLW1 20.00000000 W

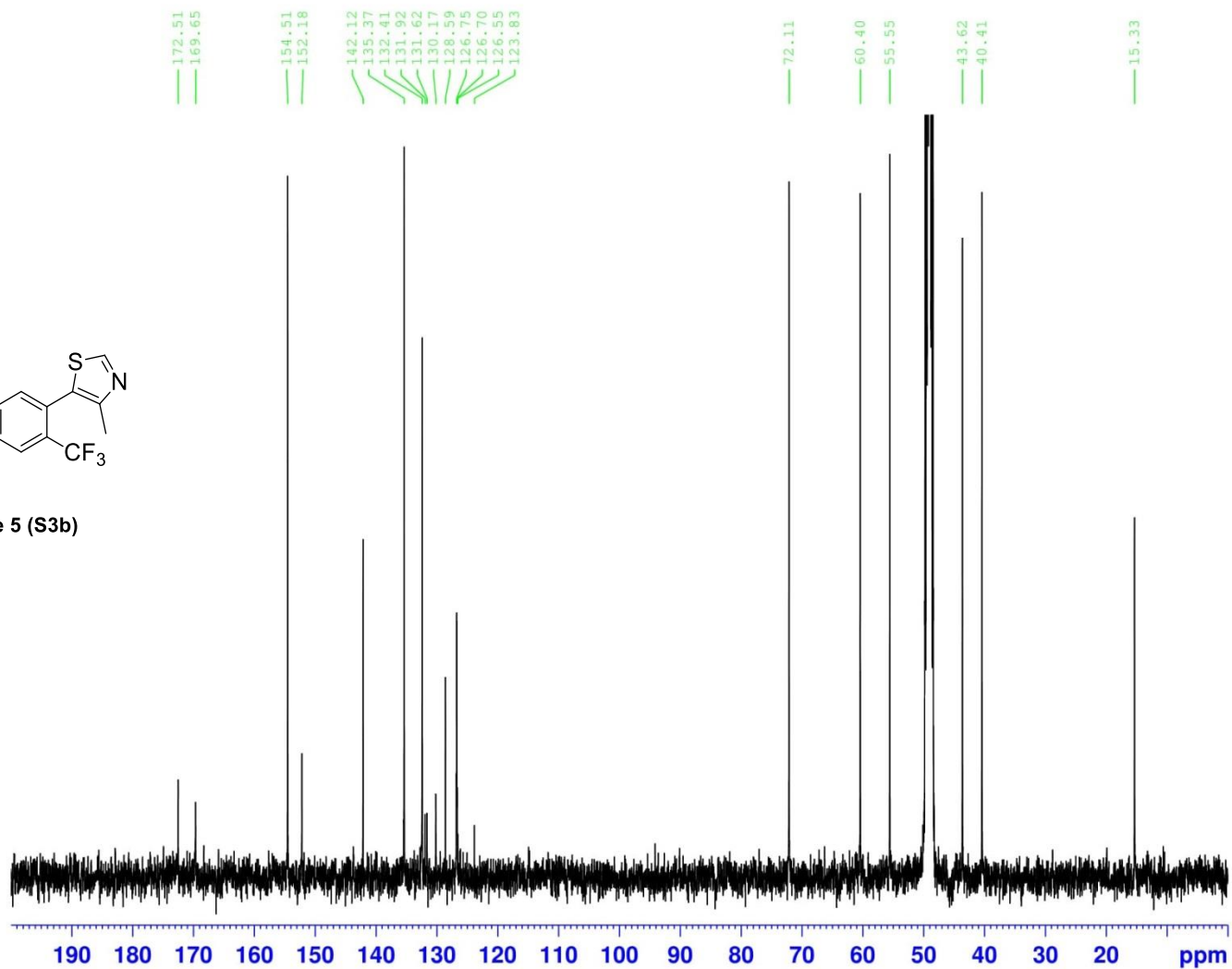
F2 - Processing parameters
 SI 131072
 SF 400.1300000 MHz
 WDW EM
 SSB 0
 LB 0.10 Hz
 GB 0
 PC 1.00

SUPPORTING INFORMATION

Spy molecule 5 (Compound S3b)



Spy molecule 5 (S3b)



```

Current Data Parameters
NAME      ALC-GC-R038
EXPNO     2
PROCNO    1

F2 - Acquisition Parameters
Date_     20160316
Time      5.10
INSTRUM   spect
PROBHD    5 mm PADUL 13C
PULPROG   udef1
TD         17996
SOLVENT   MeOD
NS         5800
DS         0
SWH        25000.000 Hz
FIDRES     1.389198 Hz
AQ         0.3599200 sec
RG         196.14
DW         20.000 usec
DE         8.66 usec
TE         298.2 K
D1         3.00000000 sec
D11        0.03000000 sec
D12        0.00002000 sec
D20        200.00000000 sec
TD0        1

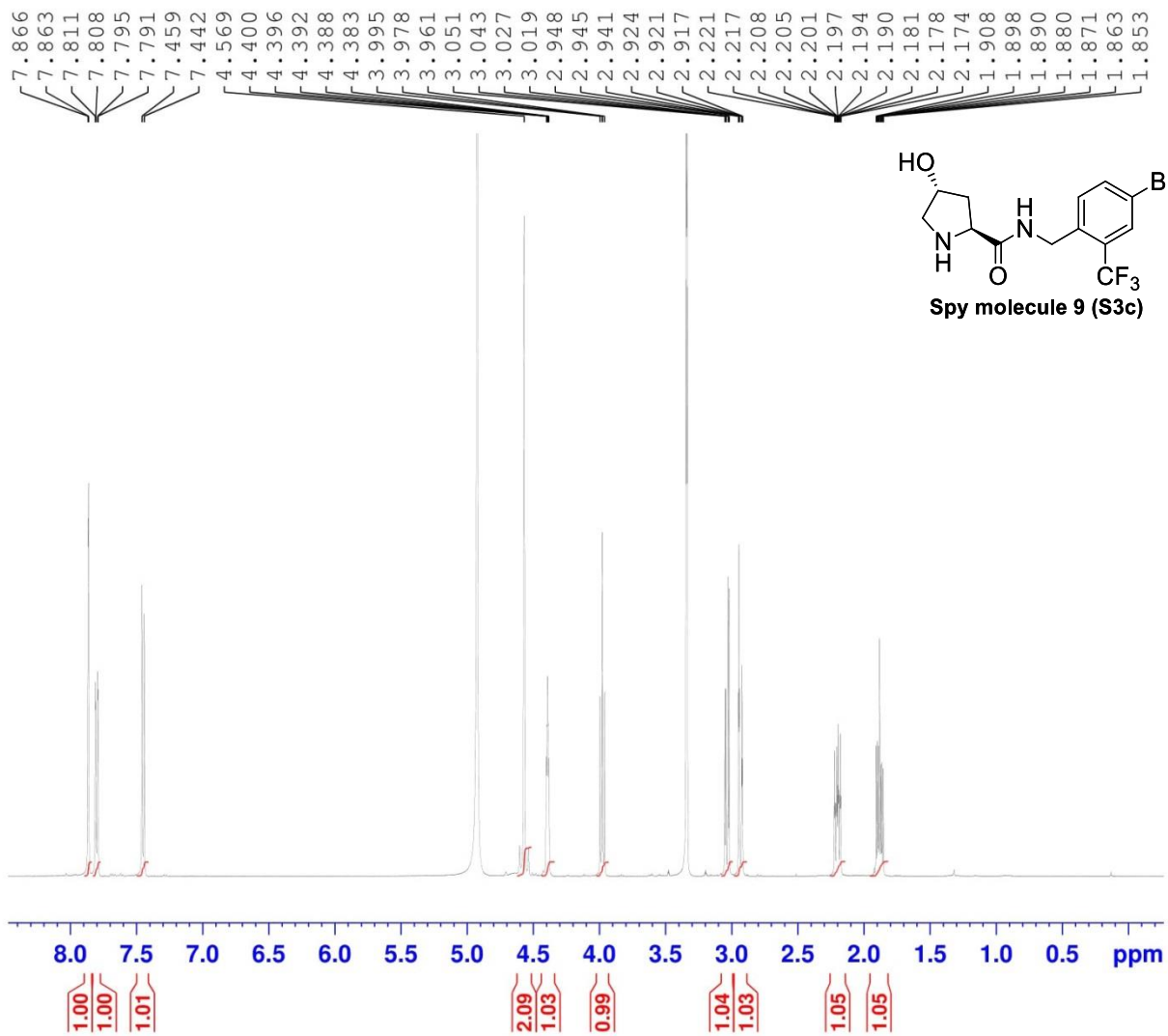
===== CHANNEL f1 =====
SF01      100.6238346 MHz
NUC1       13C
P1         10.00 usec
P13        2000.00 usec
P26         500.00 usec
PLW1       36.00000000 W
SPNAM[5]   Crp60comp.4
SPOAL5     0.500
SPOFFS5    0 Hz
SPW5       5.50040007 W
SPNAM[8]   Crp60,0.5,20.1
SPOAL8     0.500
SPOFFS8    0 Hz
SPW8       5.50040007 W

===== CHANNEL f2 =====
SF02      400.1316005 MHz
NUC2       1H
CPDPRG[2]  waltz64
PCPD2      90.00 usec
PLW2       20.00000000 W
PLW12      0.24691001 W

F2 - Processing parameters
SI         262144
SF         100.6126170 MHz
WDW        EM
SSB        0
LB         2.00 Hz
GB         0
PC         1.40
    
```


SUPPORTING INFORMATION

Spy molecule 9 (Compound S3c)



Current Data Parameters
 NAME GCF09
 EXPNO 28
 PROCNO 1

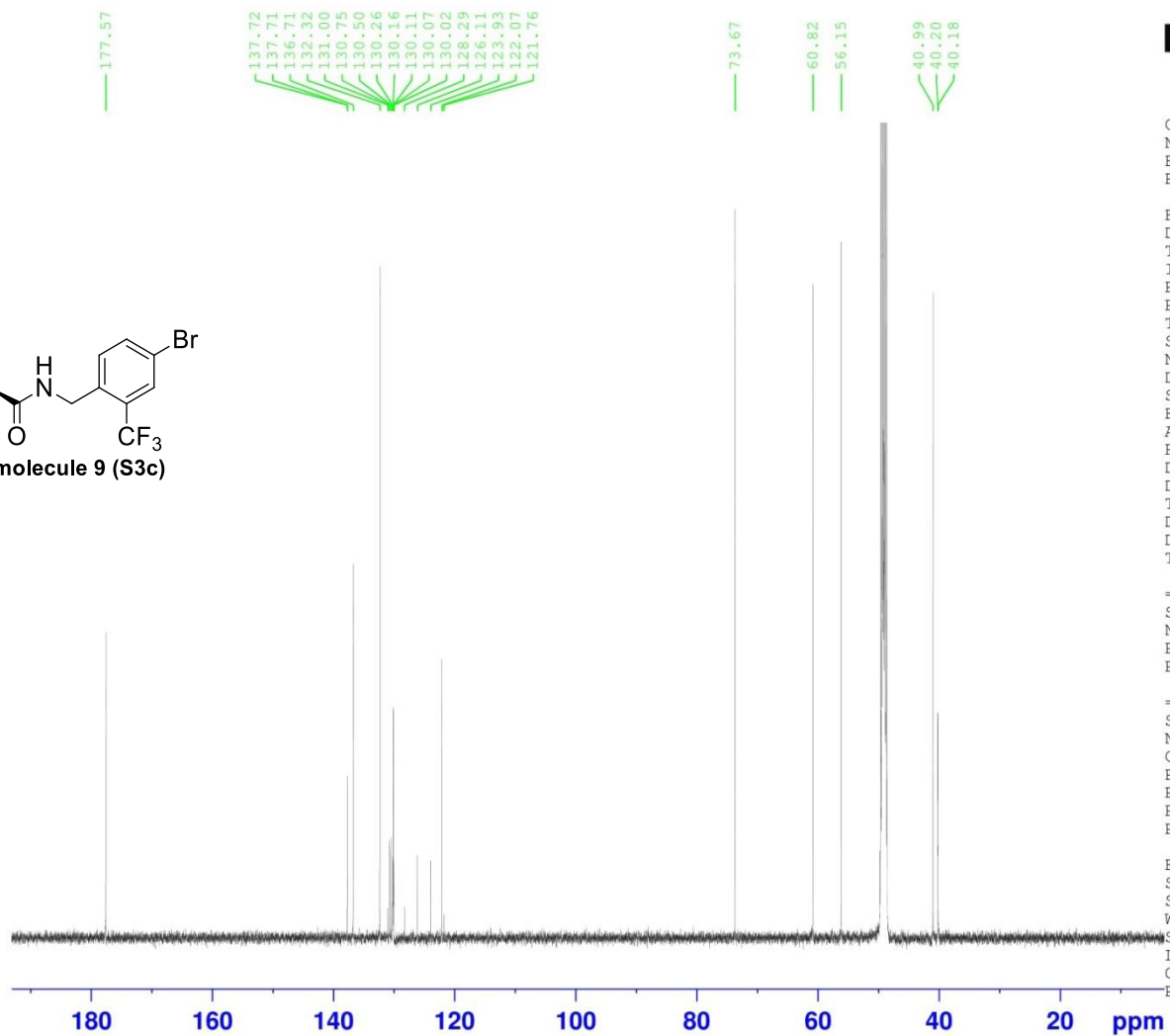
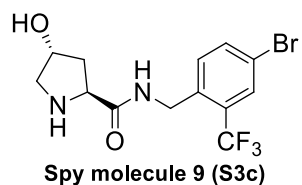
F2 - Acquisition Parameters
 Date_ 20181113
 Time 23.38
 INSTRUM spect
 PROBHD 5 mm CPQCI 1H/
 PULPROG zg30
 TD 65536
 SOLVENT MeOD
 NS 16
 DS 2
 SWH 10000.000 Hz
 FIDRES 0.152588 Hz
 AQ 3.2767999 sec
 RG 151.47
 DW 50.000 usec
 DE 10.00 usec
 TE 293.2 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 500.1330885 MHz
 NUC1 1H
 P1 11.00 usec
 PLW1 5.19999981 W

F2 - Processing parameters
 SI 131072
 SF 500.1299947 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

SUPPORTING INFORMATION

Spy molecule 9 (Compound S3c)



Current Data Parameters
 NAME GCF09
 EXPNO 31
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20181116
 Time 14.33
 INSTRUM spect
 PROBHD 5 mm CPQCI 1H/
 PULPROG zgpg30
 TD 65400
 SOLVENT MeOD
 NS 4000
 DS 4
 SWH 29761.904 Hz
 FIDRES 0.455075 Hz
 AQ 1.0987200 sec
 RG 190.78
 DW 16.800 usec
 DE 18.00 usec
 TE 293.1 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TD0 1

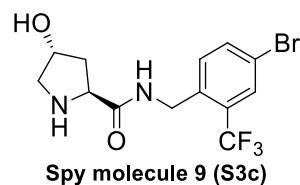
===== CHANNEL f1 =====
 SFO1 125.7703637 MHz
 NUC1 13C
 P1 12.00 usec
 PLW1 133.00000000 W

===== CHANNEL f2 =====
 SFO2 500.1320005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 100.00 usec
 PLW2 5.19999981 W
 PLW12 0.06406900 W
 PLW13 0.06406900 W

F2 - Processing parameters
 SI 131072
 SF 125.7575994 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

SUPPORTING INFORMATION

Spy molecule 9 (Compound S3c)



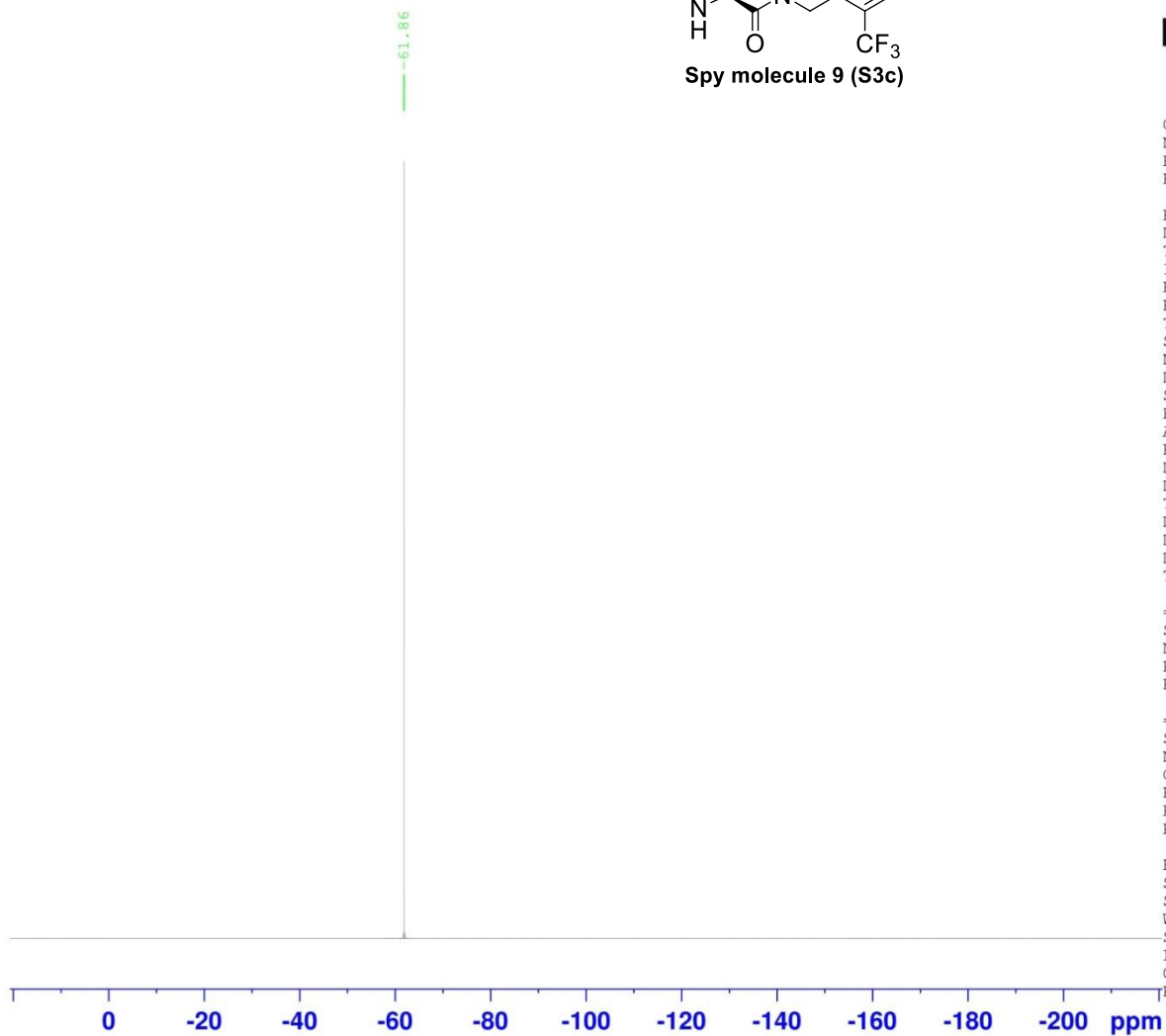
Current Data Parameters
 NAME GCF09
 EXPNO 11
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20181113
 Time 22.37
 INSTRUM spect
 PROBHD 5 mm CPQCI 1H/
 PULPROG zgfhigqn.2
 TD 131072
 SOLVENT MeOD
 NS 256
 DS 4
 SWH 113636.367 Hz
 FIDRES 0.866977 Hz
 AQ 0.5767168 sec
 RG 190.78
 DW 4.400 usec
 DE 18.00 usec
 TE 293.2 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 D12 0.00002000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 470.5453180 MHz
 NUC1 19F
 P1 12.00 usec
 PLW1 5.19999981 W

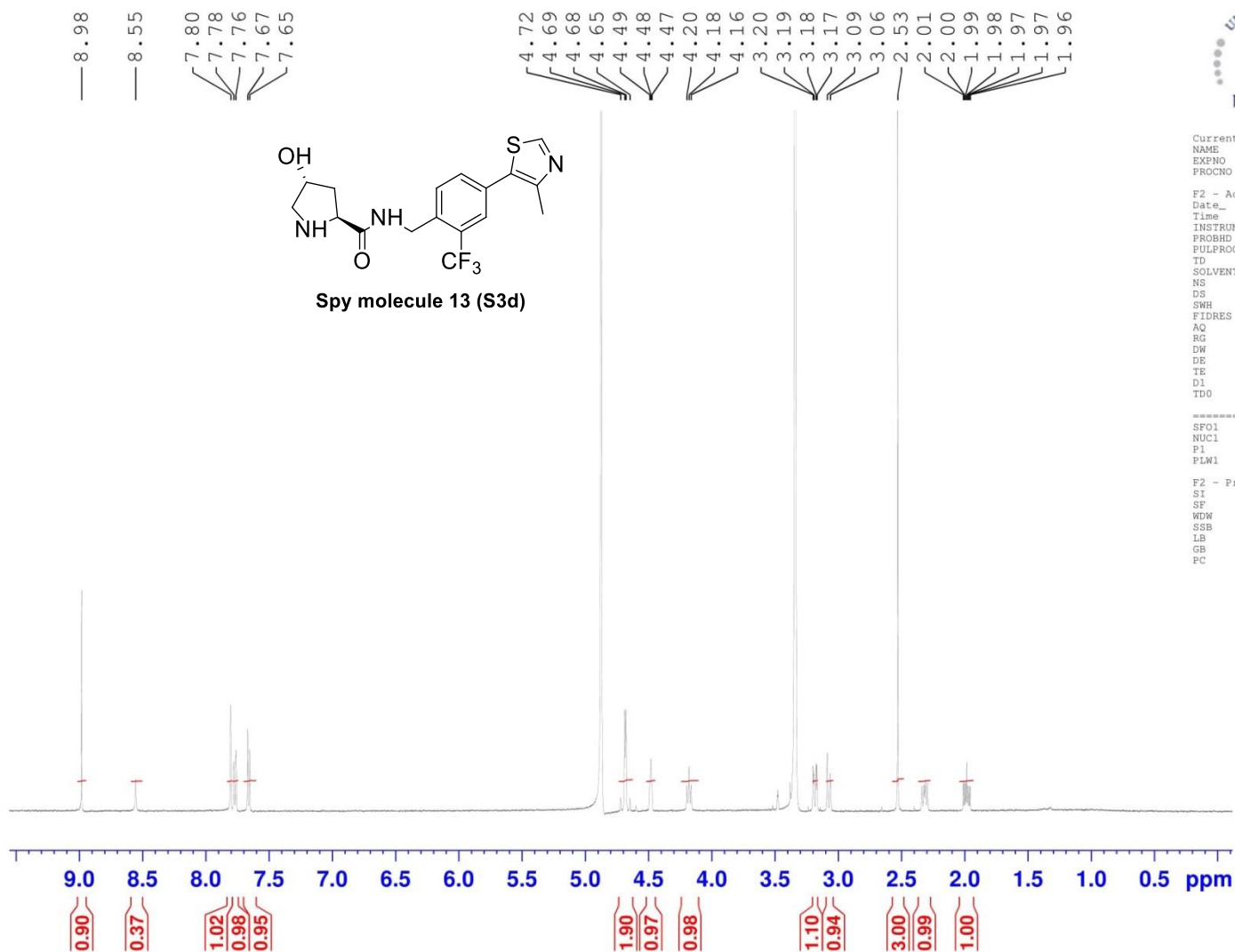
===== CHANNEL f2 =====
 SFO2 500.1320005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 100.00 usec
 PLW2 5.19999981 W
 PLW12 0.06406900 W

F2 - Processing parameters
 SI 262144
 SF 470.5923772 MHz
 WDW EM
 SSB 0
 LB 1.50 Hz
 GB 0
 PC 1.00



SUPPORTING INFORMATION

Spy molecule 13 (Compound S3d)



```

Current Data Parameters
NAME          GC-R053
EXPNO         1
PROCNO        1

F2 - Acquisition Parameters
Date_         20160530
Time          19.33
INSTRUM       spect
PROBHD        5 mm QNP 1H/13
PULPROG       zg30
TD            65536
SOLVENT       MeOD
NS            256
DS            2
SWH           10000.000 Hz
FIDRES        0.152588 Hz
AQ            3.2767999 sec
RG            32
DW            50.000 usec
DE            6.50 usec
TE            298.2 K
D1            1.00000000 sec
TD0           1

===== CHANNEL f1 =====
SF01          500.1330885 MHz
NUC1          1H
P1            10.00 usec
PLW1          25.00000000 W

F2 - Processing parameters
SI            65536
SF            500.1307361 MHz
WDW           EM
SSB           0
LB            0.30 Hz
GB            0
PC            1.00
    
```

SUPPORTING INFORMATION

Spy molecule 13 (Compound S3d)



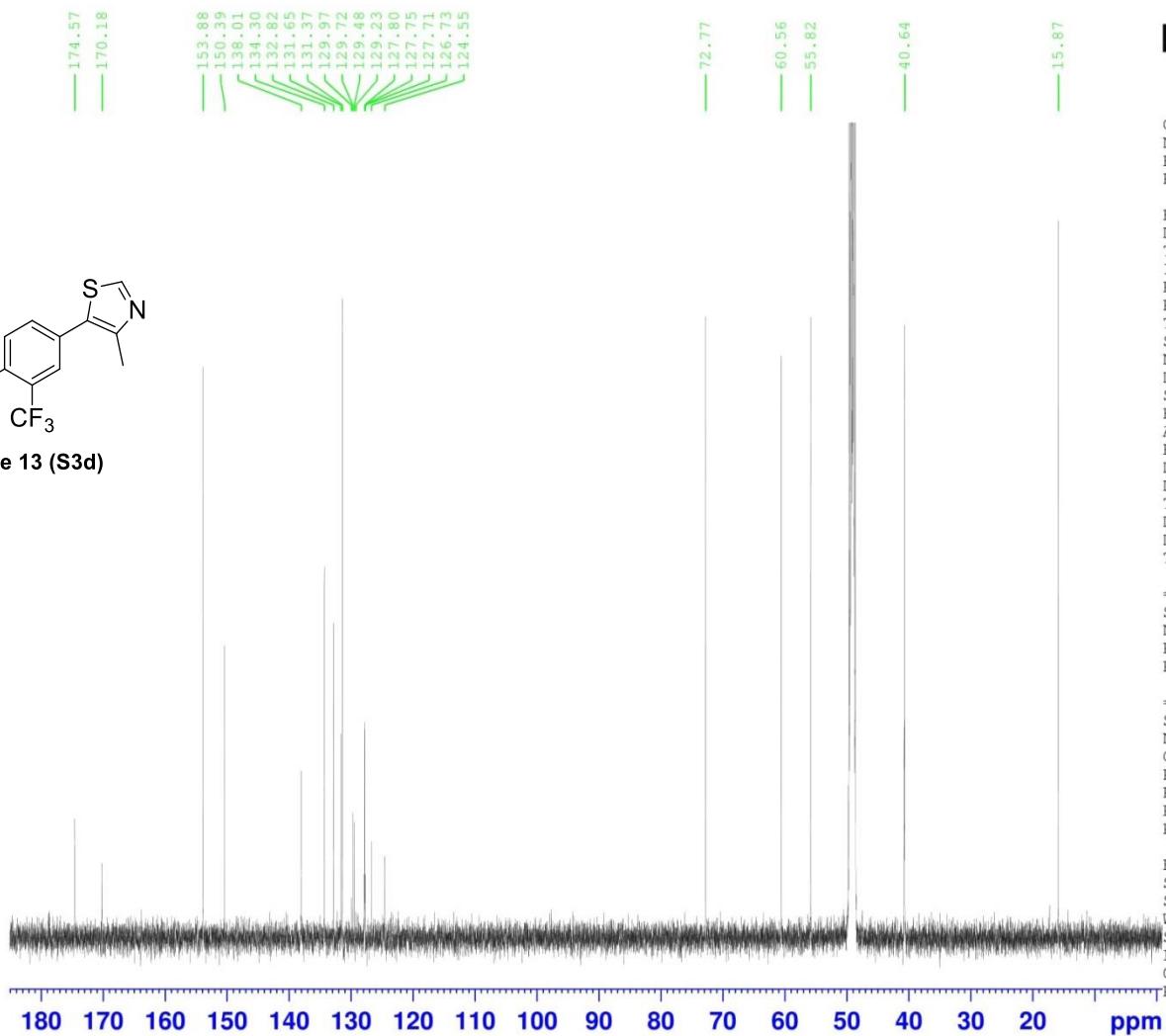
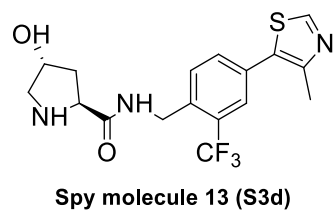
Current Data Parameters
 NAME GCF13
 EXPNO 33
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20181115
 Time 14.20
 INSTRUM spect
 PROBHD 5 mm CPQCI 1H/
 PULPROG zgpg30
 TD 65400
 SOLVENT MeOD
 NS 2048
 DS 4
 SWH 29761.904 Hz
 FIDRES 0.455075 Hz
 AQ 1.0987200 sec
 RG 190.78
 DW 16.800 usec
 DE 18.00 usec
 TE 293.2 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 125.7703637 MHz
 NUC1 13C
 P1 12.00 usec
 PLW1 133.00000000 W

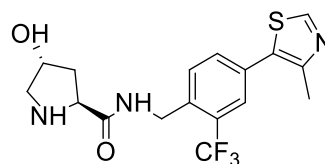
===== CHANNEL f2 =====
 SFO2 500.1320005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 100.00 usec
 PLW2 5.19999981 W
 PLW12 0.06406900 W
 PLW13 0.06406900 W

F2 - Processing parameters
 SI 131072
 SF 125.7575992 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40



SUPPORTING INFORMATION

Spy molecule 13 (Compound S3d)



Spy molecule 13 (S3d)



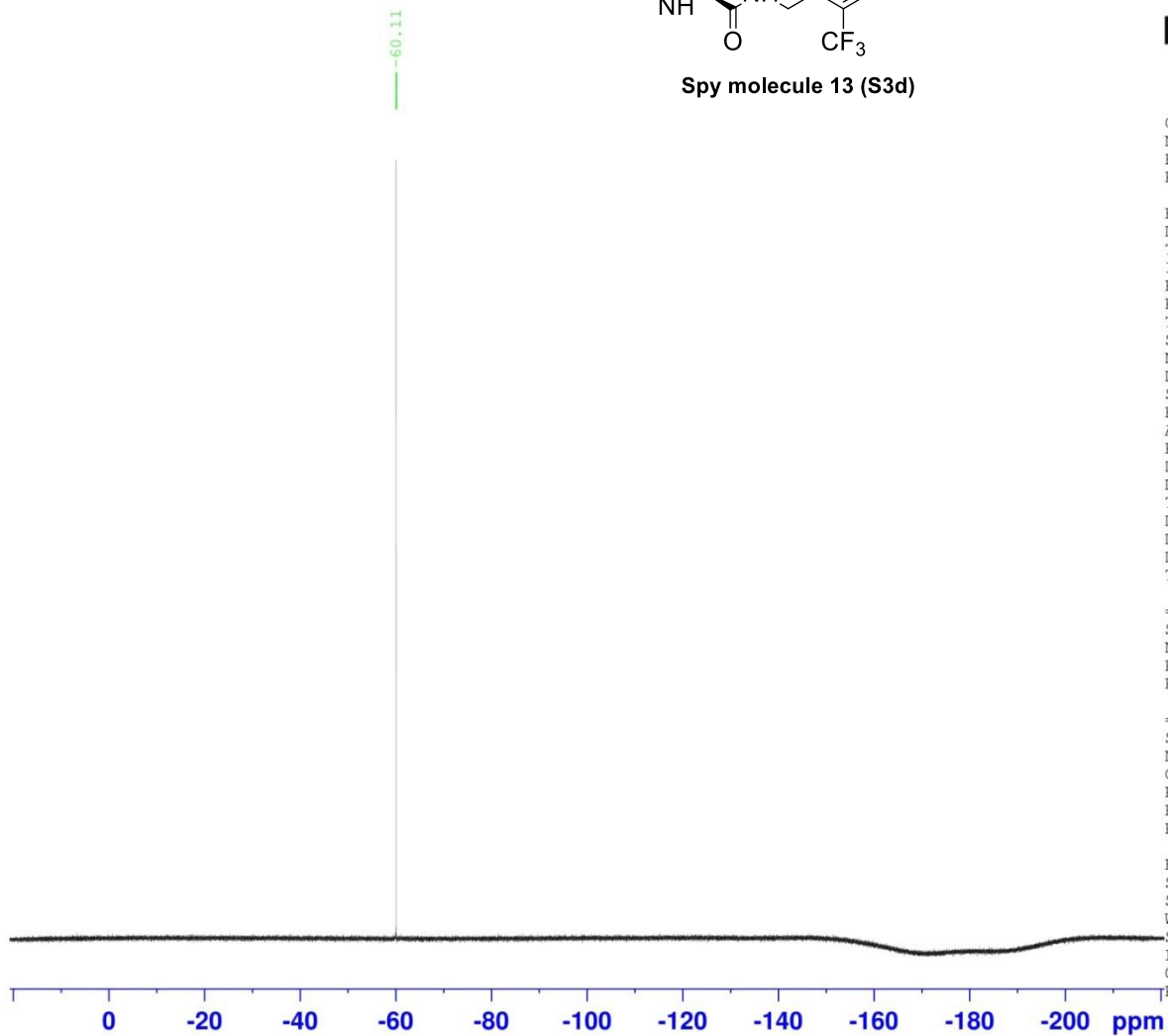
Current Data Parameters
 NAME GC-R053
 EXPNO 4
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20160530
 Time 20.31
 INSTRUM spect
 PROBHD 5 mm QNP 1H/13
 PULPROG zgfhigqn.2
 TD 131072
 SOLVENT MeOD
 NS 16
 DS 4
 SWH 113636.367 Hz
 FIDRES 0.866977 Hz
 AQ 0.5767168 sec
 RG 362
 DW 4.400 usec
 DE 6.50 usec
 TE 298.2 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 D12 0.00002000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 470.5453180 MHz
 NUC1 19F
 P1 13.50 usec
 PLW1 25.00000000 W

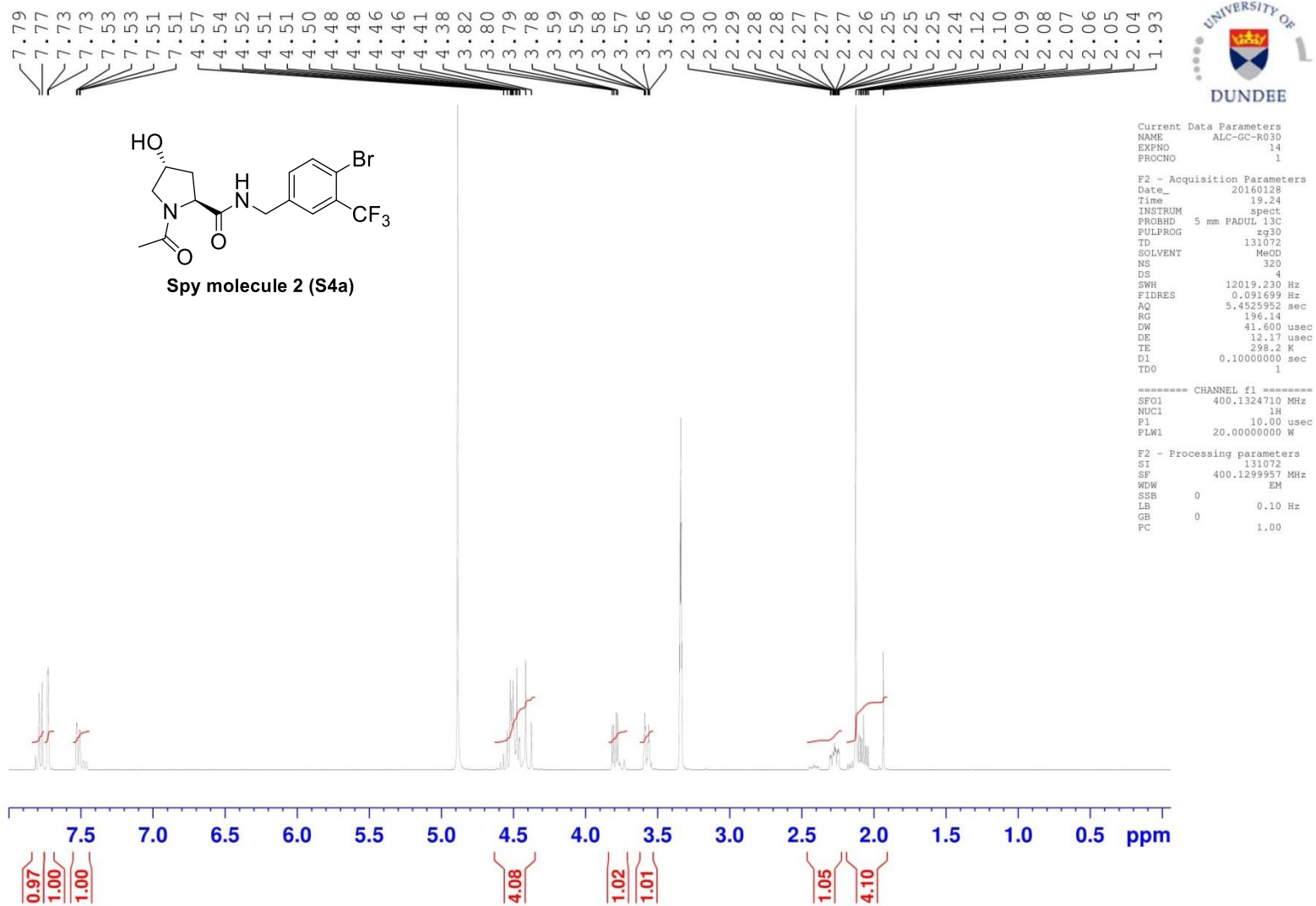
===== CHANNEL f2 =====
 SFO2 500.1320005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 80.00 usec
 PLW2 25.00000000 W
 PLW12 0.39063001 W

