

SUPPORTING INFORMATION:

Optimizing Blended Collagen-Fibrin Hydrogels for Cardiac Tissue Engineering with Human iPSC-derived Cardiomyocytes

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10 Pages, 7 Figures, 13 Tables

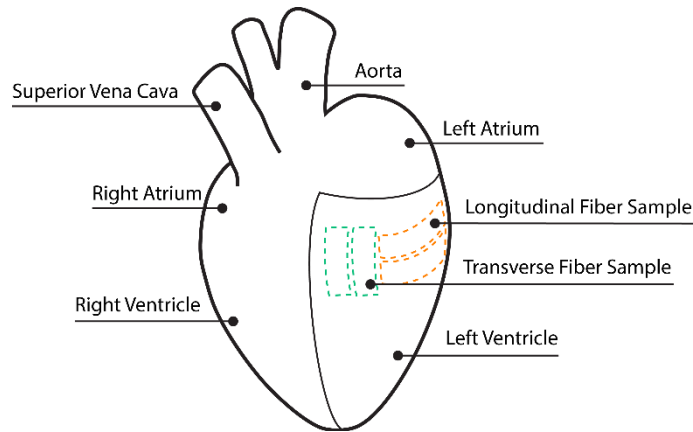


Figure S1: Native rat myocardium sample collection. Longitudinal fiber samples approximately 2 mm x 10 mm (orange) were collected from the left ventricle myocardial layer with the long axis aligned with cardiac muscle fibers. Conversely, transverse fiber samples with the same dimensions (green) were collected from the left ventricle myocardial layer with the long axis perpendicular to cardiac muscle fibers.

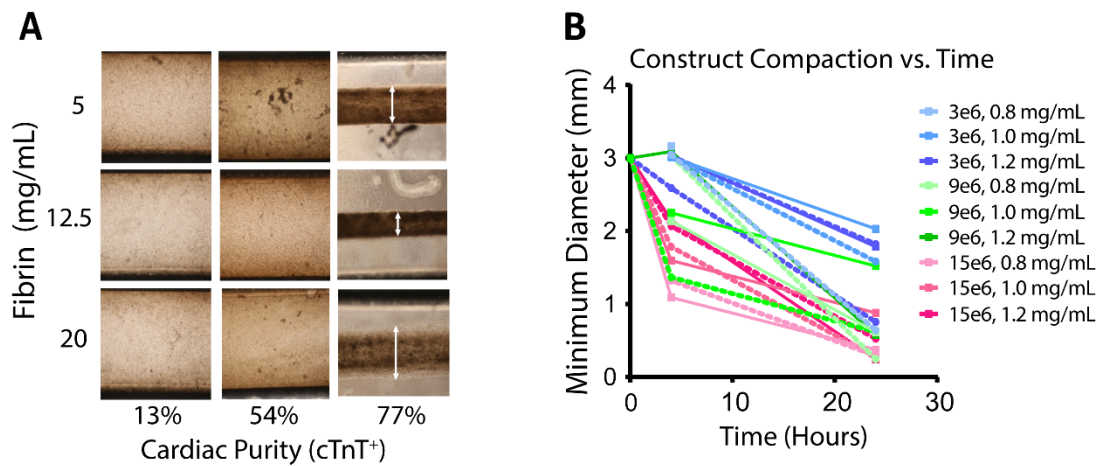


Figure S2: Compaction of cellularized hydrogel constructs over time. (A) Phase microscopy images of blended 2 mg/mL collagen and fibrin hydrogels after 144 hours of incubation under cell culture conditions. Fibrin concentration is indicated on the y-axis, and cTnT⁺ percentage (a measure of hiPSC-derived cardiomyocyte purity) is indicated on the x-axis. Double headed arrows indicate the construct midpoint diameter that was measured. No fibrinolytic inhibitor was used in this single experiment. (B) Scatterplot of collagen-only 49.71% cTnT⁺ cardiac construct diameter at the narrowest point is shown through culture time. Replicates prepared under the same conditions are paired by color and have solid and dashed lines. The legend indicates cell seeding density per mL and collagen concentration.

