

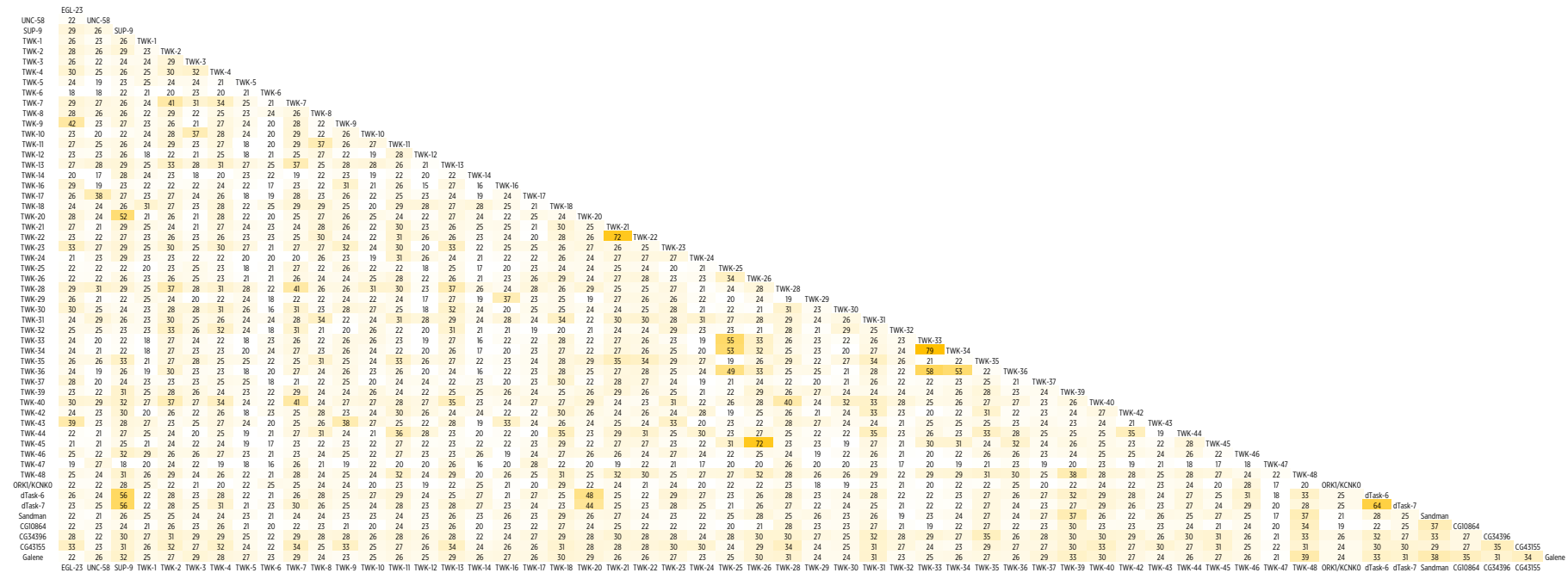
Supplementary Information

Mutation of a single residue promotes gating of vertebrate and invertebrate two-pore domain potassium channels

Ben Soussia*, El Mouridi*, *et al.*

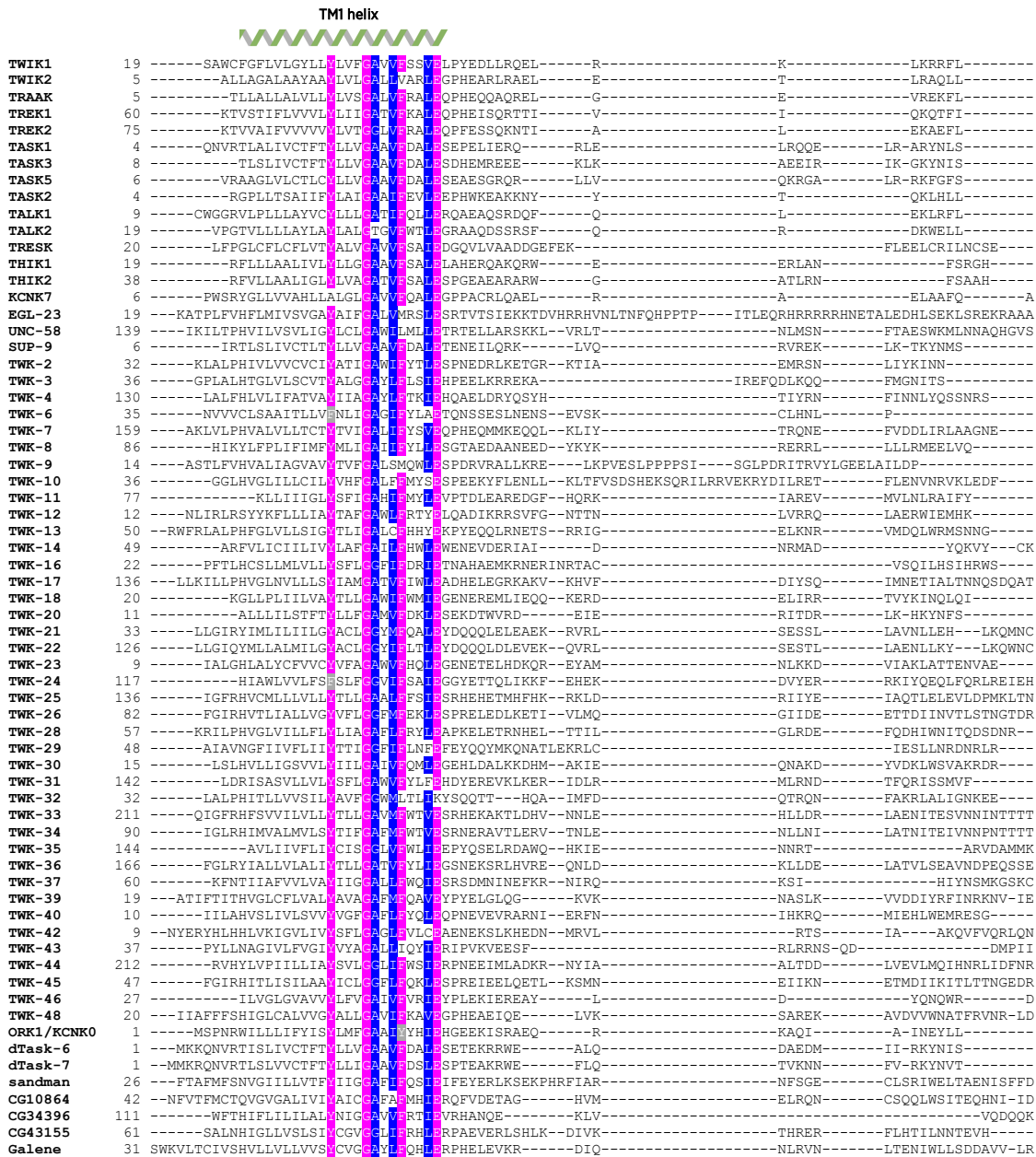
Nature Communications (2019)

