

**Table S1: Characteristics of amplicons of candidate reference genes amplified using gene specific primers**

<b>Gene Symbol</b>	<b>Product Tm (°C)</b>	<b>Amplicon length</b>	<b>Efficiency</b>	<b>Amplification efficiency</b>	<b>Correlation coefficient</b>
<i>Actin</i>	84	110	1.79	78.56	0.99
<i>18s</i>	82	100	1.85	84.58	0.97
<i>28s</i>	79	100	1.88	88.3	0.99
<i>TUB</i>	82	114	1.83	82.96	0.98
<i>GADPH</i>	82	120	1.73	72.57	0.94
<i>EF</i>	84	111	2.01	100.71	1
<i>RPL17</i>	83	103	1.95	94.97	0.95
<i>Hist3</i>	82	113	2.45	144.68	0.99
<i>UbiCE</i>	83	113	1.87	86.8	0.99
<i>TATA</i>	81	114	1.97	97.3	1
<i>E2F</i>	83	100	1.92	92.11	0.98
<i>vATPase</i>	81	103	2.75	175.1	0.79
<i>SOD</i>	84	102	1.96	95.94	0.99
<i>GSTD2</i>	80	100	1.86	86.38	0.99

**Table S2. Detailed Bestkeeper analysis for all candidate reference genes**

<b>ADULT</b>											
		<b>18S</b>	<b>28S</b>	<b>TUB</b>	<b>EF</b>	<b>RPL17</b>	<b>UBICE</b>	<b>TATA</b>	<b>E2F</b>	<b>SOD</b>	<b>GSTD2</b>
<b>geo mean</b>	<b>CP</b>	22.93	22.75	25.46	24.09	26.85	24.99	27.74	24.38	23.31	20.76
<b>AR mean</b>	<b>CP</b>	22.94	22.76	25.47	24.09	26.85	24.99	27.74	24.38	23.32	20.77
<b>Min</b>	<b>CP</b>	21.94	22.05	24.53	23.94	26.82	24.48	27.68	24.15	22.35	20.17
<b>Max</b>	<b>CP</b>	23.94	23.47	26.41	24.24	26.88	25.50	27.79	24.72	24.10	21.51
<b>Stddev</b>	<b>[± CP]</b>	0.67	0.47	0.63	0.10	0.02	0.34	0.04	0.23	0.65	0.49
<b>Cv</b>	<b>%CP</b>	2.91	2.08	2.46	0.42	0.07	1.36	0.13	0.93	2.77	2.38
<b>Min</b>	<b>x-fold</b>	-1.98	-1.63	-1.90	-1.11	-1.02	-1.42	-1.04	-1.17	-1.94	-1.51
<b>Max</b>	<b>x-fold</b>	2.02	1.64	1.93	1.11	1.02	1.43	1.04	1.27	1.73	1.68
<b>Stddev</b>	<b>[± x-fold]</b>	1.59	1.39	1.54	1.07	1.01	1.27	1.02	1.17	1.57	1.41
<b>Pearson correlation coefficient</b>											
<b>BestKeeper vs. coeff. of correlation</b>		0.997	0.997	0.997	0.001	0.997	0.997	0.997	0.001	0.994	0.969
<b>p-value</b>		0.001	0.001	0.001	0.001	0.001	0.001	0.001	0.837	0.001	0.007
<b>NYMPH</b>											
		<b>18S</b>	<b>28S</b>	<b>TUB</b>	<b>EF</b>	<b>RPL17</b>	<b>UBICE</b>	<b>TATA</b>	<b>E2F</b>	<b>SOD</b>	<b>GSTD2</b>
<b>geo Mean [CP]</b>	<b>CP</b>	24.66	24.13	24.97	25.53	25.67	28.13	25.33	25.37	23.50	20.56
<b>ar Mean [CP]</b>	<b>CP</b>	24.69	24.18	25.00	25.65	25.71	28.13	25.34	25.39	23.52	20.58
<b>min [CP]</b>	<b>CP</b>	23.84	23.09	23.46	23.24	24.49	27.91	24.41	24.28	22.53	19.77
<b>max [CP]</b>	<b>CP</b>	26.36	26.30	26.54	29.00	27.87	28.24	26.27	26.64	25.00	22.00
<b>stddev [± CP]</b>	<b>[± CP]</b>	1.12	1.41	1.03	2.24	1.44	0.15	0.62	0.83	0.98	0.94
<b>CV [% CP]</b>	<b>%CP</b>	4.52	5.85	4.12	8.72	5.59	0.52	2.46	3.27	4.18	4.59
<b>min [x-fold]</b>	<b>x-fold</b>	-1.76	-2.06	-2.85	-4.90	-2.26	-1.16	-1.89	-2.14	-1.96	-1.73
<b>max [x-fold]</b>	<b>x-fold</b>	3.25	4.49	2.97	11.05	4.60	1.08	1.92	2.40	2.83	2.71
<b>stddev [± x-fold]</b>	<b>[± x-</b>	2.17	2.66	2.04	4.71	2.71	1.11	1.54	1.78	1.98	1.92