F2 - Processing parameters
 SI 65536
 SF 470.5923772 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



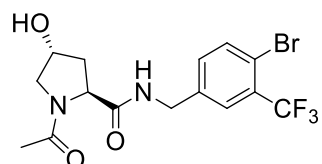
SUPPORTING INFORMATION

Spy molecule 2 (Compound S4a)

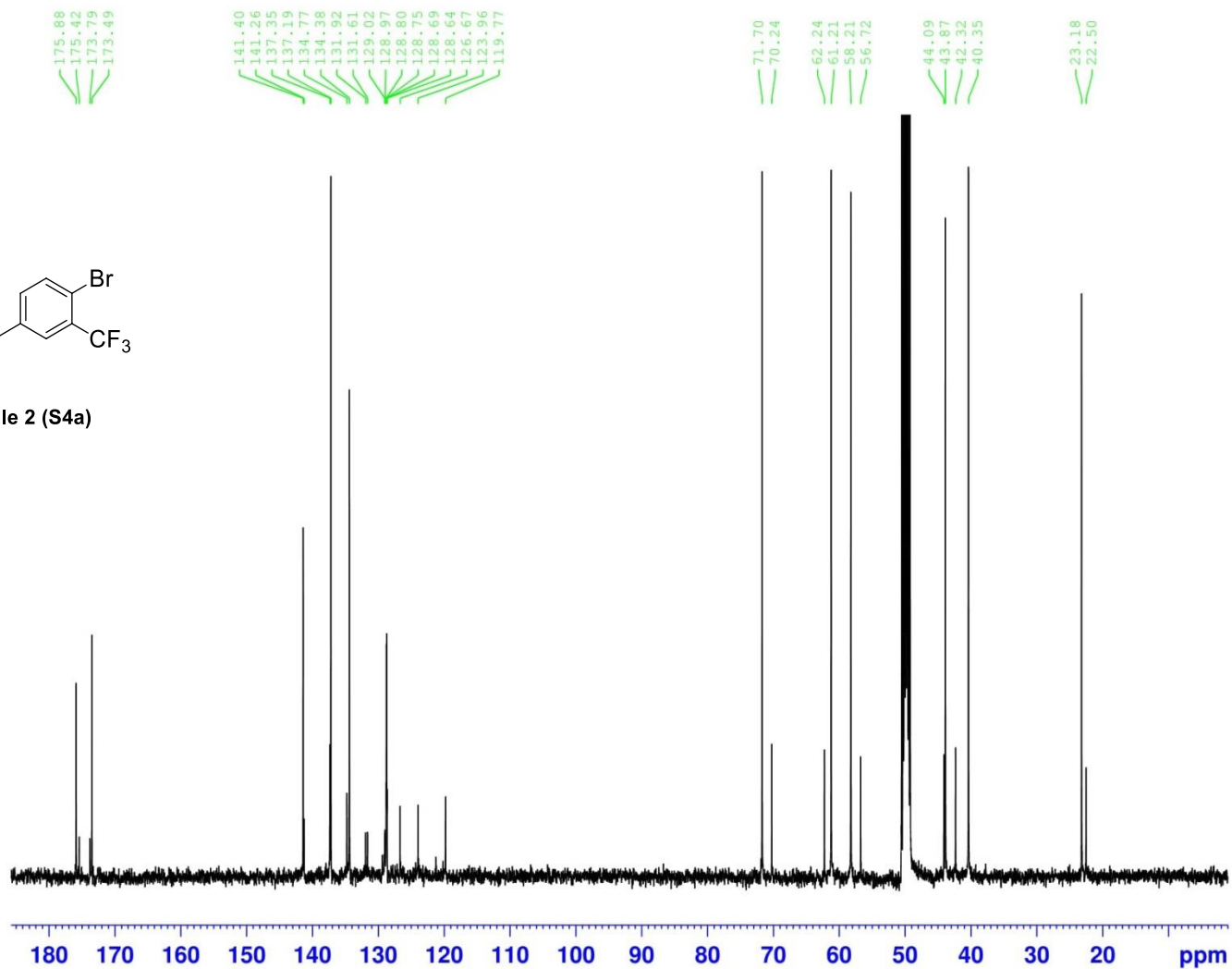


SUPPORTING INFORMATION

Spy molecule 2 (Compound S4a)



Spy molecule 2 (S4a)



```

Current Data Parameters
NAME      ALC-GC-R030
EXPNO    16
PROCNO   1

F2 - Acquisition Parameters
Date_    20160128
Time     22.27
INSTRUM  spect
PROBHD   5 mm PADUL 13C
PULPROG  udef1
TD        17996
SOLVENT  MeOD
NS        2800
DS         0
SWH       25000.000 Hz
FIDRES    1.389198 Hz
AQ         0.3599200 sec
RG         196.14
DW         20.000 usec
DE         8.66 usec
TE        298.2 K
D1         3.00000000 sec
D11        0.03000000 sec
D12        0.00002000 sec
D20        200.00000000 sec
TD0        1

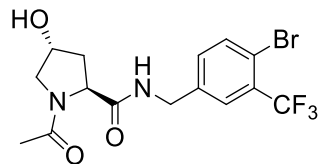
===== CHANNEL f1 =====
SF01      100.6238346 MHz
NUC1       13C
P1         10.00 usec
P13        2000.00 usec
P26         500.00 usec
PLW1       36.00000000 W
SPNAM[5]   Crp60comp.4
SPOAL5     0.500
SPOFFS5    0 Hz
SPW5       5.50040007 W
SPNAM[8]   Crp60,0.5,20.1
SPOAL8     0.500
SPOFFS8    0 Hz
SPW8       5.50040007 W

===== CHANNEL f2 =====
SF02      400.1316005 MHz
NUC2       1H
CPDPRG[2]  waltz64
PCPD2      90.00 usec
PLW2       20.00000000 W
PLW12      0.24691001 W

F2 - Processing parameters
SI         262144
SF         100.6125410 MHz
WDW        EM
SSB         0
LB          2.00 Hz
GB          0
PC          1.40
    
```


SUPPORTING INFORMATION

Spy molecule 2 (Compound S4a)



Spy molecule 2 (S4a)



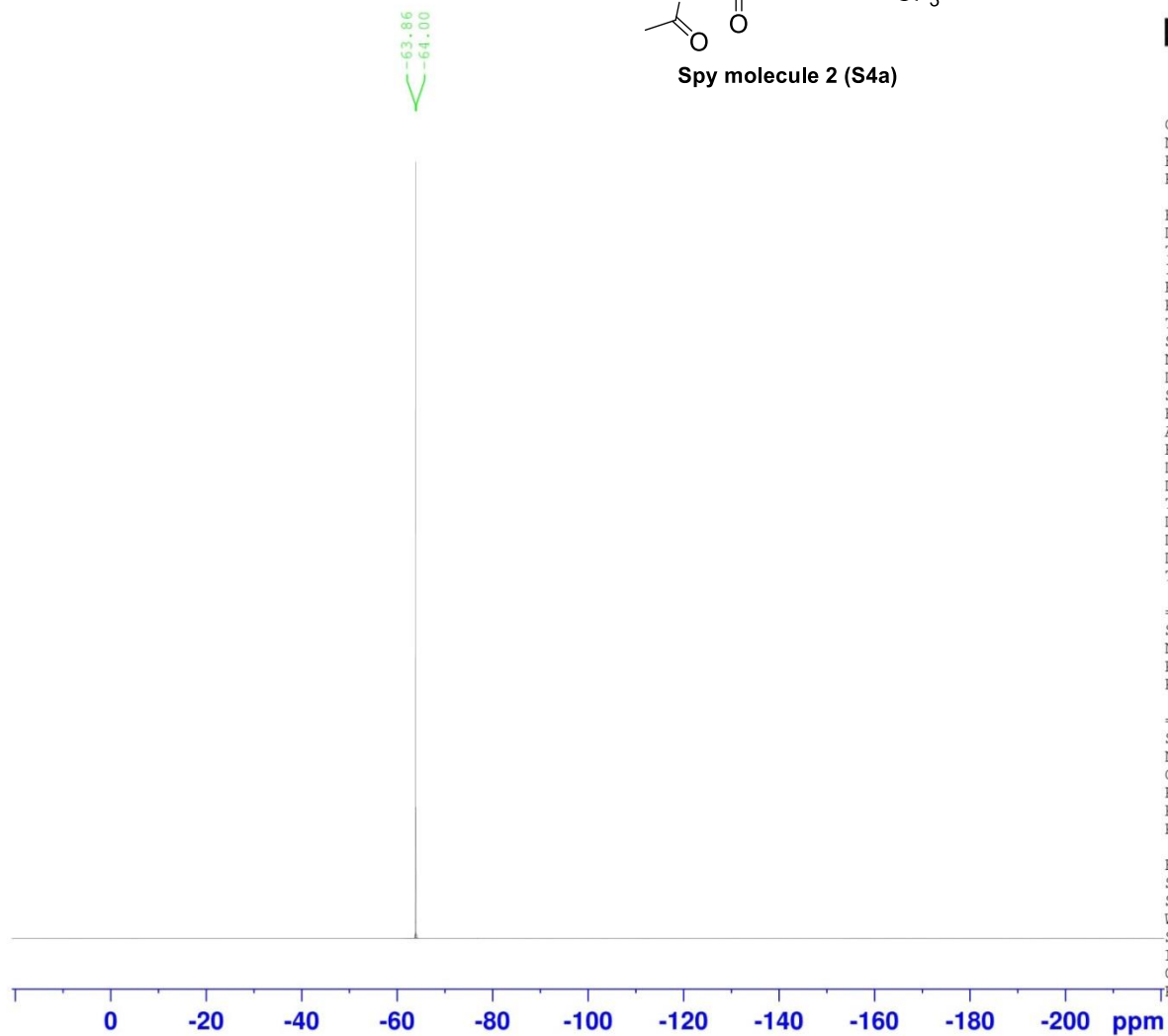
Current Data Parameters
 NAME GCF02 (Correct)
 EXPNO 100
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20181114
 Time 10.35
 INSTRUM spect
 PROBHD 5 mm CPQCI 1H/
 PULPROG zgfhigqn.2
 TD 131072
 SOLVENT MeOD
 NS 256
 DS 4
 SWH 113636.367 Hz
 FIDRES 0.866977 Hz
 AQ 0.5767168 sec
 RG 190.78
 DW 4.400 usec
 DE 18.00 usec
 TE 293.1 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 D12 0.00002000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 470.5453180 MHz
 NUC1 19F
 P1 12.00 usec
 PLW1 5.19999981 W

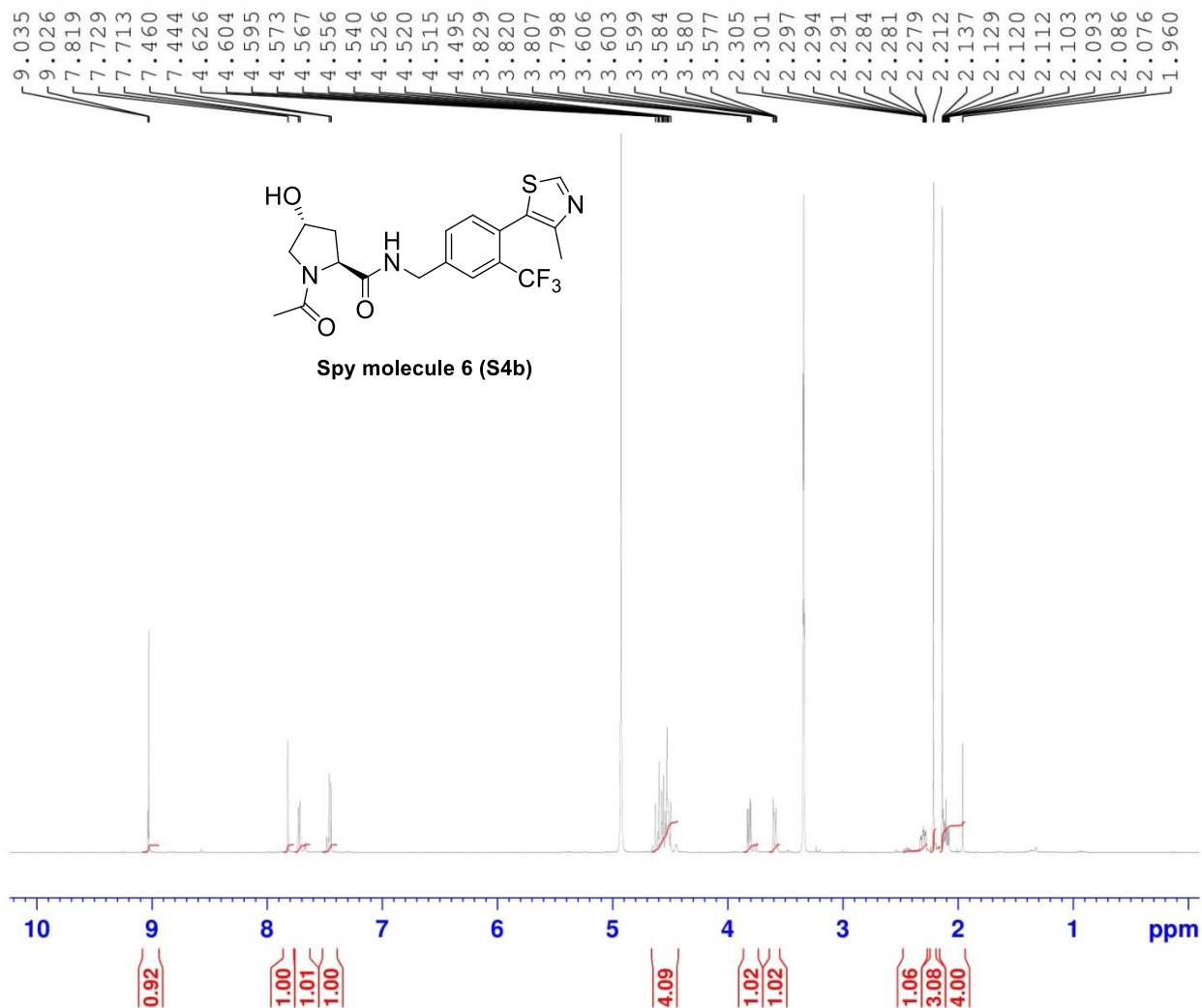
===== CHANNEL f2 =====
 SFO2 500.1320005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 100.00 usec
 PLW2 5.19999981 W
 PLW12 0.06406900 W

F2 - Processing parameters
 SI 262144
 SF 470.5923772 MHz
 WDW EM
 SSB 0
 LB 1.50 Hz
 GB 0
 PC 1.00



SUPPORTING INFORMATION

Spy molecule 6 (Compound S4b)



Current Data Parameters
 NAME GCF06
 EXPNO 11
 PROCNO 1

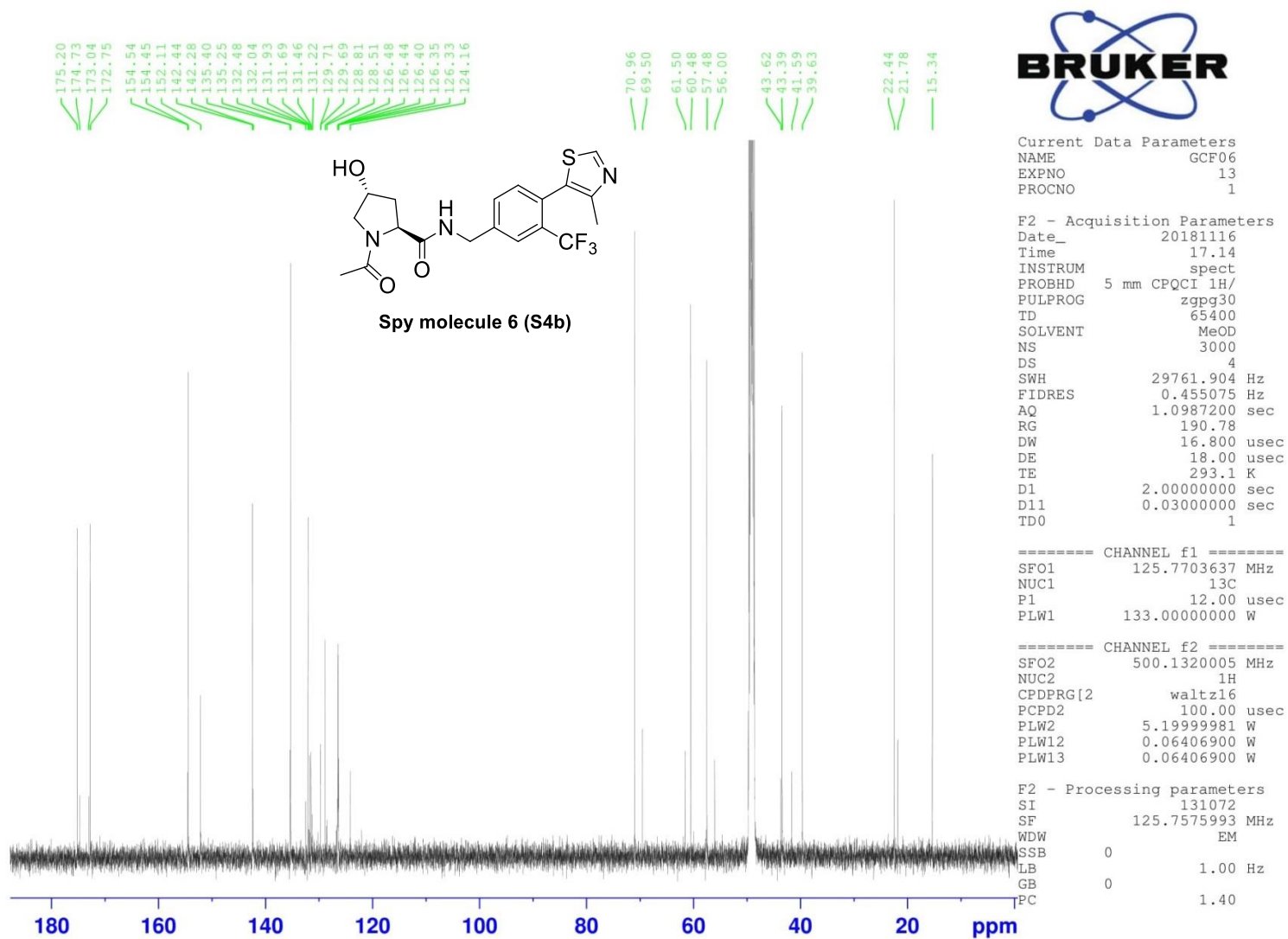
F2 - Acquisition Parameters
 Date_ 20181114
 Time 9.28
 INSTRUM spect
 PROBHD 5 mm CPQCI 1H/
 PULPROG zg30
 TD 65536
 SOLVENT MeOD
 NS 128
 DS 2
 SWH 10000.000 Hz
 FIDRES 0.152588 Hz
 AQ 3.2767999 sec
 RG 137.73
 DW 50.000 usec
 DE 10.00 usec
 TE 293.2 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 500.1330885 MHz
 NUC1 1H
 P1 11.00 usec
 PLW1 5.19999981 W

F2 - Processing parameters
 SI 131072
 SF 500.1299946 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

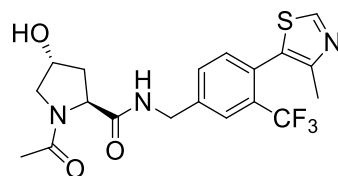
SUPPORTING INFORMATION

Spy molecule 6 (Compound S4b)



SUPPORTING INFORMATION

Spy molecule 6 (Compound S4b)



Spy molecule 6 (S4b)

-60.75
-60.87



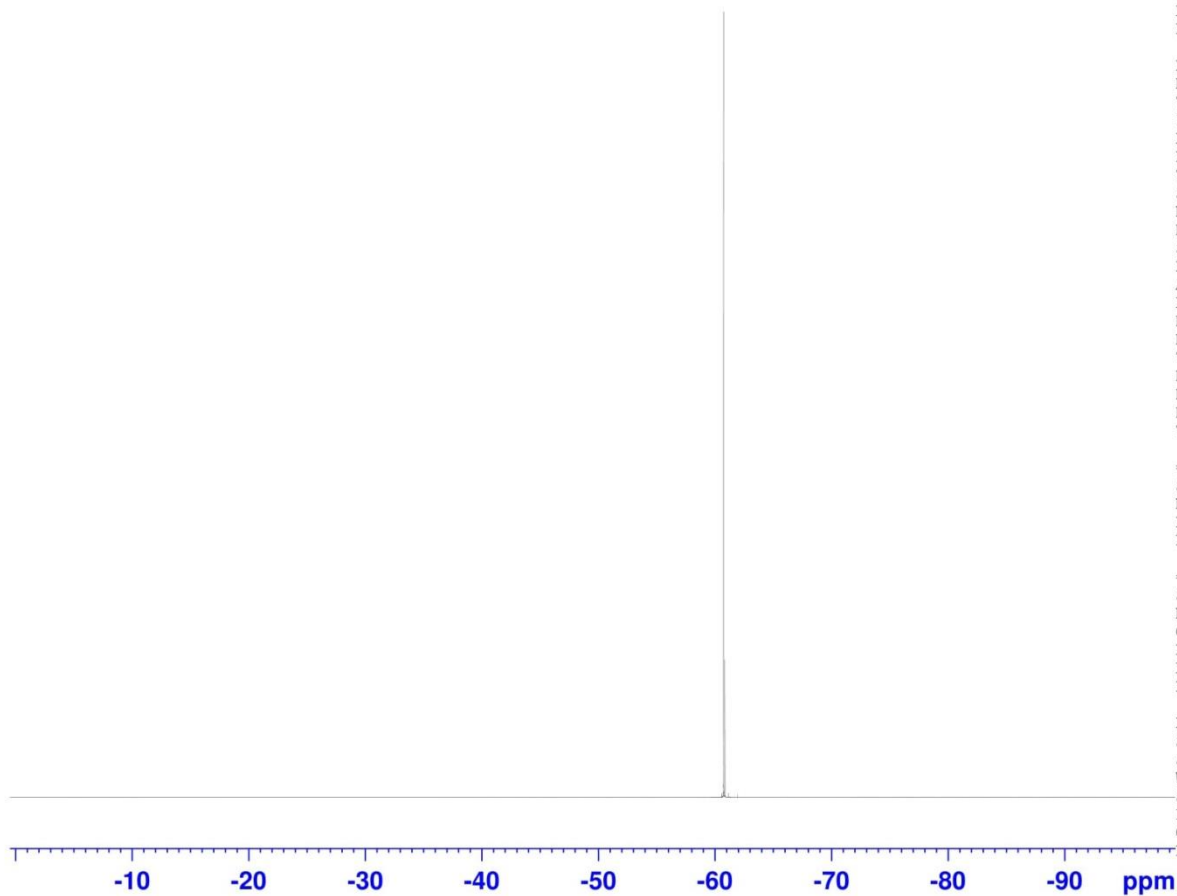
Current Data Parameters
 NAME GCF06
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20181114
 Time 9.18
 INSTRUM spect
 PROBHD 5 mm CPQCI 1H/
 PULPROG zgfhigqn.2
 TD 131072
 SOLVENT MeOD
 NS 256
 DS 4
 SWH 113636.367 Hz
 FIDRES 0.866977 Hz
 AQ 0.5767168 sec
 RG 190.78
 DW 4.400 usec
 DE 18.00 usec
 TE 293.2 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 D12 0.00002000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 470.5453180 MHz
 NUC1 19F
 P1 12.00 usec
 PLW1 5.19999981 W

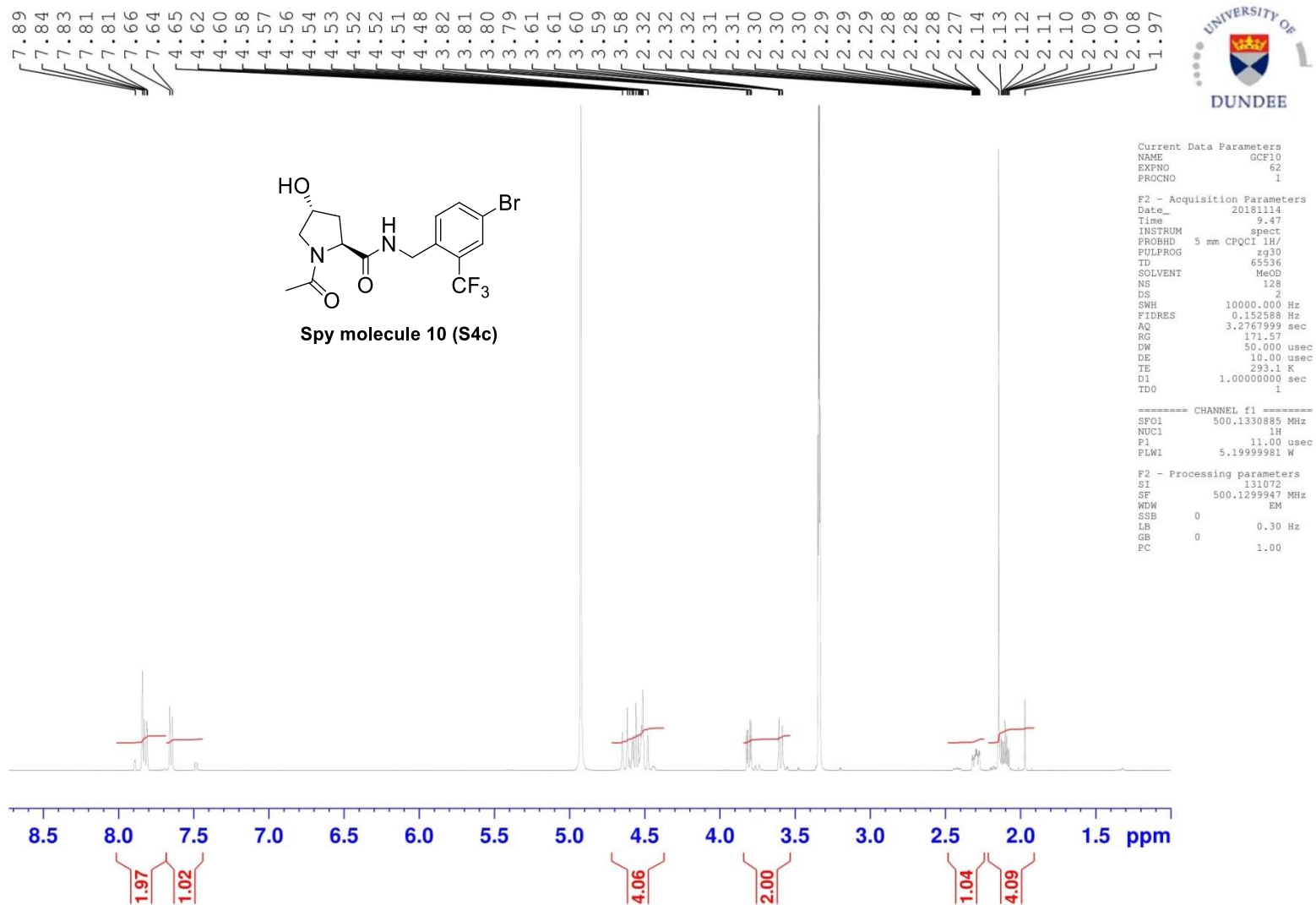
===== CHANNEL f2 =====
 SFO2 500.1320005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 100.00 usec
 PLW2 5.19999981 W
 PLW12 0.06406900 W

F2 - Processing parameters
 SI 262144
 SF 470.5923772 MHz
 WDW EM
 SSB 0
 LB 1.50 Hz
 GB 0
 PC 1.00



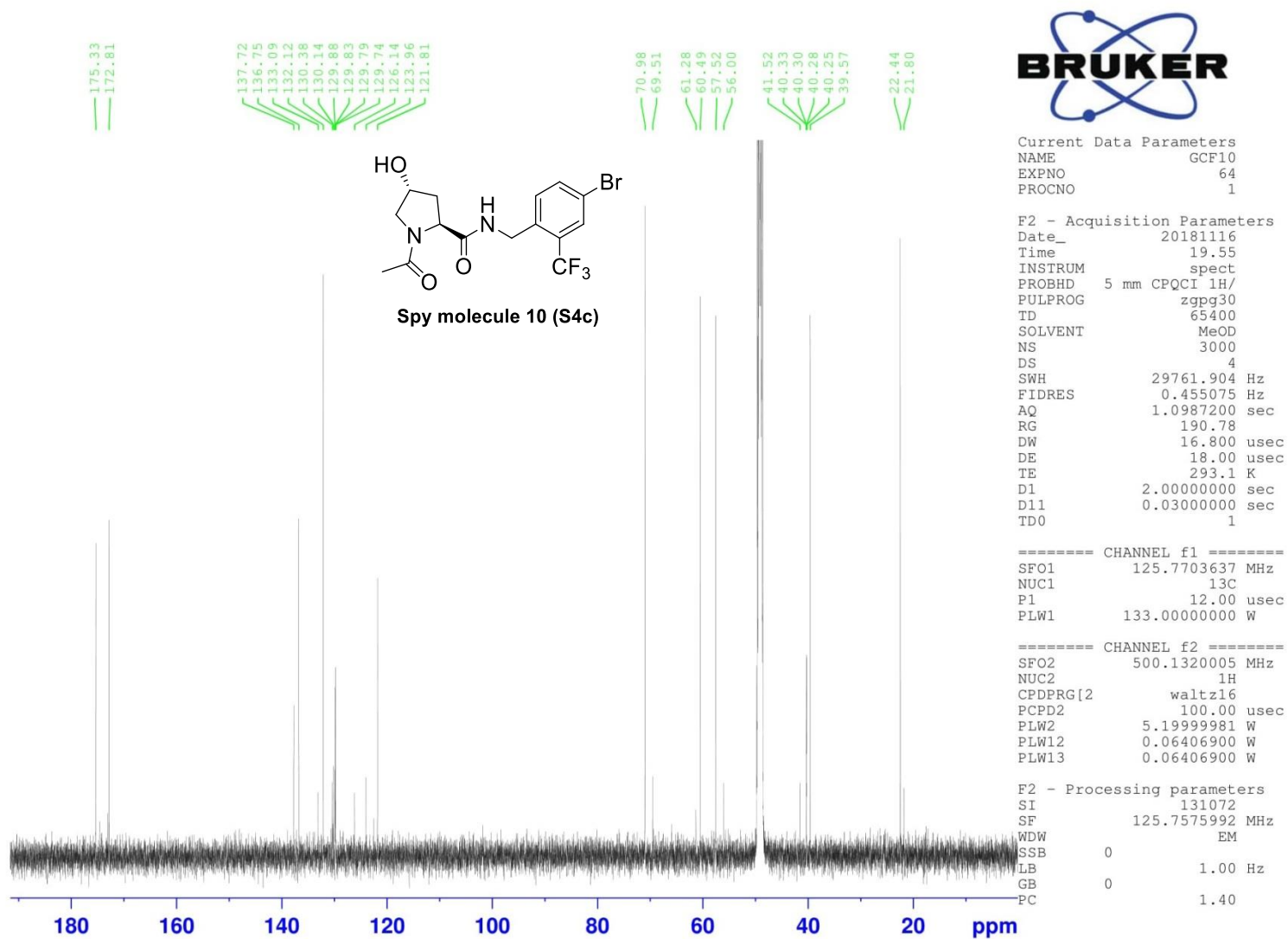
SUPPORTING INFORMATION

Spy molecule 10 (Compound S4c)



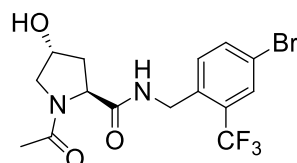
SUPPORTING INFORMATION

Spy molecule 10 (Compound S4c)

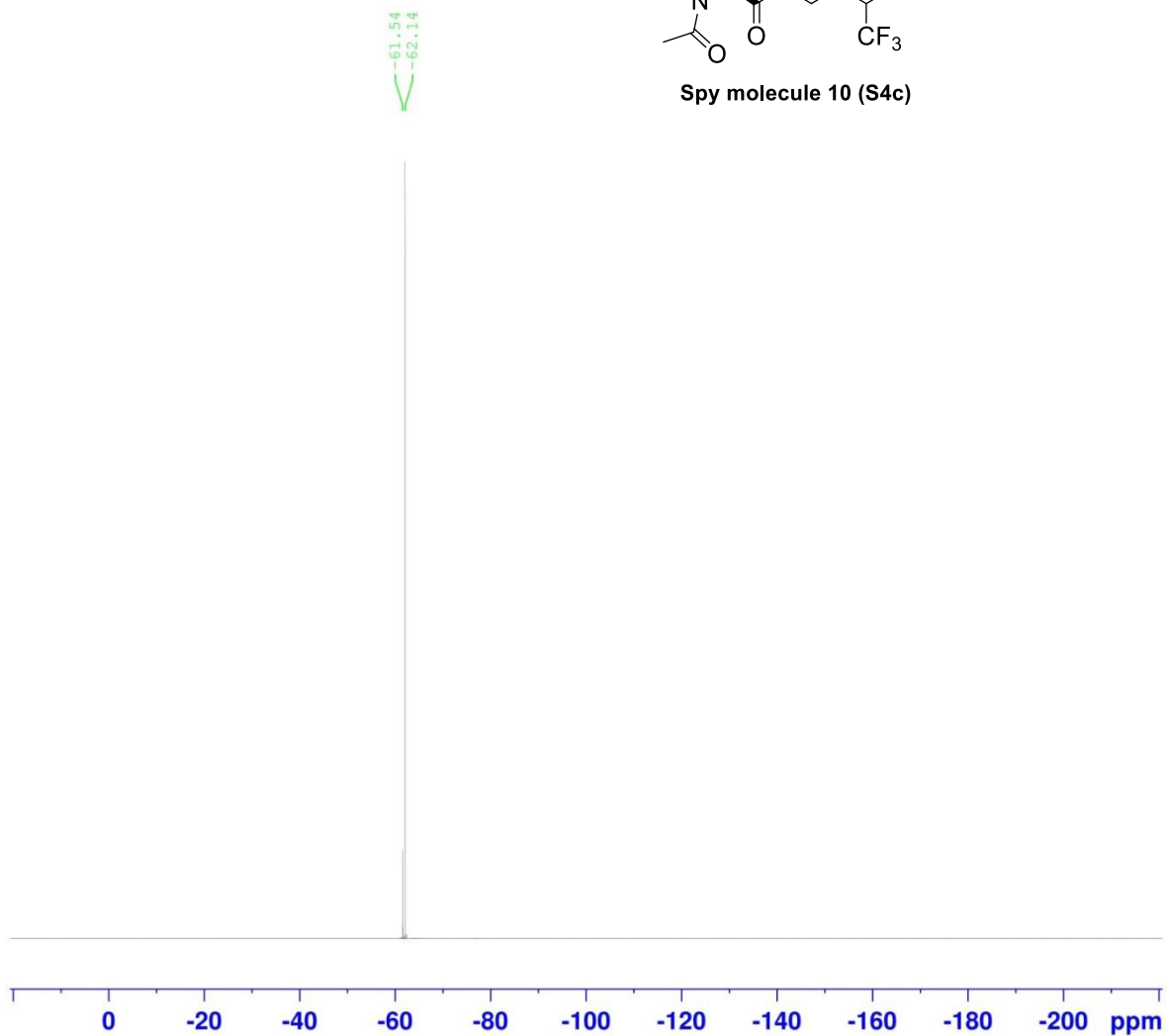


SUPPORTING INFORMATION

Spy molecule 10 (Compound S4c)



Spy molecule 10 (S4c)