Supplementary Figure 1 - continued.



TWK1, NP_002236 ; TWK2, NP_004814 ; TRAAK, NP_201567 ; TREK1, NP_001017425 ;
 TREK2, NP_612190 ; TASK1, NP_002237 ; TASK3, NP_001269463 ; TASK5, NP_071753 ;
 TASK2, NP_003731 ; TALK1, NP_001128578 ; TALK2, AAH25726 ; TRESK, NP_862823 ;
 THIK1, NP_071337 ; THIK2, NP_071338 ; KCNK7, NP_203133 ; EGL-23, NP_001255776 ;
 UNC-58, NP_741880 ; SUP-9, NP_494333 ; TWK-1, NP_492054 ; TWK-2, NP_494786 ;
 TWK-3, NP_495727 ; TWK-4, NP_001343566 ; TWK-5, NP_001021895 ; TWK-6, NP_497973 ;
 TWK-7, NP_498903 ; TWK-8, NP_001023596 ; TWK-9, NP_501724 ; TWK-10,
 NP_001300134 ; TWK-11, NP_001343632 ; TWK-12, NP_505731 ; TWK-13, NP_506091 ;
 TWK-14, NP_001256414 ; TWK-16, NP_508526 ; TWK-17, NP_001024466 ; TWK-18,
 NP_509516 ; TWK-20, NP_510284 ; TWK-21, NP_510654 ; TWK-22, NP_510655 ; TWK-23,
 NP_001257229 ; TWK-24, AAC32865 ; TWK-25, NP_502170 ; TWK-26, NP_508522 ; TWK-
 28, NP_508732 ; TWK-29, NP_001021467 ; TWK-30, NP_492381 ; TWK-31, NP_001022883
 ; TWK-32, NP_506416 ; TWK-33, NP_001309463 ; TWK-34, NP_506906 ; TWK-35,
 NP_508031 ; TWK-36, NP_507485 ; TWK-37, NP_491810 ; TWK-39, NP_001076639 ; TWK-
 40, NP_001255207 ; TWK-42, NP_507483 ; TWK-43, NP_872137 ; TWK-44, NP_509942 ;
 TWK-45, NP_001122742 ; TWK-46, NP_741678 ; TWK-47, NP_001309491 ; TWK-48,
 NP_001022681 ; ORR1/KCNKO, NP_511112 ; dTask-6, NP_001262539 ; dTask-7,
 NP_649891 ; sandman, NP_610349 ; CG10864, NP_650726 ; CG34396, NP_001097392 ;
 CG43155, NP_572720