	<b>fold]</b>										
<b>Pearson correlation coefficient ( r )</b>											
<b>BestKeeper vs.coeff. of corr. [r]</b>		0.912	0.933	0.924	0.96	0.976	0.001	0.956	0.971	0.999	0.969
<b>p-value</b>		0.031	0.021	0.025	0.01	0.004	0.028	0.011	0.006	0.001	0.006
<b>STARVATION STRESS</b>											
<b>GENE</b>		<b>18S</b>	<b>28S</b>	<b>TUB</b>	<b>EF</b>	<b>RPL17</b>	<b>UBICE</b>	<b>TATA</b>	<b>E2F</b>	<b>SOD</b>	<b>GSTD2</b>
<b>geo mean</b>	<b>CP</b>	24.57	24.90	25.33	26.16	26.41	25.72	29.21	27.59	25.24	22.64
<b>AR mean</b>	<b>CP</b>	24.57	24.92	25.33	26.16	26.43	25.72	29.21	27.59	25.27	22.65
<b>Min</b>	<b>CP</b>	24.00	23.77	25.03	25.92	25.00	25.40	28.87	27.28	24.00	22.06
<b>Max</b>	<b>CP</b>	24.97	26.06	25.63	26.59	27.39	26.34	29.69	28.00	27.18	23.76
<b>Stddev</b>	<b>[± CP]</b>	0.38	0.77	0.20	0.29	0.95	0.41	0.32	0.27	1.27	0.74
<b>Cv</b>	<b>%CP</b>	1.56	3.08	0.79	1.10	3.61	1.61	1.10	0.99	5.03	3.27
<b>Min</b>	<b>x-fold</b>	-1.48	-2.19	-1.23	-1.18	-2.66	-1.25	-1.26	-1.24	-2.36	-1.49
<b>Max</b>	<b>x-fold</b>	1.32	2.23	1.23	1.35	1.97	1.54	1.40	1.33	3.85	2.18
<b>Stddev</b>	<b>[± x-fold]</b>	1.30	1.70	1.15	1.22	1.94	1.33	1.25	1.21	2.41	1.67
<b>Pearson correlation coefficient</b>											
<b>BestKeeper vs.coeff. of correlation</b>		0.968	0.549	0.999	0.117	0.962	0.83	0.001	0.001	0.928	0.859
<b>p-value</b>		0.007	0.338	0.001	0.851	0.009	0.082	0.006	0.166	0.023	0.062

**Table S3 : Normfinder based stability values of different candidate genes**

<b>GENES</b>	<b>ADULT</b>	<b>NYMPH</b>	<b>Starvation Stress</b>
<i>18s</i>	0.164	0.083	0.007
<i>28s</i>	0.026	0.121	0.525
<i>TUB</i>	0.128	0.094	0.007
<i>EF</i>	0.210	1.451	0.162
<i>RPL17</i>	0.098	0.109	0.357
<i>UbiCE</i>	0.005	1.503	0.019
<i>TATA</i>	0.086	0.178	0.455
<i>E2F</i>	0.189	0.034	0.342
<i>SOD</i>	0.102	0.003	0.899
<i>GSTD2</i>	0.029	0.003	0.181