Current Data Parameters
 NAME GCF10
 EXPNO 60
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20181114
 Time 9.37
 INSTRUM spect
 PROBHD 5 mm CPQCI 1H/
 PULPROG zgfhigqn.2
 TD 131072
 SOLVENT MeOD
 NS 256
 DS 4
 SWH 113636.367 Hz
 FIDRES 0.866977 Hz
 AQ 0.5767168 sec
 RG 190.78
 DW 4.400 usec
 DE 18.00 usec
 TE 293.1 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 D12 0.00002000 sec
 TD0 1

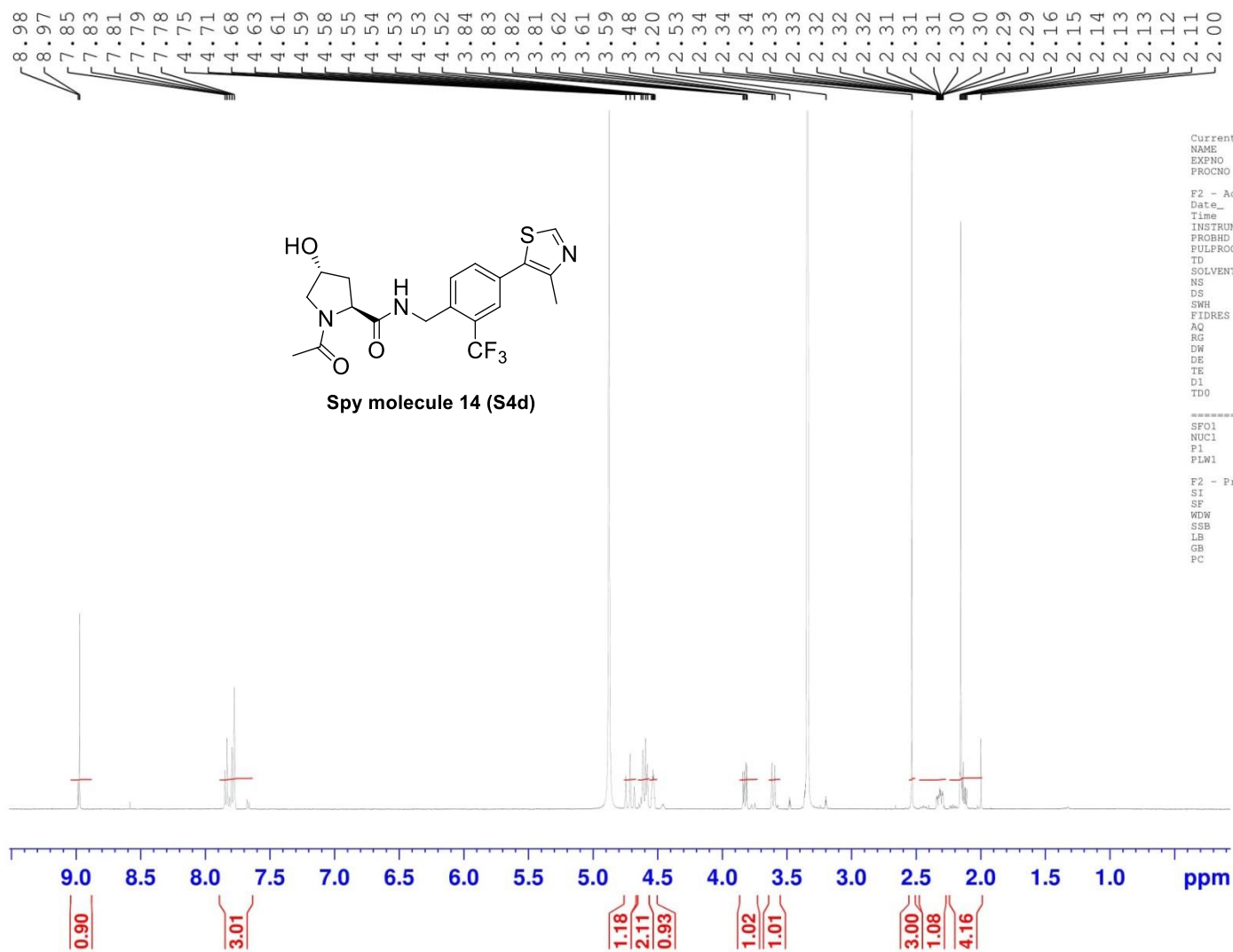
===== CHANNEL f1 =====
 SFO1 470.5453180 MHz
 NUC1 19F
 P1 12.00 usec
 PLW1 5.19999981 W

===== CHANNEL f2 =====
 SFO2 500.1320005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 100.00 usec
 PLW2 5.19999981 W
 PLW12 0.06406900 W

F2 - Processing parameters
 SI 262144
 SF 470.5923772 MHz
 WDW EM
 SSB 0
 LB 1.50 Hz
 GB 0
 PC 1.00

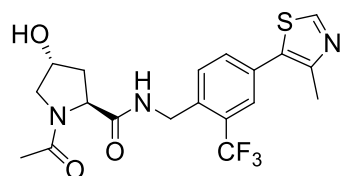
SUPPORTING INFORMATION

Spy molecule 14 (Compound S4d)

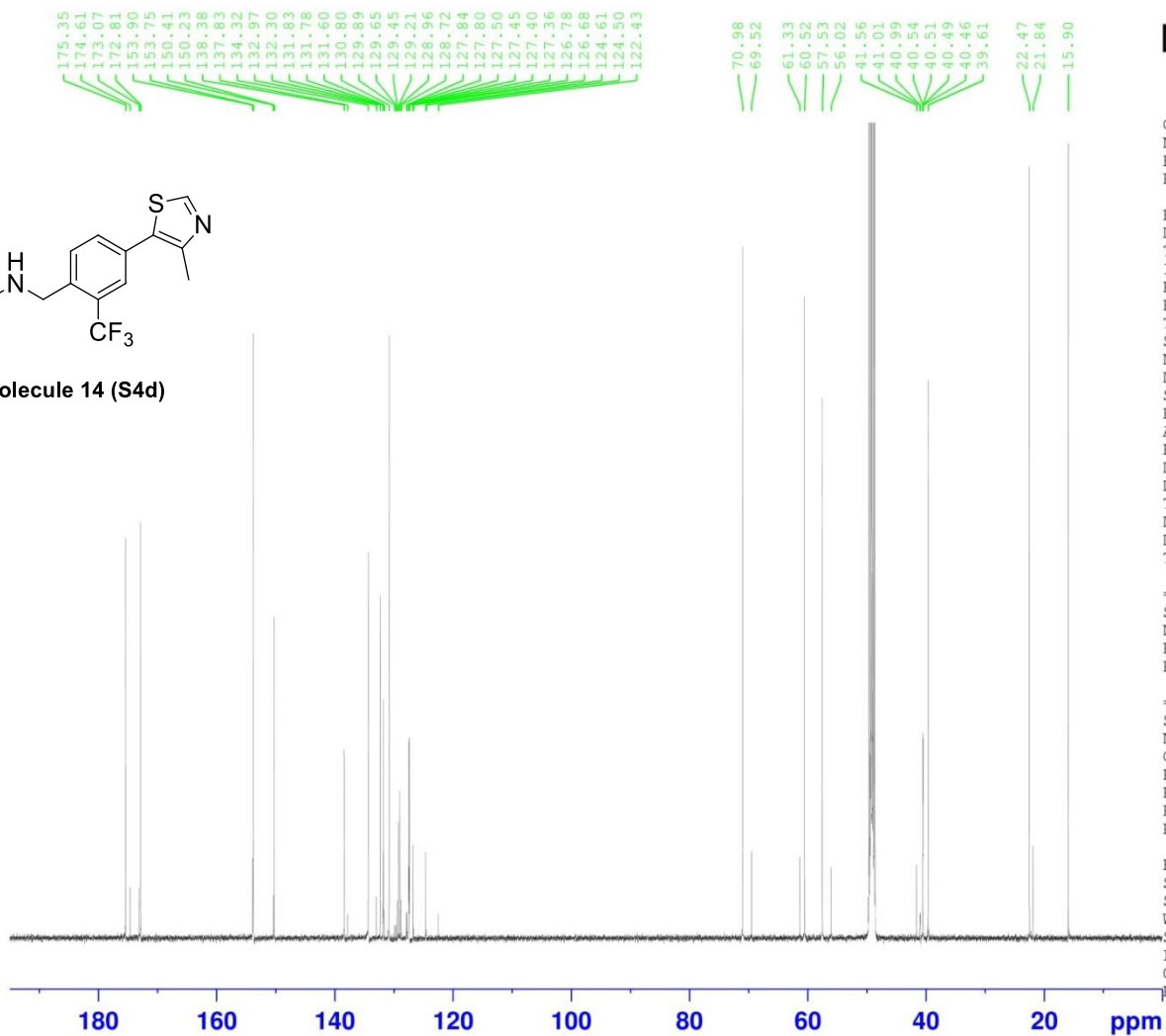


SUPPORTING INFORMATION

Spy molecule 14 (Compound S4d)



Spy molecule 14 (S4d)



Current Data Parameters
 NAME GCF14
 EXPNO 184
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20181117
 Time 3.26
 INSTRUM spect
 PROBHD 5 mm CPQCI 1H/
 PULPROG zgpg30
 TD 65400
 SOLVENT MeOD
 NS 2048
 DS 4
 SWH 29761.904 Hz
 FIDRES 0.455075 Hz
 AQ 1.0987200 sec
 RG 190.78
 DW 16.800 usec
 DE 18.00 usec
 TE 293.1 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TD0 1

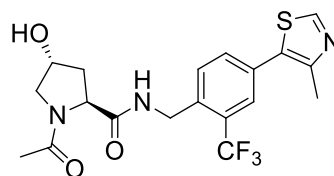
===== CHANNEL f1 =====
 SFO1 125.7703637 MHz
 NUC1 13C
 P1 12.00 usec
 PLW1 133.00000000 W

===== CHANNEL f2 =====
 SFO2 500.1320005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 100.00 usec
 PLW2 5.19999981 W
 PLW12 0.06406900 W
 PLW13 0.06406900 W

F2 - Processing parameters
 SI 131072
 SF 125.7576007 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

SUPPORTING INFORMATION

Spy molecule 14 (Compound S4d)



Spy molecule 14 (S4d)



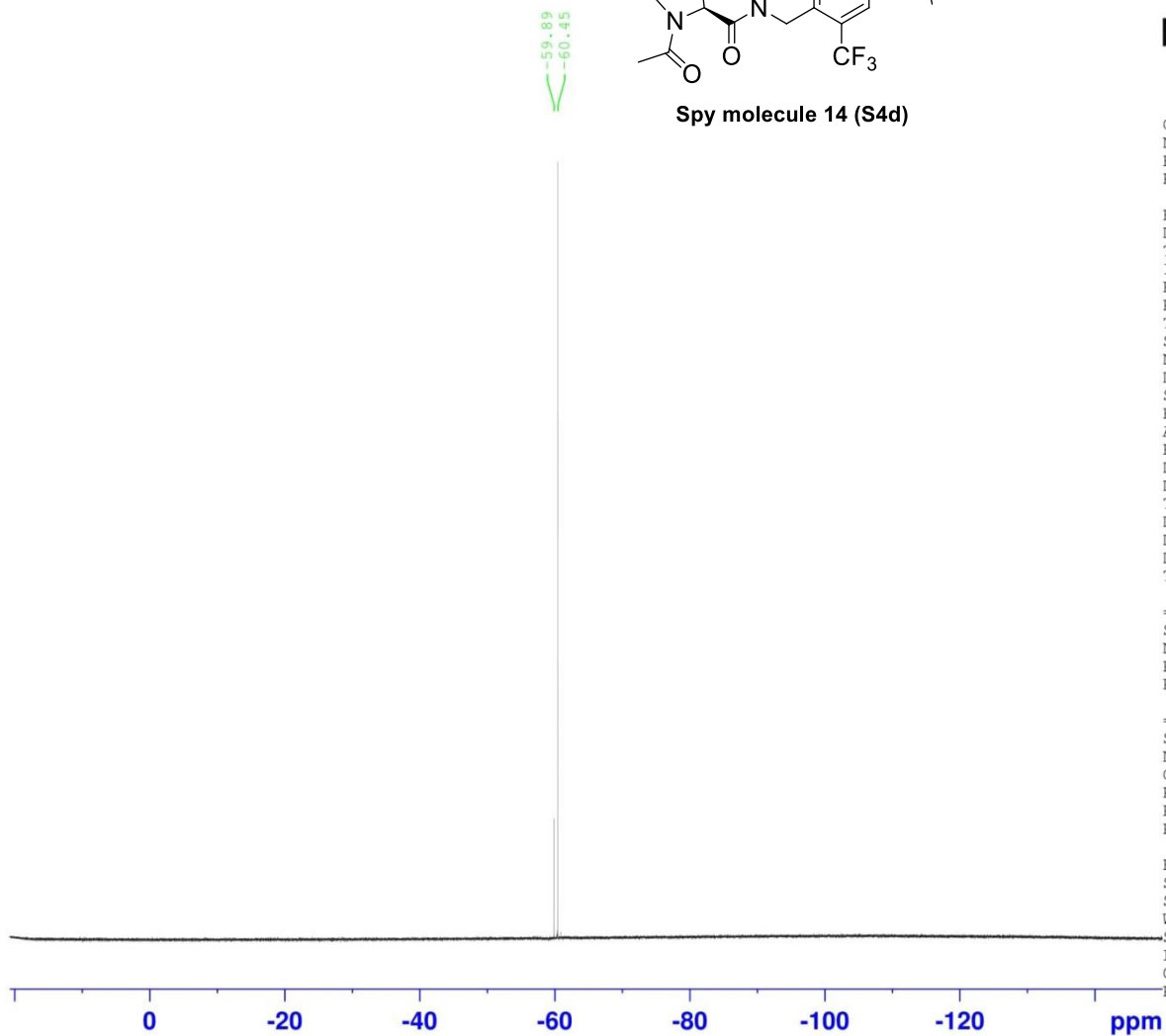
Current Data Parameters
 NAME GC-R054
 EXPNO 12
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20160530
 Time 23.11
 INSTRUM spect
 PROBHD 5 mm QNP 1H/13
 PULPROG zgfhigqn.2
 TD 131072
 SOLVENT MeOD
 NS 16
 DS 4
 SWH 113636.367 Hz
 FIDRES 0.866977 Hz
 AQ 0.5767168 sec
 RG 362
 DW 4.400 usec
 DE 6.50 usec
 TE 298.2 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 D12 0.00002000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 470.5453180 MHz
 NUC1 19F
 P1 13.50 usec
 PLW1 25.00000000 W

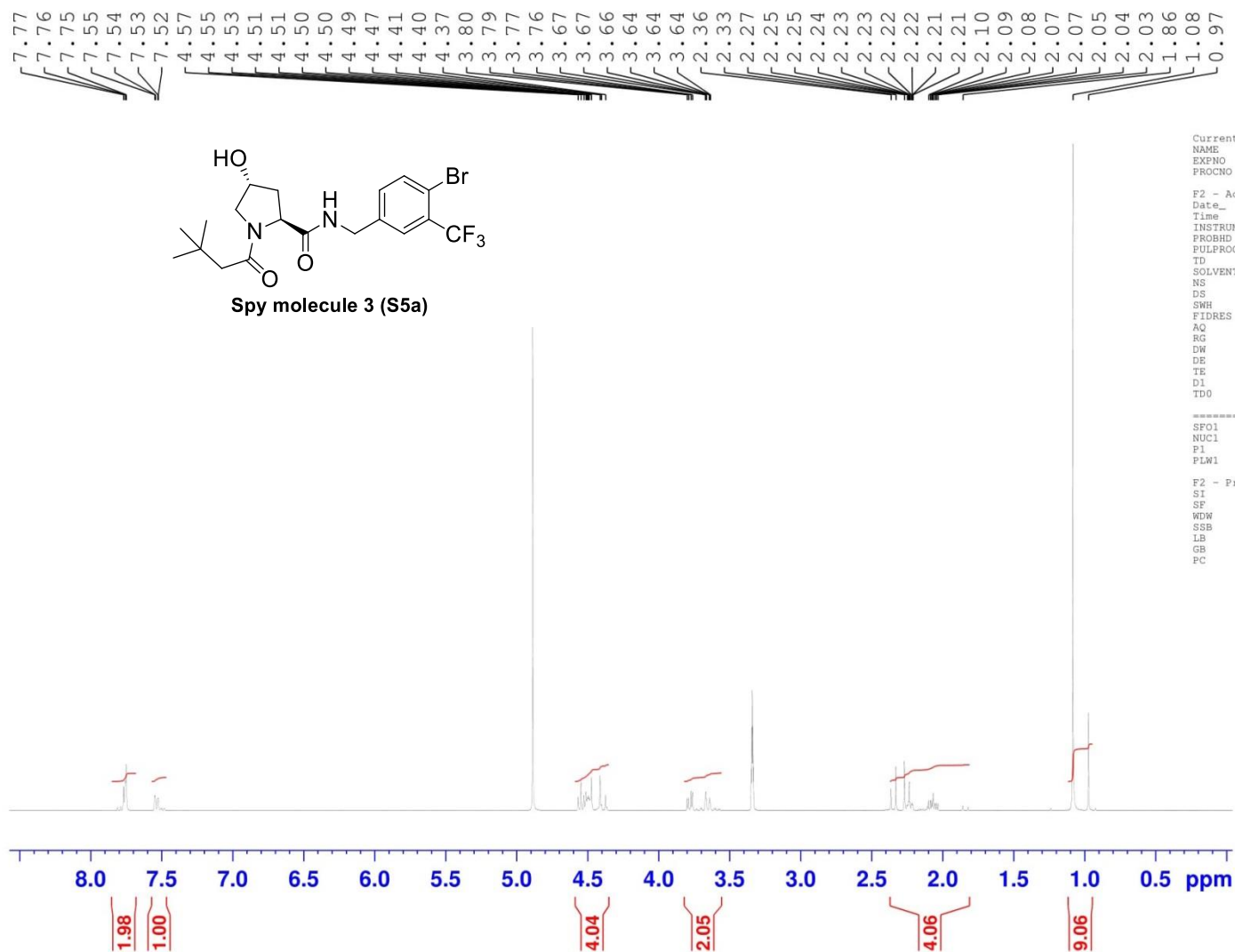
===== CHANNEL f2 =====
 SFO2 500.1320005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 80.00 usec
 PLW2 25.00000000 W
 PLW12 0.39063001 W

F2 - Processing parameters
 SI 65536
 SF 470.5923772 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



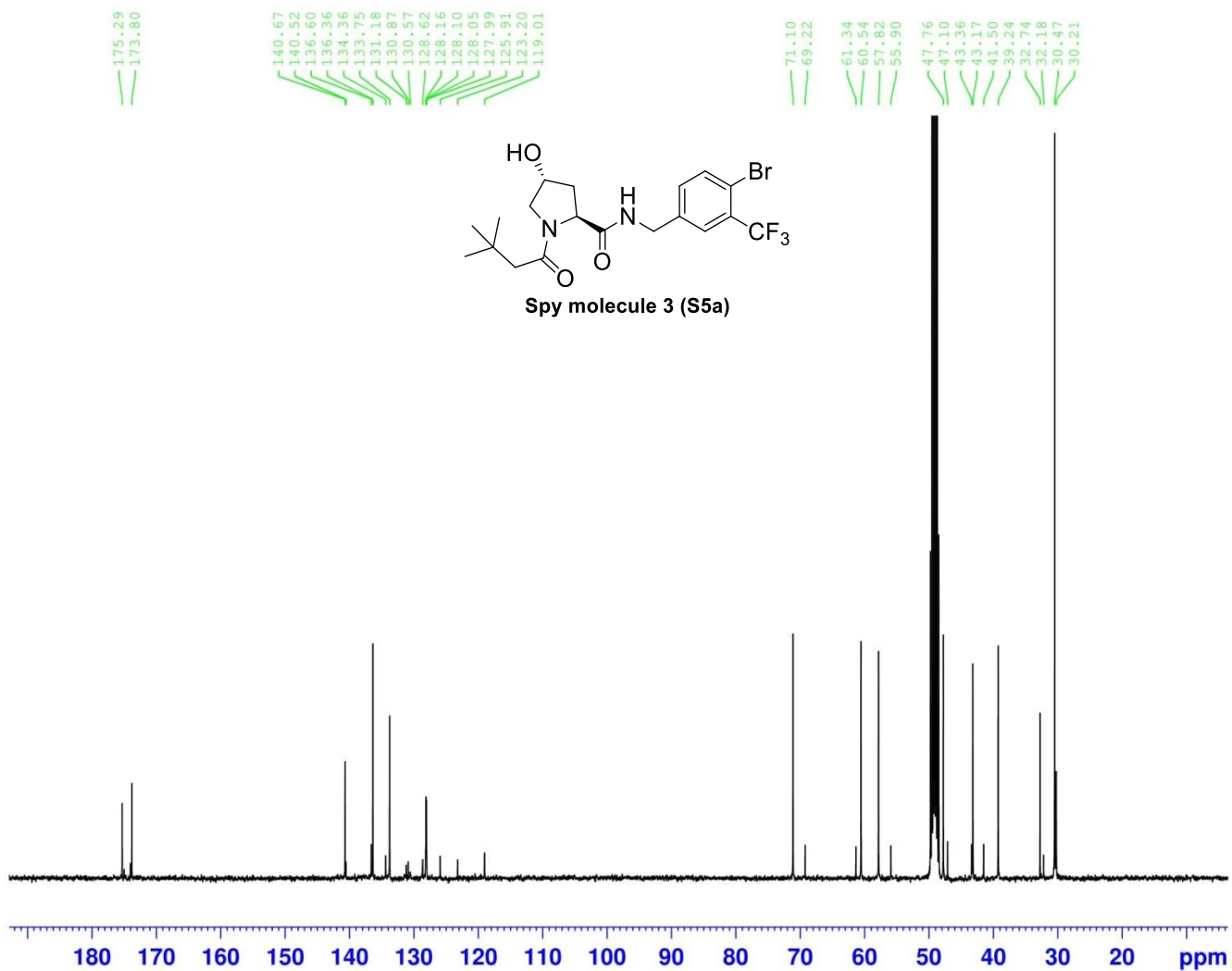
SUPPORTING INFORMATION

Spy molecule 3 (Compound S5a)



SUPPORTING INFORMATION

Spy molecule 3 (Compound S5a)



Current Data Parameters
 NAME ALC-GC-R029
 EXPNO 11
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20160129
 Time 4.40
 INSTRUM spect
 PROBHD 5 mm PADUL 13C
 PULPROG udef
 TD 17996
 SOLVENT MeOD
 NS 2800
 DS 0
 SWH 25000.000 Hz
 FIDRES 1.389198 Hz
 AQ 0.3599200 sec
 RG 196.14
 DW 20.000 usec
 DE 8.66 usec
 TE 298.2 K
 D1 3.00000000 sec
 D11 0.03000000 sec
 D12 0.00002000 sec
 D20 200.00000000 sec
 TDO 1

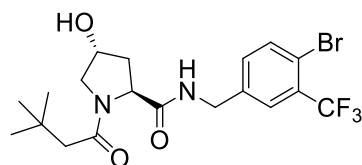
===== CHANNEL f1 =====
 SF01 100.6238346 MHz
 NUC1 13C
 P1 10.00 usec
 P13 2000.00 usec
 P26 500.00 usec
 PLW1 36.00000000 W
 SPNAM[5] Crp60comp.4
 SPOAL5 0.500
 SPOFFS5 0 Hz
 SPW5 5.50040007 W
 SPNAM[8] Crp60,0.5,20.1
 SPOAL8 0.500
 SPOFFS8 0 Hz
 SPW8 5.50040007 W

===== CHANNEL f2 =====
 SF02 400.1316005 MHz
 NUC2 1H
 CPDPRG[2] waltz64
 PCPD2 90.00 usec
 PLW2 20.00000000 W
 PLW12 0.24691001 W

F2 - Processing parameters
 SI 262144
 SF 100.6126176 MHz
 WDW EM
 SSB 0
 LB 2.00 Hz
 GB 0
 PC 1.40

SUPPORTING INFORMATION

Spy molecule 3 (Compound S5a)



Spy molecule 3 (S5a)



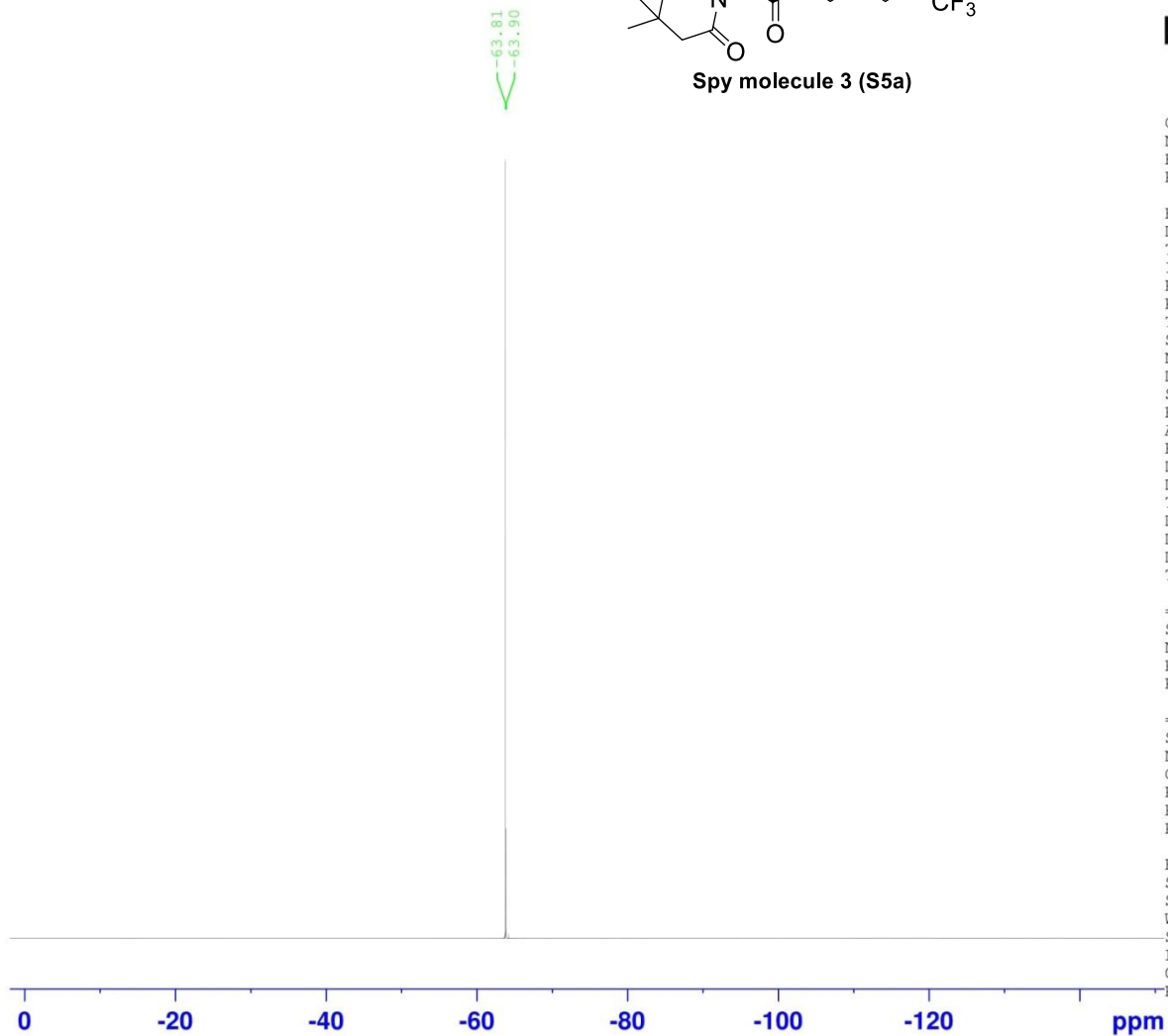
Current Data Parameters
 NAME GCF03 (Correct)
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20181114
 Time 8.58
 INSTRUM spect
 PROBHD 5 mm CPQCI 1H/
 PULPROG zgfhigqn.2
 TD 131072
 SOLVENT MeOD
 NS 256
 DS 4
 SWH 113636.367 Hz
 FIDRES 0.866977 Hz
 AQ 0.5767168 sec
 RG 190.78
 DW 4.400 usec
 DE 18.00 usec
 TE 293.2 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 D12 0.00002000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 470.5453180 MHz
 NUC1 19F
 P1 12.00 usec
 PLW1 5.19999981 W

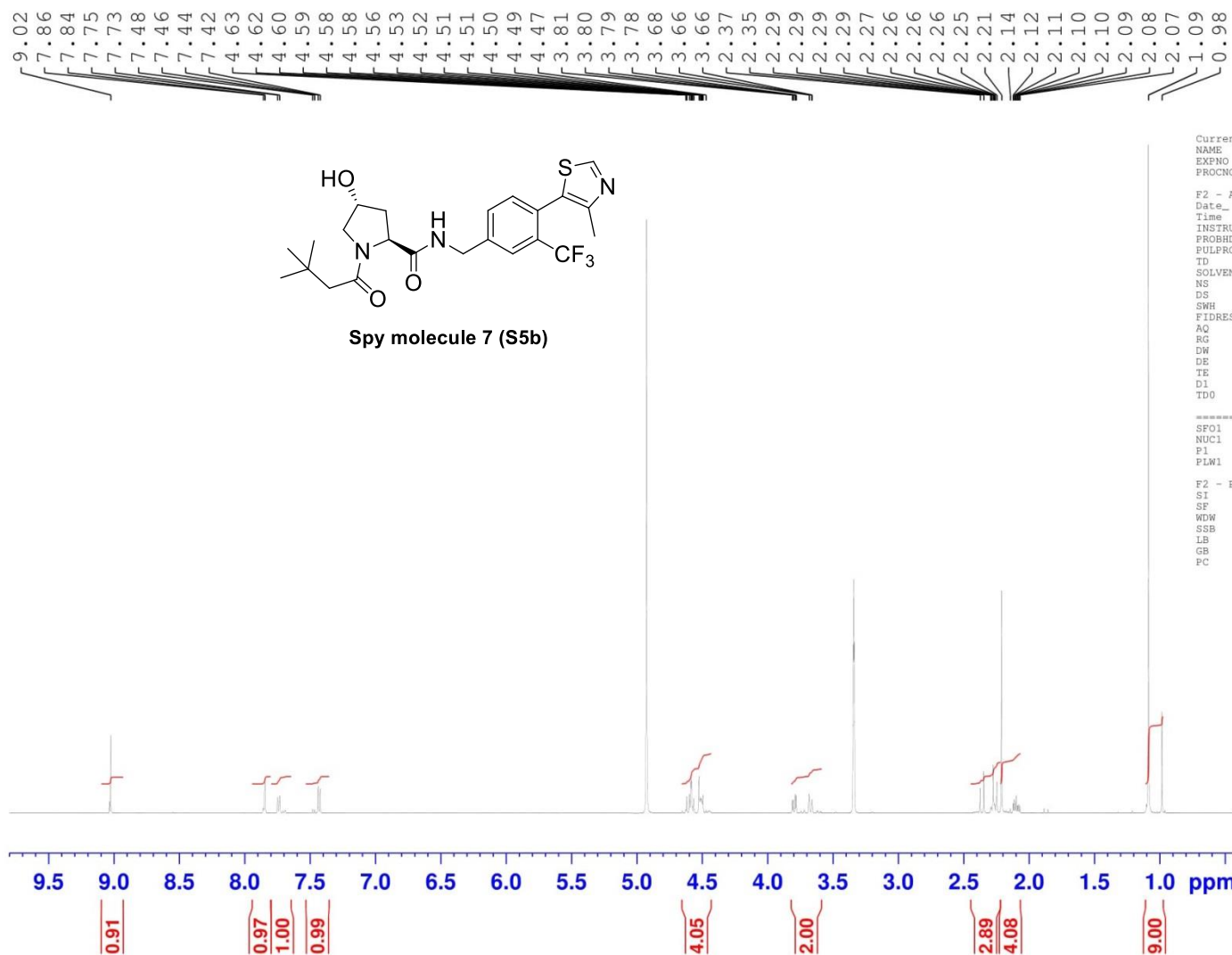
===== CHANNEL f2 =====
 SFO2 500.1320005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 100.00 usec
 PLW2 5.19999981 W
 PLW12 0.06406900 W

F2 - Processing parameters
 SI 262144
 SF 470.5923772 MHz
 WDW EM
 SSB 0
 LB 1.50 Hz
 GB 0
 PC 1.00



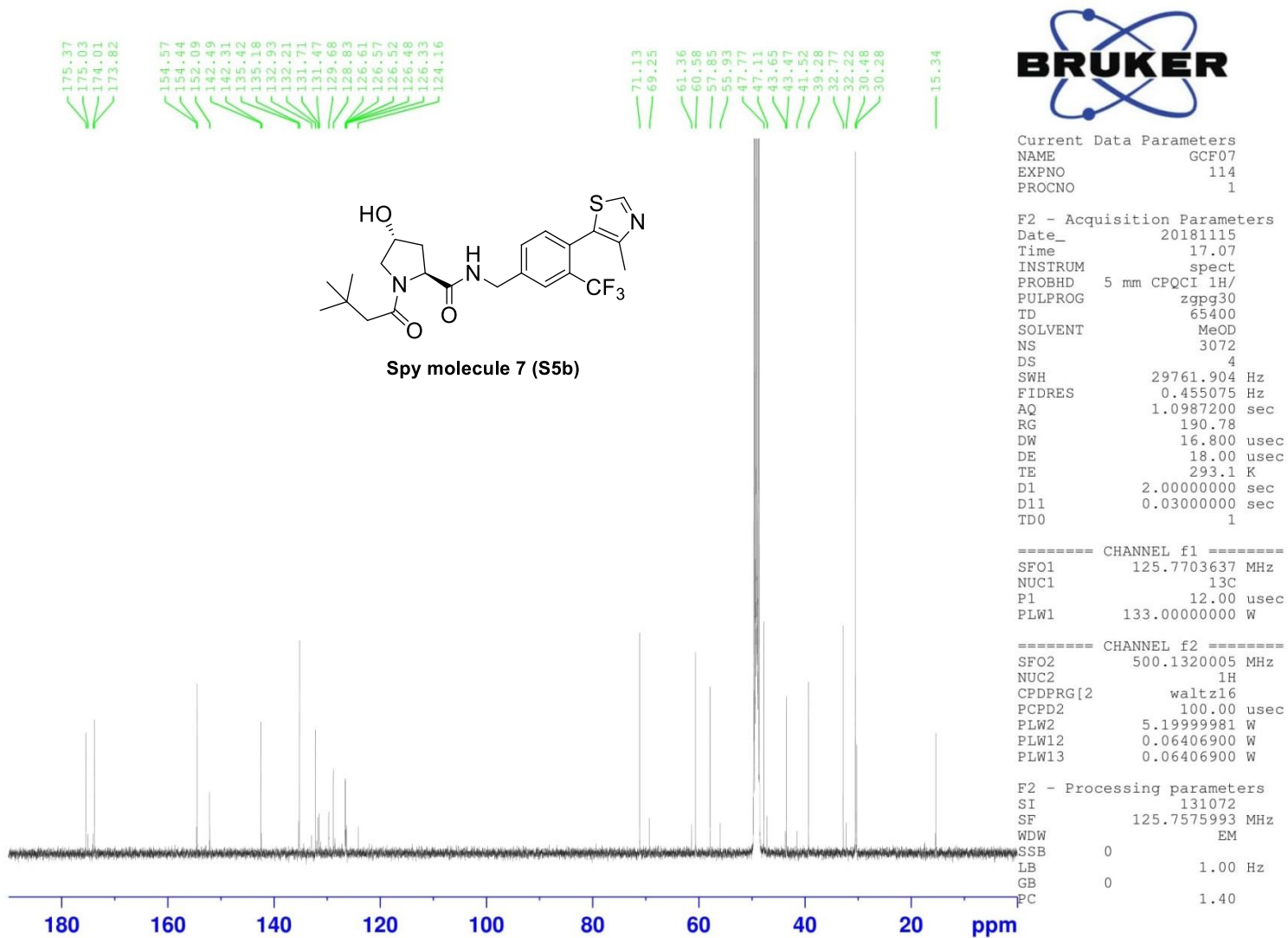
SUPPORTING INFORMATION

Spy molecule 7 (Compound S5b)



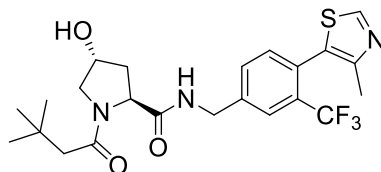
SUPPORTING INFORMATION

Spy molecule 7 (Compound S5b)

