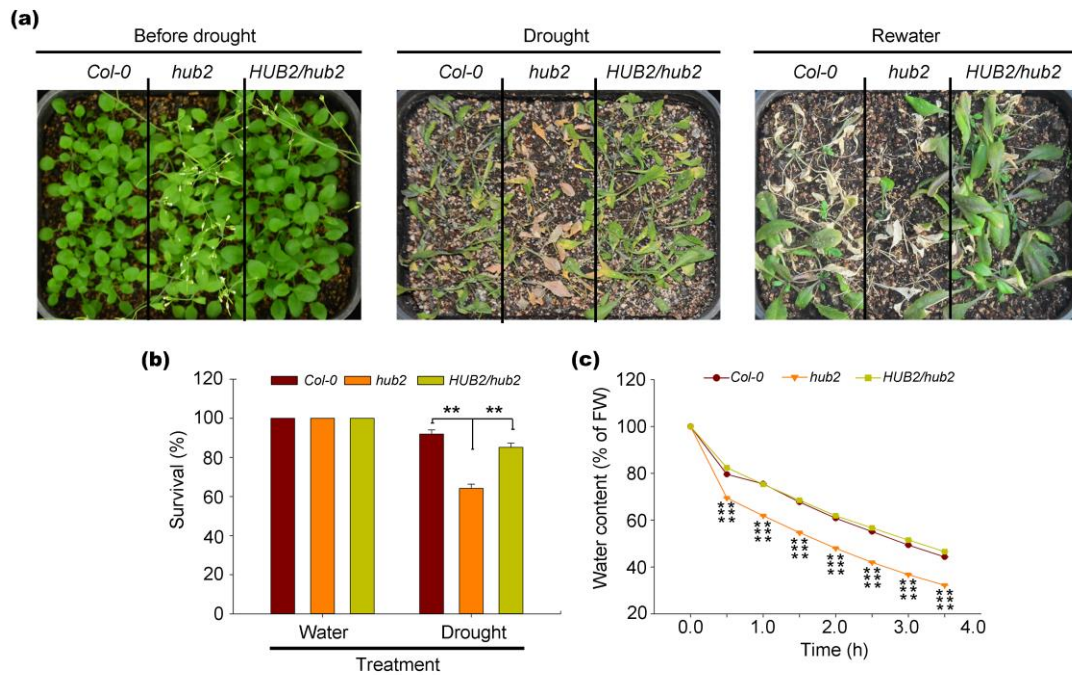


1 **Supporting Information**



2

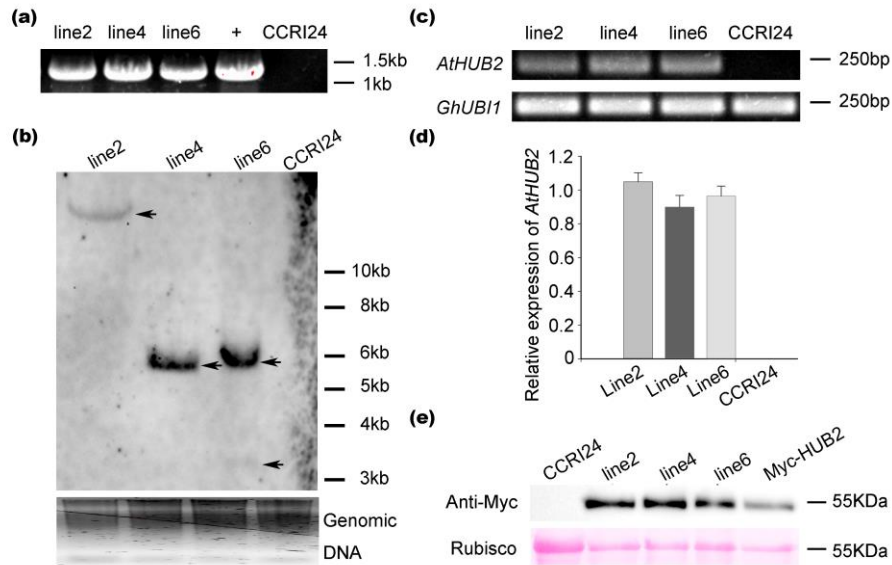
3 **Figure S1.** Response of *Athub2* mutants to dehydration stress. (a) Drought
 4 sensitivity of wild-type (WT), *hub2* mutant and *HUB2/hub2* complementation
 5 line seedlings. At 2 weeks after germination (Before drought), the plants were
 6 grown without water (Drought) for 12 days, followed by 3 days of watering for
 7 recovery (Rewater). (b) Survival percentages of the genotypes indicated in (a)
 8 after 12 days of drought treatment, followed by the 3-day recovery period. (c)
 9 The volume of water lost via air-drying from detached leaves of the indicated
 10 genotypes (4 weeks old) was measured as the percentage of change in the
 11 fresh weight (FW) of the leaves. Vertical bars represent the SD from three
 12 independent experiments (n ≥ 30) (**P < 0.01).