**Table S4 : Geometric mean ranking of candidate reference genes using RefFinder**

<b>ADULT</b>		<b>NYMPH</b>		<b>Starvation stress</b>	
<b>GENES</b>	<b>Geomean of ranking values</b>	<b>GENES</b>	<b>Geomean of ranking values</b>	<b>GENES</b>	<b>Geomean of ranking values</b>
<i>UbiCE</i>	2.34	<i>E2F</i>	2.06	<i>TUB</i>	1.19
<i>28s</i>	3.13	<i>SOD</i>	2.11	<i>18s</i>	1.78
<i>TATA</i>	3.87	<i>TATA</i>	2.51	<i>UbiCE</i>	3.57
<i>RPL17</i>	3.94	<i>GSTD2</i>	3.66	<i>EF</i>	3.72
<i>GSTD2</i>	4.21	<i>UbiCE</i>	5.62	<i>E2F</i>	4.36
<i>TUB</i>	4.45	<i>TUB</i>	5.63	<i>GSTD2</i>	5.92
<i>18s</i>	5.03	<i>EF</i>	5.63	<i>TATA</i>	6.05
<i>SOD</i>	5.33	<i>RPL17</i>	5.79	<i>RPL17</i>	7.97
<i>EF</i>	7.02	<i>28s</i>	8.18	<i>28s</i>	8.74
<i>E2F</i>	7.75	<i>18s</i>	9	<i>SOD</i>	10

**Table S5 :Listof genes associated with RNAi mechanism identified in the *T. tabaci* assembled transcriptome at  $E$ -value  $< e^{-30}$ .  
\*Similarity; #Alignment length**

Gene	CDS/Transcript Number	Annotation	S-value	S*	AL#
<b>DsRNA uptake</b>					
SID-1	CDS_15961_transcript_14627	gi 646715429 gb KDR19041.1 SID1 transmembrane family member 1 [ <i>Zootermopsisnevadensis</i> ]	5.09814E-49	77	178
	CDS_15962_transcript_14627		0	70	622
	CDS_15963_transcript_14628				72
Clathrin Heavy Chain	CDS_194_transcript_841	gi 646720852 gb KDR22435.1 Clathrin heavy chain [ <i>Zootermopsisnevadensis</i> ]	0	97	1676
	CDS_24400_transcript_25704		1.13E-77	96	137
	CDS_25967_transcript_29416		1.1E-178	94	323
	CDS_25967_transcript_29416		1.1E-178	94	323
Innexin2	CDS_13894_transcript_12670	gi 242014052 ref XP_002427712.1 Innexin inx2, putative [ <i>Pediculushumanuscorporeis</i> ]	0	86	399
	CDS_4215_transcript_4271		0	91	358
	CDS_13893_transcript_12669		0	86	399
	CDS_13894_transcript_12670		0	86	399
<b>dsRNA cleavage</b>					
Dicer-1	CDS_24712_transcript_26352	gi 240247227 emb CAX68236.1 dicer-1 [ <i>Blattellagermanica</i> ]	0	64	807
	CDS_25848_transcript_29057		5.3605E-161	78	415
	CDS_23342_transcript_23605		1.38053E-30	71	154
Dicer-2	CDS_20968_transcript_20034	gi 406034947 emb CCF23094.1 Dicer 2 [ <i>Blattellagermanica</i> ]	3.9936E-157	47	1709
	CDS_21294_transcript_20494		0	56	1653
Drosha	CDS_20526_transcript_19507	gi 646694518 gb KDR08017.1 Ribonuclease 3 [ <i>Zootermopsisnevadensis</i> ]	0	86	982
<b>Endonuclease activity</b>					
Argonaute 1	CDS_15926_transcript_14593	gi 516306731 gb AGO85968.1 argonaute 1, partial [ <i>Locustamigratoria</i> ]	0	99	403
	CDS_15921_transcript_14590		0	97	855
	CDS_15922_transcript_14590		0	97	418
	CDS_15923_transcript_14591		0	97	855