Group	Collagen (mg/mL)	Fibrin (mg/mL)	Seeding Density (1x10 ⁶ cells/mL)
1	0.8	0	12
2	0.8	4	9
3	0.8	8	12
4	1.2	0	9
5	1.2	4	12
6	1.2	8	9
7	1.6	0	12
8	1.6	4	9
9	1.6	8	12
10	0.8	4	15
11	1.2	0	15
12	1.2	8	15
13	1.6	4	15

Table S1: Response surface model groups. Collagen, fibrin, and seeding density factor levels that defined each group. Note that group 5 was considered a center point for the Box-Behnken design format and was replicated across each of the two six well plates required for each experiment.

Experiment	cTnT ⁺ (%)	SMA ⁺ (%)	cTnT ⁺ and SMA ⁺ (%)	SMA ⁺ only (%)
Unpurified 1	60.2	25.9	18.7	7.20
Unpurified 2	24.4	51.2	8.11	43.1
Unpurified 3	30.2	18.3	9.41	8.90
Lactate Purified	75.5	54.1	51.1	3.00
GCaMP-CM	52.2	25.8	7.90	17.9

Table S2: Flow cytometry analysis of iPSC-derived cardiomyocytes.

Group	Force				Calcium		
	Active Stress (kPa)	V _{up} (mN/mm ² /s)	T ₅₀ (ms)	T ₉₀ (ms)	V _{up} (Intensity/s)	T ₅₀ (ms)	T ₉₀ (ms)
4	0.02±0.01	0.08±0.04	131±24.0	237±26.7	11.5±2.99	118±66.0	195±84.7
11	0.08±0.01*	0.42±0.07*	96.7±3.18	193±8.39	8.44±2.00	86.0±16.3	159±38.9
Predicted	0.10	0.57	87.0	174	25.7	80	161

Table S3: Force and calcium kinetics of constructs prepared with hiPSC-GCaMP-derived cardiomyocytes at 15% strain. Only one replicate was evaluated for Group Predicted due to necking and breaking failure (n=3 for Groups 4 and 11). * indicates p<0.05 versus group 4.

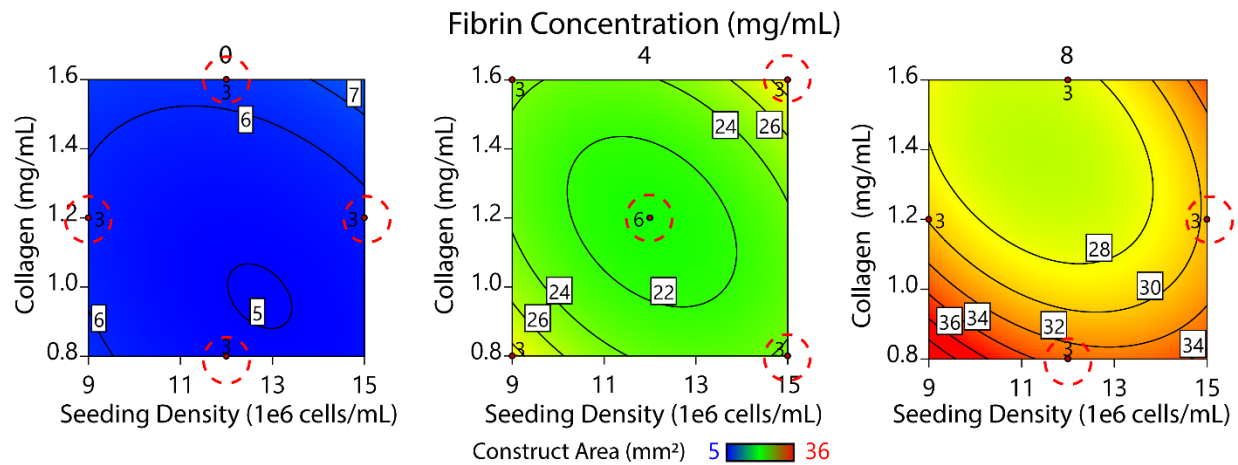


Figure S3: Contour plots describing predicted construct compaction with lactate purified cardiomyocytes (75% cTnT⁺) after 72 hours of culture with respect to fibrin concentration (major x-axis), collagen concentration (minor y-axes), and seeding density (minor x-axes). Contour labels indicate the predicted construct two-dimensional area in mm² after 72 hours of culture. Red dots and adjacent numbers indicate the number of empirical data replicates for a corresponding factor set, as prescribed by the Box-Behnken design. Red dashed circles indicate uniformly beating groups.

