

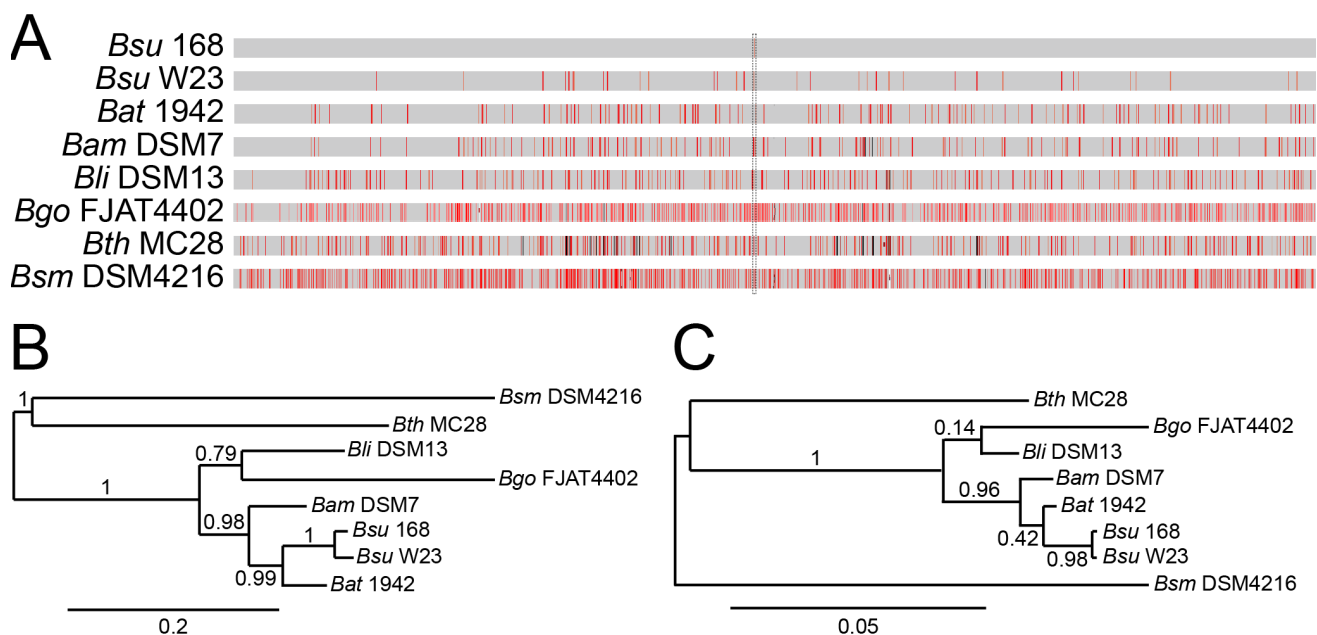
## Supplementary Material

### *Bacillus subtilis* MutS modulates RecA-mediated DNA strand exchange between divergent DNA sequences

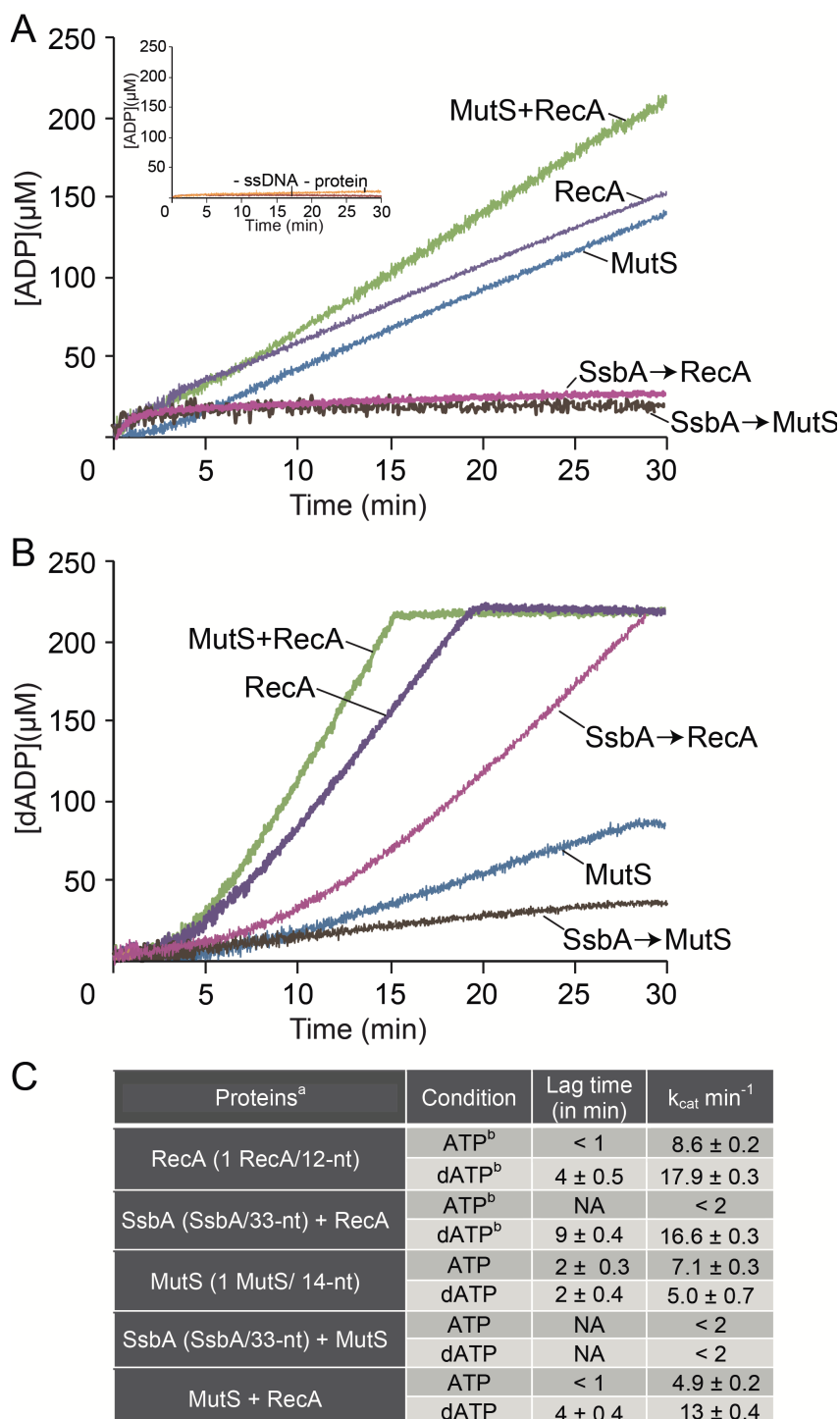
Begoña Carrasco<sup>1,†</sup>, Ester Serrano<sup>1,†</sup>, Alejandro Martín-González<sup>2</sup>, Fernando