

Supplementary Material

Single molecule tracking reveals functions for RarA at replication forks but also independently from replication during DNA repair in *Bacillus subtilis*

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Table S1A. *Bacillus subtilis* strains used

| Strains | Genotype source strain | Source/parent strain | Strains | Relevant genotype | Source |
|---------|--------------------------|----------------------|---------|-------------------|-----------|
| BG214 | + wild type ^a | | BG1067 | + ΔrarA | This work |
| BG190 | + ΔrecA | 1 | BG1555 | + ΔrecA, ΔrarA | This work |
| BG439 | + ΔrecO | 2 | BG1433 | + ΔrecO, ΔrarA | This work |
| BG129 | + recF15 | 3 | BG1055 | + recF15, ΔrarA | This work |
| BG1455 | + ΔrecD2 | 4 | BG1421 | + ΔrecD2, ΔrarA | This work |
| BG1065 | + ΔrecX | 5 | BG1371 | + ΔrecX, ΔrarA | This work |
| BG1337 | + ΔaddAB | 6 | BG1107 | + ΔaddAB, ΔrarA | This work |
| BG675 | + ΔrecJ | 7 | BG1059 | + ΔrecJ, ΔrarA | This work |
| BG705 | + ΔrecQ | 7 | BG1575 | + ΔrecQ, ΔrarA | This work |
| BG425 | + ΔrecS | 7 | BG1563 | + ΔrecS, ΔrarA | This work |
| BG855 | + ΔrecU | 8 | BG1083 | + ΔrecU, ΔrarA | This work |
| BG1131 | + ΔrecG | 9 | BG1103 | + ΔrecG, ΔrarA | This work |
| BG703 | + ΔruvAB | 10 | BG1351 | + ΔruvAB, ΔrarA | This work |
| BG1245 | + ΔradA | 11 | BG1373 | + ΔradA, ΔrarA | This work |
| BG905 | + ΔpolY1 | 12 | BG1401 | + ΔpolY1, ΔrarA | This work |
| BG907 | + ΔpolY2 | 12 | BG1403 | + ΔpolY1, ΔrarA | This work |
| BG193 | + dnaB37 | 13 | BG1687 | + dnaB37, ΔrarA | This work |
| BG196 | + dnaC30 | 13 | BG1681 | + dnaC30, ΔrarA | This work |
| BG198 | + dnaG20 | 13 | BG1661 | + dnaG20, ΔrarA | This work |
| BG199 | + dnaF33 | 13 | BG1685 | + dnaF33, ΔrarA | This work |
| BG201 | + dnaX51 | 13 | BG1659 | + dnaX51, ΔrarA | This work |
| BG1679 | + dnaE58 | This work | BG1683 | + dnaE58, ΔrarA | This work |