13



14

15 **Figure S2.** Characteristics of *AtHUB2*. Multiple sequence alignment of the four
 16 *AtHUB2* alternative splicing isoforms (AT1G55250.1, AT1G55250.2,
 17 AT1G55250.3, and AT1G55250.4). The black arrow indicates *AtHUB2* cloned
 18 by our group. The red arrow indicates key functional sites in the conserved
 19 C3HC4 RING domain.



20

21 **Figure S3.** Generation and molecular characterization of transgenic cotton
 22 lines expressing *AtHUB2*. (a) PCR analysis of *AtHUB2* in the genomic DNA of
 23 plants from the T2 generation. (b) Southern blotting analysis-positive *AtHUB2*
 24 transgenic cotton plants; the arrow indicates the target strip. The genomic DNA
 25 was as loading control. (c) Semi-quantitative RT-PCR analysis of *AtHUB2*
 26 expression in three T2 transgenic cotton lines. (d) RT-qPCR analysis of *AtHUB2*
 27 expression in three T2 transgenic cotton lines. (e) Immunoblotting analysis of
 28 the expression of the *AtHUB2* protein in three T2 transgenic cotton lines. An
 29 anti-Myc antibody was used to detect the *AtHUB2* protein, Myc-HUB2 was used
 30 as the positive control, and rubisco protein was as a loading control. The black
 31 line indicates the strip size.

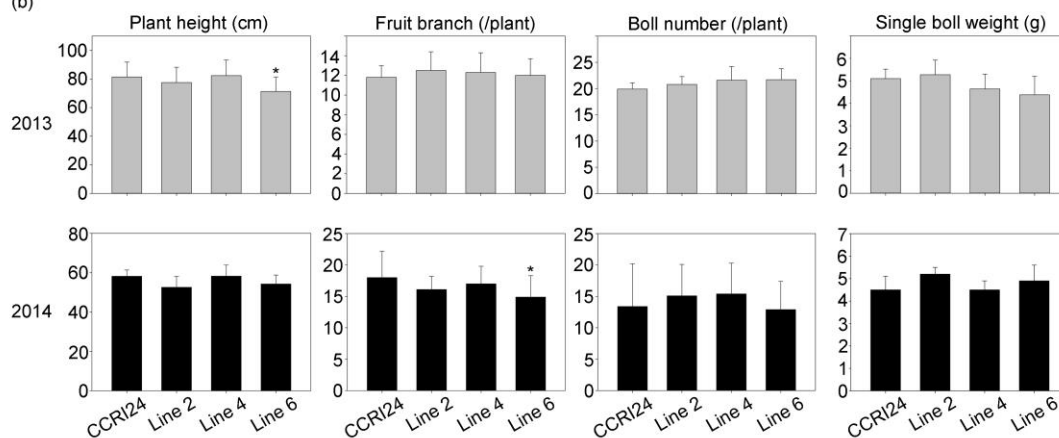
(a)



CCRI24

Transgenic line

(b)