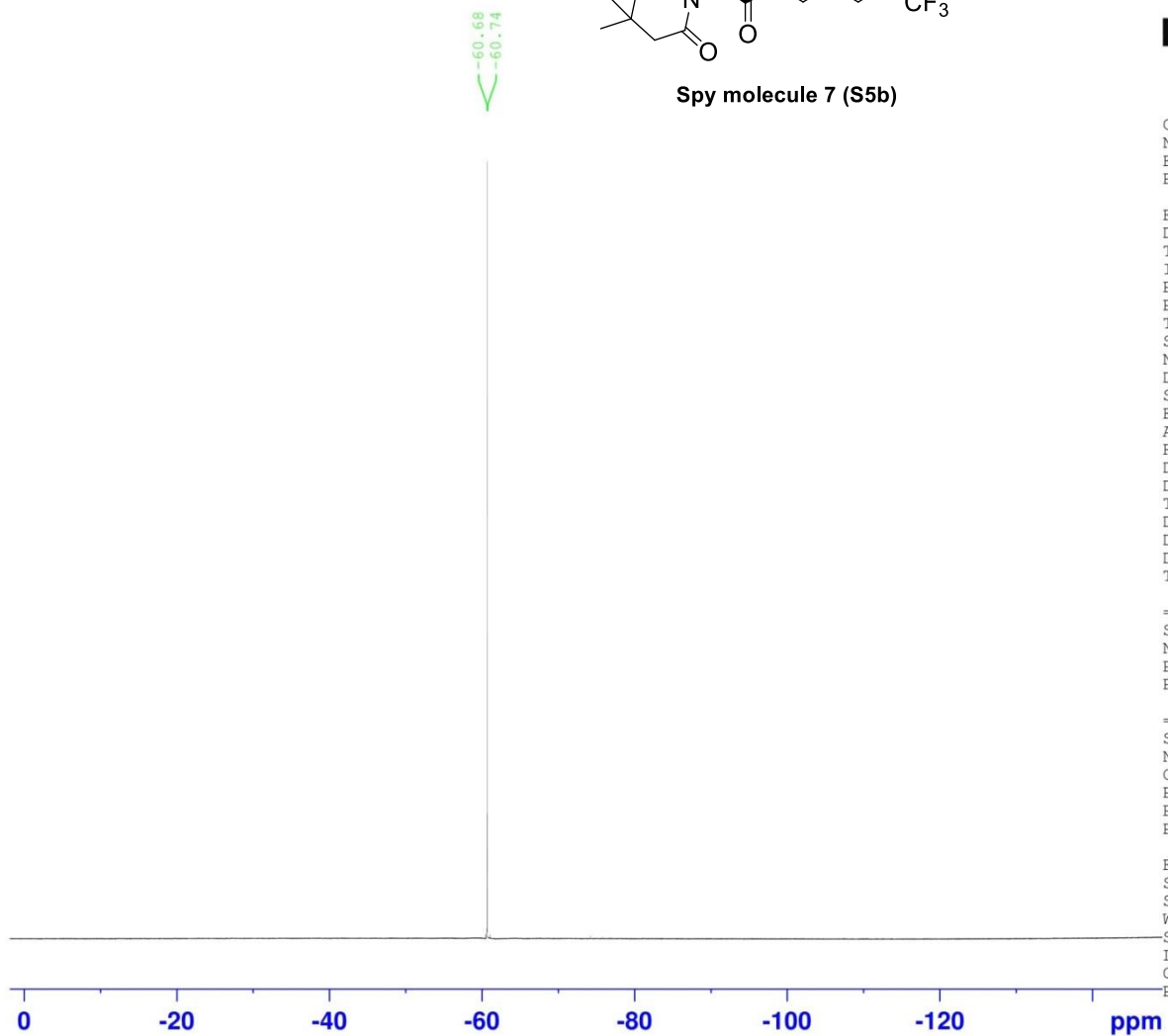


SUPPORTING INFORMATION

Spy molecule 7 (Compound S5b)



Spy molecule 7 (S5b)



Current Data Parameters
 NAME GC-GCF007
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20181122
 Time 15.25
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zgfhigqn.2
 TD 131072
 SOLVENT MeOD
 NS 16
 DS 4
 SWH 113636.367 Hz
 FIDRES 0.866977 Hz
 AQ 0.5767168 sec
 RG 322
 DW 4.400 usec
 DE 6.50 usec
 TE 303.4 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 D12 0.00002000 sec
 TD0 1

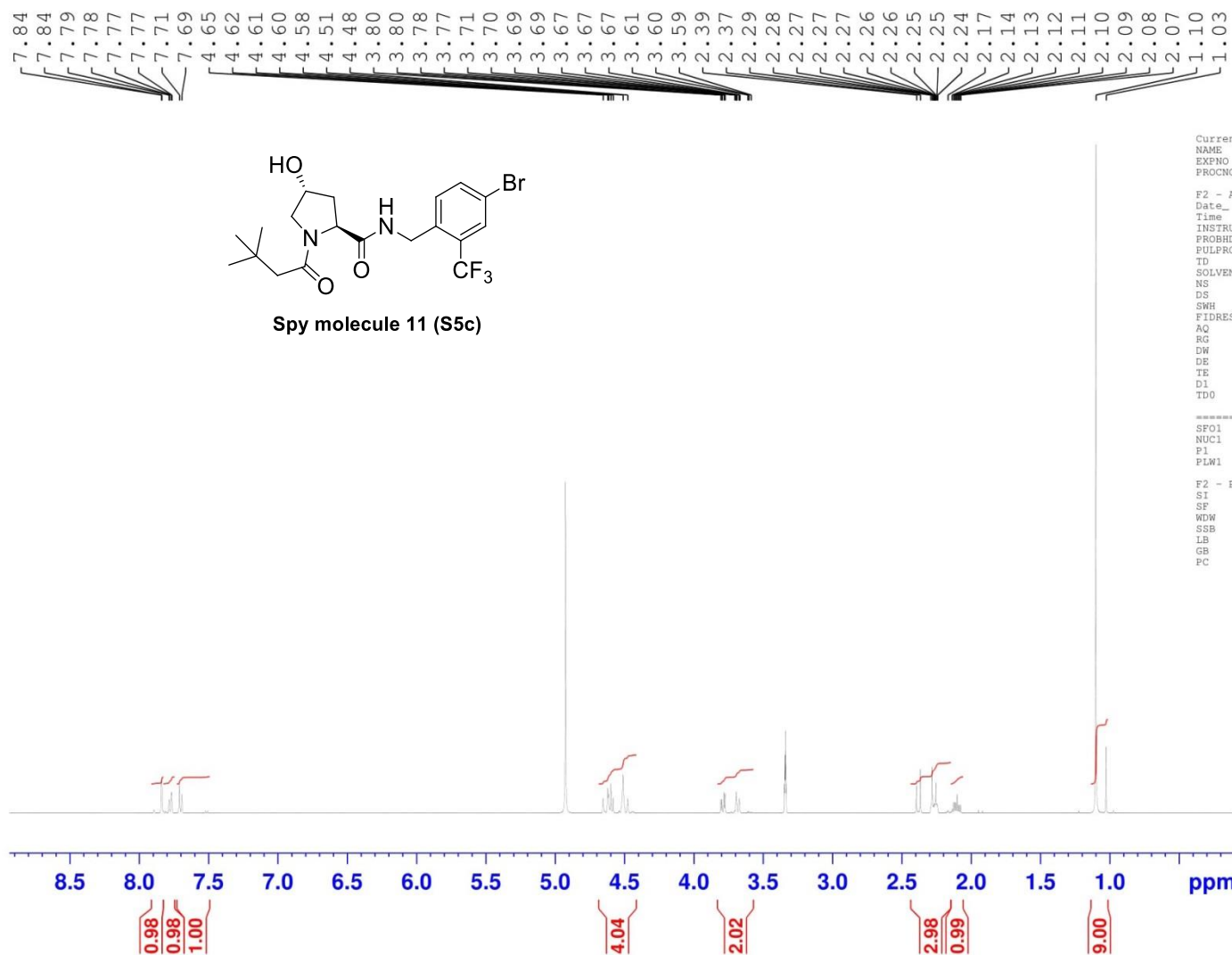
===== CHANNEL f1 =====
 SFO1 470.5453180 MHz
 NUC1 19F
 P1 14.50 usec
 PLW1 45.00000000 W

===== CHANNEL f2 =====
 SFO2 500.1320005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 80.00 usec
 PLW2 20.85000038 W
 PLW12 0.35916999 W

F2 - Processing parameters
 SI 65536
 SF 470.5923772 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

SUPPORTING INFORMATION

Spy molecule 11 (Compound S5c)



Current Data Parameters
 NAME GCF11
 EXPNO 122
 PROCNO 1

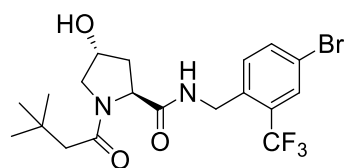
F2 - Acquisition Parameters
 Date_ 20181114
 Time 11.24
 INSTRUM spect
 PROBHD 5 mm CPQCI 1H/
 PULPROG zg30
 ID 65536
 SOLVENT MeOD
 NS 128
 DS 2
 SWH 10000.000 Hz
 FIDRES 0.152588 Hz
 AQ 3.2767999 sec
 RG 70.68
 DW 50.000 usec
 DE 10.00 usec
 TE 293.1 K
 D1 1.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 SF01 500.1330885 MHz
 NUC1 1H
 P1 11.00 usec
 PLW1 5.19999981 W

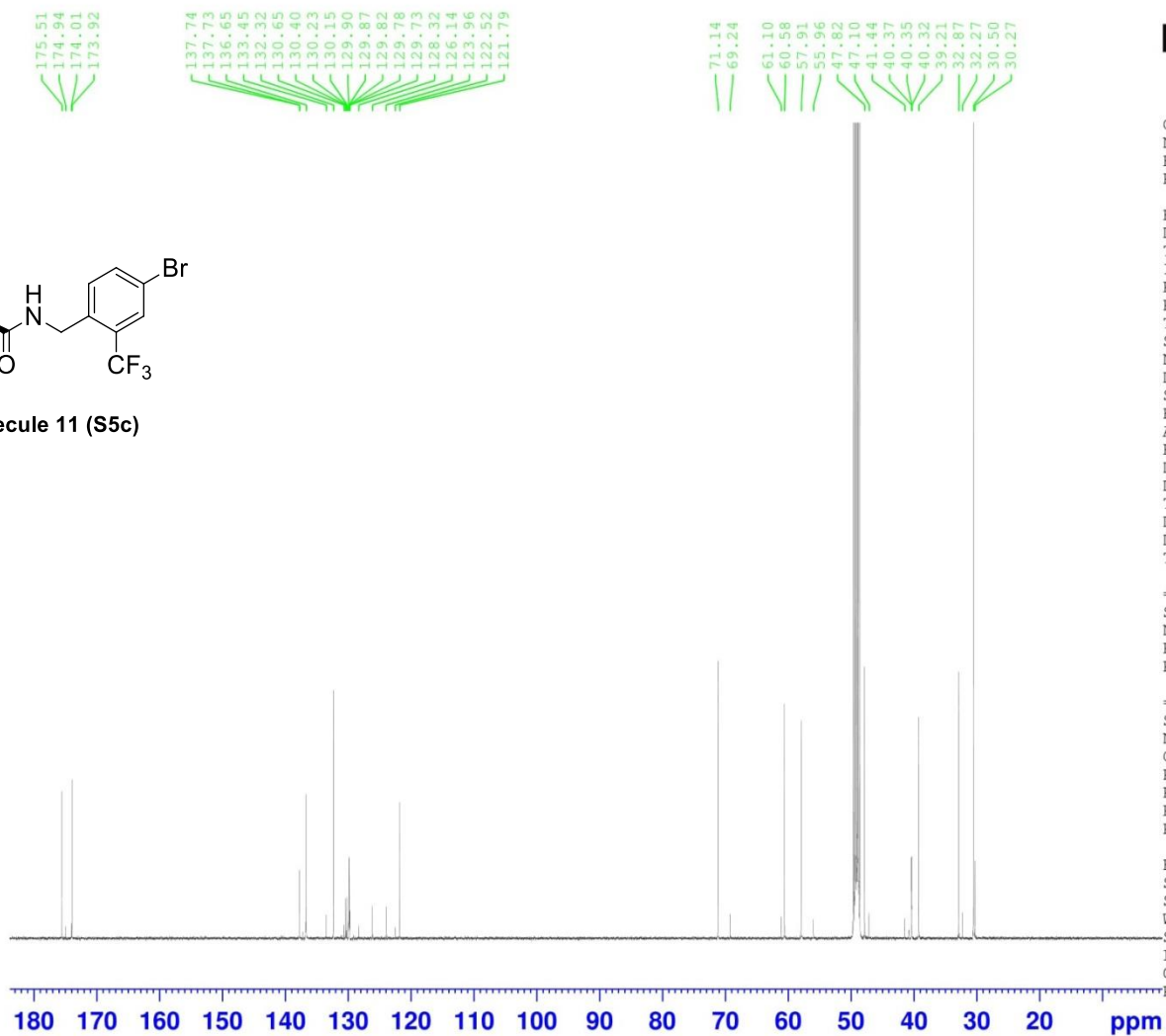
F2 - Processing parameters
 SI 131072
 SF 500.1299947 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

SUPPORTING INFORMATION

Spy molecule 11 (Compound S5c)



Spy molecule 11 (S5c)



Current Data Parameters
 NAME GCF11
 EXPNO 124
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20181116
 Time 10.58
 INSTRUM spect
 PROBHD 5 mm CPQCI 1H/
 PULPROG zgpg30
 TD 65400
 SOLVENT MeOD
 NS 3072
 DS 4
 SWH 29761.904 Hz
 FIDRES 0.455075 Hz
 AQ 1.0987200 sec
 RG 190.78
 DW 16.800 usec
 DE 18.00 usec
 TE 293.1 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TD0 1

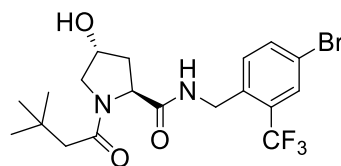
===== CHANNEL f1 =====
 SFO1 125.7703637 MHz
 NUC1 13C
 P1 12.00 usec
 PLW1 133.00000000 W

===== CHANNEL f2 =====
 SFO2 500.1320005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 100.00 usec
 PLW2 5.19999981 W
 PLW12 0.06406900 W
 PLW13 0.06406900 W

F2 - Processing parameters
 SI 131072
 SF 125.7576002 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

SUPPORTING INFORMATION

Spy molecule 11 (Compound S5c)



Spy molecule 11 (S5c)



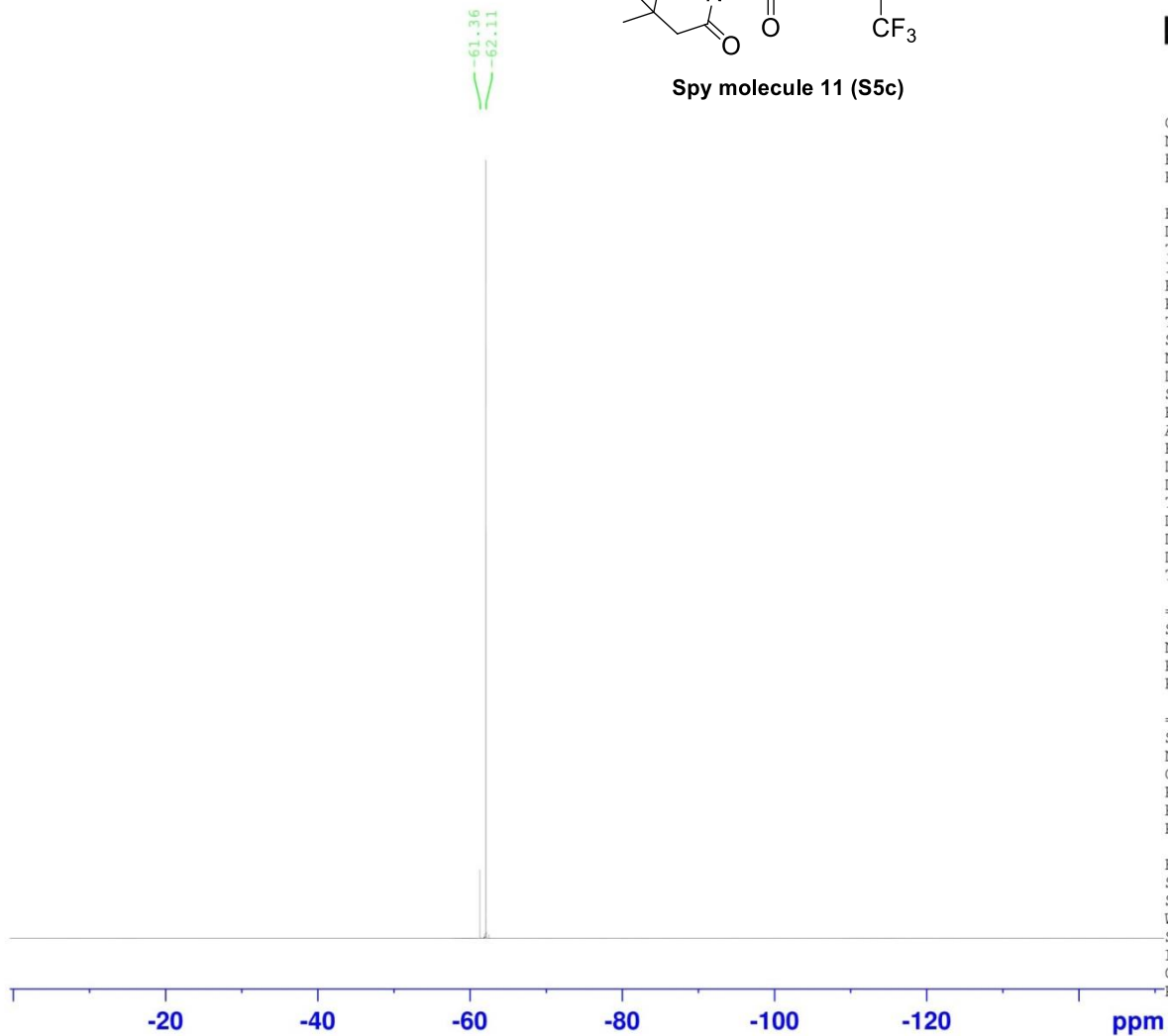
Current Data Parameters
 NAME GCF11
 EXPNO 120
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20181114
 Time 11.14
 INSTRUM spect
 PROBHD 5 mm CPQCI 1H/
 PULPROG zgfhigqn.2
 TD 131072
 SOLVENT MeOD
 NS 256
 DS 4
 SWH 113636.367 Hz
 FIDRES 0.866977 Hz
 AQ 0.5767168 sec
 RG 190.78
 DW 4.400 usec
 DE 18.00 usec
 TE 293.1 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 D12 0.00002000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 470.5453180 MHz
 NUC1 19F
 P1 12.00 usec
 PLW1 5.19999981 W

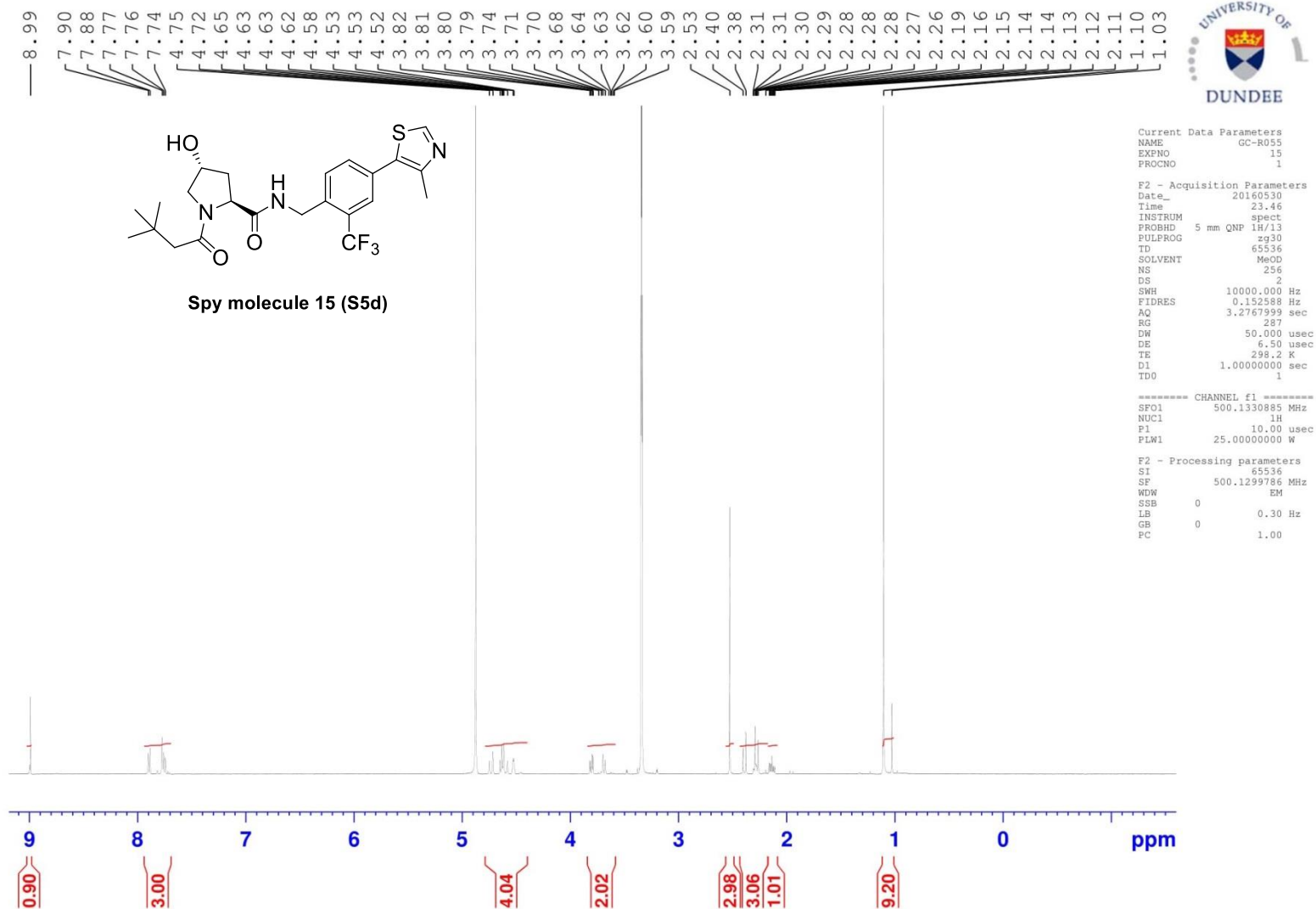
===== CHANNEL f2 =====
 SFO2 500.1320005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 100.00 usec
 PLW2 5.19999981 W
 PLW12 0.06406900 W

F2 - Processing parameters
 SI 262144
 SF 470.5923772 MHz
 WDW EM
 SSB 0
 LB 1.50 Hz
 GB 0
 PC 1.00



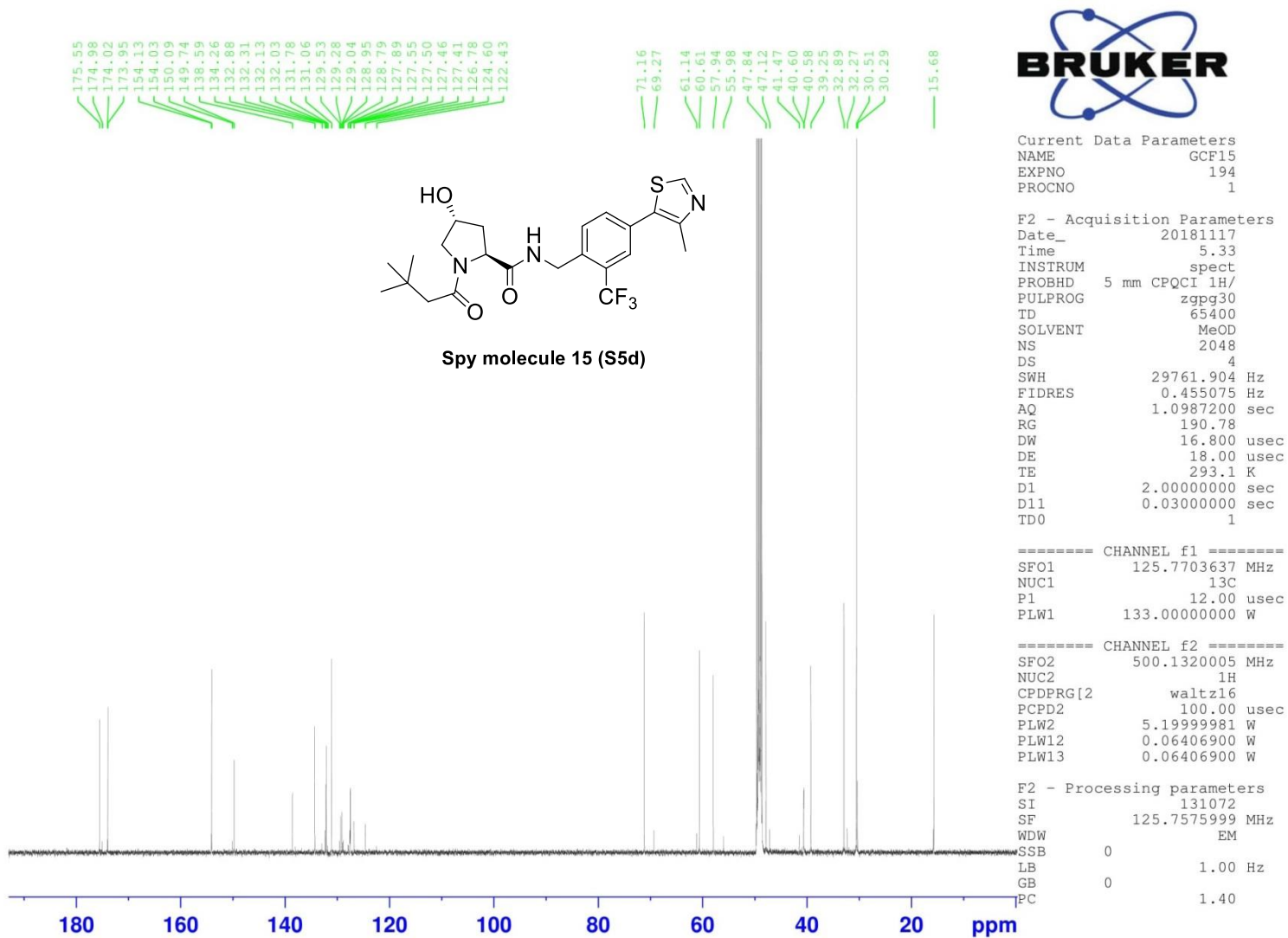
SUPPORTING INFORMATION

Spy molecule 15 (Compound S5d)



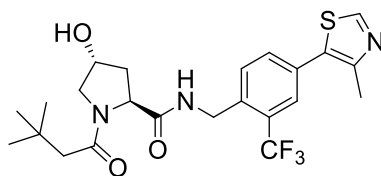
SUPPORTING INFORMATION

Spy molecule 15 (Compound S5d)

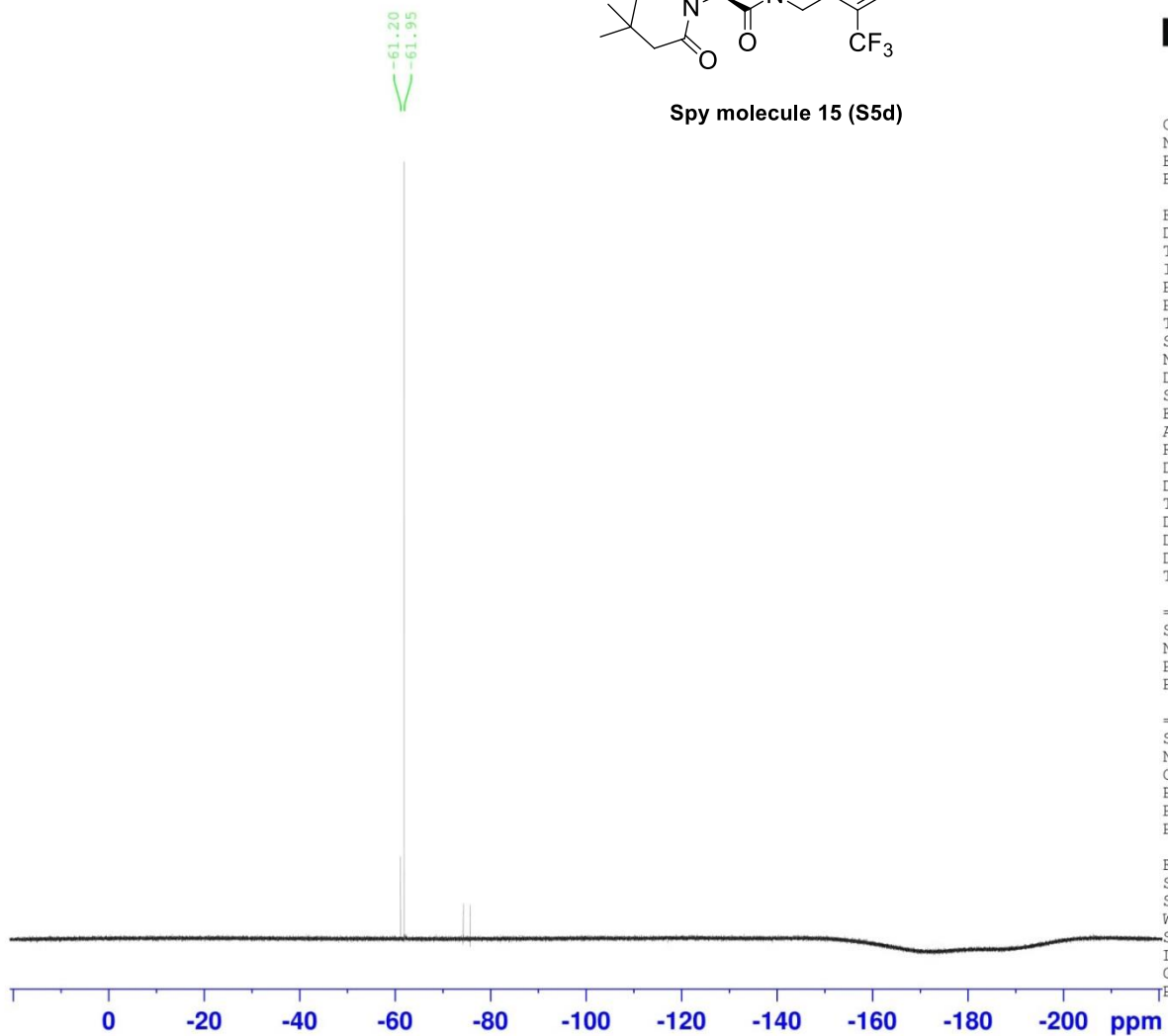


SUPPORTING INFORMATION

Spy molecule 15 (Compound S5d)



Spy molecule 15 (S5d)



Current Data Parameters
 NAME GC-R055
 EXPNO 19
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20160531
 Time 0.45
 INSTRUM spect
 PROBHD 5 mm QNP 1H/13
 PULPROG zgfhigqn.2
 TD 131072
 SOLVENT MeOD
 NS 16
 DS 4
 SWH 113636.367 Hz
 FIDRES 0.866977 Hz
 AQ 0.5767168 sec
 RG 322
 DW 4.400 usec
 DE 6.50 usec
 TE 298.2 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 D12 0.00002000 sec
 TD0 1

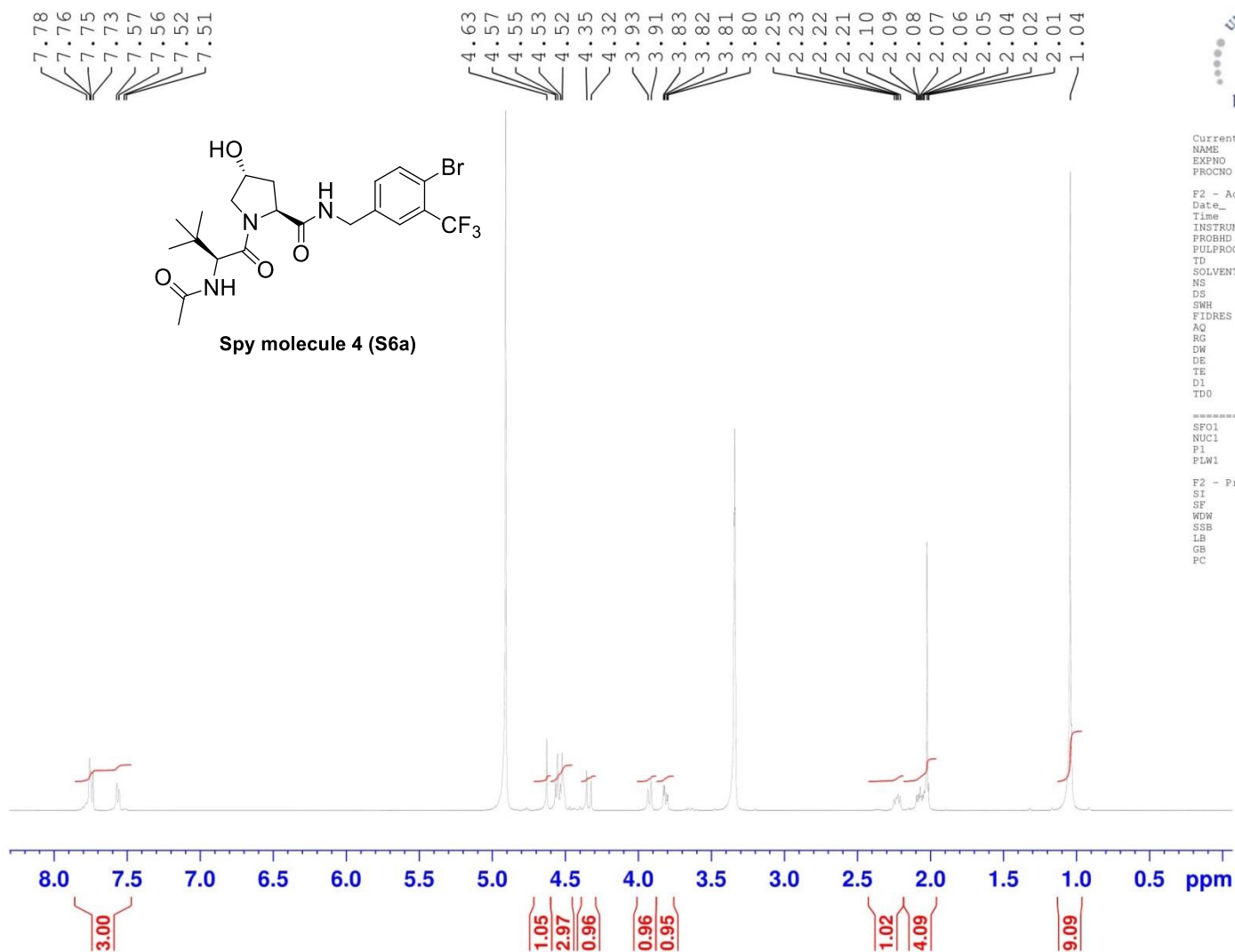
===== CHANNEL f1 =====
 SFO1 470.5453180 MHz
 NUC1 19F
 P1 13.50 usec
 PLW1 25.00000000 W

===== CHANNEL f2 =====
 SFO2 500.1320005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 80.00 usec
 PLW2 25.00000000 W
 PLW12 0.39063001 W

F2 - Processing parameters
 SI 65536
 SF 470.5923772 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

SUPPORTING INFORMATION

Spy molecule 4 (Compound S6a)