^atrpCE metA5 amyE1 ytsJ1 rsbV37 xre1 xkda1 att^{SPB} att^{ICEBs1}

Table S1B. *B. subtilis* *rarA-yfp* and its mutant variants

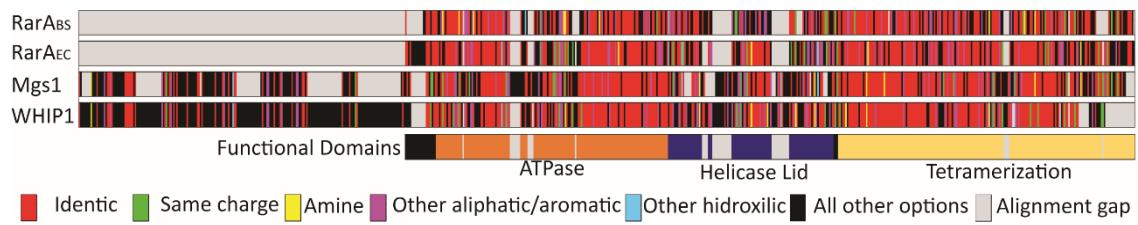
| Strains ^a | Relevant genotype | Source |
|----------------------|---|-----------|
| BG1331 | + <i>rarA-yfp</i> | This work |
| PG3171 | + <i>rarA-mVenus</i> | This work |
| BG1445 | + <i>rarA-yfp</i> , <i>ΔrecO</i> | This work |
| BG1345 | + <i>rarA-yfp</i> , <i>recF15</i> | This work |
| BG1347 | + <i>rarA-yfp</i> , <i>ΔrecD2</i> | This work |
| BG1349 | + <i>rarA-yfp</i> , <i>ΔrecX</i> | This work |
| PG3316 | + <i>rarA-yfp</i> , <i>ΔaddAB</i> | This work |
| PG3423 | + <i>rarA-yfp</i> , <i>ΔrecJ</i> | This work |
| PG3318 | + <i>rarA-yfp</i> , <i>ΔrecQ</i> | This work |
| PG3424 | + <i>rarA-yfp</i> , <i>ΔrecS</i> | This work |
| BG1443 | + <i>rarA-yfp</i> , <i>ΔrecU</i> | This work |
| PG3317 | + <i>rarA-yfp</i> , <i>ΔrecG</i> | This work |
| PG3426 | + <i>rarA-yfp</i> , <i>ΔruvAB</i> | This work |
| PG3429 | + <i>rarA-yfp</i> , <i>ΔradA</i> | This work |
| PG3427 | + <i>rarA-yfp</i> , <i>ΔpolY1</i> | This work |
| PG3428 | + <i>rarA-yfp</i> , <i>ΔpolY2</i> | This work |
| PG3174 | + <i>rarA-yfp</i> , <i>dnaX-cfp</i> | This work |
| BG1451 | + <i>rarA-yfp</i> , <i>dnaB37</i> | This work |
| BG1453 | + <i>rarA-yfp</i> , <i>dnaC30</i> | This work |
| PG3430 | + <i>rarA-yfp</i> , <i>dnaB37</i> , <i>dnaX-cfp</i> | This work |
| PG3431 | + <i>rarA-yfp</i> , <i>dnaC30</i> , <i>dnaX-cfp</i> | This work |

^aAll strains are derivatives of *B. subtilis* BG214 (*trpCE metA5 amyE1 ytsJ1 rsbV37 xre1 xkda1 att^{SPβ} att^{ICEBs1}*)

Movie S1: Exponentially growing cells expressing RarA-mVenus as sole source of the protein, images taken every 3 minutes. Shown are overlays of bright field and mVenus (variant of YFP) fluorescence (shown in yellow) images. 4 frames per second, White bar 5 μm. Note that the cells grow in short chains of cells.

Figure S1

A



B

| Species | Identity (%) | | | |
|---------------------------|-------------------------|---------------------|---------------------------|-------------------------|
| | <i>B. subtilis</i> RarA | <i>E. coli</i> RarA | <i>S. cerevisiae</i> Mgs1 | <i>H. sapiens</i> WHIP1 |
| <i>B. subtilis</i> RarA | 100 | 35.7 | 38.9 | 38.2 |
| <i>E. coli</i> RarA | 35.7 | 100 | 40.3 | 40.6 |
| <i>S. cerevisiae</i> Mgs1 | 38.9 | 40.3 | 100 | 42.0 |
| <i>H. sapiens</i> WHIP1 | 38.1 | 40.6 | 42.0 | 100 |
| amino acids number | 422 | 447 | 587 | 665 |

Figure S1. Comparison of RarA protein family (A) BLASTA for the sequences obtained in NCBI database and the functional domains predicted by Page *et al.*¹⁴ for *E. coli* RarA. Color-code represent the identities or the type of minor changes (polar with same charge, green; amine, yellow; or same kind of side chain, pink and blue). Figure is scaled to the size of proteins except of the gap needed for alignment. (B) Identity values for RarA homologues in evolution. The protein increases its size in eukaryotes, the identity is conserved