32

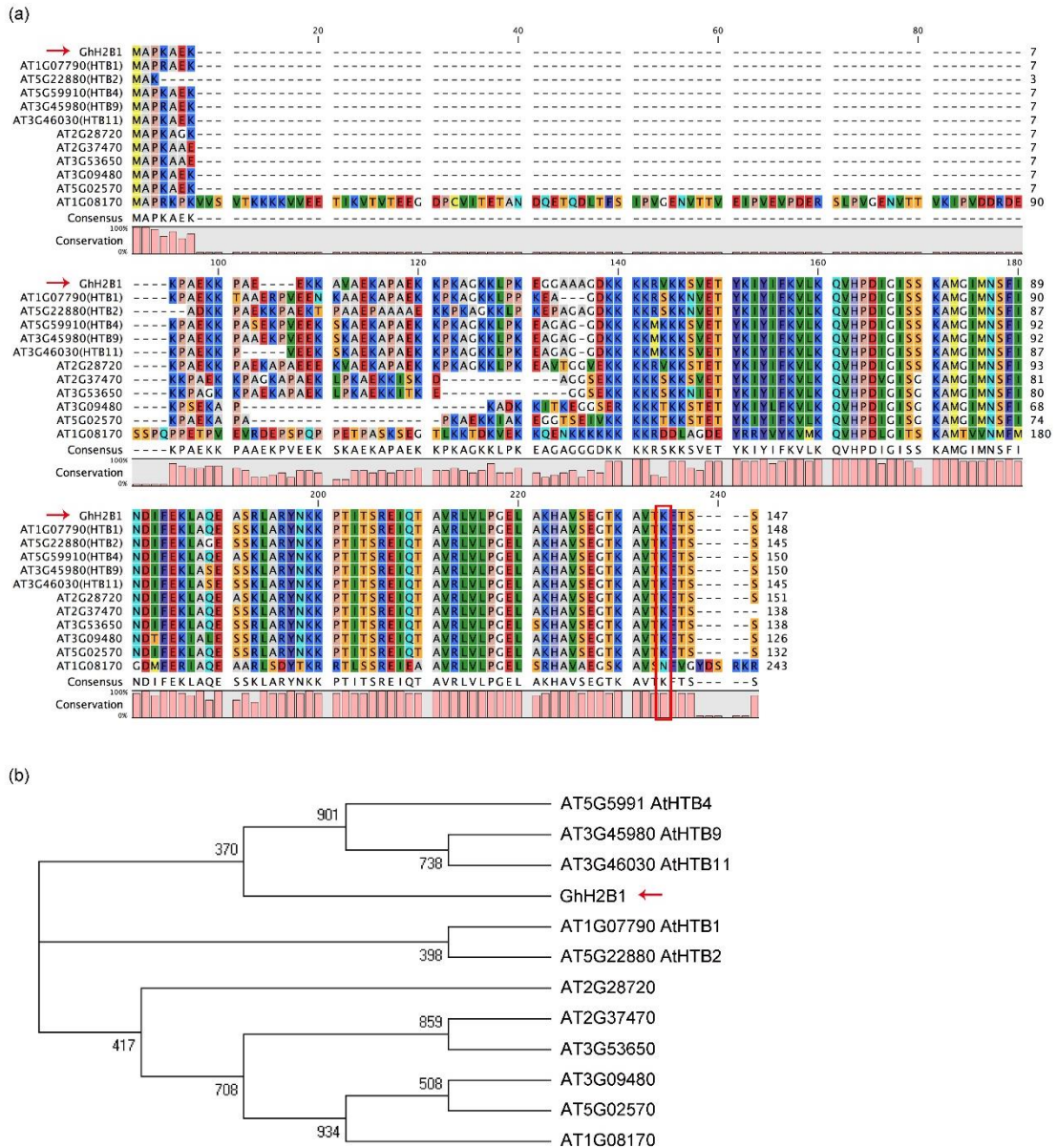
33 **Figure S4.** Agronomic traits of different *AtHUB2* transgenic lines in the field. (a)
34 Wild-type (WT) and *AtHUB2*-expressing cotton lines were grown under normal
35 conditions. Photographs were taken 90 days after germination. (b) Plant height,
36 fruit branching, boll number, and single boll weight of the control (CCRI24) and
37 three independent transgenic lines (line2, line4, and line6) grown in the field (n
38 ≥ 30). Vertical bars represented the SD. (* $P < 0.05$).

