

## **Transcriptional regulation of Hepatic Stellate Cell activation in NASH**

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### **Supplementary information:**

Supplementary figure 1

Supplementary figure 2

Supplementary figure 3

Supplementary figure 4

Supplementary table T1

Supplementary table T2

Supplementary table T3

Supplementary table T4

Supplementary table T5

## LEGENDS

### Supplementary Fig. 1

**a** IF confocal microscopy of liver sections from ctrl and CCl<sub>4</sub>-treated mice (0 and 8 weeks) stained for Desmin and  $\alpha$ SMA (scale bar 50 $\mu$ m). Background fluorescence was reduced using Huygens Software v. 16.10.

**b** IF confocal microscopy of isolated mouse HSCs transdifferentiating *in vitro* 1, 4, and 12 days post isolation and stained for  $\alpha$ SMA (scale bar 50 $\mu$ m). Background fluorescence was reduced using Huygens Software v. 16.10.

**c** Phase contrast microscopy of isolated mouse HSCs transdifferentiating *in vitro* 12 hours to 12 days post isolation (scale bar 50 $\mu$ m). Insert at 12 hours image showing 10x magnification of a single cell.

**d** Representative FACS plot showing sequential gating for vitamin A<sup>+</sup> cells, singlets, and live HSCs enriched from mouse liver cell suspension by density gradient centrifugation. Number of positive cells is indicated as fraction of parent population. Purity of the sorted cells is verified by microscopic evaluation.

**e** Coherent anti-Stokes Raman scattering (CARS) spectroscopy and second harmonic generation (SHG) microscopy of liver sections from Western diet-fed mice (scale bar 50 $\mu$ m).

**f** Hematoxylin and Eosin (H&E), sirius red staining, F4/80 IHC, and  $\alpha$ SMA IHC of FFPE liver sections from CCl<sub>4</sub>-treated mice (scale bars 100 $\mu$ m).

### Supplementary Fig. 2

**a** Cumulative distribution of RPKM values for all genes in three experimental models of HSC activation; WD feeding (24w vs. chow), CCl<sub>4</sub> treatment (8w vs. ctrl), and IVT (12d vs. ctrl). Kolmogorov–Smirnov tests were used to calculate corresponding D and p-values for pairwise comparisons of distributions. D is the maximal difference between distributions. Expression values represent averages of 3-4 biological replicates.

**b** Heat map of z-scores from 3913 genes induced or repressed  $\geq 4$ -fold (FDR < 0.05) in any experimental model and with median rlog expression across all samples of  $\geq 5$ . Samples and genes subjected to unsupervised hierarchical clustering. Scaling performed across all replicates in heatmap.

**c** PCA plot based on all regulated genes in any experiment ( $\log_2$ FC  $\neq 0$ ; FDR < 0.01, RPKM<sub>avr</sub> > 20). All replicates shown as small spheres. Bigger spheres indicate centroids of replicates.

**d** Heat map of 2484 genes, which are induced  $\geq 2$ -fold (FDR < 0.05) between any two time points in all three models, or repressed  $\geq 2$ -fold (FDR < 0.05) between any two time points in all three models. Exon counts are individually scaled for each model. Expression values represent averages of 3-4 biological replicates.

**e** Log<sub>2</sub>RPKM of HSC activation markers through all three experiments. Means within experiments not sharing a letter are significantly different (FDR < 0.05, edgeR exact test for overdispersed data).

### Supplementary Fig. 3

**a** Significance of motif enrichment for all genes changing expression ( $-2 \geq \log_2FC \geq 2$ ;  $FDR < 0.05$ ) between any two time points in our models. Motif enrichment analyses were performed on genes with reduced (red) and increased expression (green), all genes (dark) and model-*specific* genes (light). Motifs are grouped according to Pearson's correlation.

**b** Comparison of RPKM values for select hepatocyte TFs and HSC marker genes in isolated HSCs from 24-week WD- or chow-fed mice and whole liver (Liv.). RPKM values are normalized to whole liver expression ( $n=3$ ). Means not sharing a letter are significantly different ( $FDR < 0.05$ , edgeR exact test for overdispersed data).

**c** Motif correlation matrix showing known motifs from Homer motif enrichment analyses subjected to hierarchical clustering by their Pearson's correlation.

### Supplementary Fig. 4

**a**  $\log_2FC$  of the 870 *common* genes in WD and  $CCl_4$  experiments grouped by the presence of putative Ets1 and Ap1 binding sites or both (E+A). Dashed horizontal lines indicate average  $\log_2FC$  for all *common* genes in experiment. Means not sharing a letter are significantly different ( $FDR < 0.05$ , pairwise Wilcoxon rank-sum test, Benjamini-Hochberg corrected).

**b**  $\log_2$  fold enrichment of Ets1 (E), Runx1 (R), and dual motifs (ER) in promoters of genes falling into GO-SLIM categories and respective WD clusters where each GO-SLIM category is enriched. Enrichment calculated relative to all regulated genes in GO-SLIM analysis.

**c** Comparison of RPKM values for key ETS1/ETS1-*like* motif-containing genes, HSC activation markers, and proliferation-associated genes in mouse HSCs cultured for six days in the presence of 10  $\mu M$  U0126 (blue) or DMSO (red) ( $n=3$ ). WD cluster # and position of motifs relative to TSS indicated for each gene. \*  $FDR < 0.05$ , edgeR exact test for overdispersed data. RPKM values normalized to DMSO controls.

**d** qPCR analysis of primary, siRNA-transfected murine HSCs (C: Universal control siRNA, E<sub>I</sub>/E<sub>II</sub>: siEts1 I and II, R<sub>I</sub>/R<sub>II</sub>: siRunx1 I and II). Transfected cells harvested on day 6 post isolation (D6) and mRNA levels of indicated genes normalized to *Gtf2b* expression and day-1 levels (D1). Whole liver cDNA (L) was prepared from healthy, male C57BL6/J mice. Means not sharing a letter are significantly different ( $p_{adj} < 0.05$ , ANOVA, Tukey-adjusted).

