Supplementary Information

TITLE

Modular type I polyketide synthase acyl carrier protein domains share a common N-terminally extended fold.

AUTHORS

Luisa Moretto¹, Rachel Heylen², Natalie Holroyd³, Steven Vance⁴ & R. William Broadhurst^{2*}

AFFILIATIONS

¹ Department of Chemistry and Biomedical Sciences, Linnaeus University, Smålandsgatan-24, 392 34 Kalmar, Sweden.

² Department of Biochemistry, University of Cambridge, 80 Tennis Court Road, Cambridge CB2 1GA, U.K.

³ Department of Medical Physics and Bioengineering, University College London, London WC1E 6BT, U.K.

⁴ Crescendo Biologics Ltd, Meditrina Building 260, Babraham Research Campus, Cambridge CB22 3AT, U.K.

* Correspondence and requests for materials should be addressed to R.W.B. (email: rwb1002@cam.ac.uk)



Each panel displays a 4-12 % Bis-Tris gel containing Thermo Scientific PageRuler prestrained protein ladder (left lane) and the indicated protein construct after purification by size exclusion chromatography (right lane).



Deconvoluted electrospray mass spectra for samples of: (A) *apo* mACPa (9509.7 Da); (B) *apo* mH0ACPa (10598.0 Da); (C) *apo* mACPb (10224.5 Da); and (D) *apo* mH0ACPb (10512.8 Da).



Deconvoluted electrospray mass spectra for samples of: (a) *holo* mH0ACPa (10937.1 Da); and (b) β -hydroxybutyryl-mH0ACPa (11023.1 Da).



Size exclusion chromatography elution profiles for: (A) mACPa (red) and mHOACPa (blue); and (B) mACP9 (red) and mHOACP9 (blue).



1D ¹H spectra of *apo* mACPa (blue) and *apo* mH0ACPa (red). *, reference signal from 3,3,3-trimethylsilylpropionate.



[¹H,¹⁵N]-HSQC spectrum of *apo* mHOACPa, showing residue assignments for backbone amide sites. Pairs of resonances from side-chain amide sites are connected using magenta lines. Assignments for closely spaced signals are displayed in two inset panels. Add 11000 to obtain the sequence position within MLSA1.



[¹H,¹⁵N]-HSQC spectrum of *apo* mH0ACPb, showing residue assignments for backbone amide sites. Pairs of resonances from side-chain amide sites are connected using magenta lines. Assignments for closely spaced signals are displayed in the inset panel. Add 13000 to obtain the sequence position within MLSB.



Underneath a schematic defining the boundaries of the secondary structure elements in *apo* mH0ACPb, nuclear spin relaxation parameters for backbone amide sites are plotted as a function of residue number for: (A) the ¹⁵N longitudinal relaxation rate, R_1 ; (B) the ¹⁵N transverse relaxation rate, R_2 ; (C) the {¹H}-¹⁵N nuclear Overhauser effect ratio (I'/I₀, where I' is the intensity when the ¹H spectrum has been saturated and I₀ is the intensity in the reference spectrum); and (D) the Lipari-Szabo the order parameter, S^2 .



Representative circular dichroism thermal denaturation curves for *apo* mACPb (blue) and *apo* mH0ACPb (red), obtained by following the mean residue ellipticity at 222 nm.



Representative ITC thermograms (upper panels) and isotherm plots (lower panels), all with mKRb and NADPH in the cell and showing consecutive injections: (A) *apo* mACPb; (B) *holo* mACPb; (C) *apo* mH0ACPb; and (D) *holo* mH0ACPb. Thermogram traces for dilution control experiments are shown at the top of each upper panel.



Representative ITC thermograms (upper panels) and isotherm plots (lower panels), all with mKRb and NADP⁺ in the cell and showing consecutive injections: (A) *apo* mACPb; (B) *holo* mACPb; (C) *apo* mH0ACPb; and (D) *holo* mH0ACPb. Thermogram traces for dilution control experiments are shown at the top of each upper panel.