39



40

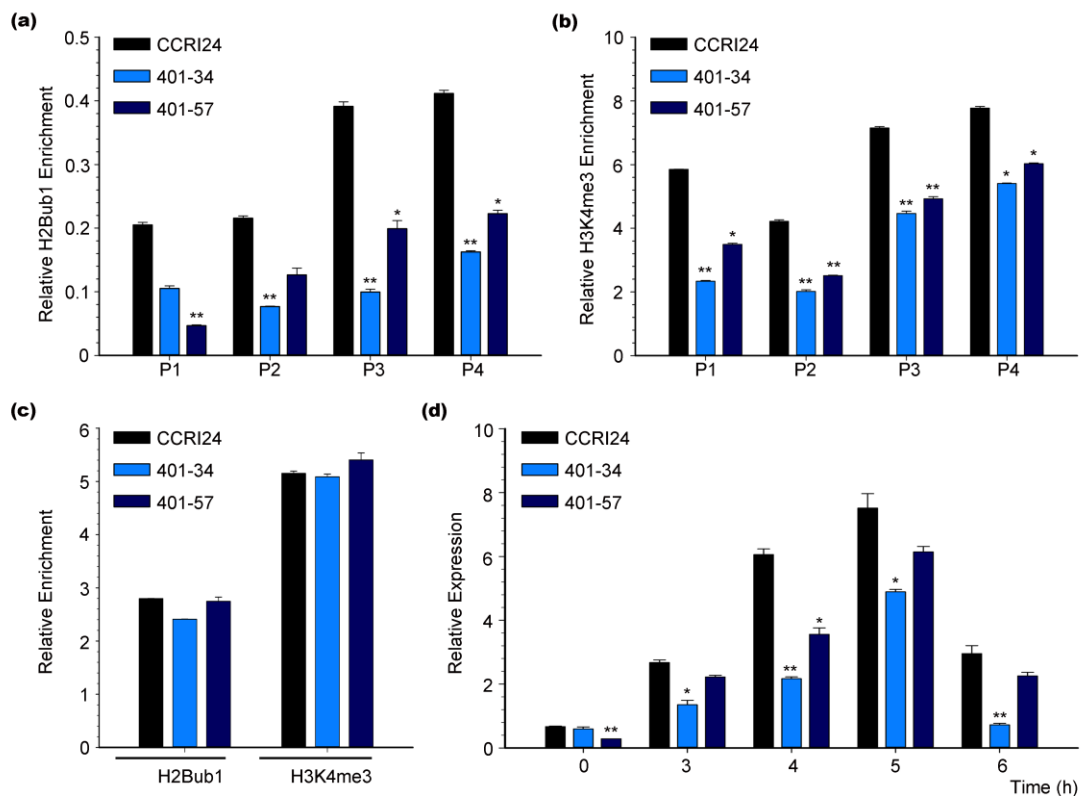
41 **Figure S5** *AtHUB2* significantly enhances the drought tolerance of transgenic
42 cotton in the greenhouse. Drought tolerance of control plants and *AtHUB2*-
43 expressing cotton plants. Photographs were taken after 35 days of drought
44 stress and after a 1-week recovery period. Bar = 7.5 cm.



45

46 **Figure S6** Multiple sequence alignment and phylogenetic tree analysis of
 47 GhH2B1 with *Arabidopsis* histone H2Bs. (a) Multiple sequence alignment was
 48 performed using CLC Sequence Viewer 7.0, and (b) the phylogenetic tree was
 49 constructed with the MEGA5 programme using the neighbour-joining method
 50 with 1,000 bootstraps. The red arrow indicates cotton histone H2B (GhH2B1).

51



52

53 **Figure S7** Detection *GhDREB* chromatin locus H2Bub1 and H3K4me3 levels
 54 in *GhHUB2*-knockdown plants. (a) H2Bub1 at the *GhDREB* locus in the CCR124
 55 and *GhHUB2*-underexpression plants. (b) H3K4me3 at the *GhDREB* locus in
 56 the CCR124 and transgenic lines. (c) The *GhUBI* gene was selected as a
 57 housekeeping gene that was not differentially expressed in the transgenic lines.
 58 (d) Expression of *GhDREB* at different time points of air drought treatment in
 59 control and *GhHUB2*-knockdown lines. Real-time RT-qPCR quantification was
 60 normalized to the expression of *GhUBI*. Vertical bars represent SD. (*P < 0.05,
 61 **P < 0.01).

62

63

64 **Table S1. The drought related indexes between transgenic cotton lines**
 65 **and controls.**

NO.	wilting	chlorosis	lodging
CCRI24	2	1	1
Line 2	1	0	0
Line 4	1	1	0
Line 6	1	1	0
ZR409	1	0	0

66 Wilting 1: part of the leaves mild wilting; 2: all of leaves mild wilting; 3: part of leaves mild wilting part

67 severe wilting; 4 most of leaves severe wilting; 5 all of leaves severe wilting

68 Chlorosis 1: part of the leaves mild chlorosis; 2:1/3 leaves chlorosis; 3: part of leaves yellow; 4: part of

69 leaves fall off; 5: leaves withered

70 Lodging 1: angle of inclination < 30°; 2: part angle of inclination >45°; 3: almost angle of inclination >45°;

71 4: part lodging; 5: all lodging

72

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77 **Table S2. Positive Clones from Library Screening.**