### Supplementary Tables

**T1** Diet nutrient and energy composition of Western and chow diets. AFE = Atwater Fuel Energy.

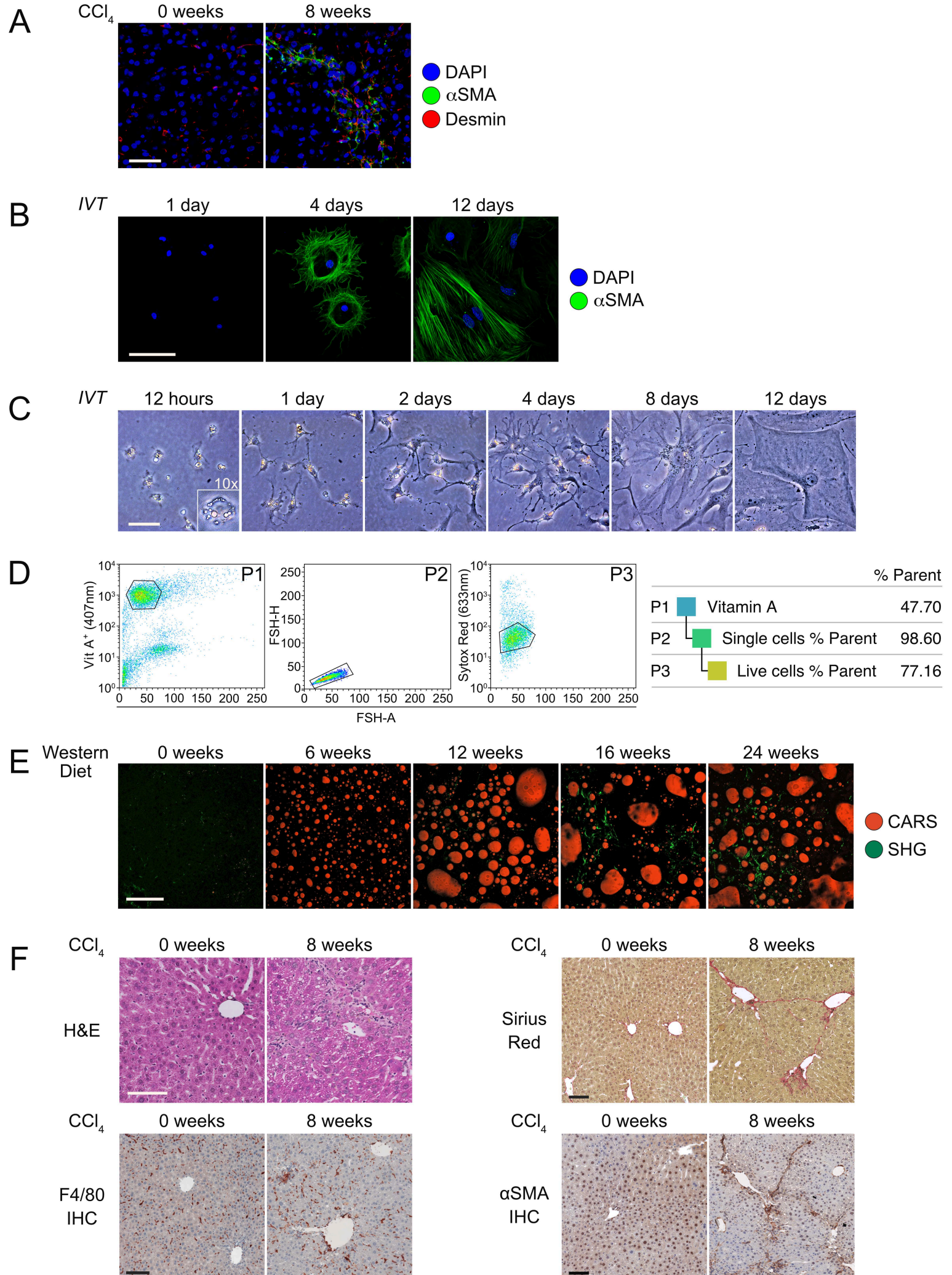
**T2** Gene cluster information from Western diet (WD) and  $CCl_4$  experiments. Pearson's correlation calculated for gene profiles within clusters and for cluster prototypes across experiments. Hypergeometric probabilities are calculated for cluster overlaps between experiments.

**T3** Homer motif logos, binding factors, and motifs enrichments in promoters of overlapping genes of WD and CCl<sub>4</sub> clusters 6. Ets-family TF, bZip/AP1, and Runx motif names shaded in blue, green and red, respectively.