Argonaute 2	CDS_6124_transcript_5961 CDS_6122_transcript_5961 CDS_6123_transcript_5961 CDS_6125_transcript_5962 CDS_6126_transcript_5962 CDS_6128_transcript_5963 CDS_6130_transcript_5965 CDS_18274_transcript_16947 CDS_25525_transcript_28247	gi 723784142 gb AIY24303.1 Argonaute 2 [ <i>Laodelphaxstriatella</i> ]	0 1.06074E-90 6.0224E-160 0 0 0 0 0	98 75 64 67 67 67 67 67	433 223 583 818 818 818 818
PCDG 6 (ALG-6)	CDS_16858_transcript_15514 CDS_24257_transcript_25393	gi 646723721 gb KDR24234.1 Programmed cell death protein 6 [ <i>Zootermopsisnevadensis</i> ]	5.7885E-114 5.7885E-114	97 97	175 175
PIWI	CDS_11559_transcript_10537 CDS_11560_transcript_10538	gi 646703429 gb KDR12118.1 Piwi-like protein 1 [ <i>Zootermopsisnevadensis</i> ]	0 0	74 75	782 773
Aub	CDS_26052_transcript_29621 CDS_24257_transcript_25393	gi 817196653 ref XP_012273923.1 PREDICTED: protein aubergine-like isoform X2 [ <i>Orussusabietinus</i> ]	8.02156E-39 8.49505E-40	64 77	148 109
Asp Spindle E	CDS_828_transcript_1424 CDS_829_transcript_1425	gi 646720534 gb KDR22214.1 Protein abnormal spindle [ <i>Zootermopsisnevadensis</i> ]	3.87747E-39 9.26456E-68	50 73	558 232
<b>Others</b>					

Tudor	CDS_9987_transcript_9161	gi 646703152 gb KDR11968.1 Tudor	2.95608E-80	58	139
	CDS_9990_transcript_9164	domain-containing protein 5	1.13593E-11	55	182
	CDS_10607_transcript_9637	[ <i>Zootermopsisnevadensis</i> ]	1.2331E-12	52	182
	CDS_10608_transcript_9638		1.20765E-12	52	182
	CDS_11871_transcript_10813		1.20765E-12	52	435
	CDS_11872_transcript_10813		1.9006E-123	66	435
	CDS_11873_transcript_10813		1.9006E-123	66	90
	CDS_11874_transcript_10814		9.21698E-09	53	188
	CDS_11875_transcript_10815		5.36742E-16	54	622
	CDS_11876_transcript_10815		1.56031E-87	53	188
	CDS_11877_transcript_10815		5.36742E-16	54	90
	CDS_11878_transcript_10816		9.21698E-09	53	188
	CDS_11879_transcript_10816		5.36742E-16	54	622
	CDS_11880_transcript_10816		1.56031E-87	53	90
	CDS_12288_transcript_11176		9.21698E-09	53	622
	CDS_14189_transcript_12964		1.56031E-87	53	188
	CDS_14190_transcript_12965		5.36742E-16	54	39
	CDS_15383_transcript_14106		2.65212E-07	69	1183
	CDS_15384_transcript_14107		4.8541E-153	50	163
	CDS_15385_transcript_14108		4.9947E-14	53	85
	CDS_15386_transcript_14109		1.21319E-07	54	832
	CDS_15387_transcript_14109		8.90154E-90	50	827
	CDS_15388_transcript_14110		2.1975E-90	50	85
	CDS_15389_transcript_14111		1.08125E-07	54	827
	CDS_15390_transcript_14111		2.1975E-90	50	832
	CDS_17731_transcript_16404		7.85433E-90	50	181
	CDS_17732_transcript_16405		3.02975E-30	61	599
	CDS_22526_transcript_22311		7.68318E-60	49	569
	CDS_23795_transcript_24504		1.52443E-60	49	570
	CDS_24864_transcript_26674		2.4689E-61	49	513
	CDS_3219_transcript_3489		2.17002E-24	44	469
	CDS_3220_transcript_3489		1.08973E-30	46	285
	CDS_3221_transcript_3489		2.58141E-26	48	