Current Data Parameters
 NAME AC-GC-R033
 EXPNO 1
 PROCNO 1

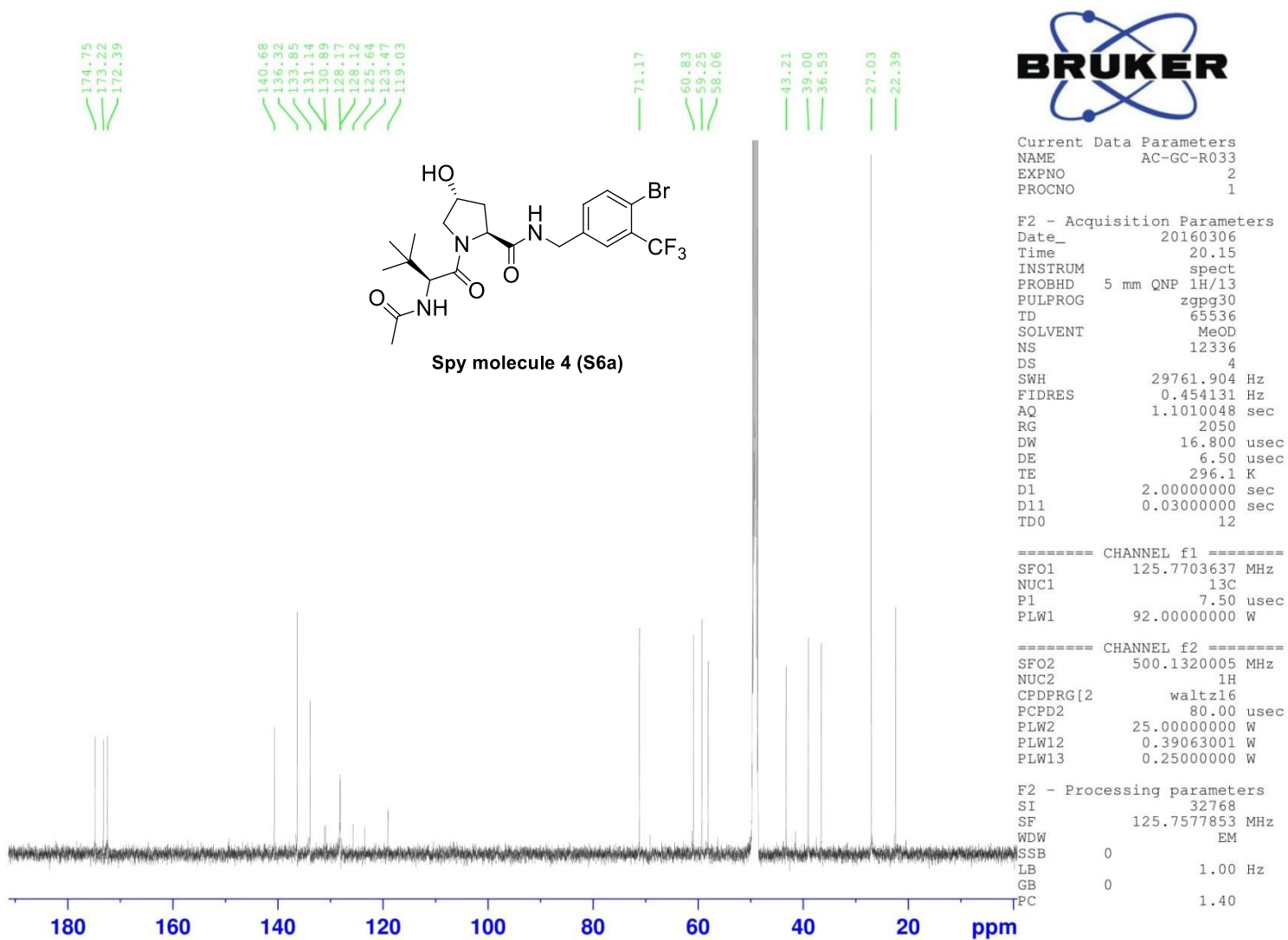
F2 - Acquisition Parameters
 Date_ 20160306
 Time 19.19
 INSTRUM spect
 PROBHD 5 mm QNP 1H/13
 PULPROG zg30
 ID 65536
 SOLVENT MeOD
 NS 1280
 DS 2
 SWH 10000.000 Hz
 FIDRES 0.152588 Hz
 AQ 3.2767999 sec
 RG 32
 DW 50.000 usec
 DE 6.50 usec
 TE 295.1 K
 D1 1.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 SF01 500.1330885 MHz
 NUC1 1H
 P1 10.00 usec
 PLW1 25.0000000 W

F2 - Processing parameters
 SI 65536
 SF 500.1307360 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

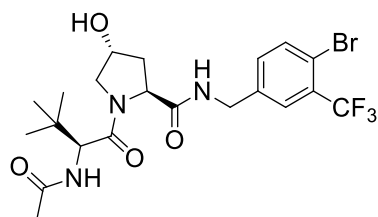
SUPPORTING INFORMATION

Spy molecule 4 (Compound S6a)



SUPPORTING INFORMATION

Spy molecule 4 (Compound S6a)



Spy molecule 4 (S6a)



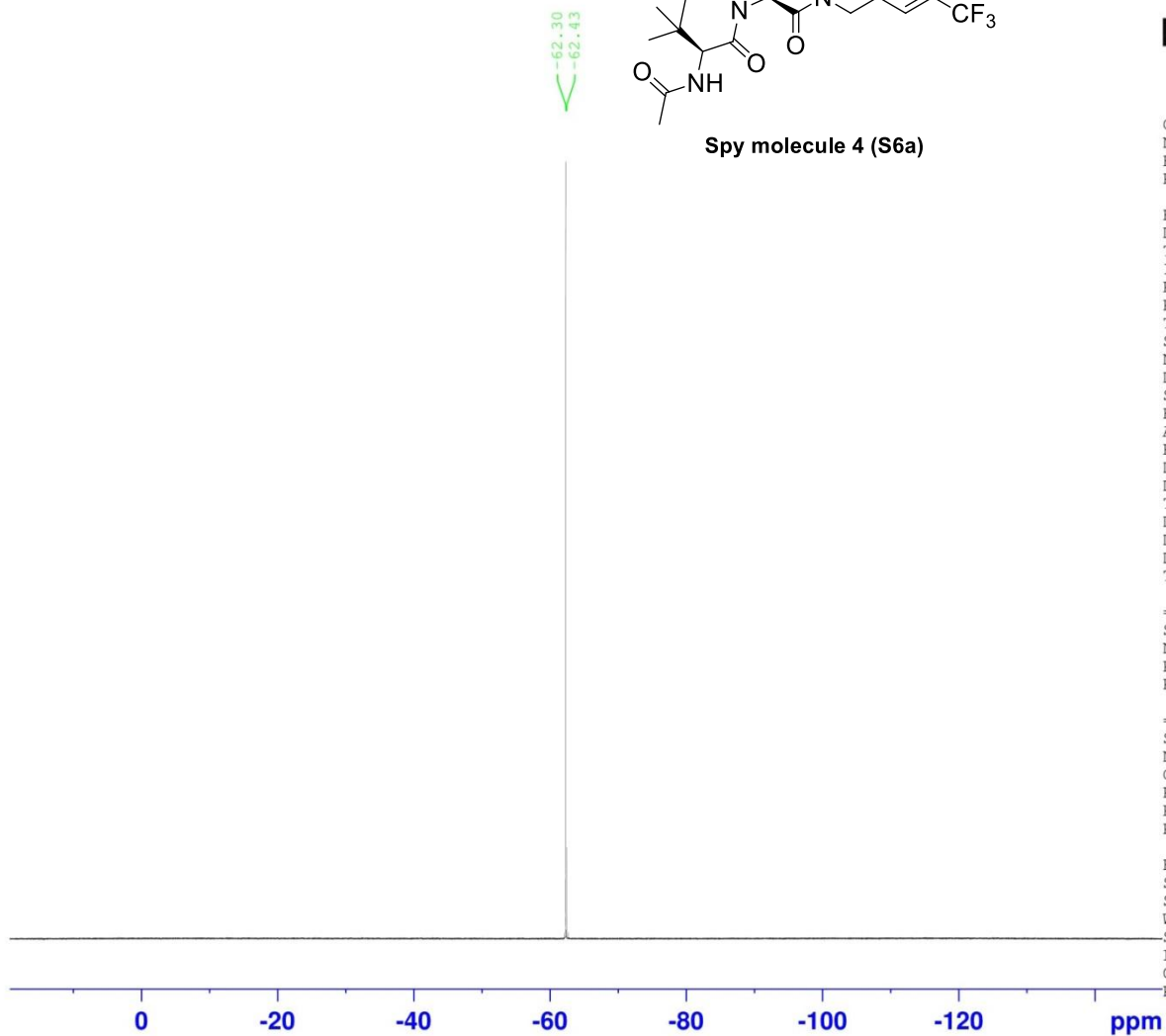
Current Data Parameters
 NAME AC-GC-R033
 EXPNO 3
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20160307
 Time 6.15
 INSTRUM spect
 PROBHD 5 mm QNP 1H/13
 PULPROG zgfhigqn.2
 TD 131072
 SOLVENT MeOD
 NS 64
 DS 4
 SWH 113636.367 Hz
 FIDRES 0.866977 Hz
 AQ 0.5767168 sec
 RG 645
 DW 4.400 usec
 DE 6.50 usec
 TE 295.8 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 D12 0.00002000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 470.5453180 MHz
 NUC1 19F
 P1 13.50 usec
 PLW1 25.00000000 W

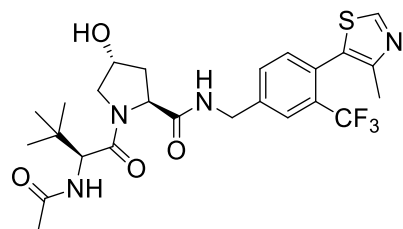
===== CHANNEL f2 =====
 SFO2 500.1320005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 80.00 usec
 PLW2 25.00000000 W
 PLW12 0.39063001 W

F2 - Processing parameters
 SI 65536
 SF 470.5923772 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

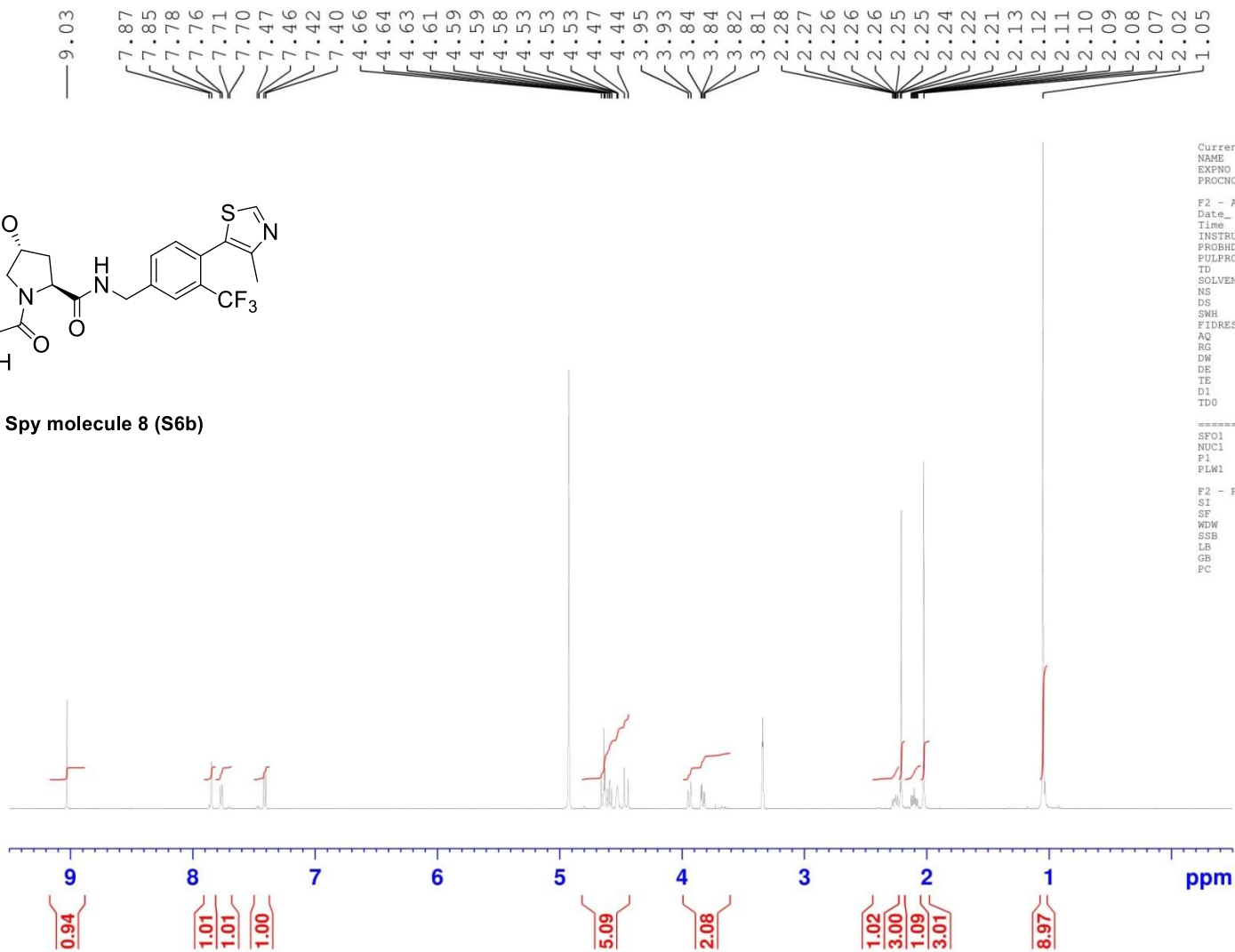


SUPPORTING INFORMATION

Spy molecule 8 (Compound S6b)



Spy molecule 8 (S6b)



```

Current Data Parameters
NAME          GCF08
EXPNO         72
PROCNO        1

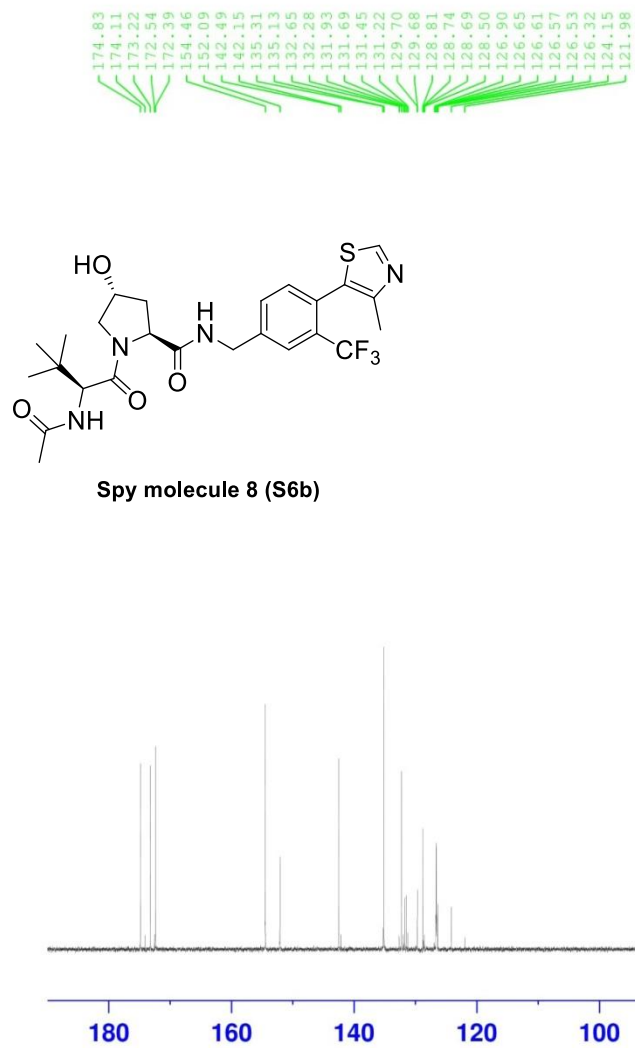
F2 - Acquisition Parameters
Date_         20181114
Time          10.06
INSTRUM       spect
PROBHD        5 mm CPOCF 1H/
PULPROG       zg30
TD            65536
SOLVENT       MeOD
NS            128
DS            2
SWH           10000.000 Hz
FIDRES        0.152588 Hz
AQ            3.2767999 sec
RG            70.68
DW            50.000 usec
DE            10.00 usec
TE            293.2 K
D1            1.00000000 sec
TDO           1

===== CHANNEL f1 =====
SFO1          500.1330885 MHz
NUC1          1H
P1            11.00 usec
PLW1          5.19999981 W

F2 - Processing parameters
SI            131072
SF            500.1299946 MHz
WDW           EM
SSB           0
LB            0.30 Hz
GB            0
PC            1.00
    
```

SUPPORTING INFORMATION

Spy molecule 8 (Compound S6b)



Current Data Parameters
 NAME GCF08
 EXPNO 74
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20181116
 Time 22.36
 INSTRUM spect
 PROBHD 5 mm CPQCI 1H/
 PULPROG zgpg30
 TD 65400
 SOLVENT MeOD
 NS 3000
 DS 4
 SWH 29761.904 Hz
 FIDRES 0.455075 Hz
 AQ 1.0987200 sec
 RG 190.78
 DW 16.800 usec
 DE 18.00 usec
 TE 293.2 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TD0 1

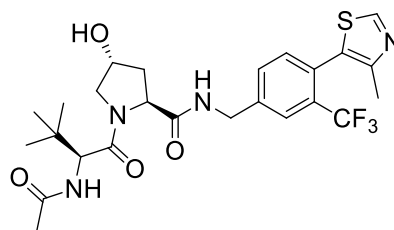
===== CHANNEL f1 =====
 SFO1 125.7703637 MHz
 NUC1 13C
 P1 12.00 usec
 PLW1 133.00000000 W

===== CHANNEL f2 =====
 SFO2 500.1320005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 100.00 usec
 PLW2 5.19999981 W
 PLW12 0.06406900 W
 PLW13 0.06406900 W

F2 - Processing parameters
 SI 131072
 SF 125.7576001 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

SUPPORTING INFORMATION

Spy molecule 8 (Compound S6b)



Spy molecule 8 (S6b)



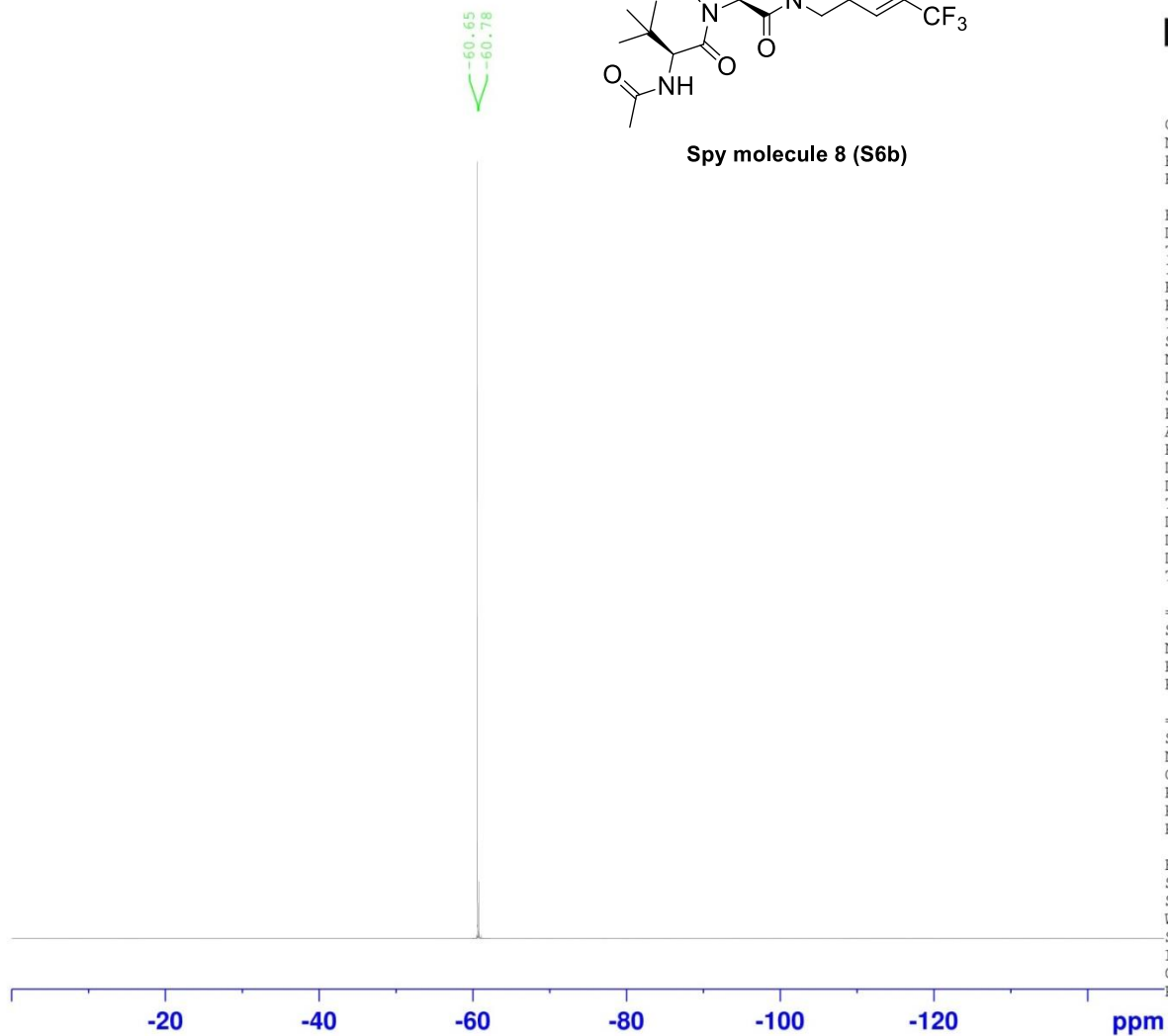
Current Data Parameters
 NAME GCF08
 EXPNO 70
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20181114
 Time 9.57
 INSTRUM spect
 PROBHD 5 mm CPQCI 1H/
 PULPROG zgfhigqn.2
 TD 131072
 SOLVENT MeOD
 NS 256
 DS 4
 SWH 113636.367 Hz
 FIDRES 0.866977 Hz
 AQ 0.5767168 sec
 RG 190.78
 DW 4.400 usec
 DE 18.00 usec
 TE 293.2 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 D12 0.00002000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 470.5453180 MHz
 NUC1 19F
 P1 12.00 usec
 PLW1 5.19999981 W

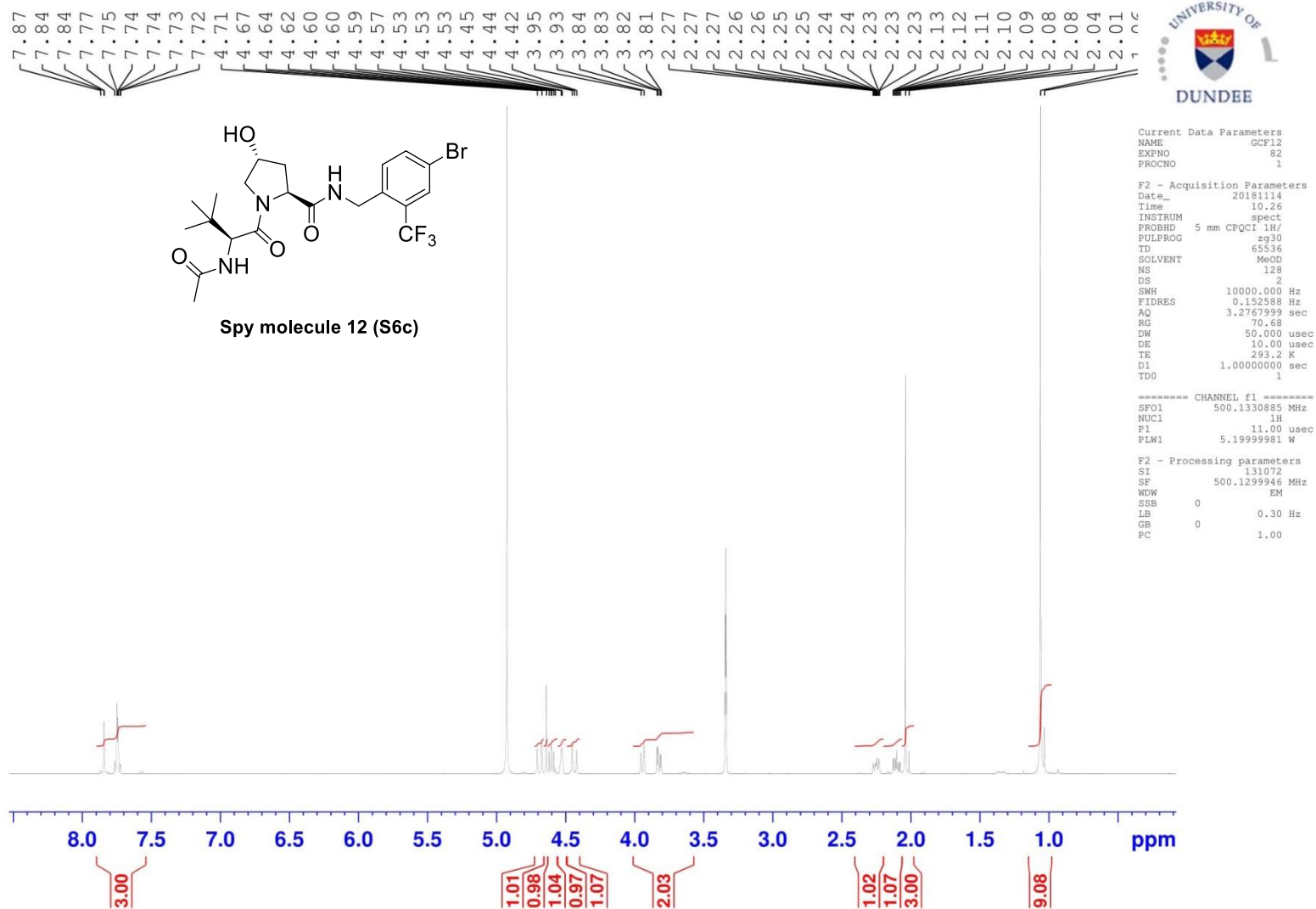
===== CHANNEL f2 =====
 SFO2 500.1320005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 100.00 usec
 PLW2 5.19999981 W
 PLW12 0.06406900 W

F2 - Processing parameters
 SI 262144
 SF 470.5923772 MHz
 WDW EM
 SSB 0
 LB 1.50 Hz
 GB 0
 PC 1.00



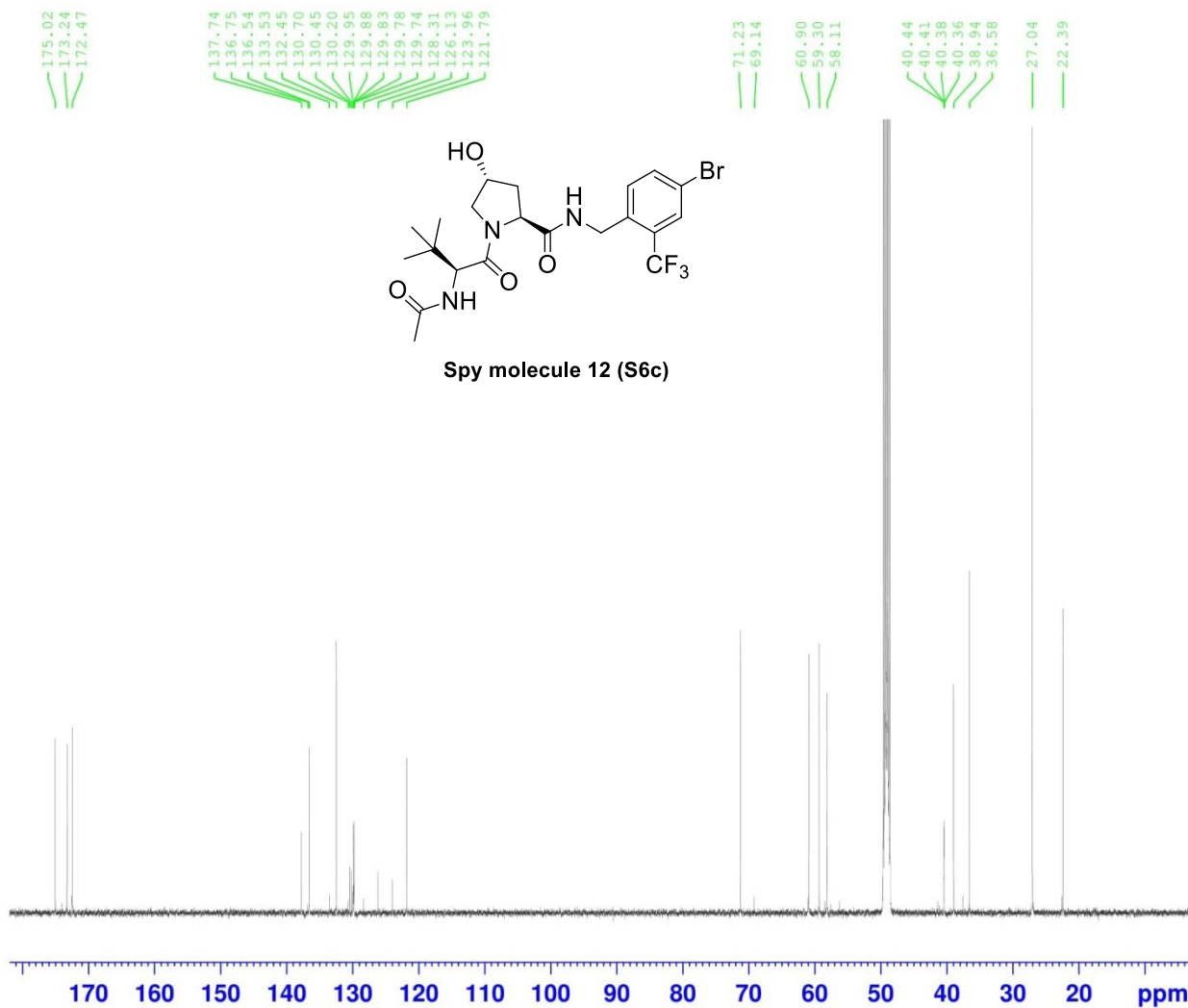
SUPPORTING INFORMATION

Spy molecule 12 (Compound S6c)



SUPPORTING INFORMATION

Spy molecule 12 (Compound S6c)



Current Data Parameters
 NAME GCF12
 EXPNO 84
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20181117
 Time 1.17
 INSTRUM spect
 PROBHD 5 mm CPQCI 1H/
 PULPROG zgpg30
 TD 65400
 SOLVENT MeOD
 NS 3000
 DS 4
 SWH 29761.904 Hz
 FIDRES 0.455075 Hz
 AQ 1.0987200 sec
 RG 190.78
 DW 16.800 usec
 DE 18.00 usec
 TE 293.1 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TD0 1

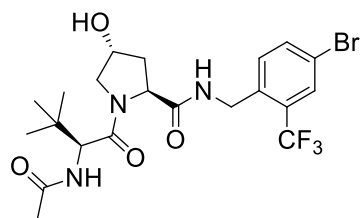
===== CHANNEL f1 =====
 SFO1 125.7703637 MHz
 NUC1 13C
 P1 12.00 usec
 PLW1 133.00000000 W

===== CHANNEL f2 =====
 SFO2 500.1320005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 100.00 usec
 PLW2 5.19999981 W
 PLW12 0.06406900 W
 PLW13 0.06406900 W

F2 - Processing parameters
 SI 131072
 SF 125.7575997 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

SUPPORTING INFORMATION

Spy molecule 12 (Compound S6c)



Spy molecule 12 (S6c)



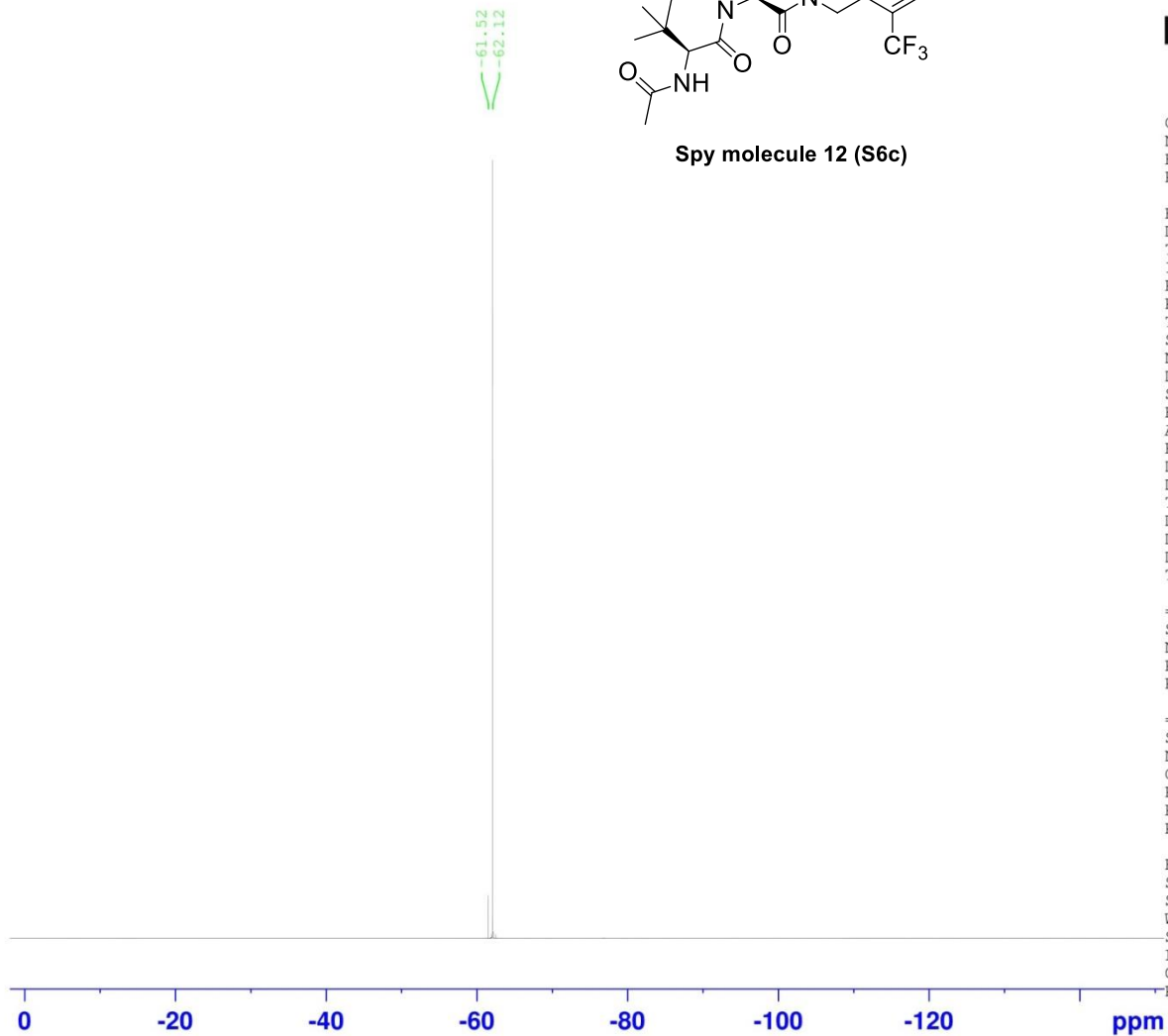
Current Data Parameters
 NAME GCF12
 EXPNO 80
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20181114
 Time 10.16
 INSTRUM spect
 PROBHD 5 mm CPQCI 1H/
 PULPROG zgfhigqn.2
 TD 131072
 SOLVENT MeOD
 NS 256
 DS 4
 SWH 113636.367 Hz
 FIDRES 0.866977 Hz
 AQ 0.5767168 sec
 RG 190.78
 DW 4.400 usec
 DE 18.00 usec
 TE 293.1 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 D12 0.00002000 sec
 TD0 1

==== CHANNEL f1 =====
 SFO1 470.5453180 MHz
 NUC1 19F
 P1 12.00 usec
 PLW1 5.19999981 W

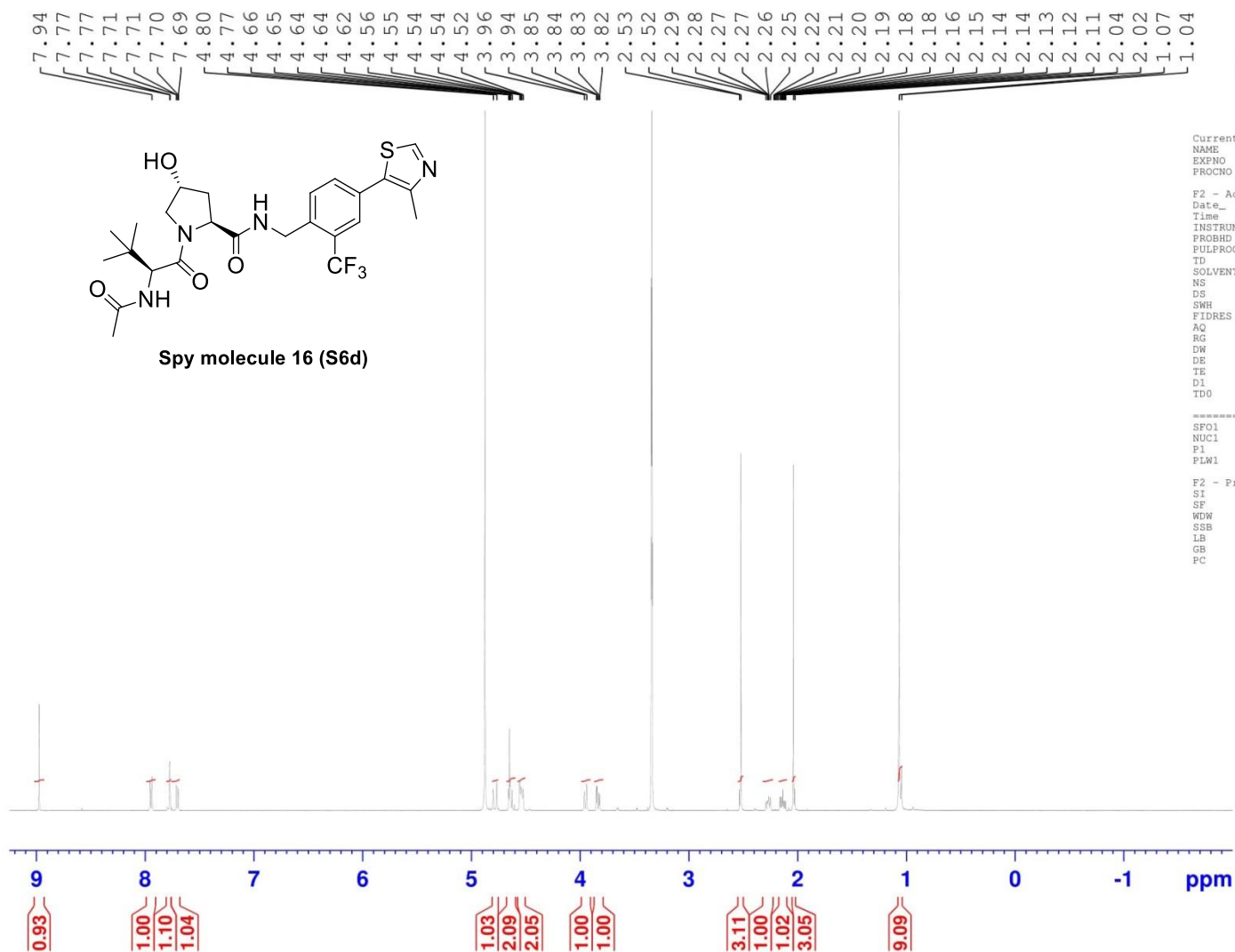
==== CHANNEL f2 =====
 SFO2 500.1320005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 100.00 usec
 PLW2 5.19999981 W
 PLW12 0.06406900 W