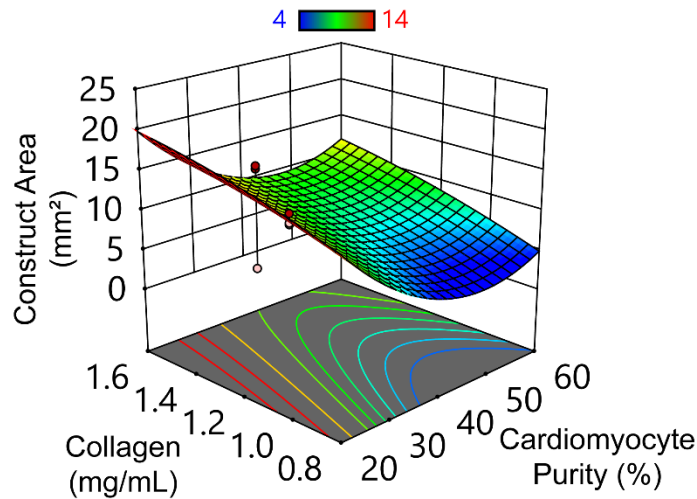


Figure S4: Response surface model for unpurified construct compaction at 72 hours with 8 mg/mL fibrin. Red dots indicate experimental sample value greater than the model prediction and pink dots indicate experimental sample values less than the model prediction. Cardiomyocyte purity is a defined as the percentage of cTnT⁺ cells.