78

NO.	Blast description	Accession	Repeats
1	Histone H2B1 [Gossypium hirsutum]	AAB97163.1	1
2	SINAH1 protein [Gossypium hirsutum]	AAD53877.1	2
3	RING/U-box superfamily protein isoform 1 [Theobroma cacao]	XP_007029018.1	3
4	Xyloglucan endotransglucosylase/hydrolase 2 [Theobroma cacao]	XP_007049742.1	3
5	Uncharacterized protein TCM_014645 [Theobroma cacao]	XP_007037985.1	2
6	hypothetical protein JCGZ_11289 [Jatropha curcas]	KDP30913.1	2
7	Histone ubiquitination proteins group [Theobroma cacao]	XP_007052228.1	4
8	Zinc transporter 11 precursor isoform 1 [Theobroma cacao]	XP_007020535.1	1
9	hypothetical protein CISIN_1g009353mg [Citrus sinensis]	KDO77403.1	1
10	PREDICTED: ubiquitin-60S ribosomal protein L40-like [Nelumbo nucifera]	XP_010271616.1	1
11	GIA/RGA-like gibberellin response modulator [Gossypium hirsutum]	AAY28970.1	1
12	putative adenine nucleotide translocase [Castanea sativa]	AAL15894.1	1
13	Cell division protein ftsH [Populus trichocarpa]	XP_002301927.1	1
14	heat shock protein 70 [Gossypium hirsutum]	ACJ11745.1	1
15	uncharacterized protein LOC100306699 [Glycine max]	NP_001235125.1	1
16	chloroplast chlorophyll A-B binding protein [Gossypium hirsutum]	ACO51067.1	1
17	Chromatin remodeling 24 [Theobroma cacao]	XP_007048407.1	1

79

80 **Supplemental Table 3. Primers used in this study.**

81

82 **For cloning**

Name		Primer sequences (5'--3')
GFP-HUB2	Forward	ATGGATCCATGCTAACAAAGGCAG
	Reverse	CGGTCGACTTACATTTTGACAAGC
LUC ^c -HUB2	Forward	CCGGGGATCCGATGCTAACAAAGGCAGACGAACAG
	Reverse	AATTGTCGACTTACATTTTGACAAGCCGGACG
HUB2-GST	Forward	CGCGGATCCATGCTAACAAAGGCAGACGAACAG
	Reverse	CCCTCGAGTTACATTTTGACAAGCCGGACGTC
HUB2-BD	Forward	CGCCATATGCTAACAA AGGCAGACGA
	Reverse	CGCGGATCCTTACATTTTGACAAGCCGGACG
HUB2(1-160)-BD	Forward	CGCCATATGCTAACAA AGGCAGACGA
	Reverse	CGCGGATCCCAACAAATGT TGGTTCTGTG
HUB2(1-260)-BD	Forward	CGCCATATGCTAACAA AGGCAGACGA
	Reverse	CGCGGATCCGGCCGACTTG AGCCATCTGA
HUB2(160-330)-BD	Forward	CGCCATATGCAGCAGGTGG CTGAGCGGGA
	Reverse	CGCGGATCCTTTGAGGATGTTTTTGCAATTC
HUB2(260-330)-BD	Forward	CGCCATATGGTGTCTTCAT CCGAAAAGGA
	Reverse	CGCGGATCCTTTGAGGATGTTTTTGCAATTC
HUB2(330-383)-BD	Forward	CGCCATATG TGTGGTGTAT GTTTTGATCG
	Reverse	CGCGGATCCTTACATTTTGACAAGCCGGACG
HUB2-pENTR2b	Forward	AGGATCCGATGCTAACAAAGGCAG
	Reverse	ACCTCGAGTTACATTTTGACAAGC
HUB2-pDEST32	Forward	GGGGACAAGTTTGTACAAAAAAGCAGGCTTGATGCTA ACAAAGGCAG
	Reverse	GGGGACCACTTTGTACAAGAAAGCTGGGTTTACATTT TGACAAGC
HUB2 mutant	Forward	GTGGTAATTGTGAAAGCTTATGCCCTG