F2 - Processing parameters
 SI 262144
 SF 470.5923772 MHz
 WDW EM
 SSB 0
 LB 1.50 Hz
 GB 0
 PC 1.00



SUPPORTING INFORMATION

Spy molecule 16 (Compound S6d)



Current Data Parameters
 NAME GC-R058
 EXPNO 22
 PROCNO 1

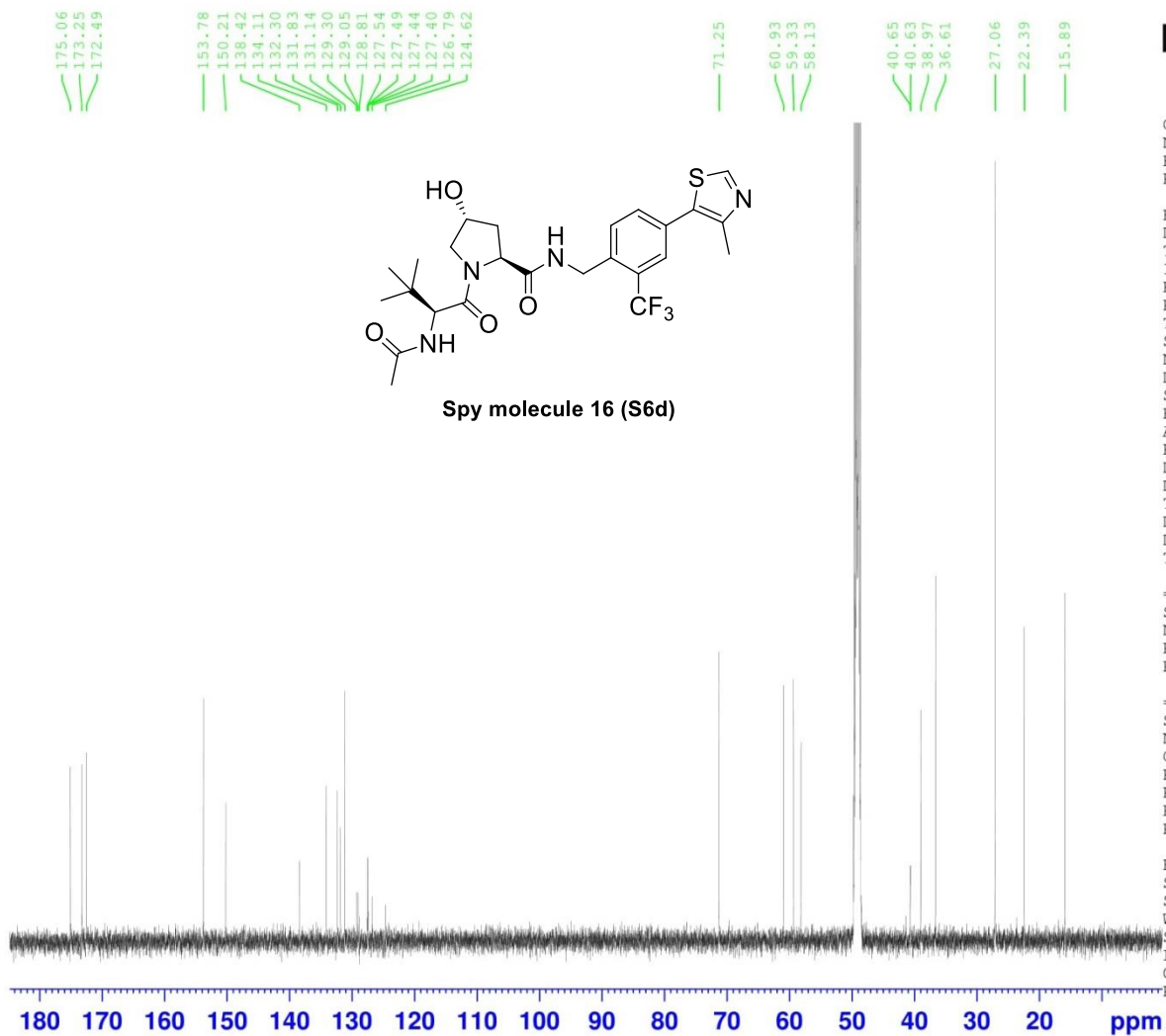
F2 - Acquisition Parameters
 Date_ 20160531
 Time 1.20
 INSTRUM spect
 PROBHD 5 mm QNP 1H/13
 PULPROG zg30
 ID 65536
 SOLVENT MeOD
 NS 256
 DS 2
 SWH 10000.000 Hz
 FIDRES 0.152588 Hz
 AQ 3.2767999 sec
 RG 32
 DW 50.000 usec
 DE 6.50 usec
 TE 298.2 K
 D1 1.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 SF01 500.1330885 MHz
 NUC1 1H
 P1 10.00 usec
 PLW1 25.0000000 W

F2 - Processing parameters
 SI 65536
 SF 500.1299795 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

SUPPORTING INFORMATION

Spy molecule 16 (Compound S6d)



Current Data Parameters
 NAME GCF16
 EXPNO 204
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20181117
 Time 7.41
 INSTRUM spect
 PROBHD 5 mm CPQCI 1H/
 PULPROG zgpg30
 TD 65400
 SOLVENT MeOD
 NS 2048
 DS 4
 SWH 29761.904 Hz
 FIDRES 0.455075 Hz
 AQ 1.0987200 sec
 RG 190.78
 DW 16.800 usec
 DE 18.00 usec
 TE 293.1 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TD0 1

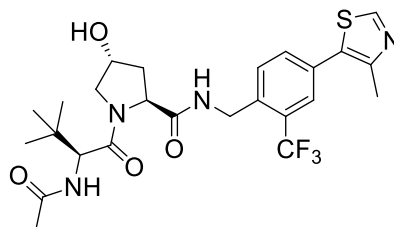
===== CHANNEL f1 =====
 SFO1 125.7703637 MHz
 NUC1 13C
 P1 12.00 usec
 PLW1 133.00000000 W

===== CHANNEL f2 =====
 SFO2 500.1320005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 100.00 usec
 PLW2 5.19999981 W
 PLW12 0.06406900 W
 PLW13 0.06406900 W

F2 - Processing parameters
 SI 131072
 SF 125.7575992 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

SUPPORTING INFORMATION

Spy molecule 16 (Compound S6d)



Spy molecule 16 (S6d)



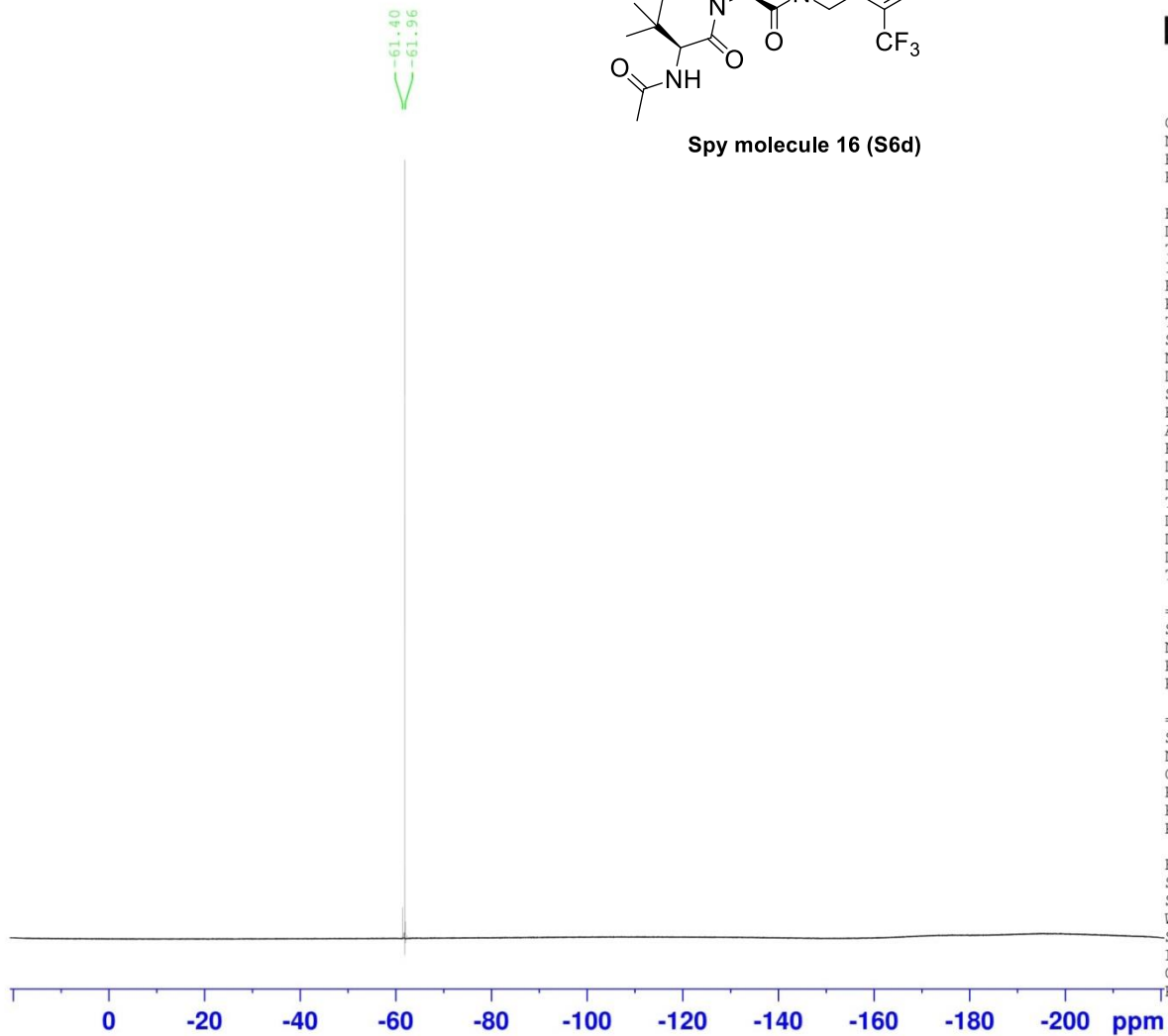
Current Data Parameters
 NAME GC-R058
 EXPNO 27
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20160531
 Time 2.19
 INSTRUM spect
 PROBHD 5 mm QNP 1H/13
 PULPROG zgfhigqn.2
 TD 131072
 SOLVENT MeOD
 NS 16
 DS 4
 SWH 113636.367 Hz
 FIDRES 0.866977 Hz
 AQ 0.5767168 sec
 RG 322
 DW 4.400 usec
 DE 6.50 usec
 TE 298.2 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 D12 0.00002000 sec
 TD0 1

==== CHANNEL f1 =====
 SFO1 470.5453180 MHz
 NUC1 19F
 P1 13.50 usec
 PLW1 25.00000000 W

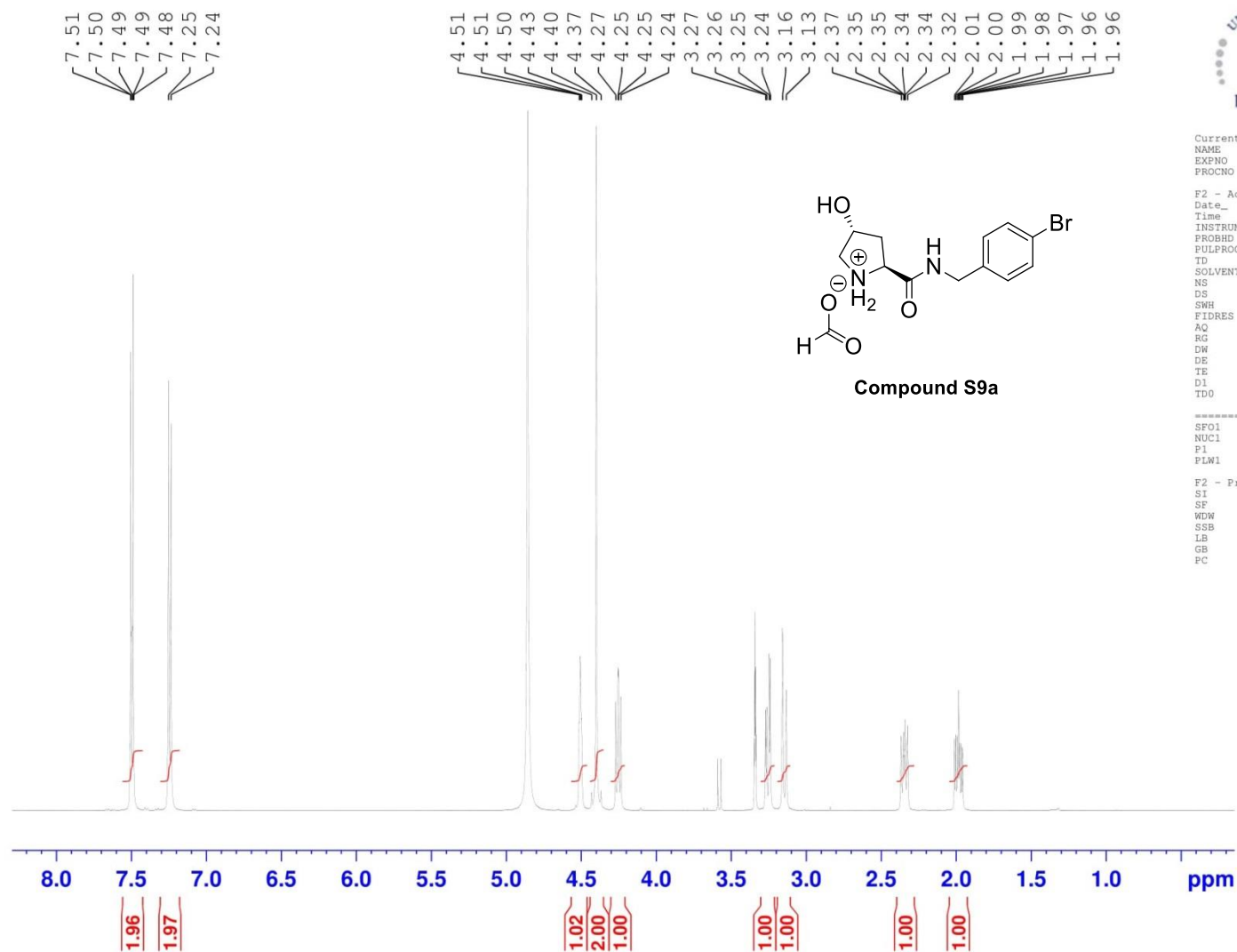
==== CHANNEL f2 =====
 SFO2 500.1320005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 80.00 usec
 PLW2 25.00000000 W
 PLW12 0.39063001 W

F2 - Processing parameters
 SI 65536
 SF 470.5923772 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



SUPPORTING INFORMATION

Compound S9a



Current Data Parameters
 NAME ALC-GC-R72B++
 EXPNO 10
 PROCNO 1

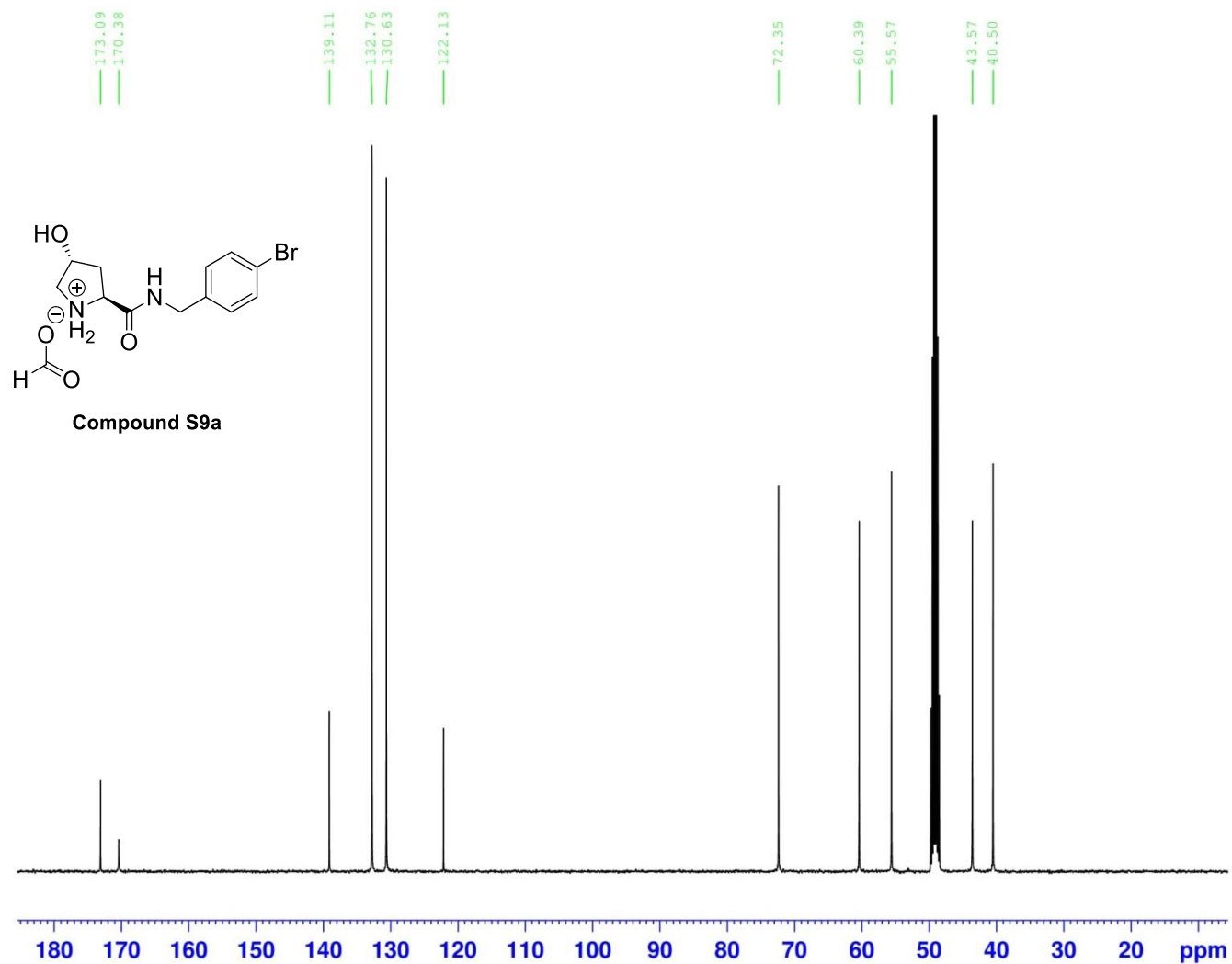
F2 - Acquisition Parameters
 Date_ 20181120
 Time 10.37
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg30
 ID 65536
 SOLVENT MeOD
 NS 16
 DS 2
 SWH 10000.000 Hz
 FIDRES 0.152588 Hz
 AQ 3.2767999 sec
 RG 96.5
 DW 50.000 usec
 DE 6.50 usec
 TE 303.1 K
 D1 1.00000000 sec
 TDO 1

===== CHANNEL f1 =====
 SF01 500.1330885 MHz
 NUC1 1H
 P1 10.00 usec
 PLW1 20.85000038 W

F2 - Processing parameters
 SI 65536
 SF 500.1299958 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

SUPPORTING INFORMATION

Compound S9a



Current Data Parameters
 NAME ALC-GC-R72B+
 EXPNO 20
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20181120
 Time 21.20
 INSTRUM spect
 PROBHD 5 mm PADUL 13C
 PULPROG udeflt
 TD 17996
 SOLVENT MeOD
 NS 2048
 DS 0
 SWH 25000.000 Hz
 FIDRES 1.389198 Hz
 AQ 0.3599200 sec
 RG 196.14
 DW 20.000 usec
 DE 8.66 usec
 TE 294.6 K
 D1 3.00000000 sec
 D11 0.03000000 sec
 D12 0.00002000 sec
 D20 200.00000000 sec
 TD0 1

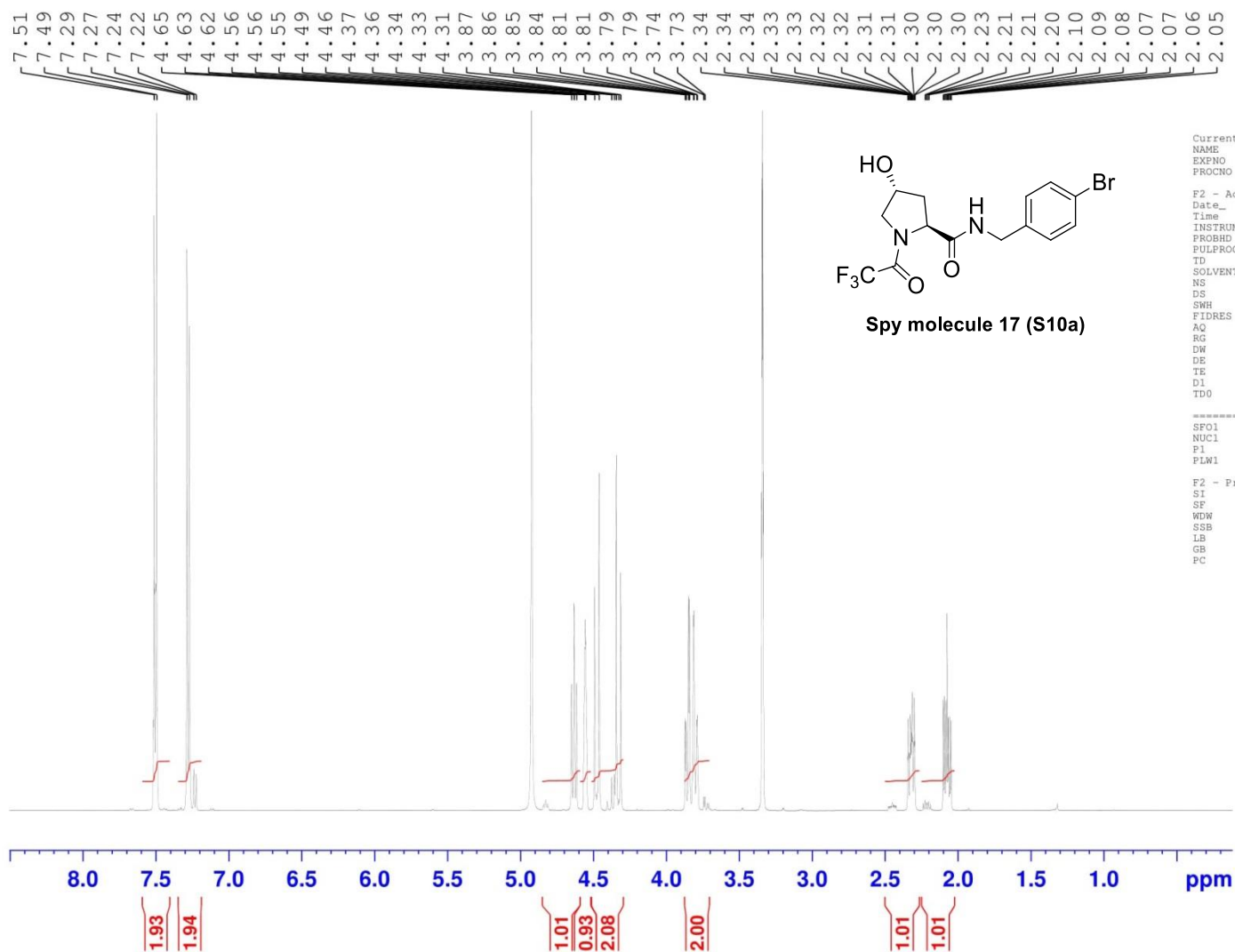
===== CHANNEL f1 =====
 SF01 100.6238346 MHz
 NUC1 13C
 P1 10.00 usec
 P13 2000.00 usec
 P26 500.00 usec
 PLW1 36.00000000 W
 SPNAM[5] Crp60comp.4
 SPOAL5 0.500
 SPOFFS5 0 Hz
 SPW5 5.50040007 W
 SPNAM[8] Crp60,0.5,20.1
 SPOAL8 0.500
 SPOFFS8 0 Hz
 SPW8 5.50040007 W

===== CHANNEL f2 =====
 SF02 400.1316005 MHz
 NUC2 1H
 CPDPRG[2] waltz64
 PCPD2 90.00 usec
 PLW2 20.00000000 W
 PLW12 0.24691001 W

F2 - Processing parameters
 SI 262144
 SF 100.6126211 MHz
 WDW EM
 SSB 0
 LB 2.00 Hz
 GB 0
 PC 1.40

SUPPORTING INFORMATION

Spy molecule 17 (Compound S10a)



```

Current Data Parameters
NAME          GCF17
EXPNO        143
PROCNO       1

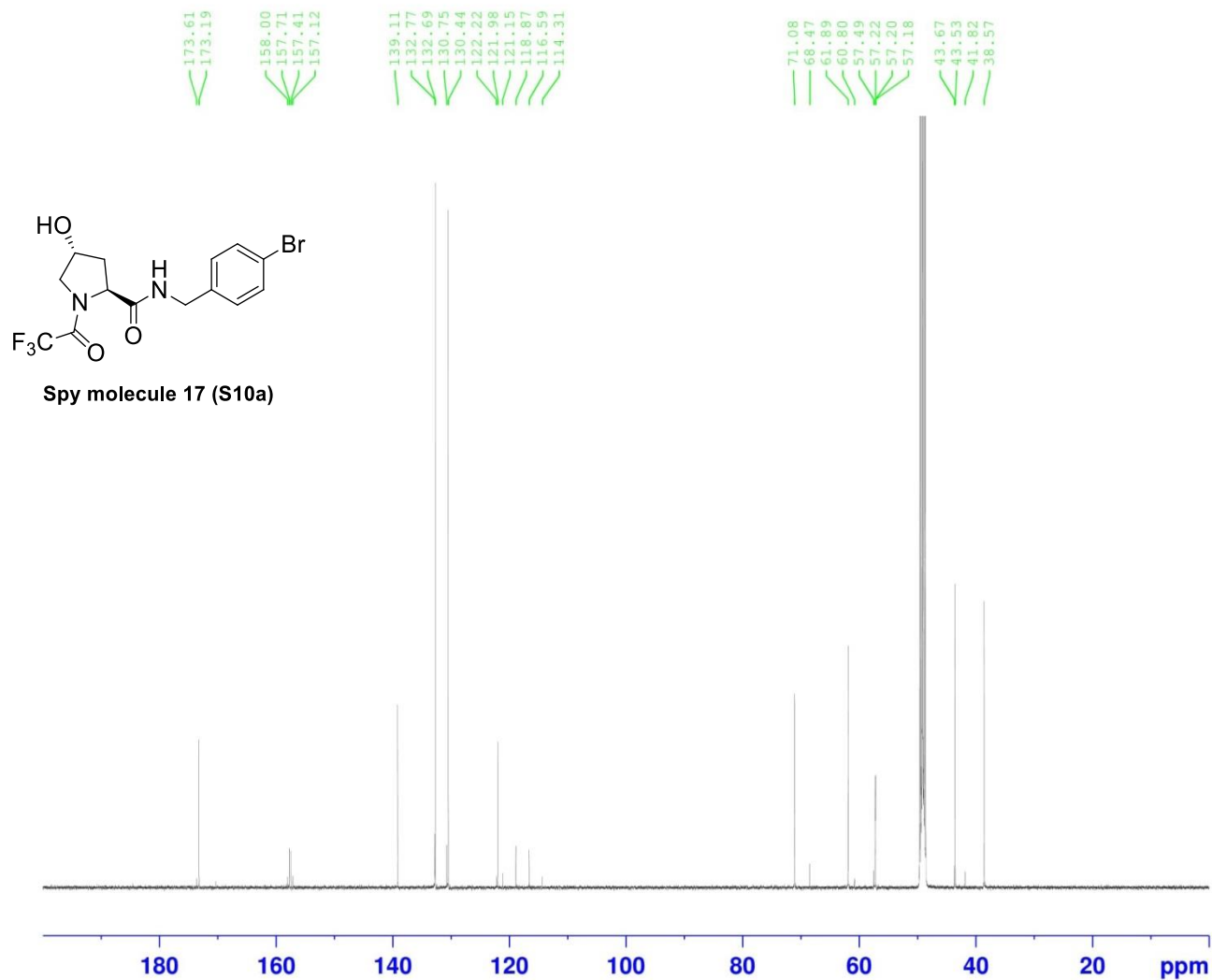
F2 - Acquisition Parameters
Date_        20181114
Time         11.36
INSTRUM     spect
PROBHD      5 mm CPQCI 1H/
PULPROG     zg30
TD          65536
SOLVENT     MeOD
NS          128
DS          2
SWH         10000.000 Hz
FIDRES      0.152588 Hz
AQ          3.2767999 sec
RG          122.56
DW          50.000 usec
DE          10.00 usec
TE          293.1 K
D1          1.00000000 sec
TD0         1

===== CHANNEL f1 =====
SF01       500.1330885 MHz
NUC1       1H
P1         11.00 usec
PLW1       5.19999981 W

F2 - Processing parameters
SI         131072
SF         500.1299946 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00
    
```

SUPPORTING INFORMATION

Spy molecule 17 (Compound S10a)



Current Data Parameters
 NAME GCF17
 EXPNO 147
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20181117
 Time 11.15
 INSTRUM spect
 PROBHD 5 mm CPQCI 1H/
 PULPROG zgpg30
 TD 65400
 SOLVENT MeOD
 NS 4000
 DS 4
 SWH 29761.904 Hz
 FIDRES 0.455075 Hz
 AQ 1.0987200 sec
 RG 190.78
 DW 16.800 usec
 DE 18.00 usec
 TE 293.1 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TD0 1

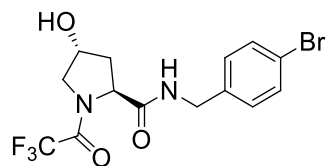
===== CHANNEL f1 =====
 SFO1 125.7703637 MHz
 NUC1 13C
 P1 12.00 usec
 PLW1 133.00000000 W

===== CHANNEL f2 =====
 SFO2 500.1320005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 100.00 usec
 PLW2 5.19999981 W
 PLW12 0.06406900 W
 PLW13 0.06406900 W

F2 - Processing parameters
 SI 131072
 SF 125.7575999 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

SUPPORTING INFORMATION

Spy molecule 17 (Compound S10a)



Spy molecule 17 (S10a)



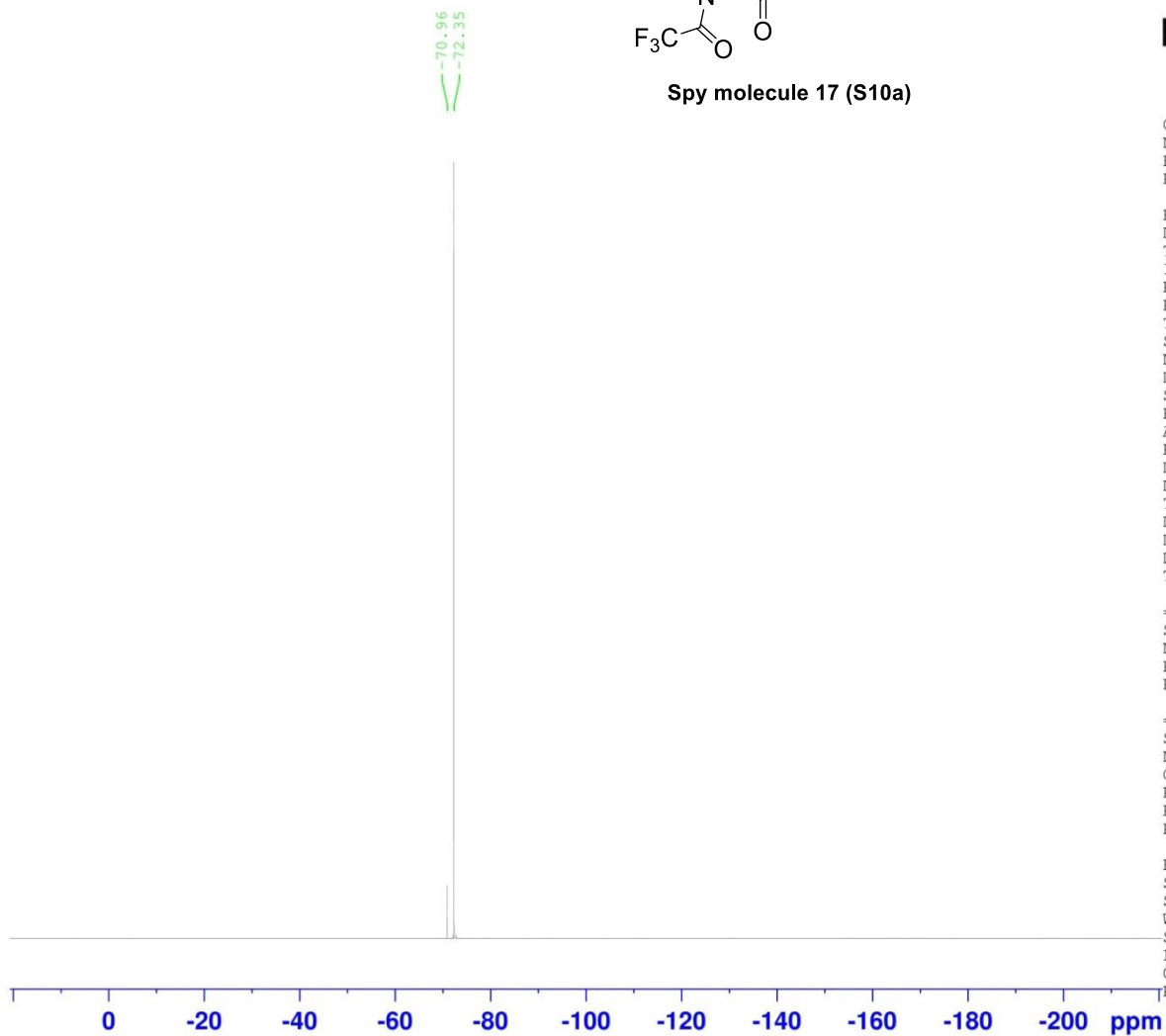
Current Data Parameters
 NAME GCF17
 EXPNO 141
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20181114
 Time 0.15
 INSTRUM spect
 PROBHD 5 mm CPQCI 1H/
 PULPROG zgfhigqn.2
 TD 131072
 SOLVENT H2O+D2O_salt
 NS 256
 DS 4
 SWH 113636.367 Hz
 FIDRES 0.866977 Hz
 AQ 0.5767168 sec
 RG 190.78
 DW 4.400 usec
 DE 18.00 usec
 TE 293.1 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 D12 0.00002000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 470.5453180 MHz
 NUC1 19F
 P1 12.00 usec
 PLW1 5.19999981 W