**T4** List of primers used for RT-qPCR.

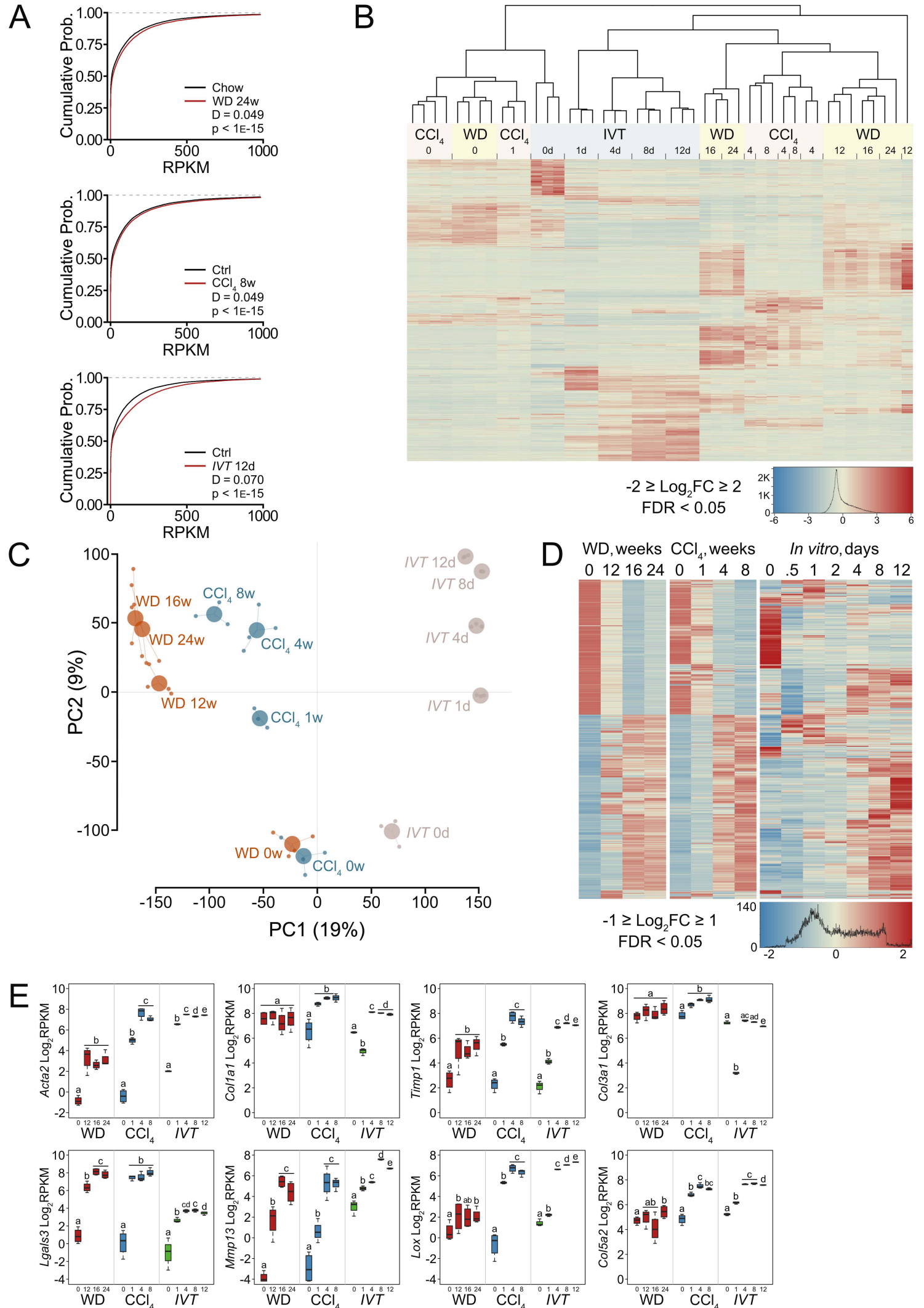
**T5** List of all *common* genes, genes of WD cluster 6 (blue), and *shared* genes between WD and CCl<sub>4</sub> clusters 6 (green).

# SUPPLEMENTARY FIGURE 1

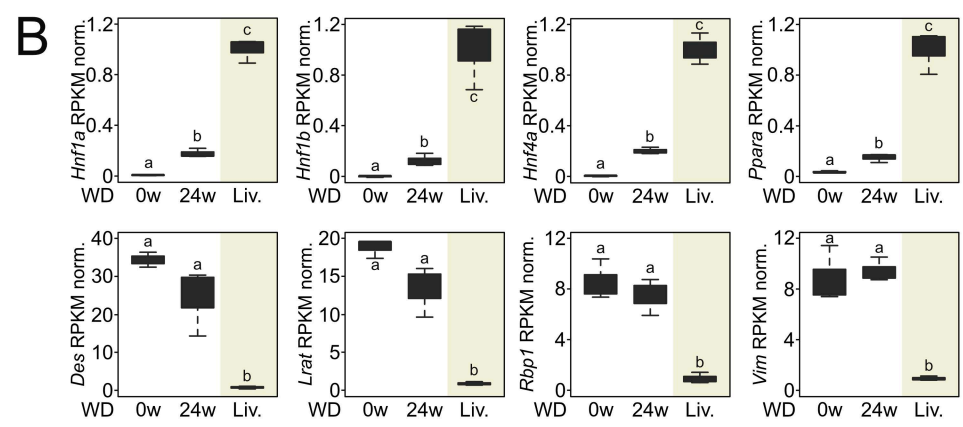
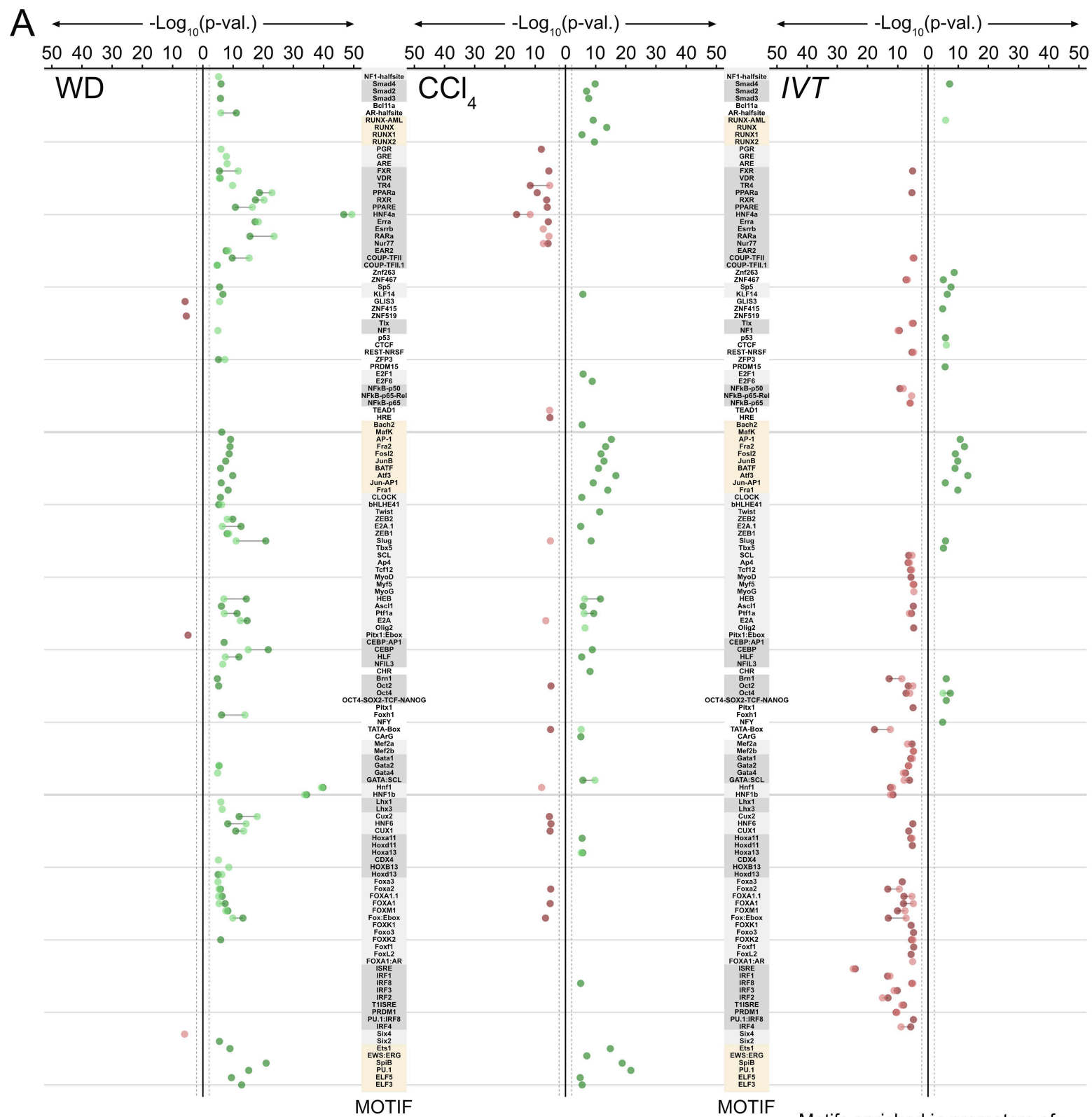




