

Table S1. Primers used for genotyping and mRNA level analysis of *sec15b-1* and *sec15b-2* mutant lines.

Primer	Sequence (5' – 3')	Purpose
SEC15BPROM-RP	GTTGAATTCCTATGGTTTC TAGAAAAC	genotyping of SALK_130663 line
SEC15BPROM-LP	AAGCATCCGTTTCAGCGTT GAC	genotyping of SALK_130663 line
LBNEW	GAACAACACTCAACCCTAT CTCGGGC	genotyping of SALK_130663 line
SEC15BRIK-RPBF	TTGGATCCATATGCAATCG TCGAAAGGA	genotyping of RATM15- 1183-1_H line
SEC15BRIK-LPBIR	CAGTAAGAGATGATTAGCC GTC	genotyping of RATM15- 1183-1_H line
DS52A	TCCGTTCCGTTTTTCGTTTT TTAC	genotyping of RATM15- 1183-1_H line
NPT_LP	TCCGAGTACGTGCTCGCTC GATGC	primers specific to kanamycin resistance gene
NPT_RP	GCTTGGGTGGAGAGGCTAT TCGGC	primers specific to kanamycin resistance gene
SEC15BRIK-RPBF	TTGGATCCATATGCAATCG TCGAAAGGA	mRNA level analysis
SEC15BRIK-LPBIR	CAGTAAGAGATGATTAGCC GTC	mRNA level analysis
ACT7-FWD	GCCGATGGTGAGGATATTC AGC	mRNA level analysis
ACT7-REV	GAAACTCACCACCACGAAC CAG	mRNA level analysis

Table S2. Formation of adventitious roots in etiolated seedlings upon transfer to light

Genotype	Plants No	Roots from hypocotyl	Roots from collet	Roots from collet (%)
WT <i>SEC15b</i>	42	51	11	18
<i>sec15b</i>	56	6	40	87
WT <i>EXO70A1</i>	21	14	1	7
<i>exo70A1</i>	39	1	28	97

5-day-old etiolated seedlings were transferred to long day conditions and the presence of emerged adventitious roots was scored after 7 days. Sister WT segregants from the last backcross were evaluated for each genotype. For each gene, the mutation significantly affects adventive root location (χ^2 test with Benjamini-Hochberg correction for multiple testing $p < 0.001$). The effect on overall adventitious root number is marginally significant for *sec15b* (χ^2 test with Benjamini-Hochberg correction $p = 0.049$) and non-significant for *exo70A1*. GLMM analysis further confirmed that the distribution of adventitious roots is significantly affected by the genotype ($p = 0.027$ for WT vs. *sec15b*, $p = 0.007$ for WT vs. *exo70A1*), as well as by the location (collet vs. hypocotyl; $p < 0.001$ for either genotype), and indicated significant interaction between genotype and location ($p < 0.001$ for either genotype).

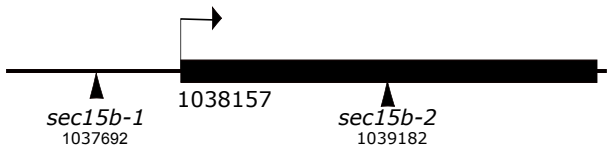
Figure S1.

(A) Graphical visualization of *sec15b-1* and *sec15b-2* insertions in *SEC15B* gene and promoter region.

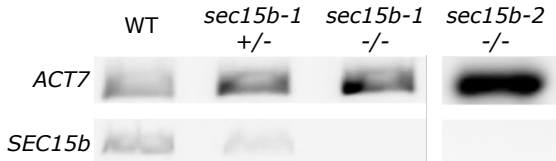
(B) RT-PCR confirmation of the absence of *SEC15B* mRNA in *sec15b-1* and *sec15b-2* mutant lines.

(A)