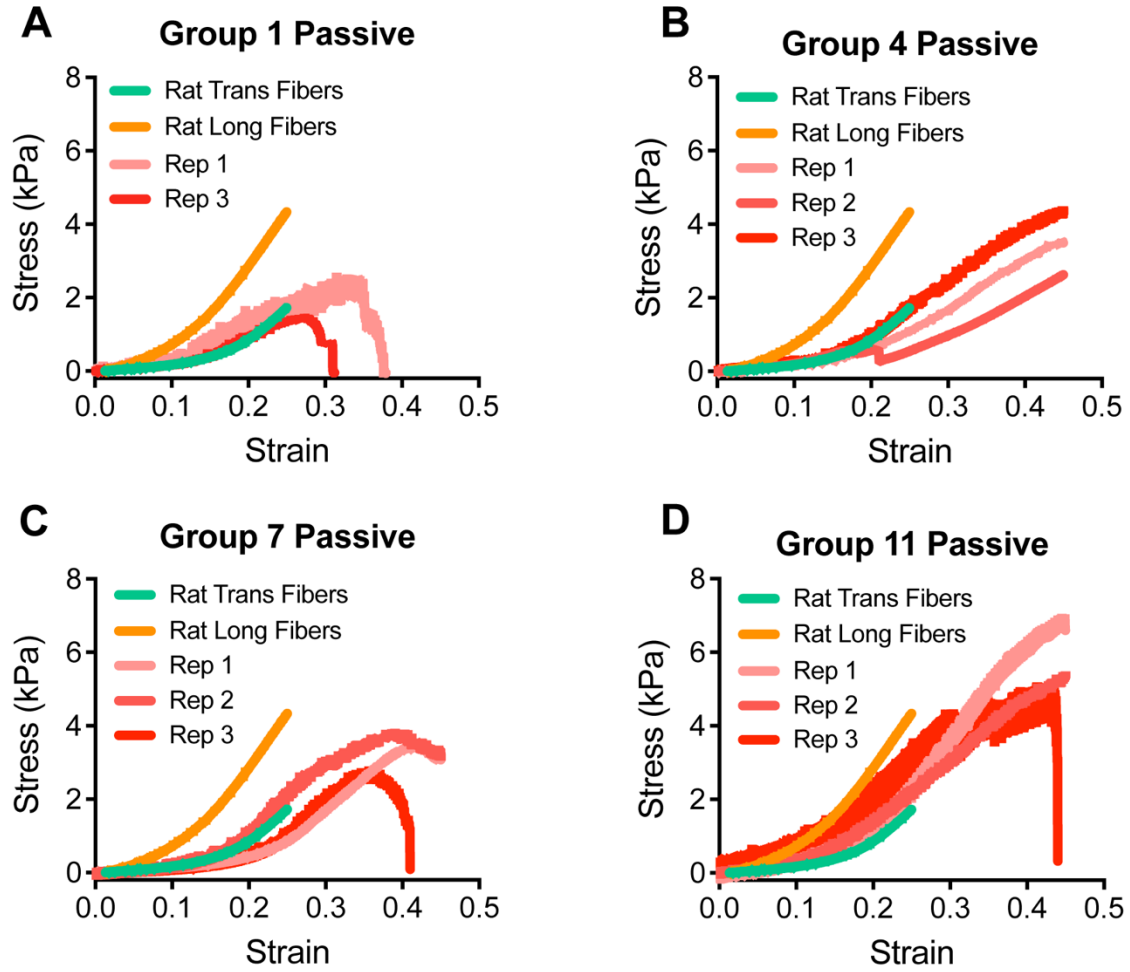


Figure S5: Passive mechanical analysis of lactate purified constructs. Green and orange traces represent stress/strain curves of native rat myocardium in the transverse and longitudinal fiber orientations (replicated from Figure 2C), respectively. Red shaded traces represent individual samples from Group 1 (A), Group 4 (B), Group 7 (C), and Group 11 (D). Increased “thickness” of some traces is a result of autonomous construct beating. All native and engineered tissues were tested at a constant strain rate of 10% strain/min.