# SUPPLEMENTARY FIGURE 2



# SUPPLEMENTARY FIGURE 3

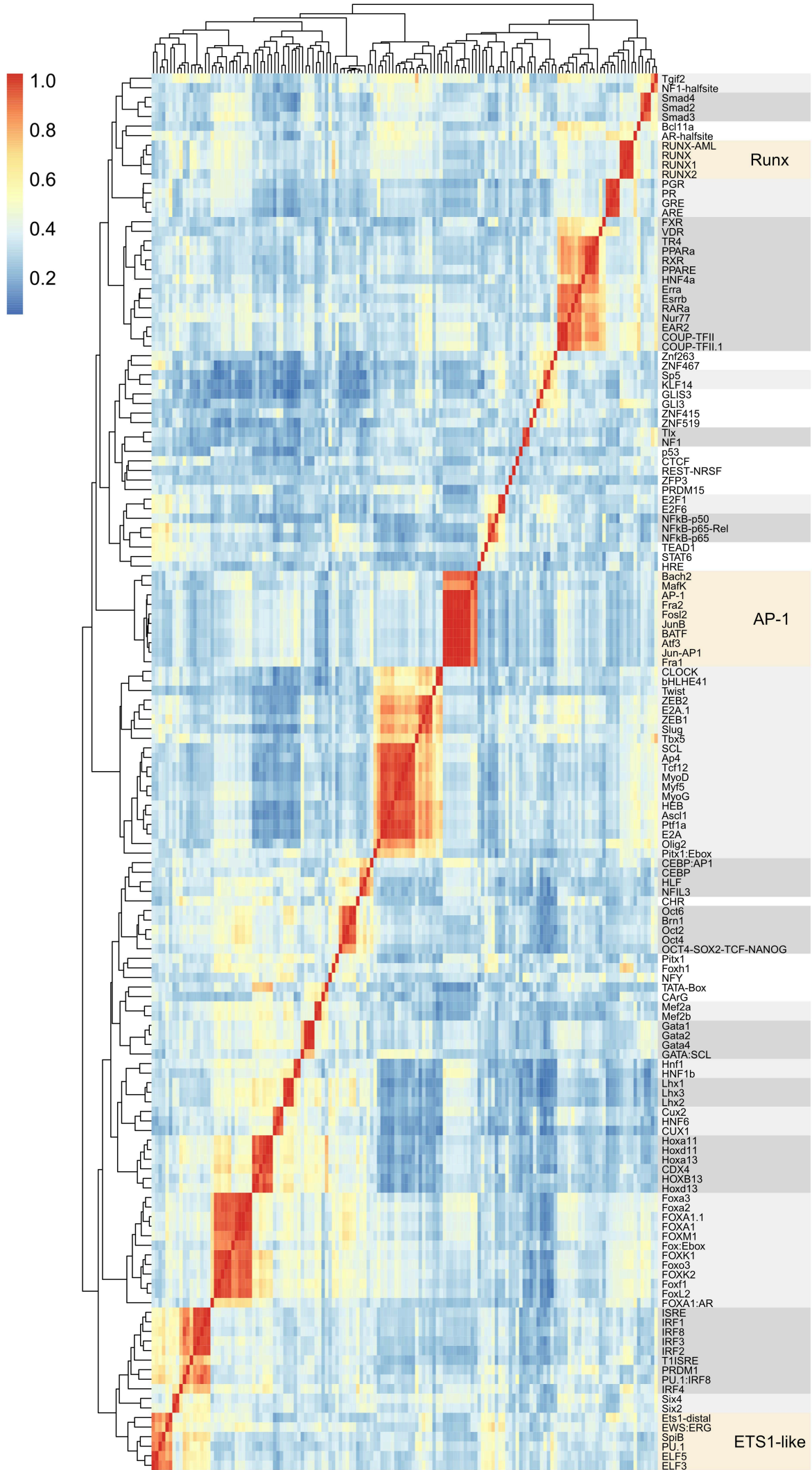


Motifs enriched in promoters of:

- Induced genes
- Specifically induced genes
- Repressed genes
- Specifically repressed genes