Moreno-Herrero<sup>2</sup> and Juan C. Alonso<sup>1,\*</sup>

<sup>1</sup>Department of Microbial Biotechnology, Centro Nacional de Biotecnología, CNB-CSIC, 28049 Madrid, Spain.

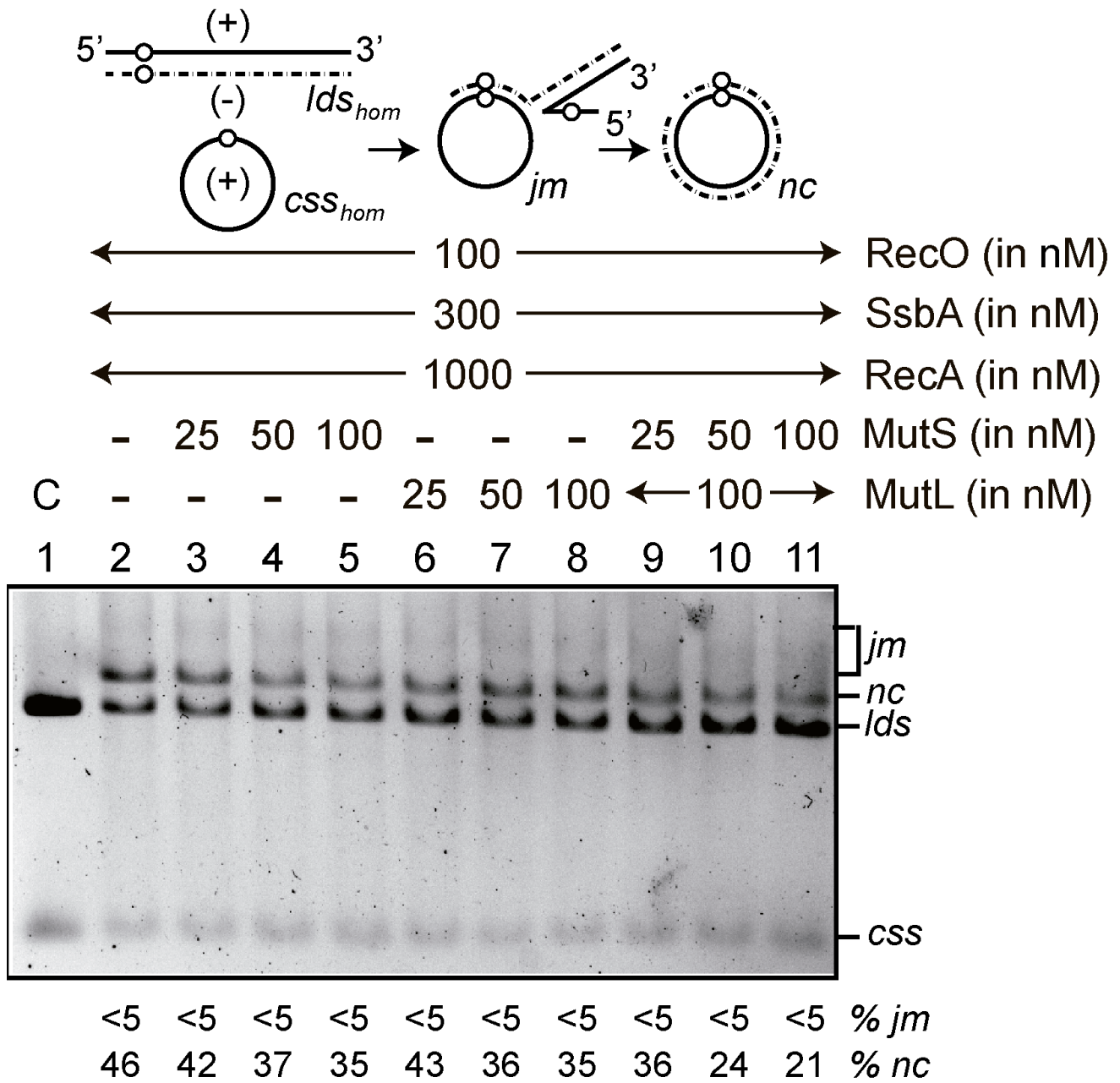
<sup>2</sup>Department of Macromolecular Structures, CNB-CSIC, 28049 Madrid, Spain