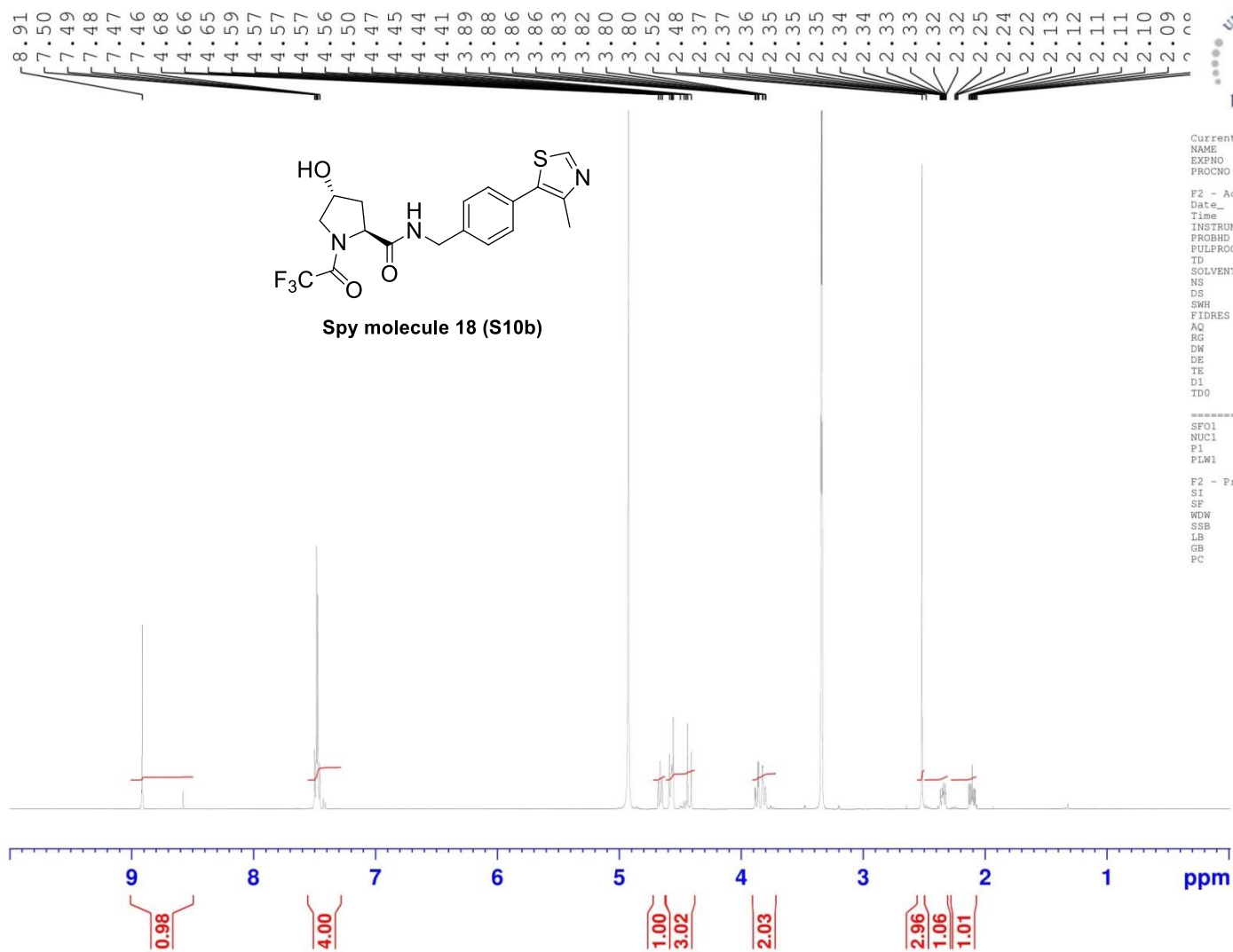
===== CHANNEL f2 =====
 SFO2 500.1320005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 100.00 usec
 PLW2 5.19999981 W
 PLW12 0.06406900 W

F2 - Processing parameters
 SI 262144
 SF 470.5923772 MHz
 WDW EM
 SSB 0
 LB 1.50 Hz
 GB 0
 PC 1.00



SUPPORTING INFORMATION

Spy molecule 18 (Compound S10b)



Current Data Parameters
 NAME GCF20
 EXPNO 144
 PROCNO 1

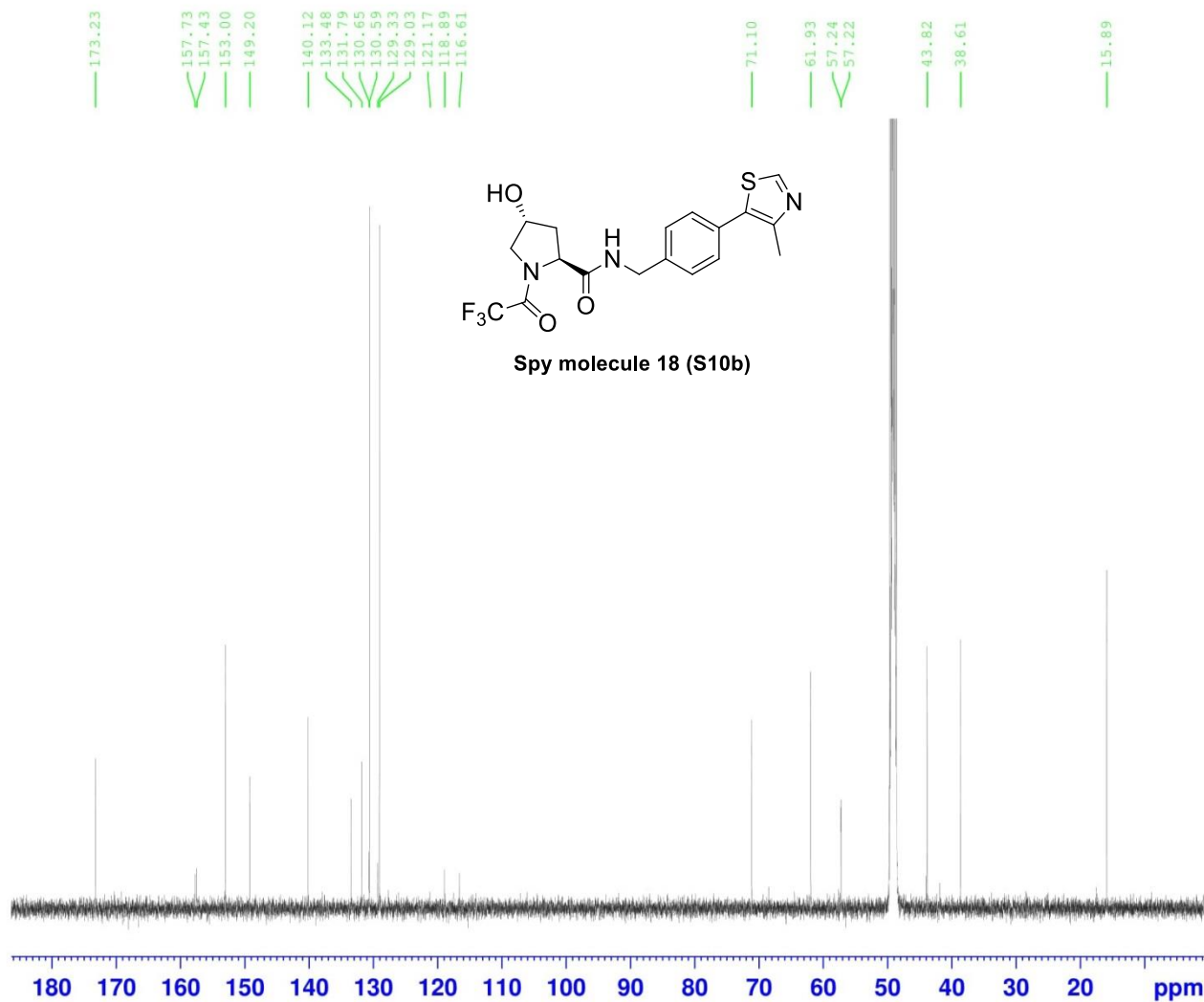
F2 - Acquisition Parameters
 Date_ 20181114
 Time 6.52
 INSTRUM spect
 PROBHD 5 mm CPQCI 1H/
 PULPROG zg30
 ID 65536
 SOLVENT MeOD
 NS 128
 DS 2
 SWH 10000.000 Hz
 FIDRES 0.152588 Hz
 AQ 3.2767999 sec
 RG 190.78
 DW 50.000 usec
 DE 10.00 usec
 TE 293.2 K
 D1 1.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 SF01 500.1330885 MHz
 NUC1 1H
 P1 11.00 usec
 PLW1 5.19999981 W

F2 - Processing parameters
 SI 131072
 SF 500.1299947 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

SUPPORTING INFORMATION

Spy molecule 18 (Compound S10b)



Current Data Parameters
 NAME GCF20
 EXPNO 148
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20181116
 Time 1.25
 INSTRUM spect
 PROBHD 5 mm CPQCI 1H/
 PULPROG zgpg30
 TD 65400
 SOLVENT MeOD
 NS 4000
 DS 4
 SWH 29761.904 Hz
 FIDRES 0.455075 Hz
 AQ 1.0987200 sec
 RG 190.78
 DW 16.800 usec
 DE 18.00 usec
 TE 293.1 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TD0 1

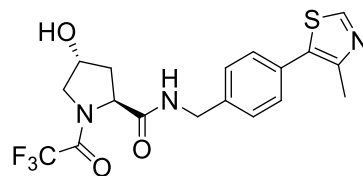
===== CHANNEL f1 =====
 SFO1 125.7703637 MHz
 NUC1 13C
 P1 12.00 usec
 PLW1 133.00000000 W

===== CHANNEL f2 =====
 SFO2 500.1320005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 100.00 usec
 PLW2 5.19999981 W
 PLW12 0.06406900 W
 PLW13 0.06406900 W

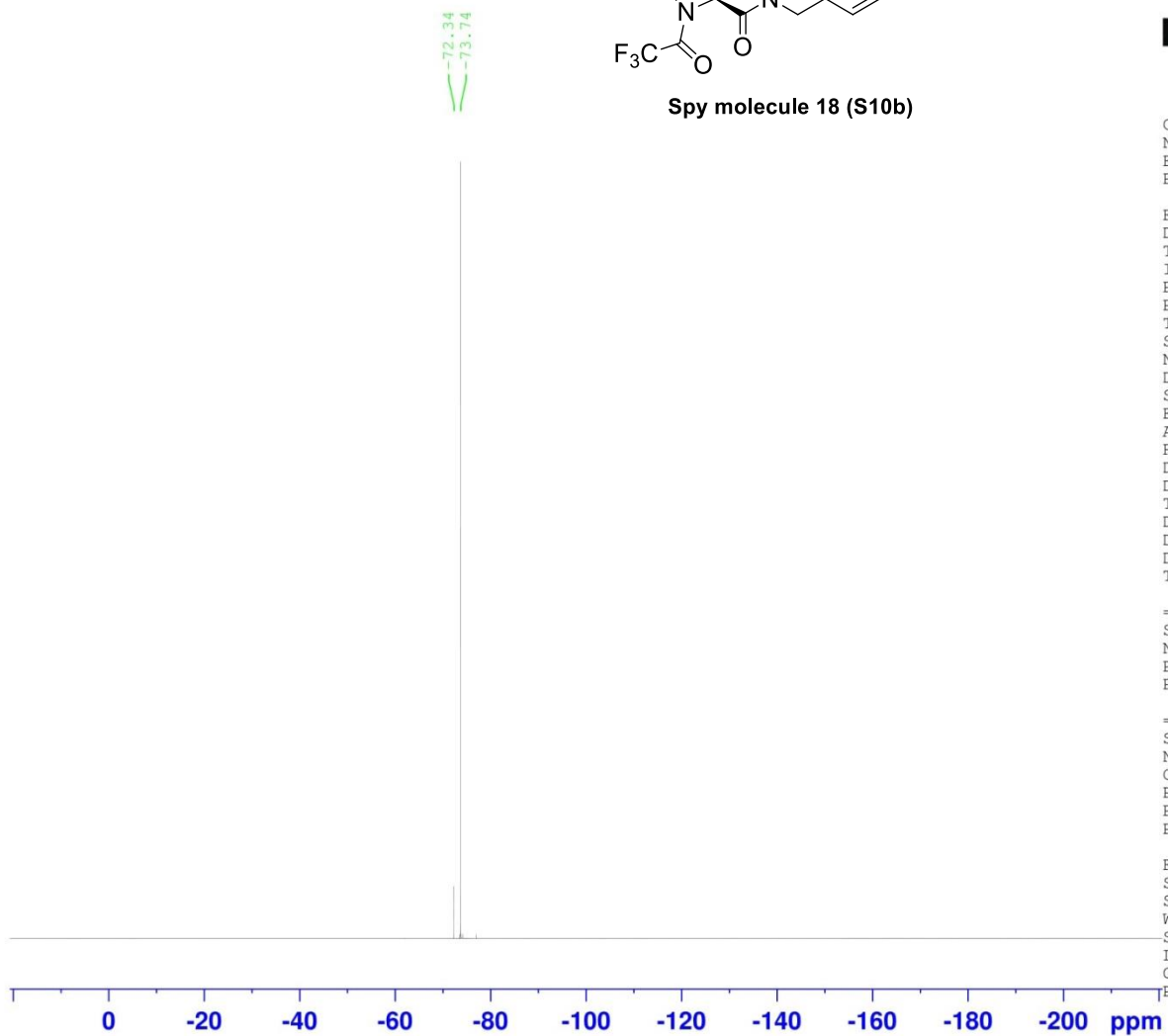
F2 - Processing parameters
 SI 131072
 SF 125.7575991 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

SUPPORTING INFORMATION

Spy molecule 18 (Compound S10b)



Spy molecule 18 (S10b)



Current Data Parameters
 NAME GCF20
 EXPNO 146
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20181114
 Time 8.48
 INSTRUM spect
 PROBHD 5 mm CPQCI 1H/
 PULPROG zgfhigqn.2
 TD 131072
 SOLVENT MeOD
 NS 256
 DS 4
 SWH 113636.367 Hz
 FIDRES 0.866977 Hz
 AQ 0.5767168 sec
 RG 190.78
 DW 4.400 usec
 DE 18.00 usec
 TE 293.2 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 D12 0.00002000 sec
 TD0 1

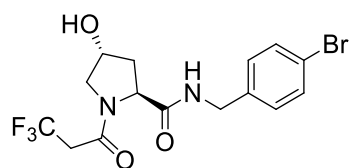
===== CHANNEL f1 =====
 SFO1 470.5453180 MHz
 NUC1 19F
 P1 12.00 usec
 PLW1 5.19999981 W

===== CHANNEL f2 =====
 SFO2 500.1320005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 100.00 usec
 PLW2 5.19999981 W
 PLW12 0.06406900 W

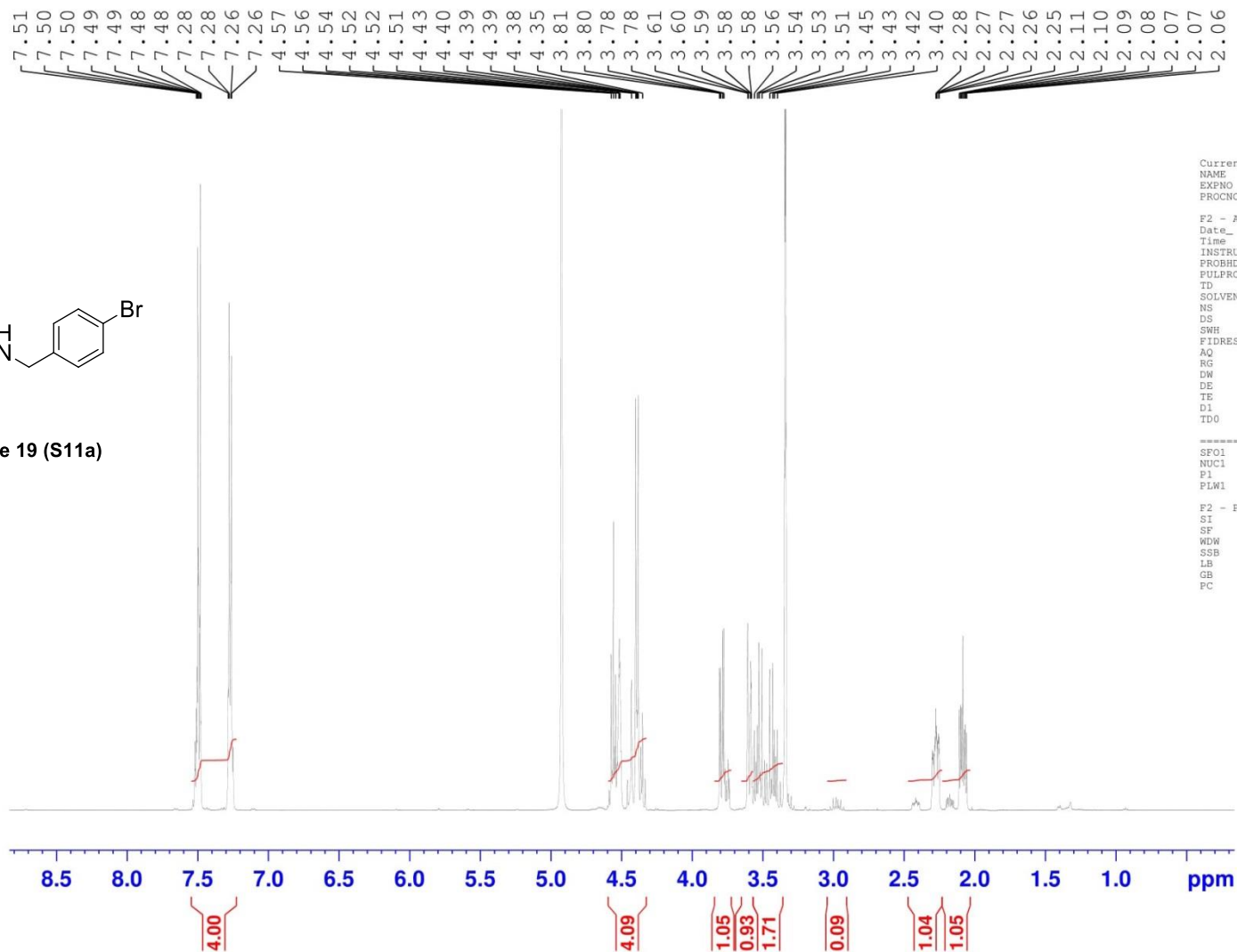
F2 - Processing parameters
 SI 262144
 SF 470.5923772 MHz
 WDW EM
 SSB 0
 LB 1.50 Hz
 GB 0
 PC 1.00

SUPPORTING INFORMATION

Spy molecule 19 (Compound S11a)



Spy molecule 19 (S11a)



```

Current Data Parameters
NAME          GCF18
EXPNO         162
PROCNO        1

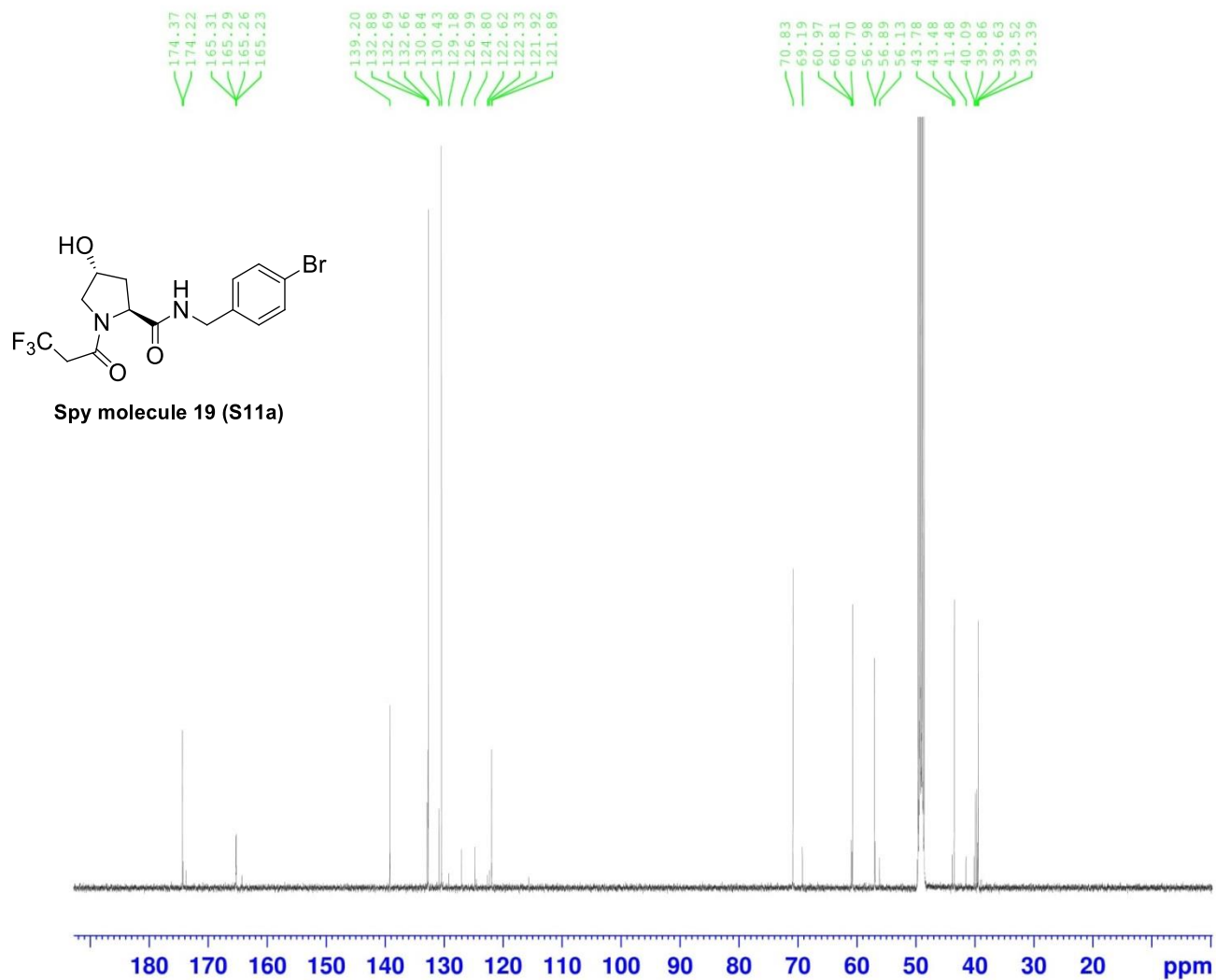
F2 - Acquisition Parameters
Date_         20181114
Time          2.44
INSTRUM       spect
PROBHD        5 mm CPQCI 1H/
PULPROG       zg30
TD            65536
SOLVENT       MeOD
NS            128
DS            2
SWH           10000.000 Hz
FIDRES        0.152588 Hz
AQ            3.2767999 sec
RG            70.68
DW            50.000 usec
DE            10.00 usec
TE            293.1 K
D1            1.00000000 sec
TD0           1

===== CHANNEL f1 =====
SFO1          500.1330885 MHz
NUC1          1H
P1            11.00 usec
PLW1          5.19999981 W

F2 - Processing parameters
SI            131072
SF            500.1299946 MHz
WDW           EM
SSB           0
LB            0.30 Hz
GB            0
PC            1.00
    
```

SUPPORTING INFORMATION

Spy molecule 19 (Compound S11a)



Current Data Parameters
 NAME GCF18
 EXPNO 165
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20181114
 Time 4.32
 INSTRUM spect
 PROBHD 5 mm CPQCI 1H/
 PULPROG zgpg30
 TD 65400
 SOLVENT MeOD
 NS 2048
 DS 4
 SWH 29761.904 Hz
 FIDRES 0.455075 Hz
 AQ 1.0987200 sec
 RG 190.78
 DW 16.800 usec
 DE 18.00 usec
 TE 293.2 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TD0 1

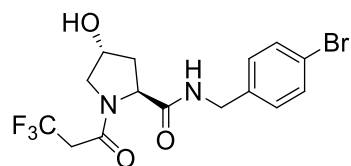
===== CHANNEL f1 =====
 SFO1 125.7703637 MHz
 NUC1 13C
 P1 12.00 usec
 PLW1 133.00000000 W

===== CHANNEL f2 =====
 SFO2 500.1320005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 100.00 usec
 PLW2 5.19999981 W
 PLW12 0.06406900 W
 PLW13 0.06406900 W

F2 - Processing parameters
 SI 131072
 SF 125.7575999 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

SUPPORTING INFORMATION

Spy molecule 19 (Compound S11a)



Spy molecule 19 (S11a)



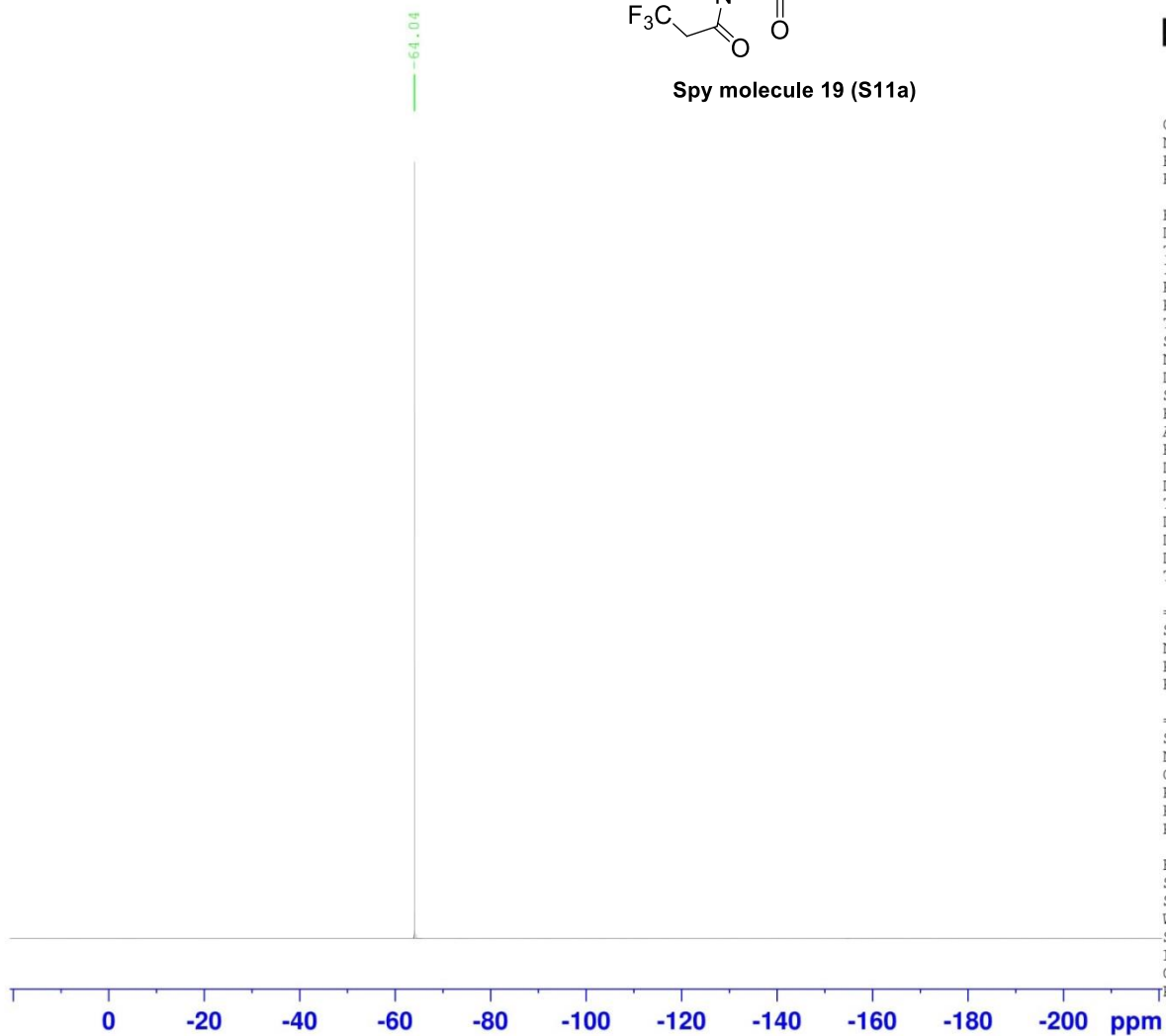
Current Data Parameters
 NAME GCF18
 EXPNO 160
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20181114
 Time 2.34
 INSTRUM spect
 PROBHD 5 mm CPQCI 1H/
 PULPROG zgfhigqn.2
 TD 131072
 SOLVENT MeOD
 NS 256
 DS 4
 SWH 113636.367 Hz
 FIDRES 0.866977 Hz
 AQ 0.5767168 sec
 RG 190.78
 DW 4.400 usec
 DE 18.00 usec
 TE 293.1 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 D12 0.00002000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 470.5453180 MHz
 NUC1 19F
 P1 12.00 usec
 PLW1 5.19999981 W

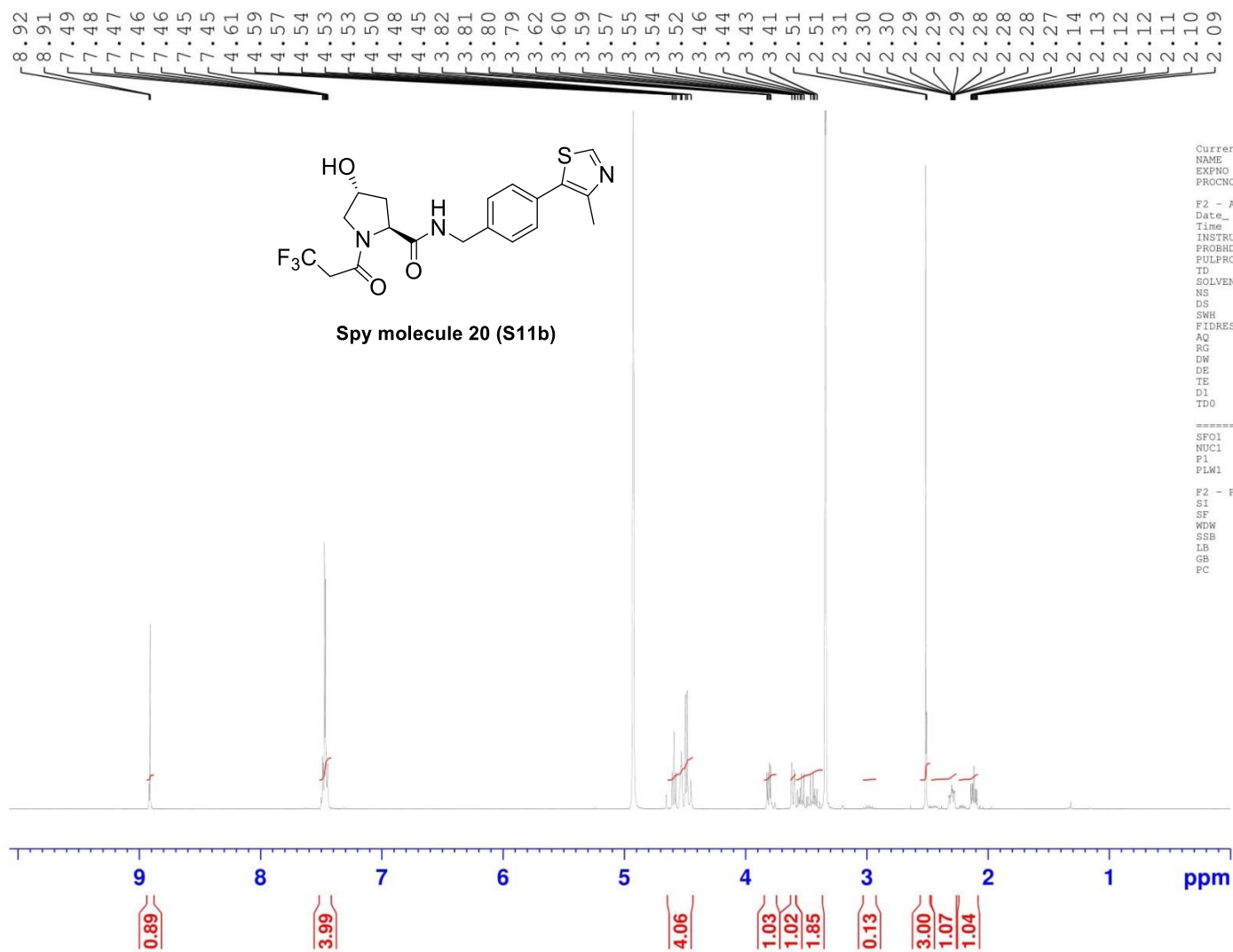
===== CHANNEL f2 =====
 SFO2 500.1320005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 100.00 usec
 PLW2 5.19999981 W
 PLW12 0.06406900 W

F2 - Processing parameters
 SI 262144
 SF 470.5923772 MHz
 WDW EM
 SSB 0
 LB 1.50 Hz
 GB 0
 PC 1.00



SUPPORTING INFORMATION

Spy molecule 20 (Compound S11b)



Current Data Parameters
 NAME GCF21
 EXPNO 172
 PROCNO 1

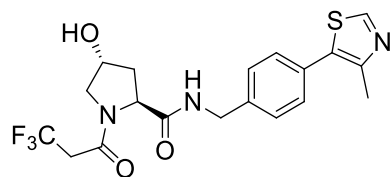
F2 - Acquisition Parameters
 Date_ 20181114
 Time 4.52
 INSTRUM spect
 PROBHD 5 mm CPOCF 1H/
 PULPROG zg30
 TD 65536
 SOLVENT MeOD
 NS 128
 DS 2
 SWH 10000.000 Hz
 FIDRES 0.152588 Hz
 AQ 3.2767999 sec
 RG 171.57
 DW 50.000 usec
 DE 10.00 usec
 TE 293.1 K
 D1 1.0000000 sec
 TD0 1

===== CHANNEL f1 =====
 SF01 500.1330885 MHz
 NUC1 1H
 P1 11.00 usec
 PLW1 5.19999981 W

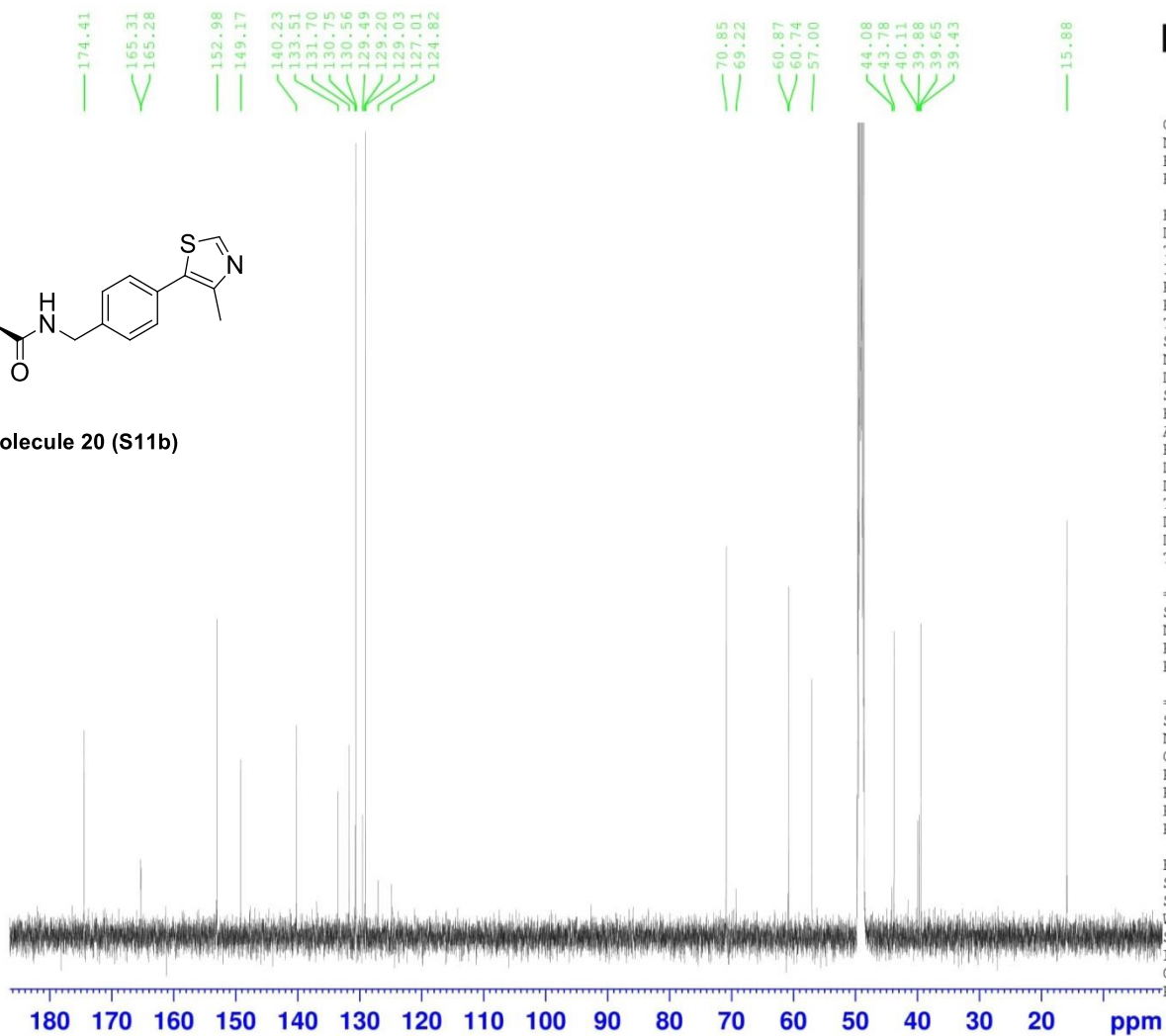
F2 - Processing parameters
 SI 131072
 SF 500.1299946 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