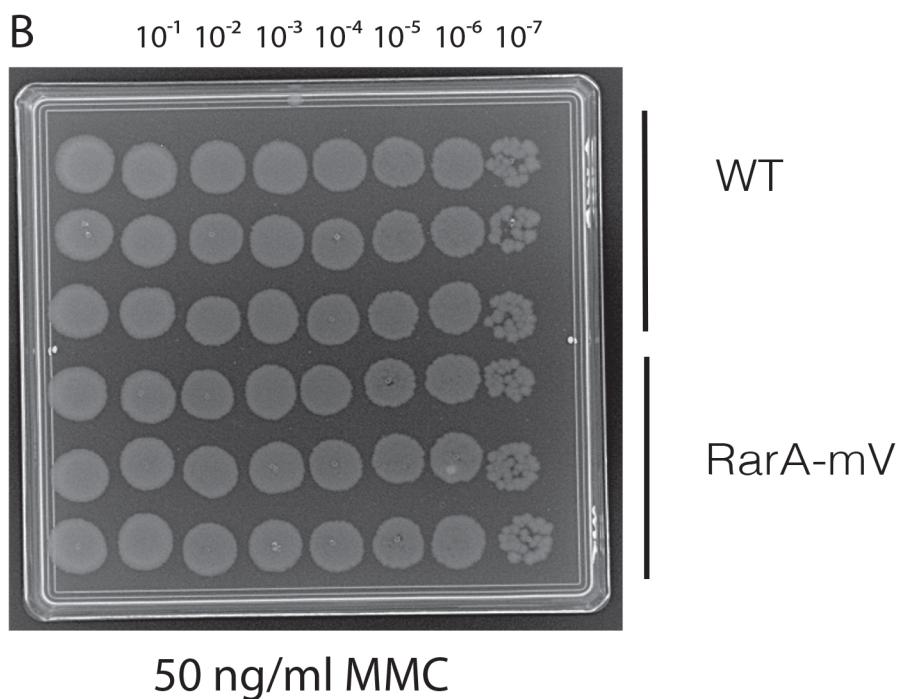
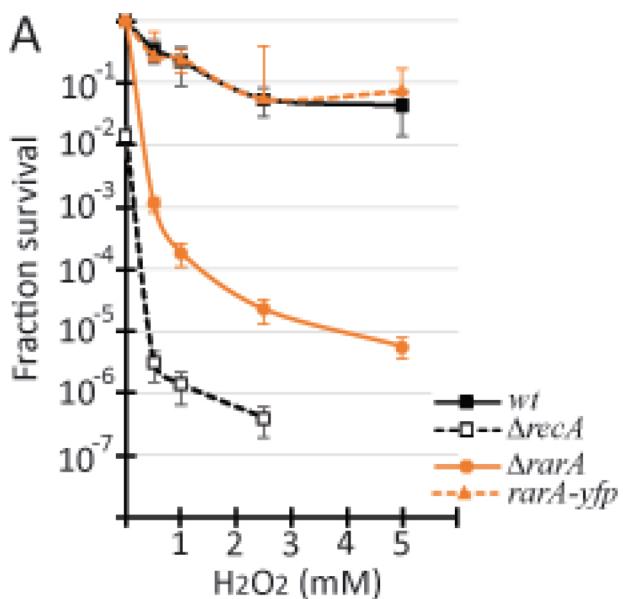


Fig. S2 Survival assays. (A) Chronic viability assay for the RarA-mVenus expressing strain. (B) Acute assay for survival of 60 minutes addition of MMC. Three independent colonies/assays are shown for wild type and for RarA-mVenus expressing cells.