Mut-7	CDS_3222_transcript_3490	gi 646704864 gb KDR12833.1 putative exonuclease mut-7-like protein [ <i>Zootermopsisnevadensis</i> ]	5.4797E-153	63	579
	CDS_3223_transcript_3490		3.36557E-10	64	62
	CDS_3224_transcript_3490		6.96901E-13	61	98
	CDS_3225_transcript_3491		5.4797E-153	63	579
	CDS_3226_transcript_3491		6.96901E-13	61	98
	CDS_3227_transcript_3492		3.36557E-10	64	62
	CDS_3228_transcript_3492		2.61635E-30	54	222
	CDS_3229_transcript_3492		5.4797E-153	63	579
	CDS_3230_transcript_3493		3.36557E-10	64	62
	CDS_3231_transcript_3493		6.96901E-13	61	98
	CDS_3232_transcript_3494		5.4797E-153	63	579
	CDS_3233_transcript_3494		3.0543E-172	59	728
	CDS_3234_transcript_3494		3.36557E-10	64	62
HPS4	CDS_18050_transcript_16726 CDS_18051_transcript_16727	gi 646704291 gb KDR12556.1 Hermansky-Pudlak syndrome 4 protein [ <i>Zootermopsisnevadensis</i> ]	0 0	57 57	929 929
Belle	CDS_2216_transcript_2724 CDS_3872_transcript_4046	gi 805814300 ref XP_012148170.1 PREDICTED: DEAD-box ATP-dependent RNA helicase 20 [ <i>Megachilerotundata</i> ]	4.07E-08 0	66 87	57 481
Gawky	CDS_1202_transcript_1771 CDS_1203_transcript_1772 CDS_1204_transcript_1773 CDS_1199_transcript_1770 CDS_1207_transcript_1774 CDS_1209_transcript_1775	gi 572313755 ref XP_006622570.1 PREDICTED: protein Gawky-like [ <i>Apis dorsata</i> ]	2.41E-51 2.41E-51 2.41E-51 2.41E-51 2.41E-51 2.41E-51	68 68 68 68 68 68	187 187 187 187 187 187
CLP1	CDS_17314_transcript_15991	gi 646701253 gb KDR11058.1 CLP1-like protein [ <i>Zootermopsisnevadensis</i> ]	0	88	414
V-type proton ATPase subunit H	CDS_9906_transcript_9079 CDS_9907_transcript_9080 CDS_9908_transcript_9080 CDS_9909_transcript_9081 CDS_9910_transcript_9082	gi 646704147 gb KDR12485.1 V-type proton ATPase subunit H [ <i>Zootermopsisnevadensis</i> ]	0 1.65118E-96 8.4095E-179 8.4095E-179 0	80 85 92 92 88	558 182 289 289 484