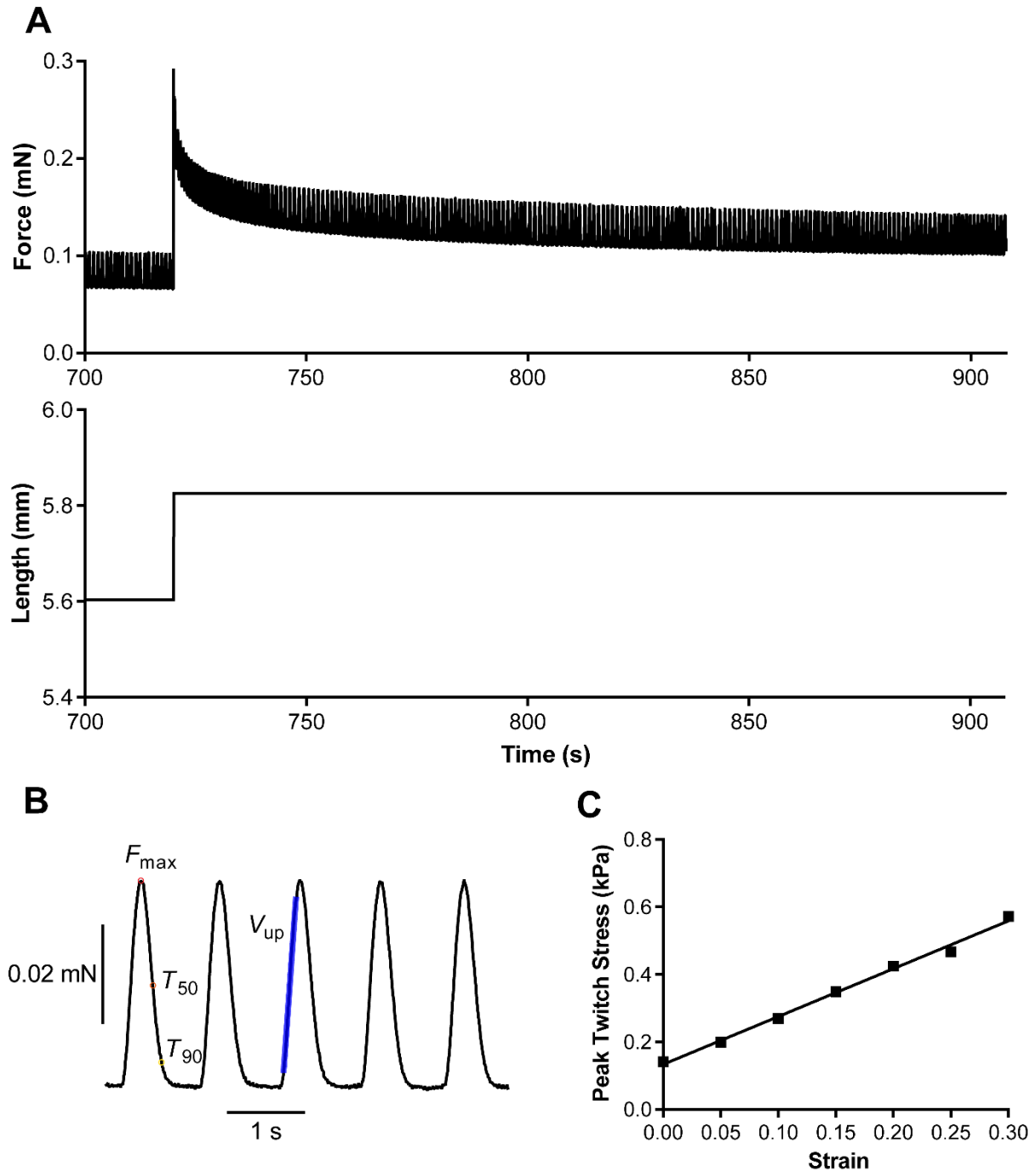


Figure S6: Representative active mechanics analysis of lactate purified groups. (A) Raw force and length vs time traces at maximum strain (30%) during 1 Hz stimulation. (B) Active contraction raw data trace shows peak force (F_{max}), which is normalized to cross-sectional area to calculate stress, upstroke velocity (V_{up}), and points used to calculate relaxation time to 50% and 90% relaxation (T_{50} and T_{90} , respectively). Values are summarized in Table 3 at maximum (30%) strain. (C) Force-length relationship for one example tissue is shown as maximum twitch stress vs. strain at each of seven length positions ($R^2 = 0.99$).

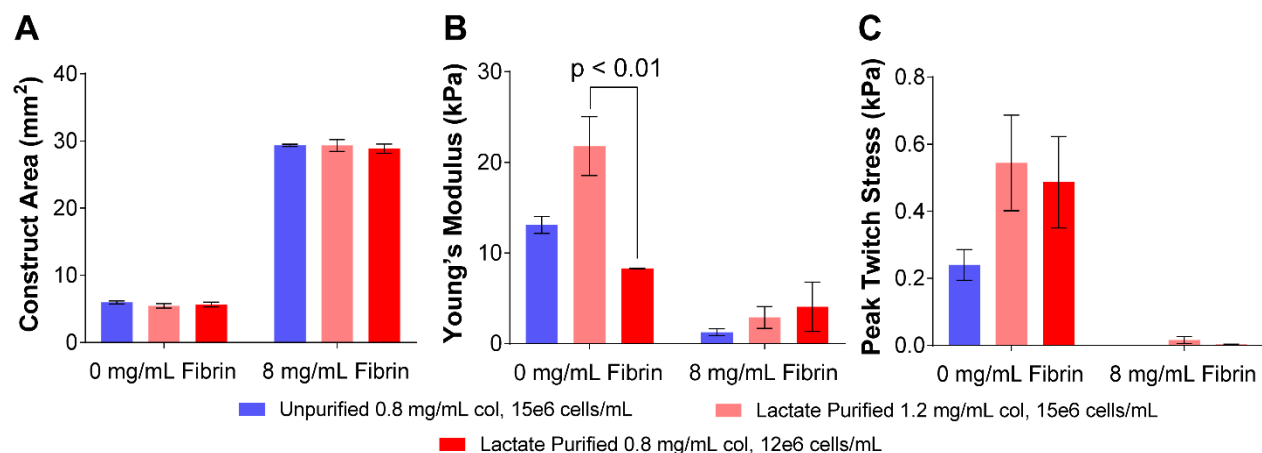


Figure S7: Constructs prepared with high purity, unpurified iPSC-cardiomyocytes (84.6% cTnT⁺) compared to select lactate purified cardiomyocyte (75.5% cTnT⁺) groups. Lactate purified Group 1, Group 3, Group 11, and Group 12 are described above by formulation factor levels to facilitate comparison. (A) Construct area after 72 hours in culture conditions. (B) Young's modulus and (C) Peak stress after 6 days in culture conditions.