Figure S3

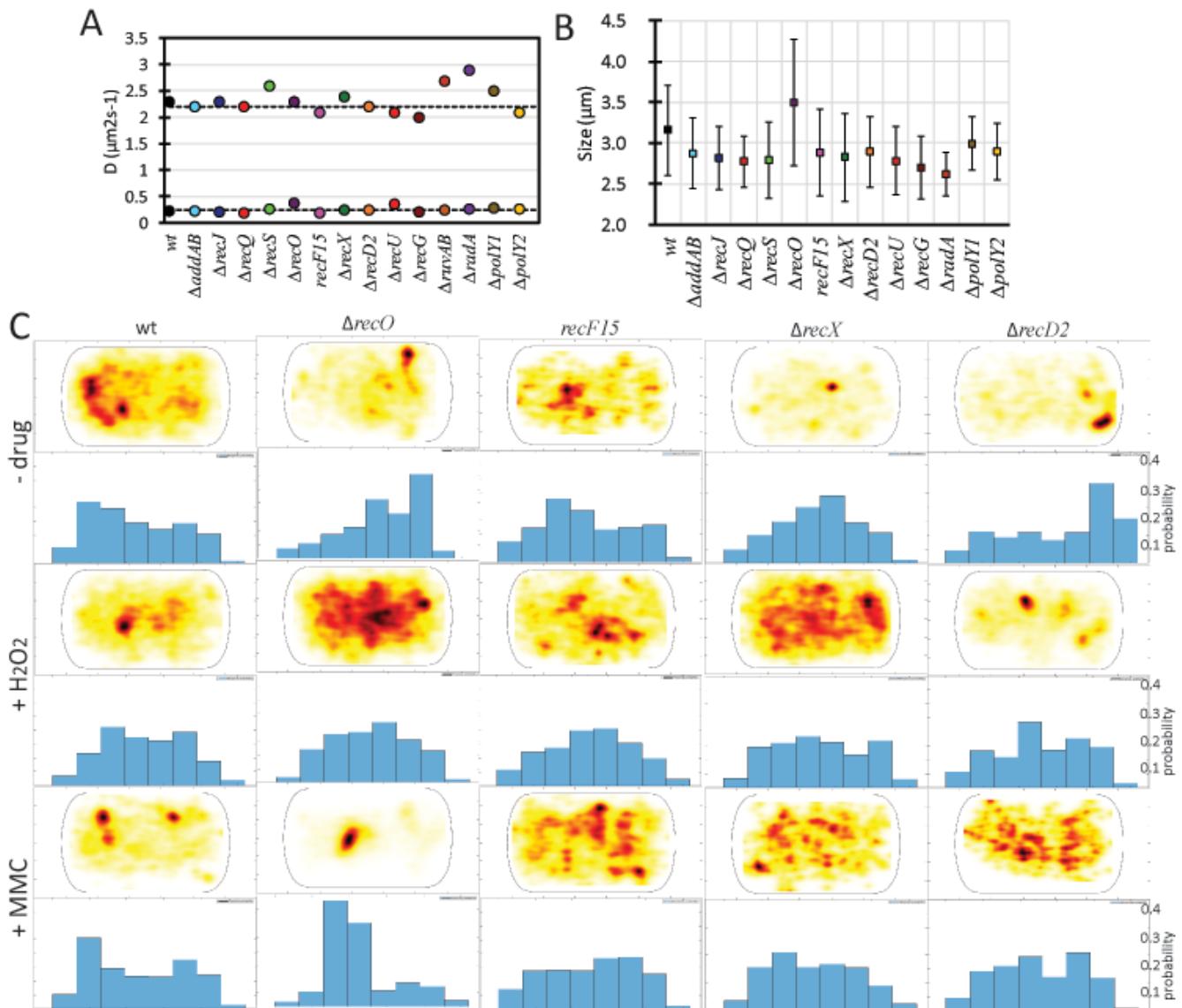


Figure S3. (A) Diffusion coefficients for static and dynamic populations calculated by Gaussian fit based in the step-size distribution. Values of each background were analysed separately, while dotted lines correspond to the analysis of all strains taken together. (B) Cell size considered prior to the normalization for the heat maps for the different backgrounds. (C) Heat maps and track distribution probability in X-axis for medium cells in the wild type, $\Delta recO$, $recF15$, $\Delta recX$ and $\Delta recD2$ in absence of DNA damage, or after induction with H₂O₂ (0.5 mM) or MMC (50 ng/ml).