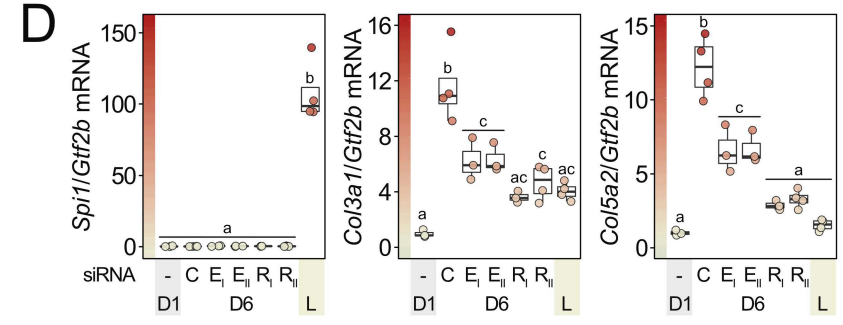
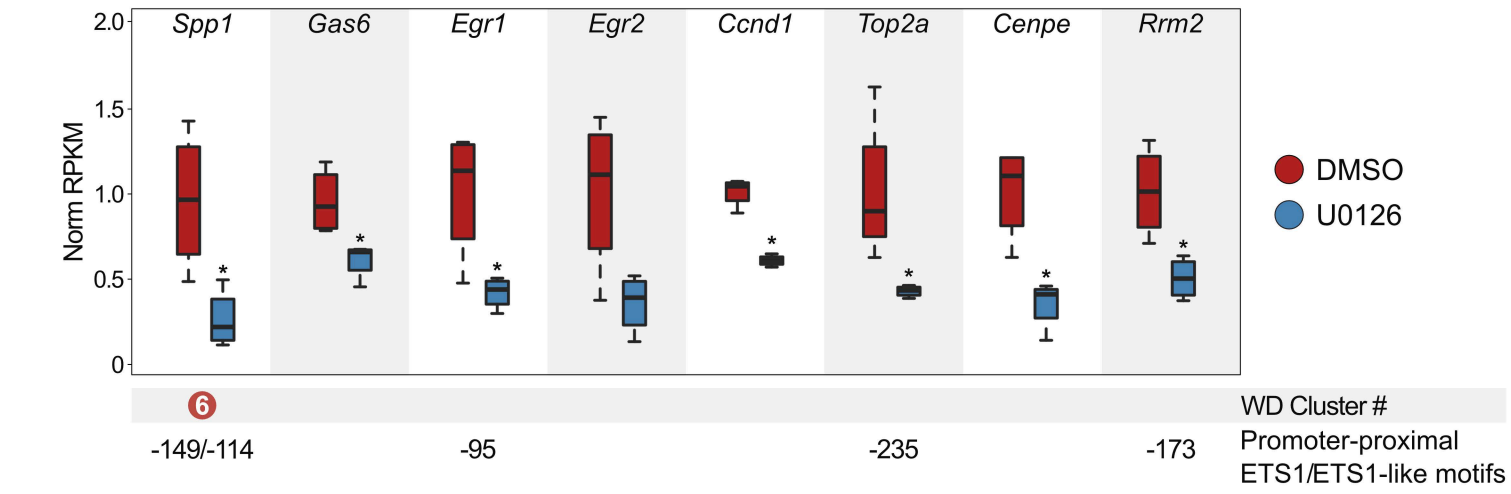
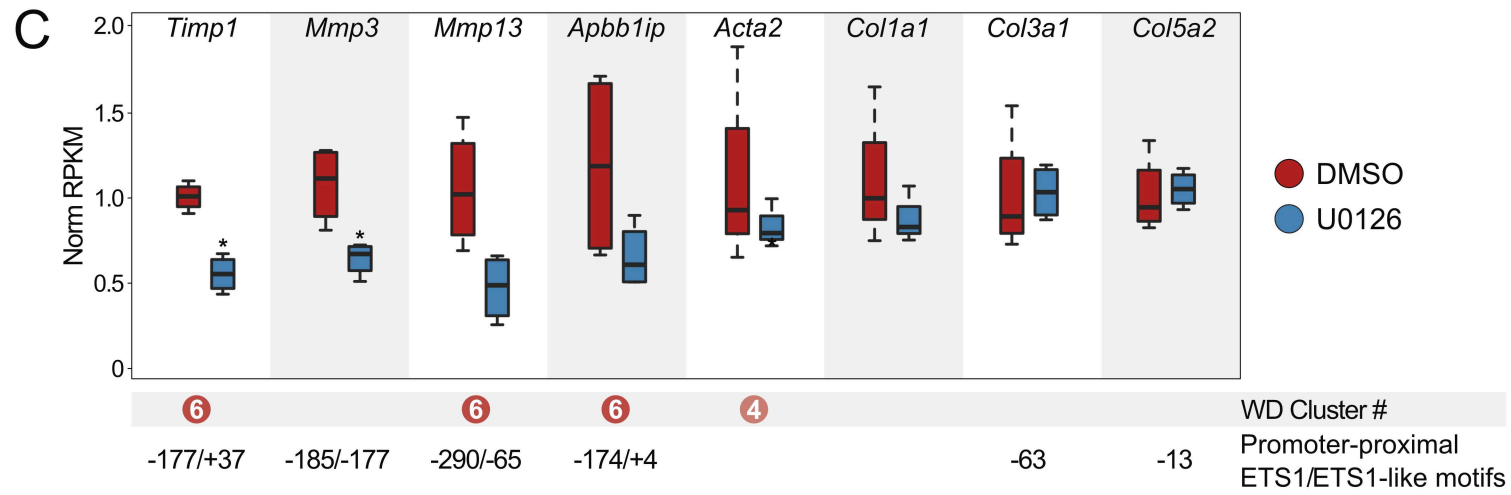
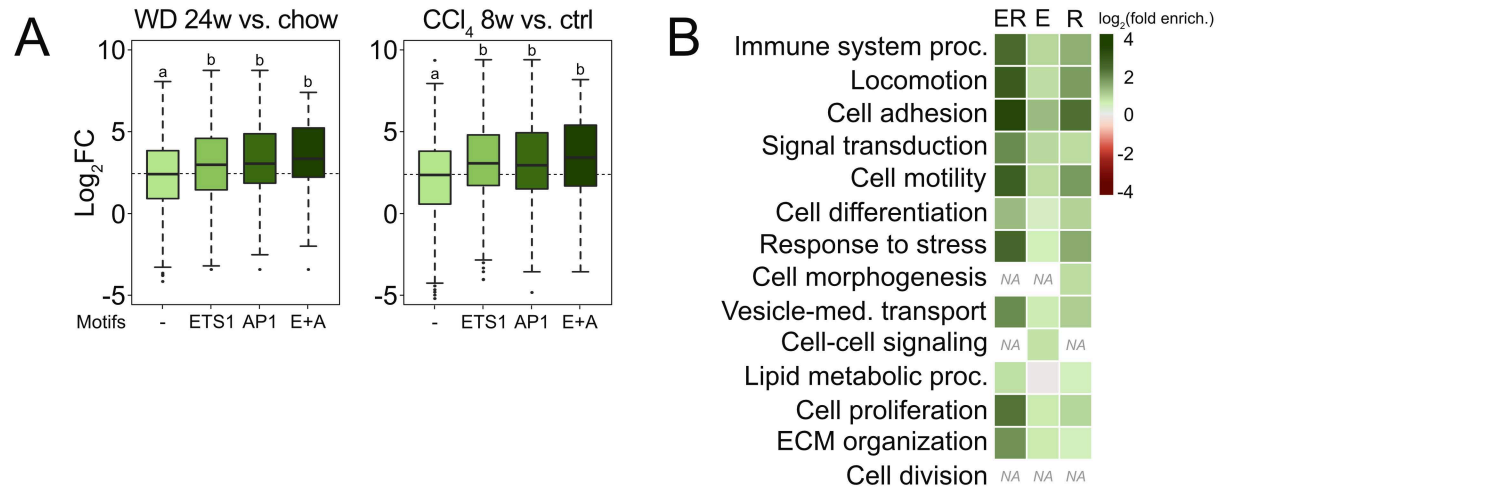
# SUPPLEMENTARY FIGURE 3 cont.