SUPPORTING INFORMATION

Spy molecule 20 (Compound S11b)



Spy molecule 20 (S11b)



Current Data Parameters
 NAME GCF21
 EXPNO 174
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20181114
 Time 6.40
 INSTRUM spect
 PROBHD 5 mm CPQCI 1H/
 PULPROG zgpg30
 TD 65400
 SOLVENT MeOD
 NS 2048
 DS 4
 SWH 29761.904 Hz
 FIDRES 0.455075 Hz
 AQ 1.0987200 sec
 RG 190.78
 DW 16.800 usec
 DE 18.00 usec
 TE 293.2 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TD0 1

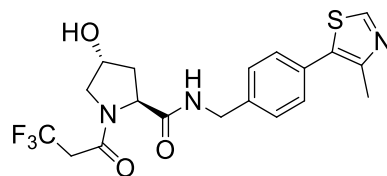
===== CHANNEL f1 =====
 SFO1 125.7703637 MHz
 NUC1 13C
 P1 12.00 usec
 PLW1 133.00000000 W

===== CHANNEL f2 =====
 SFO2 500.1320005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 100.00 usec
 PLW2 5.19999981 W
 PLW12 0.06406900 W
 PLW13 0.06406900 W

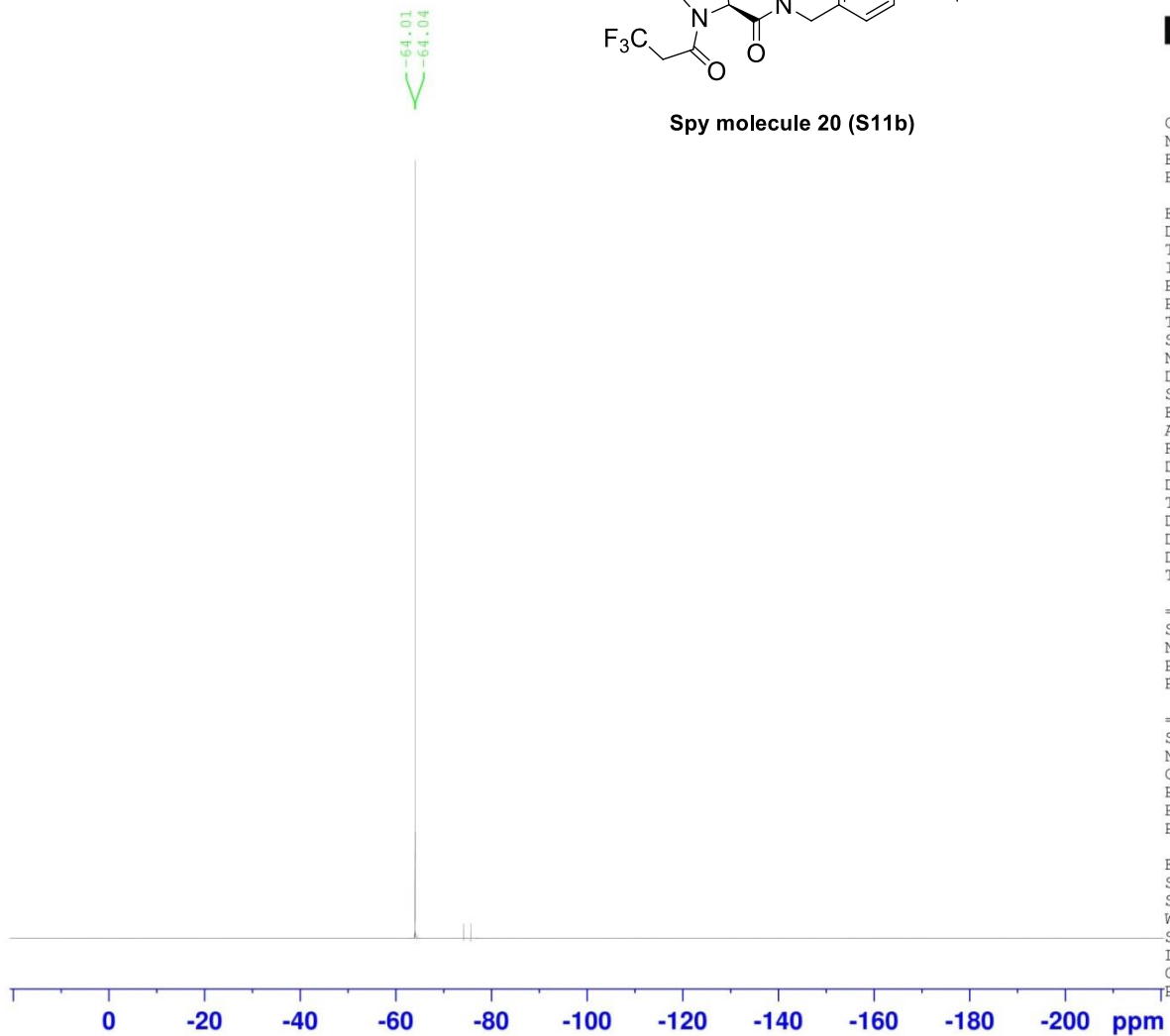
F2 - Processing parameters
 SI 131072
 SF 125.7575992 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

SUPPORTING INFORMATION

Spy molecule 20 (Compound S11b)



Spy molecule 20 (S11b)



Current Data Parameters
 NAME GCF21
 EXPNO 170
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20181114
 Time 4.42
 INSTRUM spect
 PROBHD 5 mm CPQCI 1H/
 PULPROG zgfhigqn.2
 TD 131072
 SOLVENT MeOD
 NS 256
 DS 4
 SWH 113636.367 Hz
 FIDRES 0.866977 Hz
 AQ 0.5767168 sec
 RG 190.78
 DW 4.400 usec
 DE 18.00 usec
 TE 293.1 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 D12 0.00002000 sec
 TD0 1

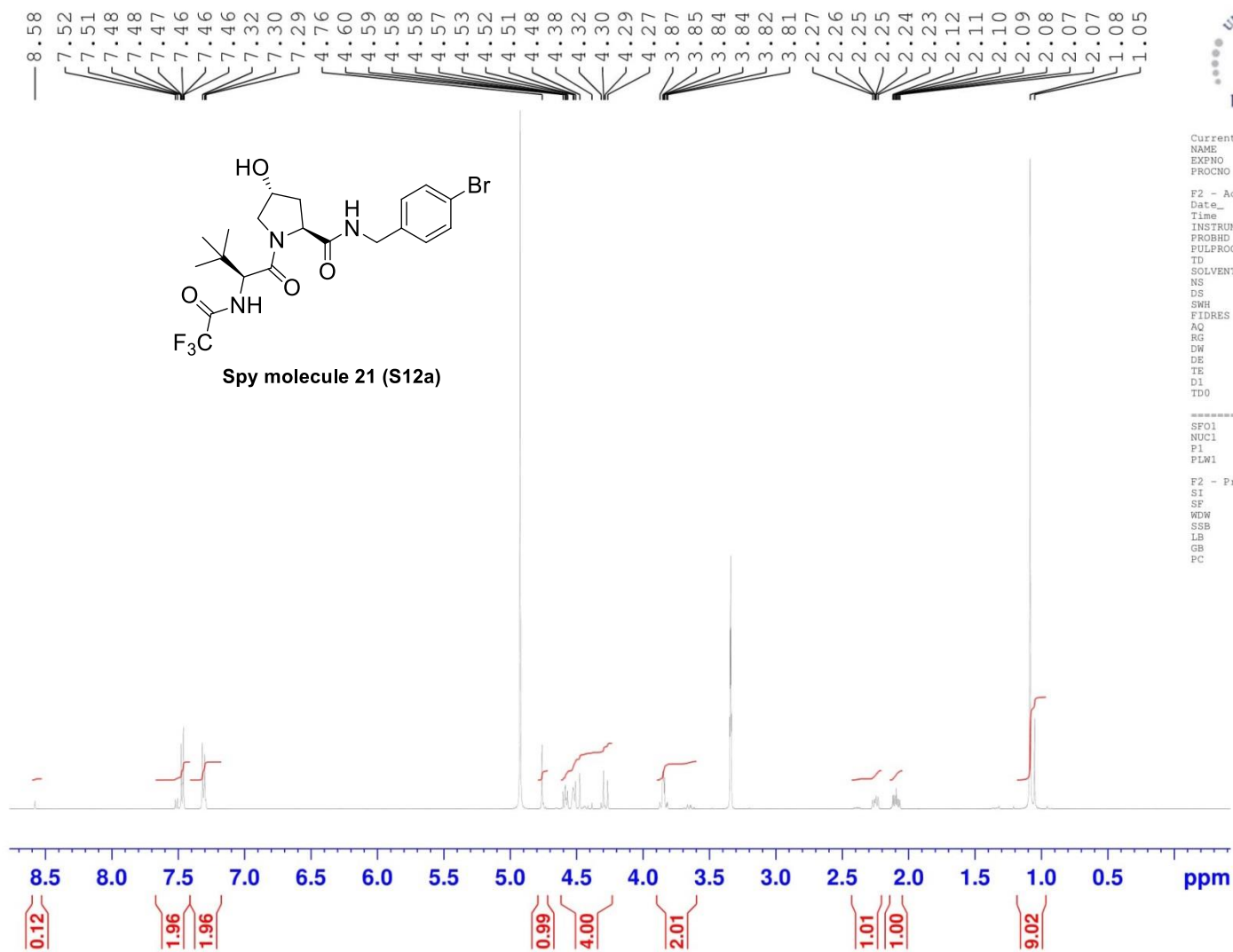
===== CHANNEL f1 =====
 SFO1 470.5453180 MHz
 NUC1 19F
 P1 12.00 usec
 PLW1 5.19999981 W

===== CHANNEL f2 =====
 SFO2 500.1320005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 100.00 usec
 PLW2 5.19999981 W
 PLW12 0.06406900 W

F2 - Processing parameters
 SI 262144
 SF 470.5923772 MHz
 WDW EM
 SSB 0
 LB 1.50 Hz
 GB 0
 PC 1.00

SUPPORTING INFORMATION

Spy molecule 21 (Compound S12a)



Current Data Parameters
 NAME GCF19
 EXPNO 152
 PROCNO 1

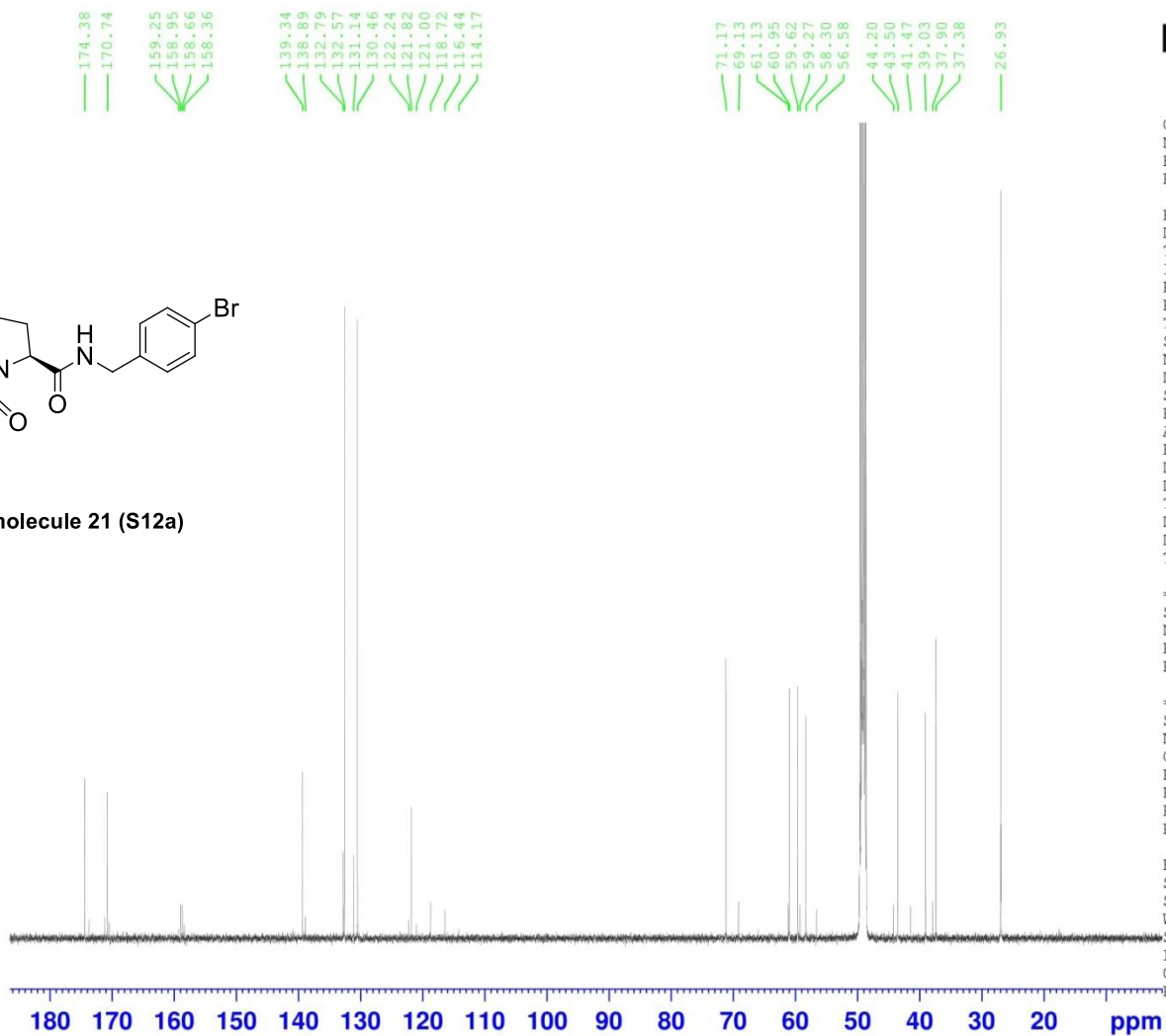
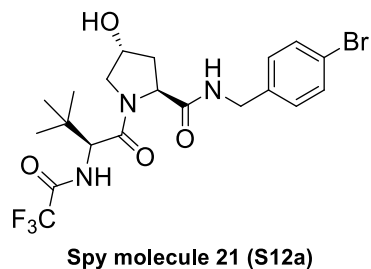
F2 - Acquisition Parameters
 Date_ 20181114
 Time 0.36
 INSTRUM spect
 PROBHD 5 mm CPQCI 1H/
 PULPROG zg30
 ID 65536
 SOLVENT MeOD
 NS 128
 DS 2
 SWH 10000.000 Hz
 FIDRES 0.152588 Hz
 AQ 3.2767999 sec
 RG 122.56
 DW 50.000 usec
 DE 10.00 usec
 TE 293.2 K
 D1 1.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 SF01 500.1330885 MHz
 NUC1 1H
 P1 11.00 usec
 PLW1 5.19999981 W

F2 - Processing parameters
 SI 131072
 SF 500.1299947 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

SUPPORTING INFORMATION

Spy molecule 21 (Compound S12a)



Current Data Parameters
 NAME GCF19
 EXPNO 155
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20181116
 Time 4.59
 INSTRUM spect
 PROBHD 5 mm CPQCI 1H/
 PULPROG zgpg30
 TD 65400
 SOLVENT MeOD
 NS 4000
 DS 4
 SWH 29761.904 Hz
 FIDRES 0.455075 Hz
 AQ 1.0987200 sec
 RG 190.78
 DW 16.800 usec
 DE 18.00 usec
 TE 293.1 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TD0 1

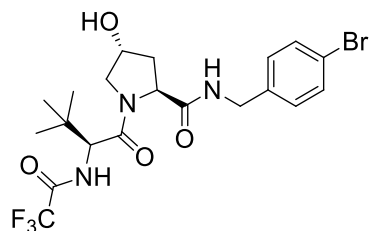
===== CHANNEL f1 =====
 SFO1 125.7703637 MHz
 NUC1 13C
 P1 12.00 usec
 PLW1 133.00000000 W

===== CHANNEL f2 =====
 SFO2 500.1320005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 100.00 usec
 PLW2 5.19999981 W
 PLW12 0.06406900 W
 PLW13 0.06406900 W

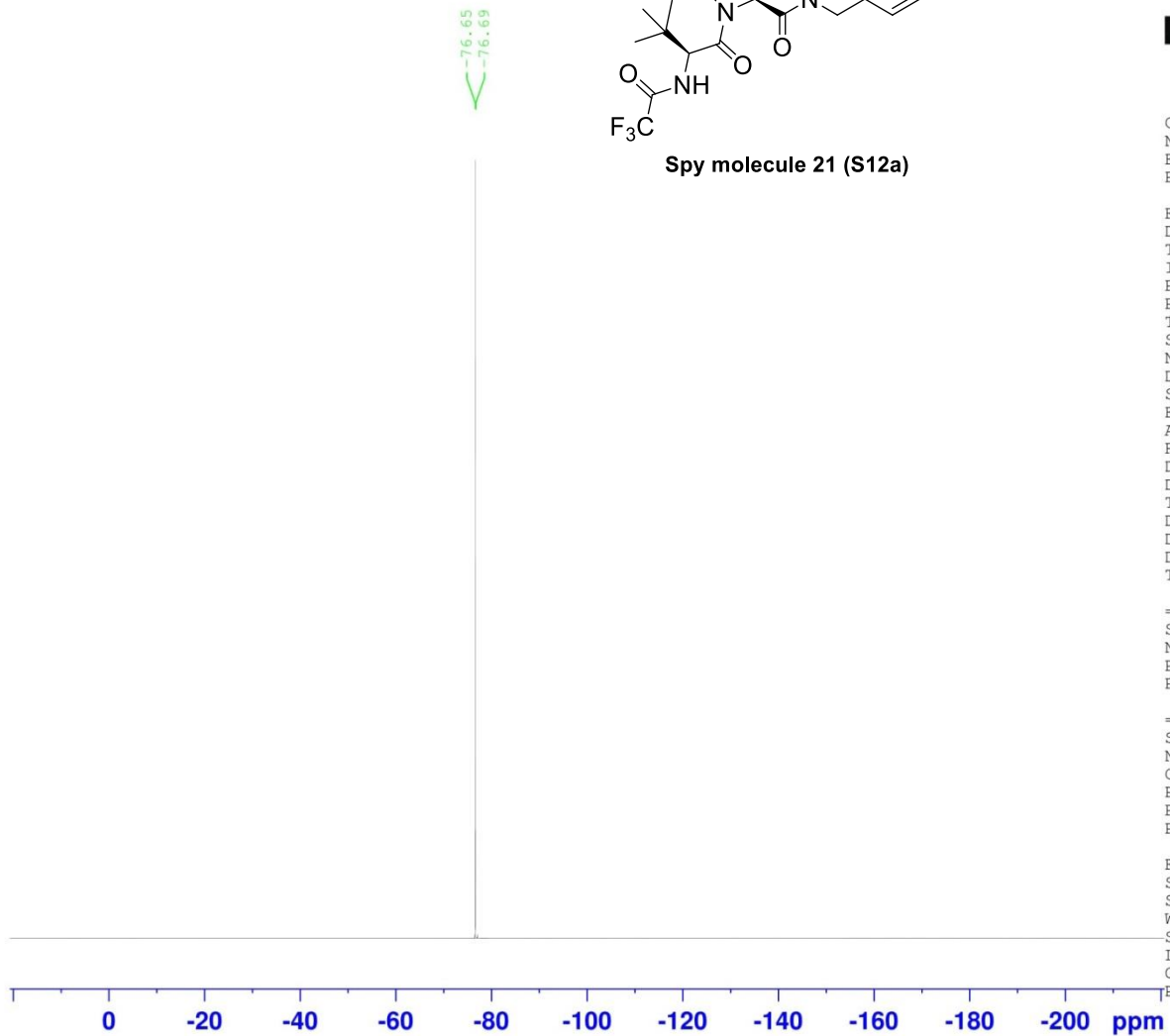
F2 - Processing parameters
 SI 131072
 SF 125.7575996 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

SUPPORTING INFORMATION

Spy molecule 21 (Compound S12a)



Spy molecule 21 (S12a)



Current Data Parameters
 NAME GCF19
 EXPNO 150
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20181114
 Time 0.26
 INSTRUM spect
 PROBHD 5 mm CPQCI 1H/
 PULPROG zgfhigqn.2
 TD 131072
 SOLVENT MeOD
 NS 256
 DS 4
 SWH 113636.367 Hz
 FIDRES 0.866977 Hz
 AQ 0.5767168 sec
 RG 190.78
 DW 4.400 usec
 DE 18.00 usec
 TE 293.1 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 D12 0.00002000 sec
 TD0 1

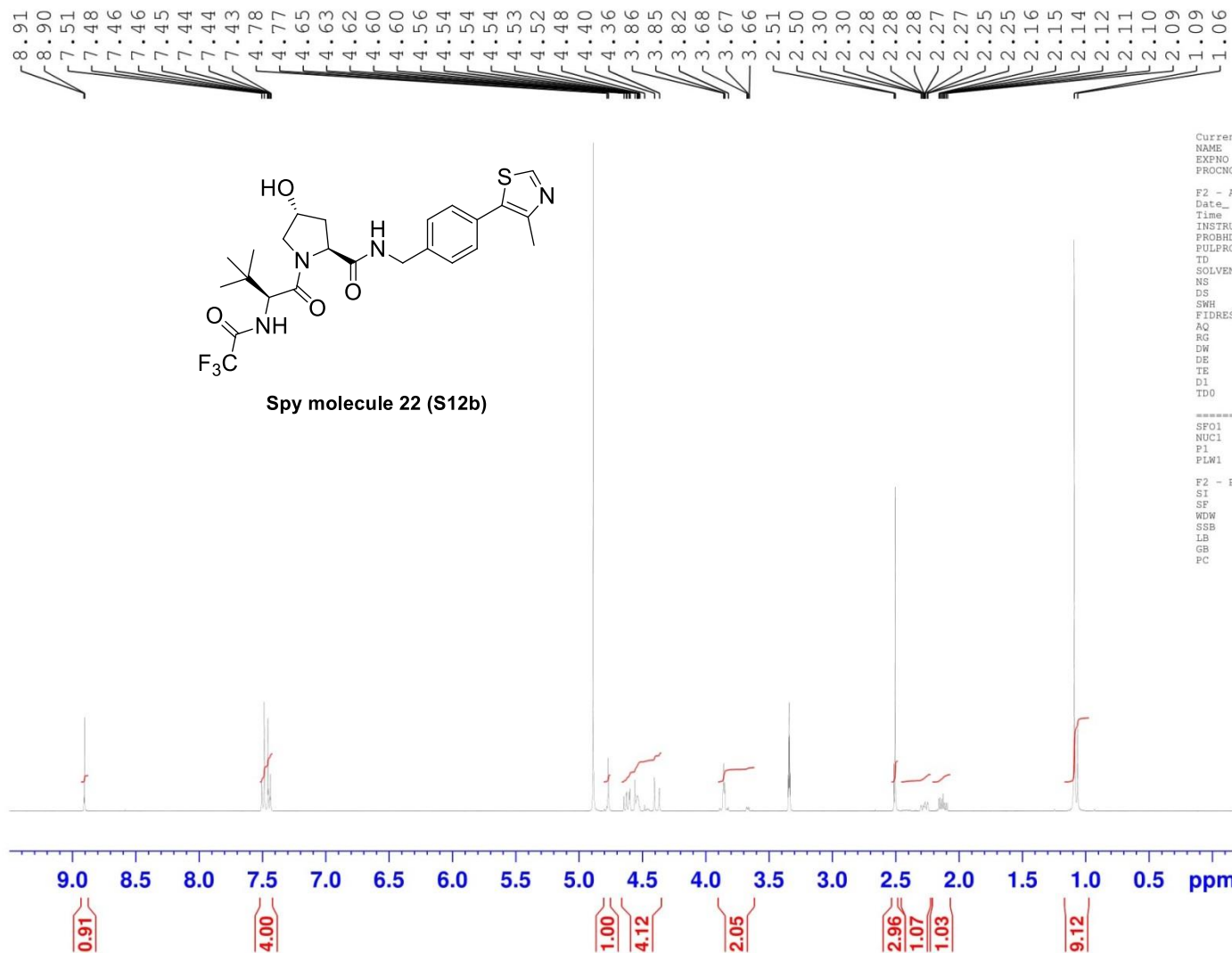
==== CHANNEL f1 =====
 SFO1 470.5453180 MHz
 NUC1 19F
 P1 12.00 usec
 PLW1 5.19999981 W

==== CHANNEL f2 =====
 SFO2 500.1320005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 100.00 usec
 PLW2 5.19999981 W
 PLW12 0.06406900 W

F2 - Processing parameters
 SI 262144
 SF 470.5923772 MHz
 WDW EM
 SSB 0
 LB 1.50 Hz
 GB 0
 PC 1.00

SUPPORTING INFORMATION

Spy molecule 22 (Compound S12b)



Current Data Parameters
 NAME ALC-GC-R090
 EXPNO 11
 PROCNO 1

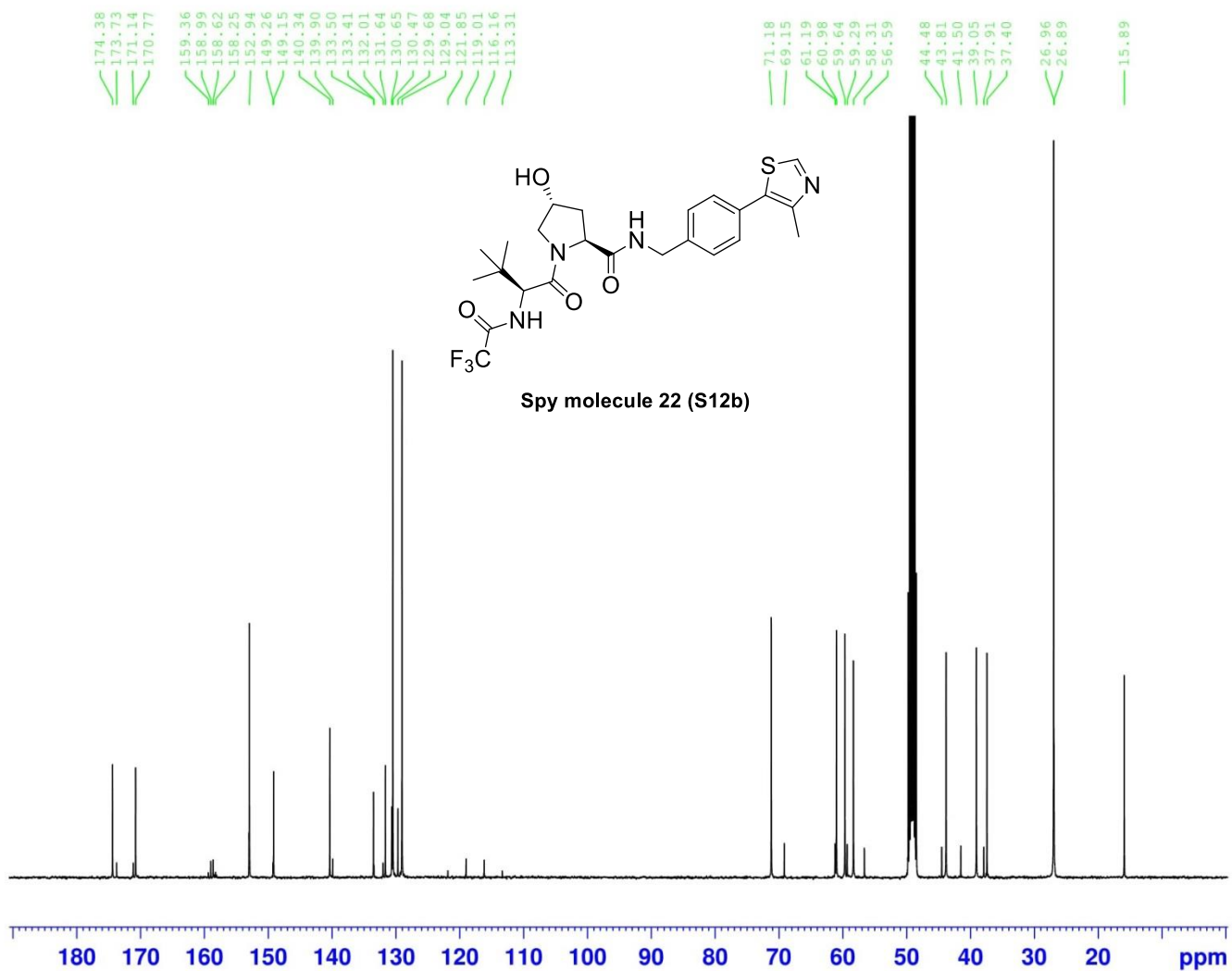
F2 - Acquisition Parameters
 Date_ 20160715
 Time 6.40
 INSTRUM spect
 PROBHD 5 mm PADUL13C
 PULPROG zg30
 TD 131072
 SOLVENT MeOD
 NS 16
 DS 4
 SWH 12019.230 Hz
 FIDRES 0.091699 Hz
 AQ 5.4525952 sec
 RG 143.61
 DW 41.600 usec
 DE 12.17 usec
 TE 298.2 K
 D1 0.1000000 sec
 TDO 1

===== CHANNEL f1 =====
 SF01 400.1324710 MHz
 NUC1 1H
 P1 10.00 usec
 PLW1 20.00000000 W

F2 - Processing parameters
 SI 131072
 SF 400.1299956 MHz
 WDW EM
 SSB 0
 LB 0.10 Hz
 GB 0
 PC 1.00

SUPPORTING INFORMATION

Spy molecule 22 (Compound S12b)



```

Current Data Parameters
NAME      ALC-GC-R090
EXPNO     10
PROCNO    1

F2 - Acquisition Parameters
Date_     20160715
Time      6.37
INSTRUM   spect
PROBHD    5 mm PADUL 13C
PULPROG   udef1
TD         17996
SOLVENT   MeOD
NS         11000
DS         0
SWH        25000.000 Hz
FIDRES     1.389198 Hz
AQ         0.3599200 sec
RG         196.14
DW         20.000 usec
DE         8.66 usec
TE         298.2 K
D1         3.00000000 sec
D11        0.03000000 sec
D12        0.00002000 sec
D20        200.00000000 sec
TD0        1

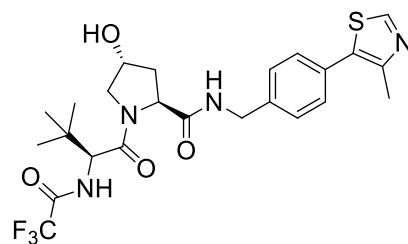
===== CHANNEL f1 =====
SF01      100.6238346 MHz
NUC1       13C
P1         10.00 usec
P13        2000.00 usec
P26        500.00 usec
PLW1       36.00000000 W
SPNAM[5]   Crp60comp.4
SPOAL5     0.500
SPOFFS5    0 Hz
SPW5       5.50040007 W
SPNAM[8]   Crp60,0.5,20.1
SPOAL8     0.500
SPOFFS8    0 Hz
SPW8       5.50040007 W

===== CHANNEL f2 =====
SF02      400.1316005 MHz
NUC2       1H
CPDPRG[2]  waltz64
PCPD2      90.00 usec
PLW2       20.00000000 W
PLW12      0.24691001 W

F2 - Processing parameters
SI         262144
SF         100.6126179 MHz
WDW        EM
SSB        0
LB         2.00 Hz
GB         0
PC         1.40
    
```

SUPPORTING INFORMATION

Spy molecule 22 (Compound S12b)



Spy molecule 22 (S12b)



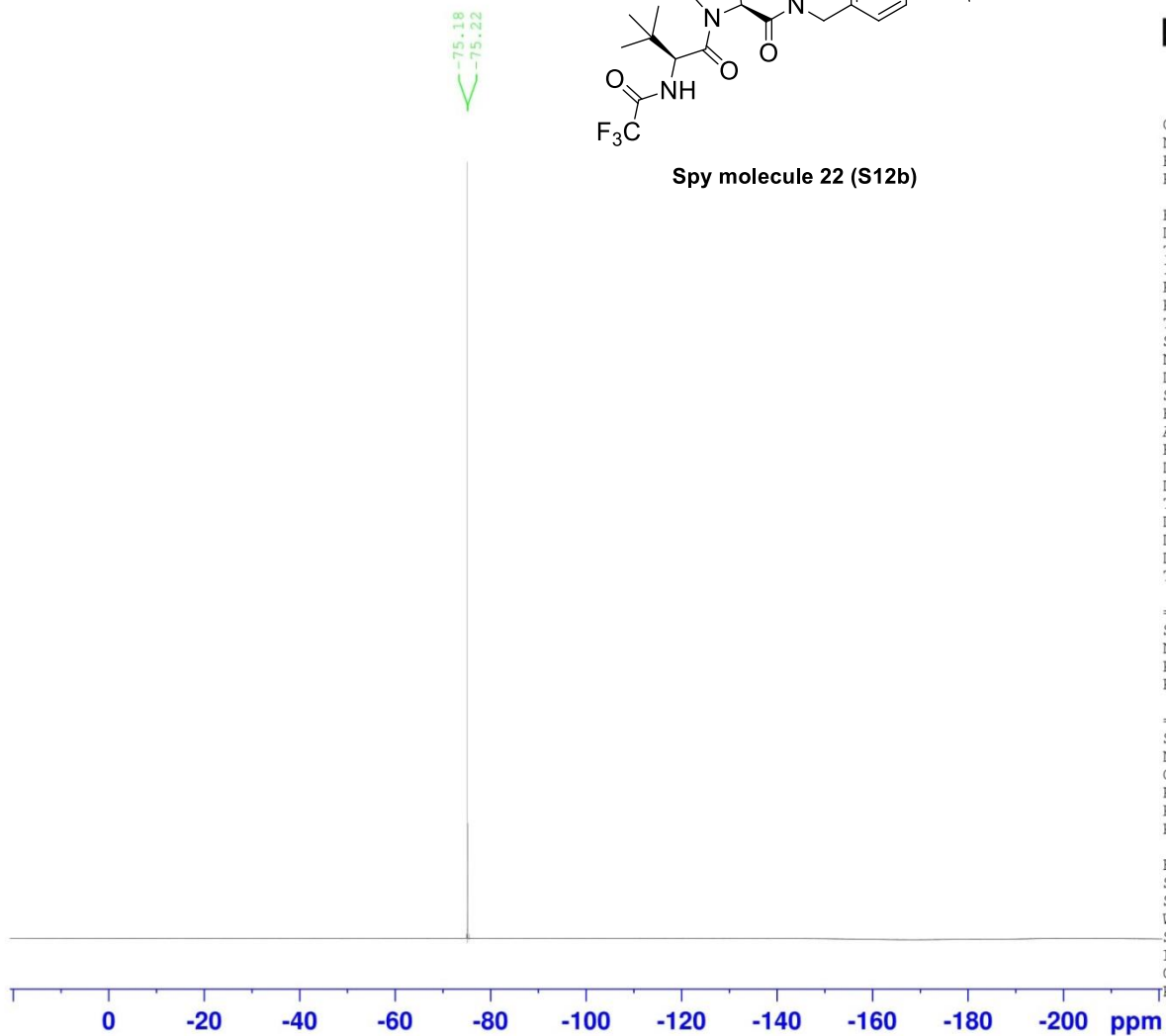
Current Data Parameters
 NAME AC-GC-R090
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20160717
 Time 14.34
 INSTRUM spect
 PROBHD 5 mm QNP 1H/13
 PULPROG zgfhigqn.2
 TD 131072
 SOLVENT MeOD
 NS 16
 DS 4
 SWH 113636.367 Hz
 FIDRES 0.866977 Hz
 AQ 0.5767168 sec
 RG 362
 DW 4.400 usec
 DE 6.50 usec
 TE 298.2 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 D12 0.00002000 sec
 TD0 1

==== CHANNEL f1 =====
 SFO1 470.5453180 MHz
 NUC1 19F
 P1 13.50 usec
 PLW1 25.00000000 W

==== CHANNEL f2 =====
 SFO2 500.1320005 MHz
 NUC2 1H
 CPDPRG[2] waltz16
 PCPD2 80.00 usec
 PLW2 25.00000000 W
 PLW12 0.39063001 W

F2 - Processing parameters
 SI 65536
 SF 470.5923772 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



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