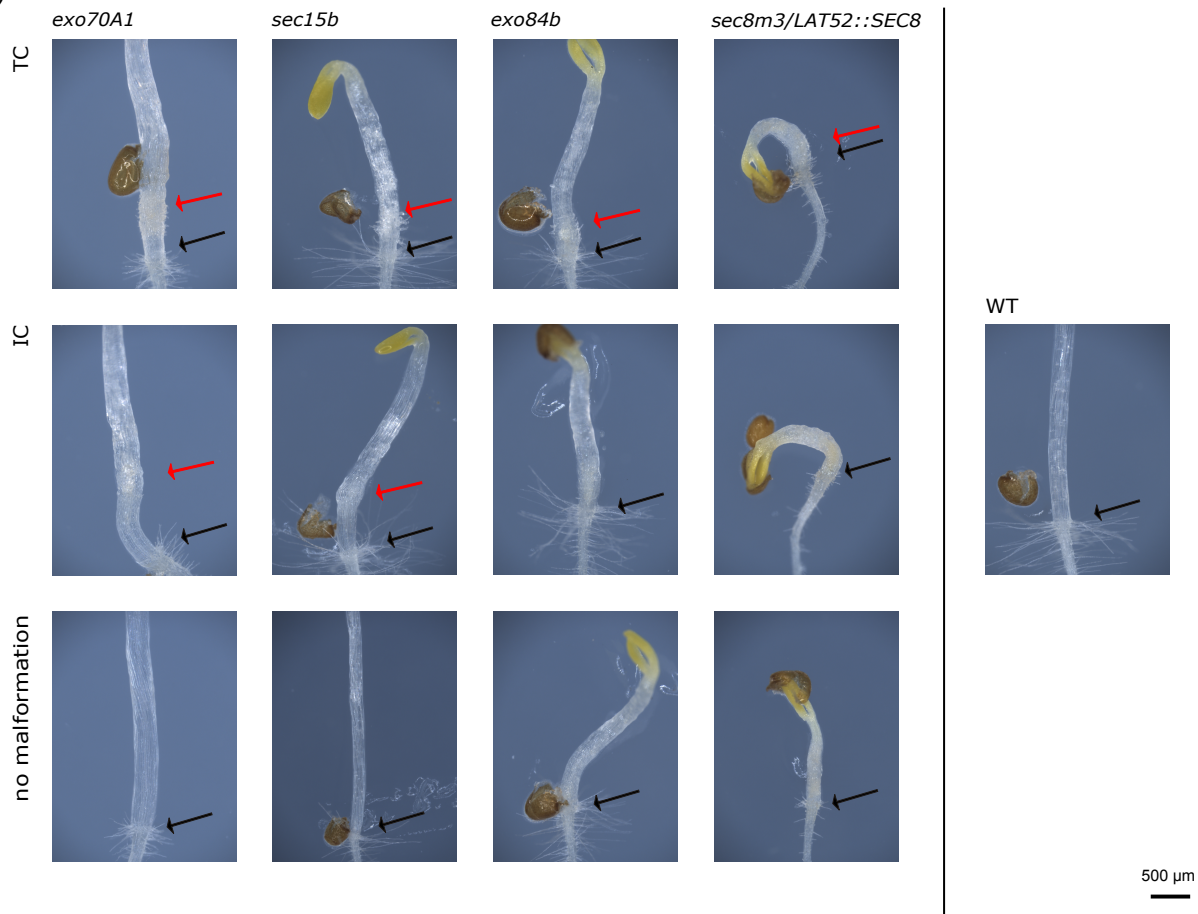
Chromosome 4, 1037194-1041693



(B)



(A)



(B)

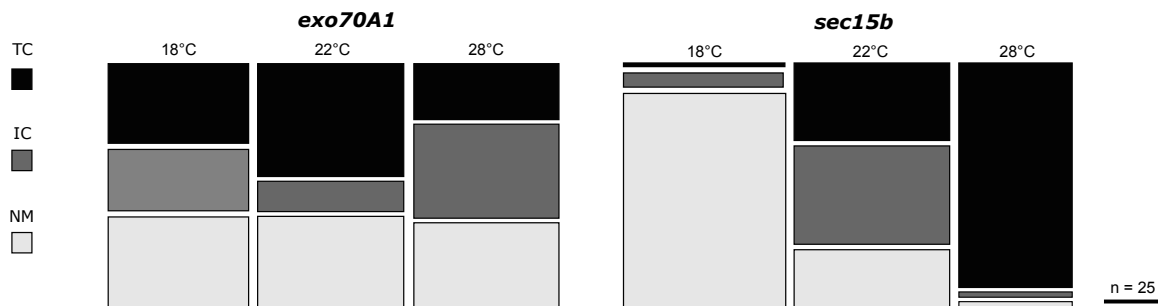


Fig. S2. Phenotypic defects of etiolated hypocotyls in exocyst mutants.

(A) Typical phenotypic deviations - ectopic collet-like structures with or without developed hairs (phenotypes TC and IC) and shortened hypocotyls without malformations in 5 days old *exo70A1*, *sec15b*, *exo84b* and *sec8m3/LAT52::SEC8* seedlings. Black arrows - collet hairs, red arrows - ectopic collet-like structures. A WT seedling is shown alongside for comparison. (B) Effect of cultivation temperature on the proportion of the phenotypic deviations in *exo70A1* and *sec15b* seedlings. Mosaic diagrams display fraction of individuals in the phenotypic categories among seedlings developed at the indicated temperature (NM – no malformation). In total 233 *exo70A1* and 452 *sec15b* mutant plants were evaluated; temperature effects were reproducibly highly significant only for the *sec15b* mutant (pairwise χ^2 test with Benjamini-Hochberg correction for multiple testing $p \ll 0.001$ for each temperature combination). Column width reflects the number of seedlings counted. Data from one experiment are shown; similar results were obtained in 3 experiments. All seedlings were 5 days old. No hypocotyl defects were ever observed in WT seedlings under these culture conditions.