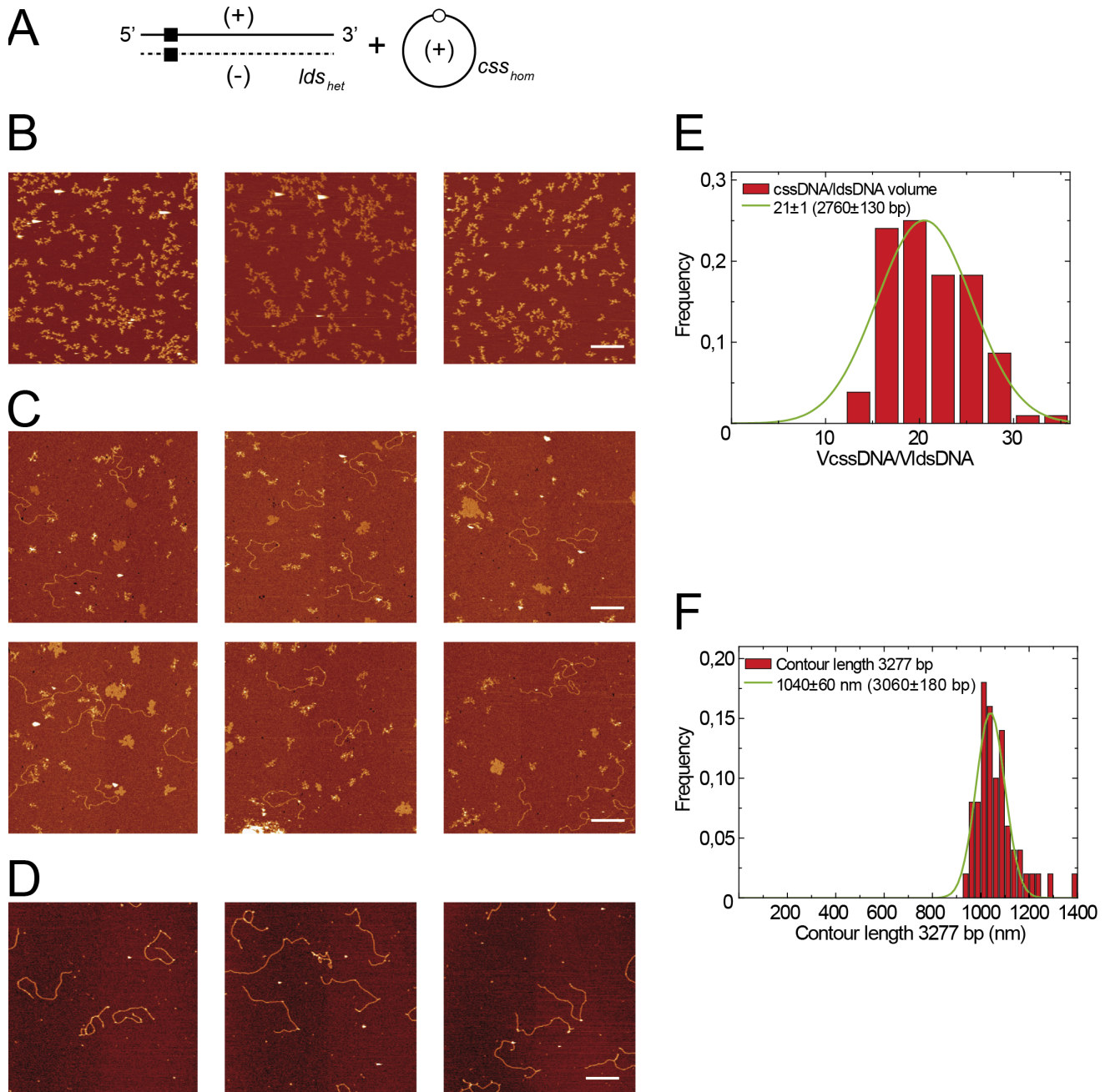
**Supplementary Figure 1.** Distribution of sequence divergence among different *Bacillus* species. **(A)** A single C-to-T transition mutation at codon 482 (framed by dotted lines) in the essential *rpoB* gene confers *Rif<sup>R</sup>* (*rpoB482* gene). The *rpoB482* DNA was derived from *B. subtilis* 168 (*Bsu* 168), *B. subtilis* W23 (*Bsu* W23), *B. atropheus* 1942 (*Bat* 1942), *B. amyloliquefaciens* DSM7 (*Bam* DSM7), *B. licheniformis* DSM13 (*Bli* DSM13), *B. gobiensis* FJAT-4402 (*Bgo* FJAT4402), *B. thuringiensis* MC28 (*Bth* MC28) and *B. smithii* DSM4512 (*Bsm* DSM4216). Mismatches between *Rif<sup>R</sup>* donor and the corresponding *rpoB* in the *Rif<sup>S</sup>* recipient strain are indicated by vertical red bars, and insertions/deletions by vertical black bars. Bar thickness represents the number of mismatches in the neighborhood. **(B and C)** Phylogenetic tree of the selected *Bacillus* species or subspecies. The tree is based on the *rpoB482* nucleotide sequence **(B)**, and of the RpoB protein sequence **(C)**. Branching confidence values are based on 1,000 bootstrap replicates. The alignment was performed using FASTA, BLAST, ClustalW, PhyML and MUSCLE 3.6.