References

- 1 Ceglowski, P., Luder, G. & Alonso, J. C. Genetic analysis of recE activities in *Bacillus subtilis*. *Mol Gen Genet* **222**, 441-445 (1990).
- 2 Fernandez, S., Kobayashi, Y., Ogasawara, N. & Alonso, J. C. Analysis of the *Bacillus subtilis* recO gene: RecO forms part of the RecFLOR function. *Mol Gen Genet* **261**, 567-573 (1999).
- 3 Alonso, J. C., Tailor, R. H. & Luder, G. Characterization of recombination-deficient mutants of *Bacillus subtilis*. *J Bacteriol* **170**, 3001-3007 (1988).
- 4 Torres, R., Romero, H., Rodriguez-Cerrato, V. & Alonso, J. C. Interplay between *Bacillus subtilis* RecD2 and the RecG or RuvAB helicase in recombinational repair. *DNA Repair (Amst)* **55**, 40-46, doi:10.1016/j.dnarep.2017.05.004 (2017).
- 5 Cardenas, P. P. et al. RecX facilitates homologous recombination by modulating RecA activities. *PLoS Genet* **8**, e1003126, doi:10.1371/journal.pgen.1003126 (2012).
- 6 Vlasic, I. et al. *Bacillus subtilis* RecA and its accessory factors, RecF, RecO, RecR and RecX, are required for spore resistance to DNA double-strand break. *Nucleic acids research* **42**, 2295-2307, doi:10.1093/nar/gkt1194 (2014).
- 7 Sanchez, H., Kidane, D., Cozar, M. C., Graumann, P. L. & Alonso, J. C. Recruitment of *Bacillus subtilis* RecN to DNA double-strand breaks in the absence of DNA end processing. *Journal of bacteriology* **188**, 353-360 (2006).
- 8 Fernandez, S., Sorokin, A. & Alonso, J. C. Genetic recombination in *Bacillus subtilis* 168: effects of recU and recS mutations on DNA repair and homologous recombination. *J Bacteriol* **180**, 3405-3409 (1998).
- 9 Sanchez, H., Carrasco, B., Cozar, M. C. & Alonso, J. C. *Bacillus subtilis* RecG branch migration translocase is required for DNA repair and chromosomal segregation. *Mol Microbiol* **65**, 920-935, doi:MMI5835 [pii]
10.1111/j.1365-2958.2007.05835.x (2007).
- 10 Sanchez, H. et al. The RuvAB branch migration translocase and RecU Holliday junction resolvase are required for double-stranded DNA break repair in *Bacillus subtilis*. *Genetics* **171**, 873-883, doi:genetics.105.045906 [pii]
10.1534/genetics.105.045906 (2005).
- 11 Gándara, C. & Alonso, J. C. DisA and c-di-AMP act at the intersection between DNA-damage response and stress homeostasis in exponentially growing *Bacillus subtilis* cells. *DNA repair* **27**, 1-8, doi:10.1016/j.dnarep.2014.12.007 (2015).
- 12 Raguse, M. et al. *Bacillus subtilis* DisA helps to circumvent replicative stress during spore revival. *DNA repair* **59**, 57-68, doi:10.1016/j.dnarep.2017.09.006 (2017).
- 13 Alonso, J. C., Stiege, C. A., Tailor, R. H. & Viret, J. F. Functional analysis of the dna (Ts) mutants of *Bacillus subtilis*: plasmid pUB110 replication as a model system. *Mol Gen Genet* **214**, 482-489 (1988).
- 14 Page, A. N., George, N. P., Marceau, A. H., Cox, M. M. & Keck, J. L. Structure and biochemical activities of *Escherichia coli* MgsA. *J. Biol. Chem.* **286**, 12075-12085, doi:10.1074/jbc.M110.210187 (2011).