Supplementary Figure 2

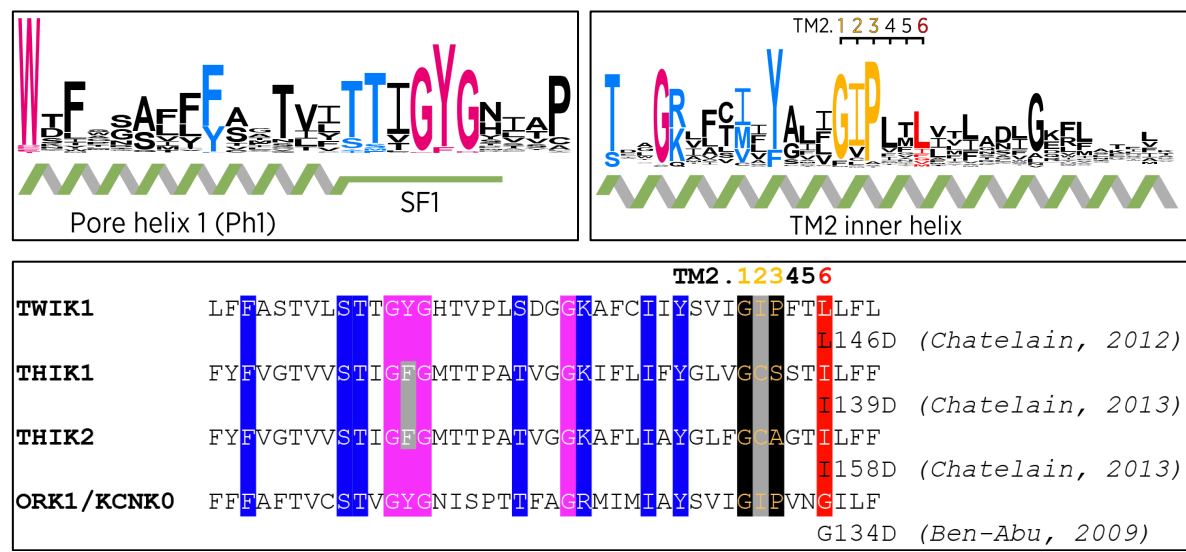
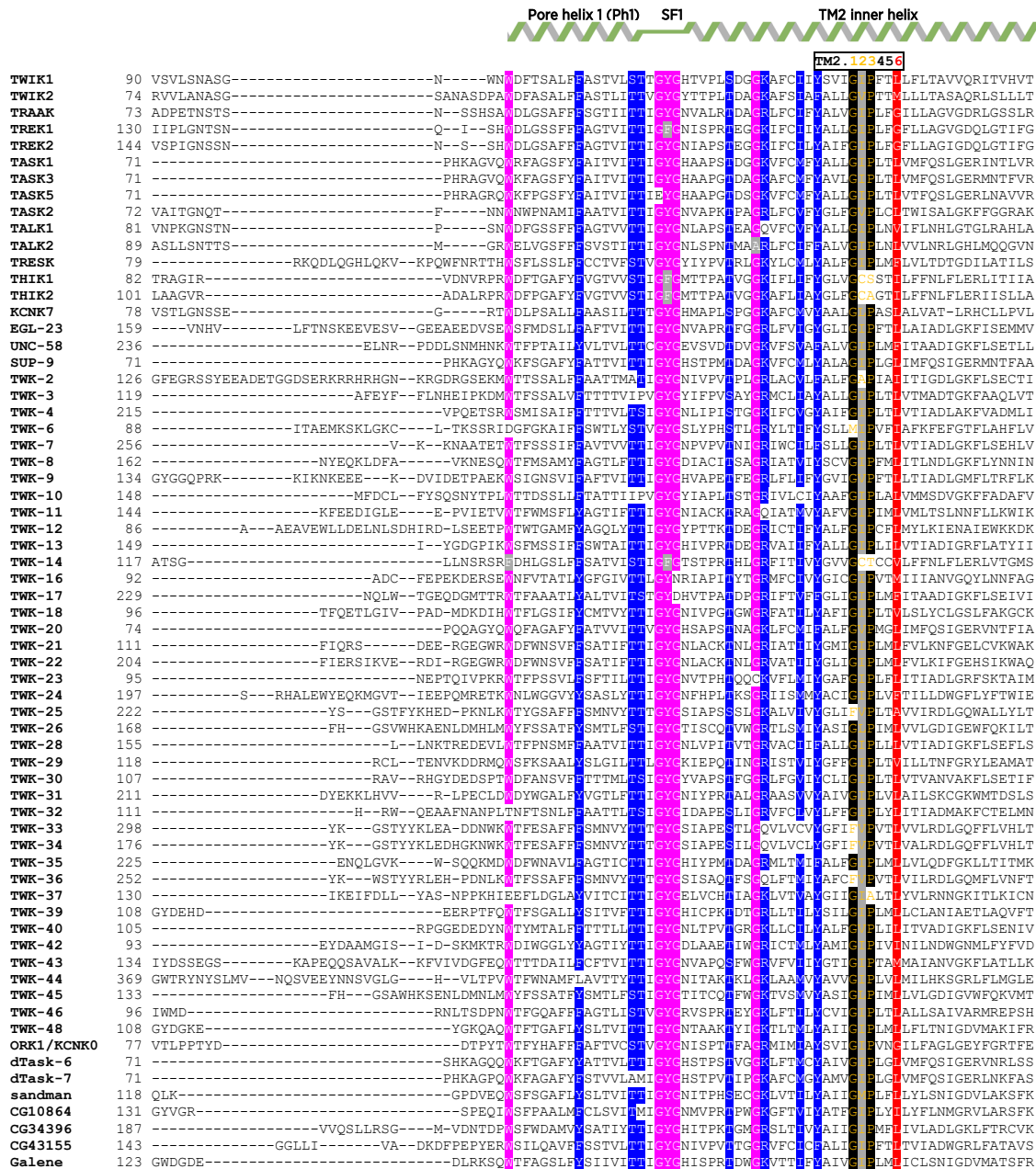


Supplementary Figure 2 - Clustal Omega alignment of human, *C. elegans*, and *D. melanogaster* two-pore domain potassium channel sequences.