C





# SUPPLEMENTARY FIGURE 4



# SUPPLEMENTARY TABLES

T1

DIET: <b>WD</b>	829100 - 'Western RD'		
SPECIFICATION	%(w/w)	kcal/g	% kcal
Crude Fat	21.4	1.93	42
Crude Protein	17.5	0.70	15
Crude Fibre	3.5		
Ash	4.1		
Carbohydrate	50.0	2.00	43
Total AFE		4.63	100
Cholesterol	0.15		

DIET: <b>Chow</b>	Altromin 1324 #30404		
SPECIFICATION	%(w/w)	kcal/g	% kcal
Crude Fat	4.1	0.37	11
Crude Protein	19.2	0.77	24
Crude Fibre	6.1		
Ash	5.9		
Carbohydrate	43.9	2.09	65
Total AFE		2.89	100

T2

WD			CCl <sub>4</sub>			Overlap			
Cluster #	Gene count	Pearson corr.	Cluster #	Gene count	Pearson corr.	Pearson corr.	Gene count	% of WD cluster	Hypergeometric probability
1	174	0.98	1	175	0.97	0.99	156	90	1.59E-119
2	50	0.97	2	67	0.94	0.97	11	22	1.58E-03
3	40	0.95	3	62	0.95	1.00	9	23	2.13E-03
4	54	0.96	4	44	0.94	0.94	5	11	1.07E-01
5	153	0.97	5	90	0.96	0.90	12	13	3.13E-03
6	236	0.98	6	217	0.97	0.98	142	65	3.36E-38
7	70	0.98	7	113	0.96	1.00	25	36	1.94E-06
Total	777		Total	768		Total	360		

T3

Rank	Motif Logo	Name	p-value	log <sub>10</sub> (p-val.)	q-value	% of genes with motif	
						shared	background
1		PU.1	1.00E-08	20.3	0	30.30%	10.95%
2		ELF5	1.00E-06	15.5	0	34.85%	16.34%
3		ELF3	1.00E-06	14.1	0.0001	31.82%	14.84%
4		ETS1-distal	1.00E-05	13.2	0.0002	18.94%	6.61%
5		SPIB	1.00E-05	12.6	0.0003	15.91%	5.06%
6		EHF	1.00E-04	1.02	0.0024	43.18%	26.83%
7		ERG	1.00E-04	9.37	0.0044	50.76%	34.41%
8		ETV2	1.00E-03	8.52	0.0091	36.36%	22.41%
9		EWS:ERG-fusion	1.00E-03	8.26	0.0105	23.48%	12.24%
10		ETS1	1.00E-03	7.98	0.0125	39.39%	25.54%
11		FLI1	1.00E-03	7.43	0.0196	43.18%	29.49%
12		AP-1	1.00E-02	6.79	0.0341	17.42%	8.74%
13		ATF3	1.00E-02	6.20	0.057	15.15%	7.48%
14		BATF	1.00E-02	6.08	0.0593	15.15%	7.55%
15		RUNX	1.00E-02	5.95	0.0635	18.94%	10.48%
16		ETV1	1.00E-02	5.58	0.0857	44.70%	33.13%
17		JUNB	1.00E-02	5.07	0.1349	12.88%	6.59%
18		JUN-AP1	1.00E-02	4.85	0.159	7.58%	3.06%
19		FRA1	1.00E-02	4.69	0.1765	12.12%	6.29%

## T4

mGtf2b F	GTTCTGCTCCAACCTTTGCCT
mGtf2b R	TGTGTAGCTGCCATCTGCACTT
mEts1 F	TTGAGATTTTAGGGTTGCGCT
mEts1 R	GGCGGGATGAGAGTAAGTTTG
mRunx1 F	GCACTCTGGTCACCGTCATG
mRunx1 R	GTTGAATCTCGCTACCTGGTTC
mTimp1 F	AACTCGGACCTGGTCATAAGG
mTimp1 R	ACGCTGGTATAAGGTGGTCTC
mSpp1 F	ATCCTTGCTTGGGTTTGCAG
mSpp1 R	CACTGCCAATCTCATGGTTCG
mCol1a1 F	CAGGGTATTGCTGGACAACG
mCol1a1 R	GAAGACCAGGGAAGCCTCTT
mActa2 F	CCTGAAGAGCATCCGACACT
mActa2 R	GGTTGGCCTTAGGGTTCAG
mSpi1 F	TCACCCCAAGGGGACTATCT
mSpi1 R	GCCAGCACAAGTTCCTGATTT
mCol3a1 F	TCTGGAAGCCAGAACCATGT
mCol3a1 R	AATGGGATCTCTGGGTTGGG
mCol5a2 F	GTGCTGAAACAGACTGAGGC
mCol5a2 R	CTGAGGTCACTGTAGCACCA