Rab-7a	CDS_24868_transcript_26695	>gi 936578738 ref XP_014204028.1 PREDI	9.8E-128	93	208
	CDS_24869_transcript_26696	CTED: ras-related protein Rab-7a	1.1E-127	93	208
	CDS_1706_transcript_2230	[Copidosomafloridanum]	7E-133	93	206
	CDS_1707_transcript_2231	>gi 936578741 ref XP_014204029.1	7E-133	93	206
	CDS_1708_transcript_2232	PREDICTED: ras-related protein Rab-7a	7E-133	93	206
	CDS_21319_transcript_20533	[Copidosomafloridanum]	4.5E-113	87	207
	CDS_21320_transcript_20534	>gi 936578744 ref XP_014204030.1	3.3E-115	72	287
		PREDICTED: ras-related protein Rab-7a			
		[Copidosomafloridanum]			
		>gi 936578747 ref XP_014204031.1			
	PREDICTED: ras-related protein Rab-7a				
	[Copidosomafloridanum]				
	>gi 936578750 ref XP_014204032.1				
	PREDICTED: ras-related protein Rab-7a				
	[Copidosomafloridanum]				
	>gi 936578753 ref XP_014204033.1				
	PREDICTED: ras-related protein Rab-7a				
	[Copidosomafloridanum]				
	>gi 936578756 ref XP_014204034.1				
	PREDICTED: ras-related protein Rab-7a				
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	>gi 936578759 ref XP_014204035.1				
	PREDICTED: ras-related protein Rab-7a				
	[Copidosomafloridanum]				
	>gi 936578762 ref XP_014204037.1				
	PREDICTED: ras-related protein Rab-7a				
	[Copidosomafloridanum]				
	>gi 936578765 ref XP_014204038.1				
	PREDICTED: ras-related protein Rab-7a				
	[Copidosomafloridanum]				
RISC	CDS_2188_transcript_2700	gi 646700117 gb KDR10415.1 putative	6.2E-23	48	353
	CDS_4702_transcript_4696	RISC-loading complex subunit	3E-149	81	331
	CDS_2185_transcript_2699	[Zootermopsisnevadensis]	6.2E-23	48	353

DEAD box protein	CDS_17843_transcript_16505	gi 242021722 ref XP_002431292.1 DEAD	5.8E-110	61	474
	CDS_3872_transcript_4046	box ATP-dependent RNA helicase, putative	0	87	481
	CDS_19922_transcript_18854	[Pediculus humanus corporis]	4.99E-06	65	84
	CDS_2216_transcript_2724	>gi 212516560 gb EEB18554.1  DEAD box	4.07E-08	66	57
		ATP-dependent RNA helicase, putative			
		[Pediculus humanus corporis]			
dsRNase 2	CDS_24848_transcript_26641	gi 599126388 gb AHN55089.1 dsRNase 2,	2.93687E-36	76	111
		partial [Schistocercagregaria]			
RDRP	CDS_2003_transcript_2493	gi 527355229 gb AGS13094.1 RdRP	0	95	330
	CDS_2006_transcript_2493	[Cowpea mild mottle virus]	2.3133E-06	44	198
	CDS_2007_transcript_2494		9.2932E-174	87	331
	CDS_2009_transcript_2494		0	95	330
	CDS_2010_transcript_2494		0	57	1027
	CDS_2012_transcript_2495		4.7604E-153	95	255
	CDS_2013_transcript_2495		9.2932E-174	87	331
	CDS_2017_transcript_2495		0	57	1027
	CDS_2018_transcript_2496		0	95	330
	CDS_2020_transcript_2496		9.2932E-174	87	331
	CDS_2021_transcript_2496		0	57	1027
	CDS_2023_transcript_2497		9.2932E-174	87	331
	CDS_2024_transcript_2497		2.3721E-156	56	833
	CDS_2026_transcript_2498		0	95	330
	CDS_2027_transcript_2498		0	74	512
	CDS_2028_transcript_2498		2.22675E-31	45	477
	CDS_2030_transcript_2498		9.2932E-174	87	331
CDS_2032_transcript_2499		0	57	1027	

**Table S6: Primer sequences for dsRNA template of potential RNAi machinery**

<b>Genes</b>	<b>dsRNA Primer Sequence</b>
<i>Dicer-2</i>	Fwd: 5'TAATACGACTCACTATAGGGCCATCACCTACTCATTC3' Rev: 5'TAATACGACTCACTATAGGGGTTAGTAGAGCCTTCAG3'
<i>Staufen</i>	Fwd: 5'TAATACGACTCACTATAGCAGGAGCCTCTTTATAC 3' Rev: 5'TAATACGACTCACTATAGCCTTAGGAAAGTCTGAG3'
<i>Aubergine</i>	Fwd: 5'TAATACGACTCACTATAGGGCTAAAGCTGAGAAGAGG3' Rev: 5'TAATACGACTCACTATAGGGCACCTCCGTATTTACTC3'