Genbank identifiers for aligned sequences are listed below. N- and C-terminal cytoplasmic portions were truncated before alignment. Cartoon representations on top mark the four transmembrane helix regions (TM1, TM2, TM3, and TM4), two pore helix domains (Ph1 and Ph2), and two selectivity filters (SF1 and SF2). Sequence logos were generated using WebLogo 3 (Crooks et al., 2004). Highly conserved residues are indicated in *hotpink* and well conserved residues in *maroon*, as in Figure 1.

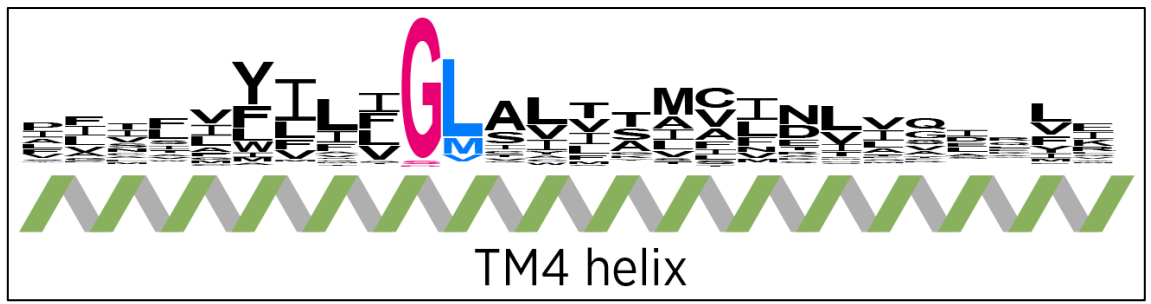
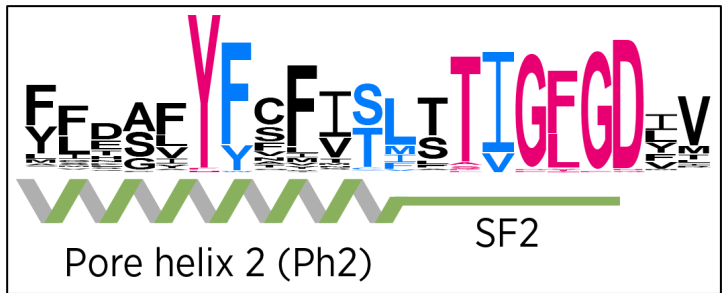
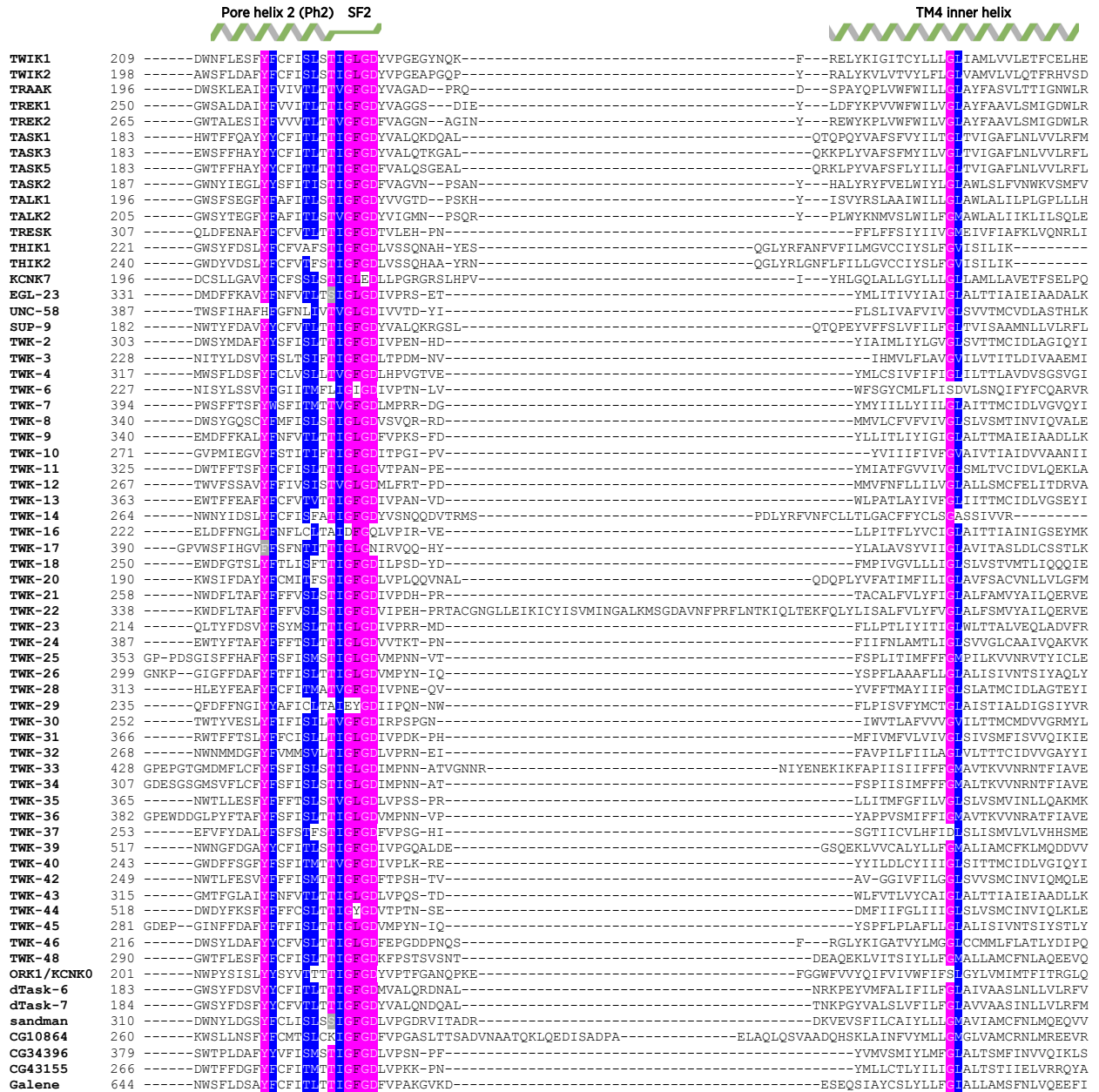
TWIK1, NP_002236 ; TWIK2, NP_004814 ; TRAAK, NP_201567 ; TREK1, NP_001017425 ; TREK2, NP_612190 ; TASK1, NP_002237 ; TASK3, NP_001269463 ; TASK5, NP_071753 ; TASK2, NP_003731 ; TALK1, NP_001128578 ; TALK2, AAH25726 ; TRESK, NP_862823 ; THIK1, NP_071337 ; THIK2, NP_071338 ; KCNK7, NP_203133 ; EGL-23, NP_001255776 ; UNC-58, NP_741880 ; SUP-9, NP_494333 ; TWK-2, NP_494786 ; TWK-3, NP_495727 ; TWK-4, NP_001343566 ; TWK-6, NP_497973 ; TWK-7, NP_498903 ; TWK-8, NP_001023596 ; TWK-9, NP_501724 ; TWK-10, NP_001300134 ; TWK-11, NP_001343632 ; TWK-12, NP_505731 ; TWK-13, NP_506091 ; TWK-14, NP_001256414 ; TWK-16, NP_508526 ; TWK-17, NP_001024466 ; TWK-18, NP_509516 ; TWK-20, NP_510284 ; TWK-21, NP_510654 ; TWK-22, NP_510655 ; TWK-23, NP_001257229 ; TWK-24, AAC32865 ; TWK-25, NP_502170 ; TWK-26, NP_508522 ; TWK-28, NP_508732 ; TWK-29, NP_001021467 ; TWK-30, NP_492381 ; TWK-31, NP_001022883 ; TWK-32, NP_506416 ; TWK-33, NP_001309463 ; TWK-34, NP_506906 ; TWK-35, NP_508031 ; TWK-36, NP_507485 ; TWK-37, NP_491810 ; TWK-39, NP_001076639 ; TWK-40, NP_001255207 ; TWK-42, NP_507483 ; TWK-43, NP_872137 ; TWK-44, NP_509942 ; TWK-45, NP_001122742 ; TWK-46, NP_741678 ; TWK-48, NP_001022681 ; ORK1/KCNK0, NP_511112 ; dTask-6, NP_001262539 ; dTask-7, NP_649891 ; sandman, NP_610349 ; CG10864, NP_650726 ; CG34396, NP_001097392 ; CG43155, NP_572720 ; Galene, NP_612084.