## T5

0610040J01Rik  
 1600010M07Rik  
 1700024P16Rik  
 1700063D05Rik  
[2310040G24Rik](#)  
 2810408I11Rik  
 2810417H13Rik  
 3110039I08Rik  
[3300005D01Rik](#)  
[4632428N05Rik](#)  
[4930486L24Rik](#)  
[4930506M07Rik](#)  
 4931408D14Rik  
 4933416M07Rik  
 5033406O09Rik  
 5430416O09Rik  
[5430427O19Rik](#)  
[5430435G22Rik](#)  
 5730559C18Rik  
 9130008F23Rik  
 9130019P16Rik  
 9930014A18Rik

**870 Common genes**

**236 WD Cluster 6 genes**

**142 Shared genes**

A630033H2ORik

A830082N09Rik

[AA467197](#)

Abca4

Abcb1a

[Abcc3](#)

Abcg3

Abi3bp

Ablim3

Ace

Ace2

Ackr1

Ackr2

Ackr4

Acot1

Acot4

[Acp5](#)

Acta2

Acvr1c

[Acvrl1](#)

Adamts15

Adamts7

Adamtsl4

[Adap1](#)

Adcy4

Adcy5

[Adcy7](#)

[Adgb](#)

[Adora2b](#)

Adora3

Adra1a

[Adrb1](#)

[Adrb2](#)

[Adssl1](#)

[Afp](#)

Ahrr

[AI504432](#)

[AI607873](#)

Aim2

Akap12

Akr1c19

Aldh1a3

Aldh1a7

Aldh1b1

Aldh3a1

[Aldoc](#)

Alox5

Aloxe3

Als2cr12  
Amica1  
[Amz1](#)  
Anapc15  
Angpt2  
[Ank2](#)  
Ankrd1  
Ankrd33b  
Ano1  
Ano2  
Anpep  
[Ap1s3](#)  
[Apbb1ip](#)  
Apoa4  
Apol9a  
Aqp8  
Arc  
Arhgap11a  
Arhgap19  
Arhgap22  
Arhgap27  
Arhgap44  
[Arhgap9](#)  
Arhgef37  
[Arl11](#)  
Armc2  
[Asb2](#)  
Asf1b  
Asns  
Astn2  
Atp13a2  
[Atp1a3](#)  
Atp2a3  
Atp2b2  
[Atp6v0d2](#)  
[Atp8b4](#)  
Avpr1a  
B230217O12Rik  
[B3galnt1](#)  
B3gnt3  
B3gnt7  
B4galnt1  
[B4galt6](#)  
Baiap2l1  
Basp1  
Batf3  
Bdh1  
Bdh2



Bdnf  
Bend6  
Birc5  
Bmp4  
Bmp6  
C1ql1  
C230037L18Rik  
C5ar1  
C5ar2  
C77080  
C920009B18Rik  
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Cadm4  
Car13  
Car9  
Caskin1  
Casp1  
Cav1  
Ccdc125  
Ccdc136  
Ccdc148  
Ccdc183  
Ccl17  
Ccl6  
Ccnjl  
Ccr1  
Ccrn4l  
Cd14  
Cd200  
Cd24a  
Cd276  
Cd300lb  
Cd36  
Cd44  
Cd53  
Cd55  
Cd59a  
Cd80  
Cd83  
Cd9  
Cd93  
Cda  
Cdca7l  
Cdh1  
Cdh4  
Cdh5  
Cdkn2a  
Cdkn3

Cenpp  
[Cers6](#)  
Ces2e  
Cgnl1  
Ch25h  
Chaf1b  
Chek2  
Chn2  
Chst11  
Cit  
Clec14a  
[Clec4d](#)  
Clec4e  
Clhc1  
Clmn  
Clspn  
Cndp2  
Cobl  
Col23a1  
Col25a1  
Colec12  
Colgalt2  
Colq  
[Comp](#)  
Comtd1  
[Coro1a](#)  
Coro2a  
[Cox6a2](#)  
Cox8b  
Cpe  
[Creg2](#)  
Crlf1  
Crtam  
Crybb3  
Crym  
[Csf1r](#)  
Cthrc1  
Ctla2a  
[Ctla2b](#)  
Ctsh  
Cux2  
[Cx3cr1](#)  
Cxadr  
Cxcl3  
[Cxcr4](#)  
Cyb561  
Cyp7b1  
[Cystm1](#)

[Cyth4](#)

D6Ertd527e

Dbnnd1

Dcc

Dchs1

Dclk1

[Dctd](#)

Dctpp1

Dennd1c

Dennd2c

Depdc7

Dera

Derl3

Dhh

Dhrs13

Dhrs9

Disp2

Dlgap3

Dll4

Dna2

Dnajb13

Dnase2a

Dnph1

Doc2b

[Dock2](#)

[Dock5](#)

[Dok2](#)

Dsp

Dtl

Dusp13

Dusp14

Dyrk3

Dysf

[E2f1](#)

E2f8

Ece2

Edn1

Edn3

Efcc1

Efhd2

Efna1

Egfem1

Egfl6

Egln3

[Elmo1](#)

Elovl3

[Elovl7](#)

[Emb](#)

Emilin2  
Enpp6  
Entpd1  
Epb4.1l3  
Epb4.1l4b  
Epha4  
Erc2  
Esam  
Esr2  
Etv2  
Evi2a-evi2b  
Evi2b  
Exoc3l  
F13a1  
F630111L10Rik  
Fabp4  
Fabp5  
Fabp7  
Fads6  
Fam107b  
Fam110b  
Fam110c  
Fam129a  
Fam131c  
Fam167b  
Fam198b  
Fam19a1  
Fam20c  
Fam213a  
Fam65b  
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Fhad1  
Fhl2  
Fkbp11  
Fmn1  
Fmnl1  
Fmo2  
Fndc1