	Reverse	CTGGCAGAACAGGGCATAAGCTTT
pC1305.1-HUB2	Forward	<u>ATCCATGGTAGAACAGAAGTTGATTTCCGAAGAAGAC</u> <u>CTCATGCTAACA AAGGCAGACG</u>
	Reverse	<u>AAACTAGTGTTACATTTTGACAAGCCGGAC</u>
CaMV 35S promoter	Forward	GCTCCTACAAATGCCATCA
HUB2	Reverse	GTTACATTTTGACAAGCCGGAC
GFP-GhH2B1	Forward	<u>GGATCCATGGCACCAAAGCTGAAAA</u>
	Reverse	<u>GTCGACTCAAGATGAA GTAAACTTGG</u>
GhH2B1-LUC ⁿ	Forward	AATT <u>GAGCTCATGGCACCAAAGCTGAAAAAA</u>
	Reverse	TTAAG <u>TCGACAAAAGATGAAGTAACTTGGTCAC</u>
GhH2B1-His	Forward	CGC <u>GGATCCATGGCACCAAAGCTGAAAA</u> AAA
	Reverse	CCCTCGAGAAAAGATGAAGTAACTTGGTCAC
GhH2B1-AD	Forward	CCGGAATTCATGGCACCAA AAGCTGAAAA AAA
	Reverse	CGC <u>GGATCCTCAAGATGAAGTAACTTGGTCAC</u>
GFP-AtH2B	Forward	<u>GGATCCATGGCGCCGA GAGCAGAGAA</u>
	Reverse	<u>GTCGACTCAAGAGCTT GTGAATTTGG</u>
AtH2B-LUC ⁿ	Forward	CCGGG <u>GATCCGATGGCGCCGAGAGCAGAGAAGAAG</u>
	Reverse	TTAAG <u>TCGACTTTAGAGCTTGTGAATTTGGTAA</u>
AtH2B-His	Forward	CGC <u>GGATCCATGGCGCCGAGAGCAGAGAA</u> GAAG
	Reverse	CCCTCGAGTTTAGAGCTTGTGAATTTGGTAA
AtH2B-AD	Forward	CCGGAATTCATGGCGCCGAGAGCAGAGAA GAAG
	Reverse	CGC <u>GGATCCTCAAGAGCTTGTGAATTTGGTAA</u>
AtHUB2 southern blot	Forward	CCGAAGAAGACCTCATGCTAACAAAGGCAGAC
	Reverse	GATGAAGACACGGCCGACTTGAGCCATC

83

84 **For real-time PCR**

AtHUB2	Forward	GAAGCTGCAATTGTGAGGCTC
	Reverse	ATTTCCGGTGTTCGGATCTCT
GhUBI1	Forward	CTGAATCTTCGCTTTCACGTTA

	Reverse	GGGATGCAAATCTTCGTGAAA
GhP5CS	Forward	AGATGGATTCTGAAAGGAAG
	Reverse	CCTTTGACGACCAAGACCA
GhPP2C	Forward	ACGCGGTTCAAAACTCAAAA
	Reverse	TCCATATCTCGTCTCCTTCCA
GhRD22	Forward	AACAAACCCCAATGCAAAAG
	Reverse	AGCACCCCTCTAAAGGAACCA
GhDREB	Forward	GTTGATTTCTGGGTCGGTTT
	Reverse	AAGCCAAATCCTCGACTTCTT
GhPIP2;7	Forward	TACTGCACTGCCGGTATCTC
	Reverse	CAAGGCAGTTCCTTGTTGT
GhMKK2	Forward	TTGGTGTGAGTCGGATCTTG
	Reverse	GGCCCAATCACCTTGTCTT
XP_016725369.1	Forward	CGGAGGATGGTTTTATTGCT
	Reverse	ACCCCTTTTTGGAACCTTTG
XP_016729746.1	Forward	TCTACTACGGCTACGGCAAG
	Reverse	CCTCGTTAAAAGCCATCCA
XP_016722486.1	Forward	TCGTCTTTGTTTCCGGTTTC
	Reverse	TACCCTTCATGCACCCTTTC
XP_016700483.1	Forward	CAAGAAAGGGTGCATGAAGG
	Reverse	GTCACCGTACATGGCTTTGG
GhDREB P1	Forward	GTCCAAGCAAGAAATTAGGTCTGTATG
	Reverse	GAGCTTCAAATAAAACACACACGCC
GhDREB P2	Forward	CAGCCAAGTGGGTAATAACTG
	Reverse	GTTTCTGGAACTATAACTTGTAAGCC
GhDREB P3	Forward	CATTCCACTACCAAAAAGGAAGCCC
	Reverse	GGTGGCTAGTTGGCAAGTCAAG
GhDREB P4	Forward	CGGAGAGAAGCGAAAGCTGCA
	Reverse	GTAGAAAACGGCTGTGTCATAAGC