Supplementary Figure 2

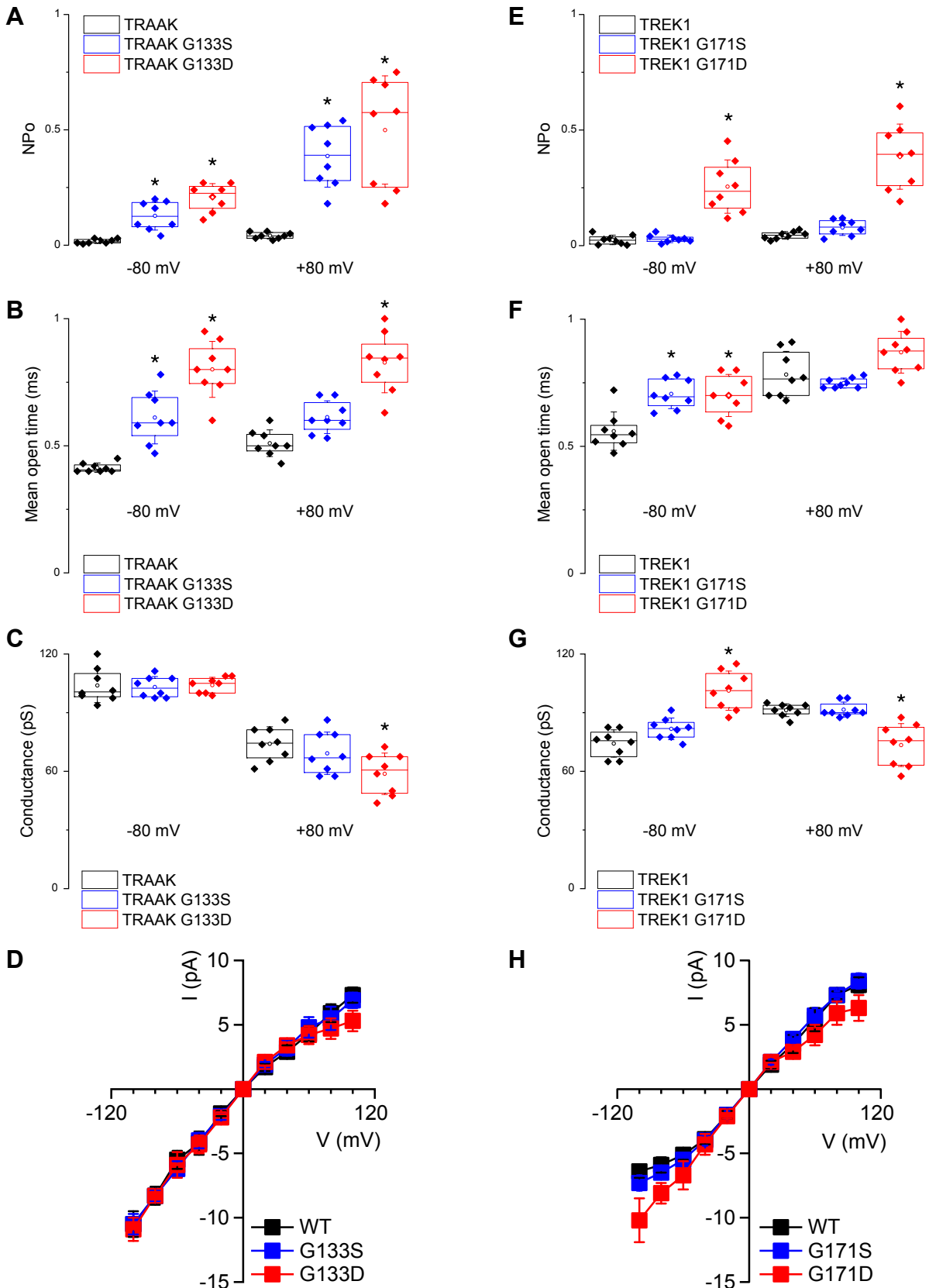


Previously published gain-of-function mutants of TM2.6 (see manuscript for references).

Supplementary Figure 2



Supplementary Figure 3



Single channel properties of TRAAK and TREK1 TM2.6 mutants

(A) TRAAK, TRAAK G133S, TRAAK G133D, (E) TREK1, TREK1 G171S, and TREK1 G171D channel activity determined by NPo analysis at -80 and +80 mV.

(B), (F) Mean open times were obtained from channel openings recorded at -80 mV and +80 mV.

(C), (G) Unitary conductances obtained from channel openings recorded at -80 mV and +80 mV.

(D), (H) Current-voltage relationships of single channels.

Center lines, medians; open circles, means; box limits, 25th and 75th percentiles; whiskers, standard deviation.

Kruskal-Wallis, Dunn's multiple comparison test, * $p < 0.05$.

Supplementary Table 1: Two-electrode voltage-clamp experiments (Figure 1B)

Channel	TM2.6	cRNA	Day	E_{rev} (mV)	I (μ A) at 0 mV
hTWIK1	WT	10 ng	1	-38.1 ± 2.4	0.2
	L146N			-88.4 ± 1	2.7 ± 0.2
rTWIK2	WT	10 ng	1	-33.1 ± 1.1	0.1
	M135D			-66.4 ± 2.2	0.4
hTASK1	WT	0.8 ng	3	-63.4 ± 2.5	0.7 ± 0.1
	L122N			-71.0 ± 1.1	13.4 ± 2
hTASK3	WT	50 pg	1	-73.4 ± 4.3	1.8 ± 0.3
	L122N			-77.6 ± 2.5	5 ± 0.8
mTREK1	WT	1 ng	1	-66.1 ± 1.5	0.7 ± 0.1
	G171D			-72.0 ± 0.6	8.8 ± 0.8
mTREK2	WT	1 ng	2	-52.5 ± 2	0.5 ± 0.1
	G196D			-72.1 ± 0.9	1.7 ± 0.2
mTRAAK	WT	1 ng	2	-55.6 ± 1.5	0.3
	G133D			-65.8 ± 0.8	32.9 ± 7.2
mTRESK	WT	1 ng	1	-70.8 ± 1.1	0.9 ± 0.1
	F156N			-68.9 ± 2	24.3 ± 3.6
mTASK2	WT	5 ng	1	-87.5 ± 2.8	3.8 ± 0.6
	L127N			-90.3 ± 6.6	14.2 ± 1.6
hTALK1	WT	20 ng	1	-80.5 ± 3.9	1 ± 0.1
	V137N			-93.6 ± 2.2	2.9 ± 0.2
hTALK2	WT	20 ng	1	-42.4 ± 2.1	0.3
	L145N			-51.4 ± 3.5	1.4 ± 0.1
hTHIK2	WT	50 ng	2	-31.1 ± 5.1	0.1
	I158D			-83.3 ± 0.9	1
hTASK2	WT	10 ng	1	-77.4 ± 2.7	0.4
	L127N			-93.4 ± 0.6	6.8 ± 0.2
rTWIK2 LY	WT	10 ng	1	-84.3 ± 0.8	2.2 ± 0.1
	M135D			-79.9 ± 0.8	8.5 ± 0.2
hTWIK1 AA	WT	10 ng	1	-71.7 ± 4	0.4
	L146N			-85.1 ± 0.9	9.3 ± 0.4
hTHIK2 5RA	WT	15 ng	1	-31.2 ± 1.4	0.2
	I158N			-76.5 ± 1	1.7 ± 0.2