**Supplementary Figure 2.** MutS effects on RecA nucleation and polymerization onto ssDNA. The 3276-nt *css<sub>hom</sub>* DNA (10 μM in nt) was incubated with MutS (0.7 μM), RecA (0.8 μM), or both MutS and RecA, or preformed SsbA-ssDNA complexes (SsbA, 0.3 μM) were incubated with MutS or RecA in buffer A containing 5 mM ATP (**A**) or dATP (**B**); the ATPase/dATPase activity was then measured (30 min, 37°C). Inset in (**A**) is a control experiment in the absence of ssDNA or any RecA or MutS. (**C**) Lag times and maximal rate of nucleotide hydrolysis are shown for different experimental conditions. NA: Not applicable. Results shown as mean ± SEM of ≥3 independent experiments.



**Supplementary Figure 3.** RecA·ATP-mediated DNA strand exchange in the presence of MutSL. The scheme shows the three-strand exchange reaction between homologous *css<sub>hom</sub>* (+ strand) and the homologous (- strand) *lds<sub>hom</sub>* (open circle) substrates. The 3276-nt *css<sub>hom</sub>* DNA (10 μM in nt) was preincubated with SsbA and RecO (5 min, 37°C) in buffer A containing 5 mM ATP, followed by RecA, (MutS, MutL or MutSL) and the 3276-bp *lds<sub>hom</sub>* DNA (20 μM in nt) substrate. The reaction was incubated (60 min, 37°C) and separated by 0.8% agarose gel electrophoresis. The positions of the bands corresponding to substrates (*css*, *lds*), intermediates (joint molecule, *jm*) and products (nicked circular, *nc*) are indicated. Lane 1, the *css* and *lds* substrates (termed C). The percentage of recombination intermediates (*jm*) and products (*nc*) are shown beneath the gel. Results shown as mean ± 5% SEM of ≥3 independent experiments. The minus (-) symbol denotes absence of MutS or MutL.



**Supplementary Figure 4.** AFM control experiments of *lds<sub>het</sub>* and *cSS<sub>hom</sub>* DNA. (A) drawing of the *lds<sub>het</sub>* (black square) and *cSS<sub>hom</sub>* (open circle) DNA substrates. DNA images of *cSS<sub>hom</sub>* (B), *cSS<sub>hom</sub>* and *lds<sub>het</sub>* (C) and *lds<sub>het</sub>* (D). The volume and the length histogram fitted to a Gaussian function for *cSS<sub>hom</sub>* (E) and *lds<sub>het</sub>* (F) are shown. Scale bar = 500 nm.