Source	Sum of squares	df	Mean Square	F-value	p-value	
Model	34.36	6	5.73	13.40	< 0.0001	significant
A-Fibrin	3.84	1	3.84	8.99	0.0046	
B-Collagen	1.57	1	1.57	3.67	0.0624	
D-Polymerization Temp	2.28	1	2.28	5.34	0.0260	
E-Time in Culture Conditions	19.15	1	19.15	44.80	< 0.0001	
AE	5.79	1	5.79	13.54	0.0007	
DE	1.73	1	1.73	4.05	0.0507	
Residual	17.52	41	0.4274			
Lack of Fit	2.96	9	0.3292	0.7234	0.6841	not significant
Pure Error	14.56	32	0.4550			
Cor Total	51.89	47				

Table S4: Acellular compression model fit. df, degrees of freedom.

Factor	Coefficient Estimate	df	Standard Error	95% CI Low	95% CI High
Intercept	1.42	1	0.0944	1.23	1.61
A-Fibrin	0.2830	1	0.0944	0.0924	0.4735
B-Collagen	0.1808	1	0.0944	-0.0098	0.3714
D-Polymerization Temp	0.2180	1	0.0944	0.0275	0.4086
E-Time in Culture Conditions	0.6316	1	0.0944	0.4410	0.8222
AE	0.3472	1	0.0944	0.1567	0.5378
DE	0.1900	1	0.0944	-0.0006	0.3806

Table S5: Acellular compression model coefficients.

Source	Sum of squares	df	Mean Square	F-value	p-value	
Model	863.75	14	61.70	9.69	< 0.0001	significant
A-Cell Number	29.85	1	29.85	4.69	0.0326	
B-Collagen Concentration	251.80	1	251.80	39.54	< 0.0001	
C-Fibrin Concentration	80.62	1	80.62	12.66	0.0006	
D-Cell Purity	320.60	1	320.60	50.34	< 0.0001	
AB	8.11	1	8.11	1.27	0.2616	
AC	5.10	1	5.10	0.8016	0.3726	
AD	25.60	1	25.60	4.02	0.0474	
BC	31.26	1	31.26	4.91	0.0288	
BD	1.20	1	1.20	0.1883	0.6652	
CD	1.79	1	1.79	0.2806	0.5974	
A ²	67.26	1	67.26	10.56	0.0015	
B ²	12.80	1	12.80	2.01	0.1591	
C ²	20.99	1	20.99	3.30	0.0722	
D ²	150.49	1	150.49	23.63	< 0.0001	
Residual	687.75	108	6.37			
Lack of Fit	150.06	24	6.25	0.9768	0.5041	not significant
Pure Error	537.68	84	6.40			
Cor Total	1551.49	122				

Table S6: Unpurified 72 hour compaction model fit.

Factor	Coefficient Estimate	df	Standard Error	95% CI Low	95% CI High
Intercept	7.35	1	0.9650	5.43	9.26
A-Cell Number	-0.6576	1	0.3037	-1.26	-0.0556
B-Collagen Concentration	1.92	1	0.3051	1.31	2.52
C-Fibrin Concentration	-1.08	1	0.3049	-1.69	-0.4804
D-Cell Purity	-3.18	1	0.4478	-4.07	-2.29
AB	0.4820	1	0.4271	-0.3645	1.33
AC	-0.3825	1	0.4272	-1.23	0.4643
AD	-0.7637	1	0.3809	-1.52	-0.0088
BC	0.9470	1	0.4274	0.0998	1.79
BD	0.1668	1	0.3845	-0.5952	0.9289
CD	-0.2045	1	0.3861	-0.9699	0.5608
A ²	1.54	1	0.4747	0.6018	2.48
B ²	-0.6732	1	0.4748	-1.61	0.2680
C ²	0.8620	1	0.4748	-0.0791	1.80
D ²	5.60	1	1.15	3.31	7.88

Table S7: Unpurified 72 hour compaction model coefficients.