Supplementary Table 2: Strain list

Strain	Genotype	Description
JIP1658	<i>egl-23(bln309) V</i>	EGL-23-wrmScarlet
JIP1506	<i>egl-23(bln309bln334) V</i>	EGL-23(L229N)-wrmScarlet
JIP1559	<i>egl-23(bln309bln359) V</i>	EGL-23(L229S)-wrmScarlet
JIP1620	<i>unc-58(bln448) X</i>	UNC-58(F294S)
JIP1621	<i>unc-58(bln449) X</i>	UNC-58(F294T)
JIP1622	<i>unc-58(bln450) X</i>	UNC-58(F294N)
JIP1623	<i>twk-18(bln451) X</i>	TWK-18(V158N)
JIP1636	<i>twk-18(bln458) X</i>	TWK-18(V158S)
JIP1706	<i>twk-18(bln491) X</i>	TWK-18(V158T)
JIP1677	<i>sup-9(bln476) II</i>	SUP-9(L122S)

Supplementary Table 3: List of single strand oligonucleotides (DNA and RNA) for CRISPR/Cas9 genome engineering

	Sequence (5' to 3')	Description
crRNA <i>unc-58</i>	gcggcagugauagacauuag	Use to generate UNC-58 (F294x)
oPT271	cggaaagggttttctcagtagcattcgcgcttggttggtataccactAatgtCTatCacTgcCgccgatattggtaaa tttttatctgaaacattactccagtttgtgagct	Repair template (Ultramer) for UNC-58 (F294S)
oPT273	tcggaaagggttttctcagtagcattcgcgcttggttggtataccactAatgACAATacTgcCgccgatattggtaa atttttatctgaaacattactccagtttgtgagctt	Repair template (Ultramer) for UNC-58 (F294T)
oPT275	tcggaaagggttttctcagtagcattcgcgcttggttggtataccactAatgAAATacTgcCgccgatattggtaa atttttatctgaaacattactccagtttgtgagctt	Repair template (Ultramer) for UNC-58 (F294N)
crRNA <i>egl-23</i>	ugguauuccguuuacacugc	Use to generate EGL-23 (L229x)
oSEM402	ttcgcctccaccatcatttctgatatgaattttccgagatctgcaattgcTagACTtgtaaacggaataccaatta gaccataaccaatgacaaatagacggccac	Repair template (Ultramer) for EGL-23 (L229S)
oSEM356	ttcgcctccaccatcatttctgatatgaattttccgagatctgcaattgcTagGTTtgtaaacggaataccaatta gaccataaccaatgacaaatagacggccac	Repair template (Ultramer) for EGL-23 (L229N)
crRNA <i>twk-18</i>	uaaagacugagcacugugag	Use to generate TWK-18 (V158x)
oSEM435	ctggatggggtcgttttgctactatcctttacgcattcattggaattccaTtGacaAACTTAAgtTtGtactgtct tggtagtctttttgccaagggatg	Repair template (Ultramer) for TWK-18 (V158N)
oSEM436	ctggatggggtcgttttgctactatcctttacgcattcattggaattccaTtGacaAGTTTAAgtTtGtactgtct tggtagtctttttgccaagggatg	Repair template (Ultramer) for TWK-18 (V158S)
oSEM437	ctggatggggtcgttttgctactatcctttacgcattcattggaattccaTtGacaACTTTAAgtTtGtactgtct tggtagtctttttgccaagggatg	Repair template (Ultramer) for TWK-18 (V158T)
crRNA <i>sup-9</i>	cgcacuggcgggaauuccau	Use to generate SUP-9 (L122x)
oNZ161	tggaactcctggaaaattttcagcaactcaccaatactctgaaacatgataTTtccTaGtggaaattcccgcagtg cgtagagcatgcagaaaacttttccgg	Repair template (Ultramer) for SUP-9 (L122N)
oMG798	tggaactcctggaaaattttcagcaactcaccaatactctgaaacatgatGCTGccTaGtggaaattcccgcagtg cgtagagcatgcagaaaacttttccgg	Repair template (Ultramer) for SUP-9 (L122S)

Supplementary Table 4: Genotyping primer combinations for *C. elegans* mutants

Allele	Primer	Sequence (5' to 3')	Description
<i>unc-58</i> F294X	oTB522	cggggtagtaatatggatgaagg	338bp PCR product; verify by sequencing.
	oTB508	ggaccacccggttgattgtaa	
<i>egl-23</i> L229X	oSEM357	cccaccattaaccgagtttg	487bp PCR product digested by FspBI; WT: 134/353bp Mutant: 104/134/249bp
	oSEM358	tcccagattataagcaggcaat	
<i>twk-18</i> V158X	oSEM430	tgcggcgcttaagtatttg	673bp PCR product digested by RsaI; WT: 673bp Mutant: 384/289bp
	oSEM431	ggagatggggaaagacaaca	
<i>sup-9</i> L122X	oMG850	caactgtgataacgacgatcgg	PCR oMG850-852-856: WT: 986/324bp L122N&L122S: 986bp PCR oMG850-852-855 WT: 986bp L122N: 986/324bp PCR oMG850-852-853 WT: 986bp L122S: 986/324bp
	oMG852	caccgcgtcgaaatatgtcc	
	oMG853	cgggaattccaCtAggCAGC	
	oMG855	gcgggaattccaCtAggaAA	
	oMG856	gcgggaattccaTtGggACT	