Folr1  
Foxj1  
Foxl1  
Frem3  
Frmd4b  
Frrs1  
Fst  
Fuca2  
Fxyd2  
Fyb  
Fzd9  
Gale  
Galnt12  
Galnt14  
Galnt3  
Galnt6  
Gas7  
Gata5  
Gatm  
Gcnt1  
Gcnt2  
Gda  
Gdf15  
Gdpd5  
Gfra2  
Ggta1  
Gjb3  
Gla  
Glp2r  
Glrp1  
Glrx  
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Gm5464  
Gm7694  
Gm9899



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Gna15  
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Gp49a  
Gpnmb  
Gpr114  
Gpr116  
Gpr137b  
Gpr137b-ps  
Gpr17  
Gpr176  
Gpr35  
Gpr39  
Gpr55  
Gpr56  
Gpr68  
Gprasp2  
Grhl3  
Gria3  
Grid1  
Grid2ip  
Grin2c  
Gsg1l  
Gsta1  
Gsta2  
Gsta3  
H19  
H2-Ab1  
H2-DMa  
H2-DMb1  
H2-Eb1  
Havcr2  
Hck  
Hcls1  
Hdac9  
Hebp1  
Hhex  
Hhip  
Hif3a  
Hist1h1d  
Hist1h2ae  
Hist1h4i  
Hmcn1  
Hmga2-ps1  
Hmha1  
Homer2

Hrh1  
Hs6st2  
Hs6st3  
Hspa12a  
Htra4  
Icos  
Ifi30  
Igf1  
Igf2  
Igfbp5  
Igsf10  
Igsf11  
Igsf21  
Ikzf4  
Il11  
Il13ra2  
Il17ra  
Il1rn  
Il2rg  
Il33  
Il7r  
Ildr2  
Inf2  
Inhbb  
Inpp5d  
Insc  
Insl3  
Irs3  
Irx3  
Itga2b  
Itga4  
Itga6  
Itgam  
Itgb2  
Jam2  
Kcnb1  
Kcng4  
Kcnj10  
Kcnj2  
Kcnk3  
Kcnn1  
Kcnn4  
Kctd15  
Kif19a  
Kif1a  
Kif5a  
Kif5c  
Kif9

Klhdc7a  
Klhdc8a  
Klk8  
Klra2  
Krt18  
Krt80  
Lacc1  
Lamb3  
Lamc3  
Laptm5  
Lbp  
Lcn2  
Lcn8  
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Lgi2  
Lilrb4  
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Liph  
Lmcd1  
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Lrrc27  
Lrrc4c  
Lrrc66  
Lrrn3  
Lrtm1  
Lsp1  
Lst1  
Ltbp2  
Ly9  
Lypd1  
Lyplal1  
Lyve1  
Lyz1  
Lyz2  
M1ap  
Mab21l3  
Mafa  
Mal2  
Mamdc2  
Map3k9  
Map4k1

Mapk8ip2  
Mast1  
Mb  
Mbp  
Mcf2  
Mcf2l  
Mcoln2  
Megf6  
Met  
Mfsd2a  
Mfsd6  
Mgam  
Mgat4a  
Mgst1  
Mis18bp1  
Mmd  
Mmp12  
Mmp13  
Mmp19  
Mmp27  
Mmp8  
Mpzl3  
Mrgpre  
Msi1  
Mt3  
Mybpc3  
Mycl  
Mycn  
Myct1  
Myh14  
Mylk3  
Myo1e  
Myof  
N28178  
Naip5  
Naip6  
Naip7  
Ncapg2  
Nceh1  
Ncf2  
Nckap1l  
Nckap5  
Nek6  
Ngef  
Nhirc4  
Nipal3  
Nlrc3  
Nlrp10

Npl  
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Nrap  
Nrep  
Nrn1  
Nrp2  
Nrros  
Nrxn1  
Nt5e  
Ntrk2  
Ntrk3  
Nxpe4  
Nxph3  
Nynrin  
Oas3  
Ociad2  
Odf3  
Olr1  
Onecut2  
Oprd1  
P2rx1  
P2rx7  
P2ry12  
Pacsin1  
Pak1  
Papln  
Parm1  
Parvg  
Pcbd1  
Pcdh15  
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Pcolce2  
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Piezo2  
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Pik3cb  
Pik3cd  
Pik3r5



Pik3r6  
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Pla2g4a  
Pla2g7  
Plagl1  
Plat  
Plau  
Plaur  
Plbd1  
Plcg2  
Plcx2  
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Plxna2  
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Ppef2  
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Prkch  
Prss46  
Prune2  
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Psd4  
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Ptgs2  
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Ptplad2  
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Ptpn6  
Ptprb  
Ptpre  
Ptprn  
Ptprt  
Ptprv  
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Rab11fip4

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Rab38  
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Rasgrp1  
Rassf6  
Rbm47  
Rcan2  
Reck  
Reps2  
Rftn1  
Rgcc  
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Rgs19  
Rgs2  
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Rhou  
Ripk3  
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Rufy4  
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Schip1  
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Scn8a  
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Sdsl  
Sectm1a

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Serpina3h  
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[Sh3bgrl2](#)  
Sh3kbp1  
Shc2  
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Ska1  
Slamf9  
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Slfn2  
Slfn3  
Slit3  
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Snx20  
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Spc25  
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Specc1  
Spire2  
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Syce1  
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Syng3  
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Tenm3

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Tns4  
Tox2  
Tpd52  
Trem2  
Trib3  
Tril  
Trim16  
Trpc4  
Trpm2

Trpm6  
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Tspan5  
Tspan8  
Ttc7  
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Tulp2  
Uhrf1  
Umps  
Unc13a  
Ung  
Vav3  
Vdr  
Vgl3  
Vill  
Vldlr  
Vnn1  
Wbscr17  
Wdr86  
Wfdc3  
Wfdc6a  
Wisp2  
Wnt2  
Wscd1  
Xylt1  
Ydjc  
Zcchc3  
Zfhx2  
Zfp423  
Zfp612