Source	Sum of squares	df	Mean Square	F-value	p-value	
Model	20.90	9	2.32	86.24	< 0.0001	significant
A-Cell Number	0.0000	1	0.0000	0.0008	0.9779	
B-Collagen Concentration	0.0006	1	0.0006	0.0234	0.8794	
C-Fibrin Concentration	16.42	1	16.42	609.76	< 0.0001	
AB	0.0798	1	0.0798	2.96	0.0948	
AC	0.0008	1	0.0008	0.0306	0.8621	
BC	0.1295	1	0.1295	4.81	0.0357	
A ²	0.1260	1	0.1260	4.68	0.0381	
B ²	0.1243	1	0.1243	4.62	0.0394	
C ²	3.34	1	3.34	124.22	< 0.0001	
Residual	0.8616	32	0.0269			
Lack of Fit	0.1652	3	0.0551	2.29	0.0988	not significant
Pure Error	0.6963	29	0.0240			
Cor Total	21.76	41				

Table S8: Lactate purified 72 hour compaction model fit.

Factor	Coefficient Estimate	df	Standard Error	95% CI Low	95% CI High
Intercept	3.04	1	0.0670	2.90	3.18
A-Cell Number	-0.0009	1	0.0335	-0.0692	0.0673
B-Collagen Concentration	0.0051	1	0.0335	-0.0631	0.0733
C-Fibrin Concentration	0.8271	1	0.0335	0.7589	0.8953
AB	0.0815	1	0.0474	-0.0149	0.1780
AC	0.0083	1	0.0474	-0.0882	0.1048
BC	-0.1039	1	0.0474	-0.2004	-0.0074
A ²	0.1146	1	0.0530	0.0067	0.2224
B ²	0.1138	1	0.0530	0.0059	0.2217
C ²	-0.5903	1	0.0530	-0.6981	-0.4824

Table S8: Lactate purified 72 hour compaction model coefficients.

Source	Sum of squares	df	Mean Square	F-value	p-value	
Model	1206.24	3	402.08	15.76	< 0.0001	significant
A-Cell Number	77.57	1	77.57	3.04	0.0895	
B-Collagen Concentration	409.18	1	409.18	16.04	0.0003	
C-Fibrin Concentration	670.54	1	670.54	26.29	< 0.0001	
Residual	943.72	37	25.51			
Lack of Fit	361.74	9	40.19	1.93	0.0880	not significant
Pure Error	581.98	28	20.79			
Cor Total	2149.96	40				

Table S10: Lactate purified Young's modulus model fit.

Factor	Coefficient Estimate	df	Standard Error	95% CI Low	95% CI High
Intercept	8.60	1	0.7896	7.00	10.20
A-Cell Number	1.80	1	1.03	-0.2910	3.89
B-Collagen Concentration	4.22	1	1.05	2.09	6.36
C-Fibrin Concentration	-5.41	1	1.05	-7.54	-3.27

Table S11: Lactate purified Young's modulus model coefficients.

Source	Sum of squares	df	Mean Square	F-value	p-value	
Model	73.44	6	12.24	30.85	< 0.0001	significant
A-Cell Number	5.39	1	5.39	13.60	0.0008	
B-Collagen Concentration	0.6841	1	0.6841	1.72	0.1977	
C-Fibrin Concentration	49.49	1	49.49	124.73	< 0.0001	
AC	0.6395	1	0.6395	1.61	0.2126	
BC	0.9568	1	0.9568	2.41	0.1295	
C ²	16.27	1	16.27	41.01	< 0.0001	
Residual	13.89	35	0.3968			
Lack of Fit	2.16	6	0.3597	0.8893	0.5154	not significant
Pure Error	11.73	29	0.4045			
Cor Total	87.33	41				

Table S12: Lactate purified peak active stress model fit.

Factor	Coefficient Estimate	df	Standard Error	95% CI Low	95% CI High
Intercept	-4.18	1	0.1485	-4.49	-3.88
A-Cell Number	0.4741	1	0.1286	0.2131	0.7352
B-Collagen Concentration	-0.1688	1	0.1286	-0.4299	0.0922
C-Fibrin Concentration	-1.44	1	0.1286	-1.70	-1.18
AC	-0.2308	1	0.1818	-0.6000	0.1383
BC	0.2824	1	0.1818	-0.0868	0.6515
C ²	1.26	1	0.1964	0.8590	1.66

Table S13: Lactate purified peak active stress model coefficients.