Supplementary Table 5: List of plasmids

Plasmid/ gene	Primer	Sequence (5' to 3')	Isothermal Ligation with EcoRV digested pTB207 and			
pTB301/ <i>TWK-18</i> <i>WT</i>	oTB553	ccactagtaacggccgcccagtgctggaa ttctgcagatatggcgattggtgcaag	oTB553- 554	from pOX_twk -18 ¹		
	oTB554	cgggccccccctcgagcggccgcccagtgctg atggatctagatgtcatgctctagat				
pNZ19/ <i>TWK-18</i> (V158D)	oNZ52	tgtgagtggaaattccaatga	oTB553- oNZ52 oTB554- oNZ53			
	oNZ53	tgctactatcctttacgcattcattggaat tccactcacaGACctcagtccttactgtct tgg				
pTB314/ <i>TWK-18</i> (M280I)	oTB571	actccaacaattggGATgaaatcgtaatcc catattgataggactcgtgtattctccac	oTB553- oTB571 oTB554- oTB572			
	oTB572	ttgtgtcaat				
pOA23/ <i>hTASK1</i> <i>WT</i>	oOA21	ccactagtaacggccgcccagtgctggaa ttctgcagatatgaagcggcagaacgtgcg	oOA21-22		from hTASK- 1-pSGEM ²	
	oOA22	agctcgggccccccctcgagcggccgcccag tgtgatggattcacacggagctcctgcgct				
pOA24/ <i>hTASK1</i> (L122N)	oOA23	ctggaacatgacATTcgtgagcgggatgcc	oOA21-23 oOA24-22			
	oOA24	ggcatcccgcctcacgAATgtcatgttccag				
pOA21/ <i>hTASK3</i> <i>WT</i>	oOA18	ccactagtaacggccgcccagtgctggaa ttctgcagatatgaagaggcagaacgtgcg	oOA18- oIS17	from hTASK- 3-pSGEM ²		
	oIS17	agctcgggccccccctcgagcggccgcccag tgtgatggatctaaacggacttccggcggt				
pOA22/ <i>hTASK3</i> (L122N)	oOA26	gctctggaacatgacATTtgtcagcgggat	oOA18-26 oIS17- oOA25			
	oOA25	atcccgcctgacaAATgtcatgttccagagc				
pOA25/ <i>mTASK2</i> <i>WT</i>	oNZ56	ccactagtaacggccgcccagtgctggaa ttctgcagatatggtggaccggggtccttt	oNZ56-57			from pTLN mTASK-2 ³
	oNZ57	agctcgggccccccctcgagcggccgcccag tgtgatggattcacgtgcccctgggggta				
pNZ20/ <i>mTASK2</i> (L127N)	oNZ55	gcacagcggcaccggaaga	oNZ56-55 oNZ54-57			
	oNZ54	ctgtgtcttctacggcctcttcgggggtgcc gctgtgcAACacatggatcagtgccctggg				
pOA26/ <i>mTREK2</i> <i>WT</i>	oOA30	ccactagtaacggccgcccagtgctggaa ttctgcagatatgaaatttccaatcgagac	oOA30-31		from mTREK2c -pEXO	
	oOA31	agctcgggccccccctcgagcggccgcccag tgtgatggatttagtttctgtcttcaagta				
pOA27/ <i>mTREK2</i> (G196D)	oOA43	tccagccaataagaaATCaaaaagcgggat	oOA30-43 oOA42-31			
	oOA42	atcccgcctttttGATtcttattggctgga				
pOA28/ <i>mTRAAK</i> <i>WT</i>	oOA34	ccactagtaacggccgcccagtgctggaa ttctgcagatatgcgagcaccacactcct	oOA34-35	from mTRAAK- pEXO		
	oOA35	agctcgggccccccctcgagcggccgcccag tgtgatggatctacaccggcacggccttgt				
pOA29/ <i>mTRAAK</i> (G133D)	oOA45	tcccgcagcagcatATCgaacagtgggat	oOA34-45 oOA44-35			
	oOA44	atcccactgttcGATatgctgctggcggga				

pOA30/ <i>mTRESK</i> <i>WT</i>	oOA38	ccactagtaacggccgcccagtgctggaa ttctgcagatatggaggctgaggagccacc	oOA38-39	from <i>mTRESK</i> - pEXO
	oOA39	agctcgggccccccctcgagcggccgcccag tgtgatggatttaccaggtagcgaactt		
pOA31/ <i>mTRESK</i> (<i>F156N</i>)	oOA41	gtctgtgaggaccagATTcattagaggat	oOA38-41 oOA40-39	
	oOA40	atccctctaataatgAATctggtcctcacagac		
	oTB			

¹ purchased from addgene

² kindly provided by V.K. Renigunta

³ kindly provided by F.V. Sepúlveda