bits ∾ of the amphotericin PKS; Ery4, module 4 of the 6DEB PKS; mH0a, module 5 of MLSA1; mH0b, module 7 of MLSB. Keatinge-Clay, A.T. (2012) Supplementary Figure S12 ACP fold at the bottom. ACP domain sources: Os11, module 11 of oxazolomycin PKS; Spn2, module 2 of the spinosyn PKS; Am16, module 16 structure predictions of helices from SSPRO (cyan), experimental heliical regions (magenta), and secondary structure annotation for the HO-ACP domains displaying the domain definitions suggested by Keatinge-Clay (2012) labelled at the top and in underlined italics, secondary Nat. Prod. Rep., 29, 1050-1073 (A) Sequence logo plot for an alignment of 349 ACP domains from modular type I polyketide synthase systems. (B) Sequence alignment of Spn2 Ery4 Am16 0s11mH0a mH0b 10 2 3 4 5 $a {\tt RAGLAERLAVLPEEQRLPF} vvdlv {\tt RAEAAT} vlghgs a da v da {\tt REFRGLGF} ds lta i e l {\tt RNRLGKA} s glt t l ta t l v f d {\tt PTPQ} la e h l l de l l ga a da v da {\tt RAGLAERLAVLPEEQRLPF} vvdl v {\tt RAEAAT} v {\tt RAGLAERLA} v da {\tt RAGLAERLAVLPEEQRLPF} vvdl v {\tt RAEAAT} v {\tt RAGLAERLA} v da {\tt RAGLAERLA} v {\tt RAGLAERLA}$ ${\tt DAESLRKRL} GRLp {\tt DAEQHRI} {\tt LLDL} {\tt VRMH} {\tt VAAV} {\tt GFAGSQE} {\tt ITADGTFKVL} {\tt GFDSL} {\tt TVVELRNR} {\tt INGATGLR} {\tt ATL} {\tt VFN} {\tt PTP} {\tt DALAAHLVTAL} {\tt SAD} {\tt CACCUTAL} {\tt CACUUAL} {\tt CACUUAL}$ DTGGGLDGLAGLPEDEQRAR VMELVRRQ VAAVLVVD-PDDVGPDTGFVELGLDSLTAVELRNRLGRATGLRLPVTLVFDHPSPGALAGYLRQR AATDLAARLNGLSPQQQQQTLATLVAAATATVLGHHTPESISPATAFKDLGIDSLTALELRNTLTHNTGLDLPPTLIFDHPTPHALTQHLHTRLTQS HO "helix 0" ωOD H2 / 3<) <٢ canonical ACP domain H2 НЗ′ weblogo.berkeley.edu LTTA τ 100 റ

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(a) Contact map for mH0ACPa (top right) and contacts predicted using evolutionary covariance analysis (bottom left; contacts across the H0-H1 hinge are in the red box). (b) Location of backbone amide N atoms of residues identified in the H0-H1 hinge identified by GREMLIN.

A mACPa

GGLTATEQRA VTRKLVLDQA ASVLGYASTE SLDTHESFKD LGFDSLTALE LRDHLQTATG LNLSSTLIFD HPTPHAVAEH LLEQIPGIG

B mHOACPa

GSTATLLTSK LAGLTATEQR AVTRKLVLDQ AASVLGYAST ESLDTHESFK DLGFDSLTAL ELRDHLQTAT GLNLSSTLIF DHPTPHAVAE HLLEQIPGIG

C mACPb

GSHMRLNGLS PQQQQQTLAT LVAAATATVL GHHTPESISP ATAFKDLGID SLTALELRNT LTHNTGLDLP PTLIFDHPTP HALTQHLHTR LTQSH

D mHOACPb

GAASAATDLA ARLNGLSPQQ QQQTLATLVA AATATVLGHH TPESISPATA FKDLGIDSLT ALELRNTLTH NTGLDLPPTL IFDHPTPHAL TQHLHTRLTQ SH

E mKRb

GDSLITRPLT TATGSAPATT AAGLLHLSWP PHPDTTTDTD TDTDALRYQV IAEPTQQLPR YLHDLHTSTD LHTSTTEADV VVWPVPVPSN EELQAHQASD TAVSSRIHTL TRQTLTVVQD WLTHPDTTGT RLVIVTRHGV STSAHDPVPD LAHAAVWGLI RSAQNEHPGR FTLLDTDDNT NSDTLTTALT LPTRENQLAI RRDTIHIPRL TRHSSDGALT APVVVDPEGT VLITGGTGTL GALFAEHLVS AHGVRHLLLT SRRGPQAHGA TDLQQRLTDL GAHVTITACD ISDPEALAAL VNSVPTQHRL TAVVHTAAVL ADTPVTELTG DQLDQVLAPK IDAAWQLHQL TYEHNLSAFI MFSSMAGMIG SPGQGNYAAA NTALDALADY RHRLGLPATS LAWGYWQTHT GLTAHLTDVD LARMTRLGLM PIATSHGLAL FDAALATGQP VSIPAPINTH TLARHARDNT LAPILSALIT TPRRR

Supplementary Figure S14

Amino acid sequences are shown for the following protein fragments: mACPa (A), mH0ACPa (B), mACPb (C), mH0ACPb (D) and mKRb (E).