

SUPPORTING INFORMATION

Proteomic evaluation of the acute radiation syndrome of the gastrointestinal tract in a murine total-body irradiation model.

Weiliang Huang*, Jianshi Yu*, Jace W. Jones*, Claire L. Carter*, Keely Pierzchalski*, Gregory Tudor†, Catherine Booth†, Thomas J. MacVittie‡, Maureen A. Kane* §.

*University of Maryland, School of Pharmacy, Department of Pharmaceutical Sciences, Baltimore, MD; †Epistem Ltd, Manchester, UK; ‡ University of Maryland, School of Medicine, Department of Radiation Oncology, Baltimore, MD

§Correspondence:

Maureen A. Kane
University of Maryland, School of Pharmacy
Department of Pharmaceutical Sciences
20 N. Pine Street, Room 723
Baltimore, MD 21201
Phone: (410) 706-5097
Fax: (410) 706-0886
Email: mkane@rx.umaryland.edu

Retinoid analysis.

Retinoid levels were determined by liquid chromatography-multistage tandem mass spectrometry (LC-MRM³) which is an LC-MS/MS method utilizing two distinct fragmentation events for enhanced selectivity (Jones et al. 2015). Preparation of mouse intestinal tissue included flushing out contents with PBS. The jejunum and ileum were isolated from the small intestine and snap frozen with liquid nitrogen and stored at -80 °C until extraction. Tissues were homogenized in saline and extraction of retinoids was performed under yellow lights using a two-step liquid-liquid extraction that has been described in detail previously using 4,4-dimethyl-RA as an internal standard (Kane et al. 2005; Kane et al. 2008b; Kane and Napoli 2010; Jones et al. 2015). Levels of RA were measured using a Shimadzu Prominence UFLC XR liquid chromatography system (Shimadzu, Columbia, MD) coupled to an AB Sciex 5500 QTRAP hybrid triple quadrupole mass spectrometer (AB Sciex, Framingham, MA) using atmospheric pressure chemical ionization (APCI) operated in positive ion mode as previously described (Jones et al. 2015). Retinol and RE were quantified via HPLC-UV according to previously published methodology (Kane et al. 2008a; Kane and Napoli 2010). Retinoid levels were measured using n=5 per condition except for control (0) which is an average of n=15 un-irradiated control mice euthanized at day -6 (n=5), day 0 (n=5), or day 6 (n=5). The statistical analysis relied on the unpaired Student's *t*-test between groups and the data was represented as means ± standard deviation. Retinoid data are shown in **Supplemental Fig. 11** and **Supplemental Fig. 12**.

References.

- Jones JW, Pierzchalski K, Yu J, Kane MA. Use of fast HPLC multiple reaction monitoring cubed for endogenous retinoic acid quantification in complex matrices. *Anal Chem* 87: 3222-30; 2015.
- Kane MA, Chen N, Sparks S, Napoli JL. Quantification of endogenous retinoic acid in limited biological samples by LC/MS/MS. *Biochem J* 388: 363-9; 2005.
- Kane MA, Folias AE, Napoli JL. HPLC/UV quantitation of retinal, retinol, and retinyl esters in serum and tissues. *Anal Biochem* 378: 71-9; 2008a.
- Kane MA, Folias AE, Wang C, Napoli JL. Quantitative profiling of endogenous retinoic acid in vivo and in vitro by tandem mass spectrometry. *Anal Chem* 80: 1702-8; 2008b.
- Kane MA, Napoli JL. Quantification of endogenous retinoids. *Methods Mol Biol* 652: 1-54; 2010.

Supplemental Figures

Supplementary Figure 1. Expression of proteins most changed after radiation. Minimum FC > 10 of expression for at least one condition and FDR adjusted ANOVA $p < 0.01$. Data shown as a function of dose on a given day after dose. Data shown as a function of time after dose at a given radiation dose is shown in **Fig. 1**.

Supplementary Figure 2. Canonical pathways altered by radiation. (2a) Canonical pathways altered after radiation where criteria for pathway changes was activation z-score > 2 for at least one condition and Fisher's exact test $p < 0.01$. **(2b)** Protein changes associated with the top three pathways altered by radiation. Minimum FC > 2 for at least one condition with a FDR corrected ANOVA $p < 0.05$. Data shown as a function of dose on a given day after dose. Data shown as a function of time after dose at a given radiation dose is shown in **Fig. 2**.

Supplementary Figure 3. Upstream regulators altered by radiation. Criteria for transcription regulators was absolute activation z-score > 2 for at least one condition and a Fisher's exact test $p < 0.01$. Data shown as a function of dose on a given day after dose. Data shown as a function of time after dose at a given radiation dose is shown in **Fig. 3**.

Supplemental Figure 4. Venn diagram of differential protein expression showing the effect of day after 8 Gy irradiation.

Supplemental Figure 5. Venn diagram of differential protein expression showing the effect of day after 10 Gy irradiation.

Supplemental Figure 6. Venn diagram of differential protein expression showing the effect of day after 12 Gy irradiation.

Supplemental Figure 7. Venn diagram of differential protein expression showing the effect of day after 14 Gy irradiation.

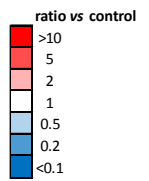
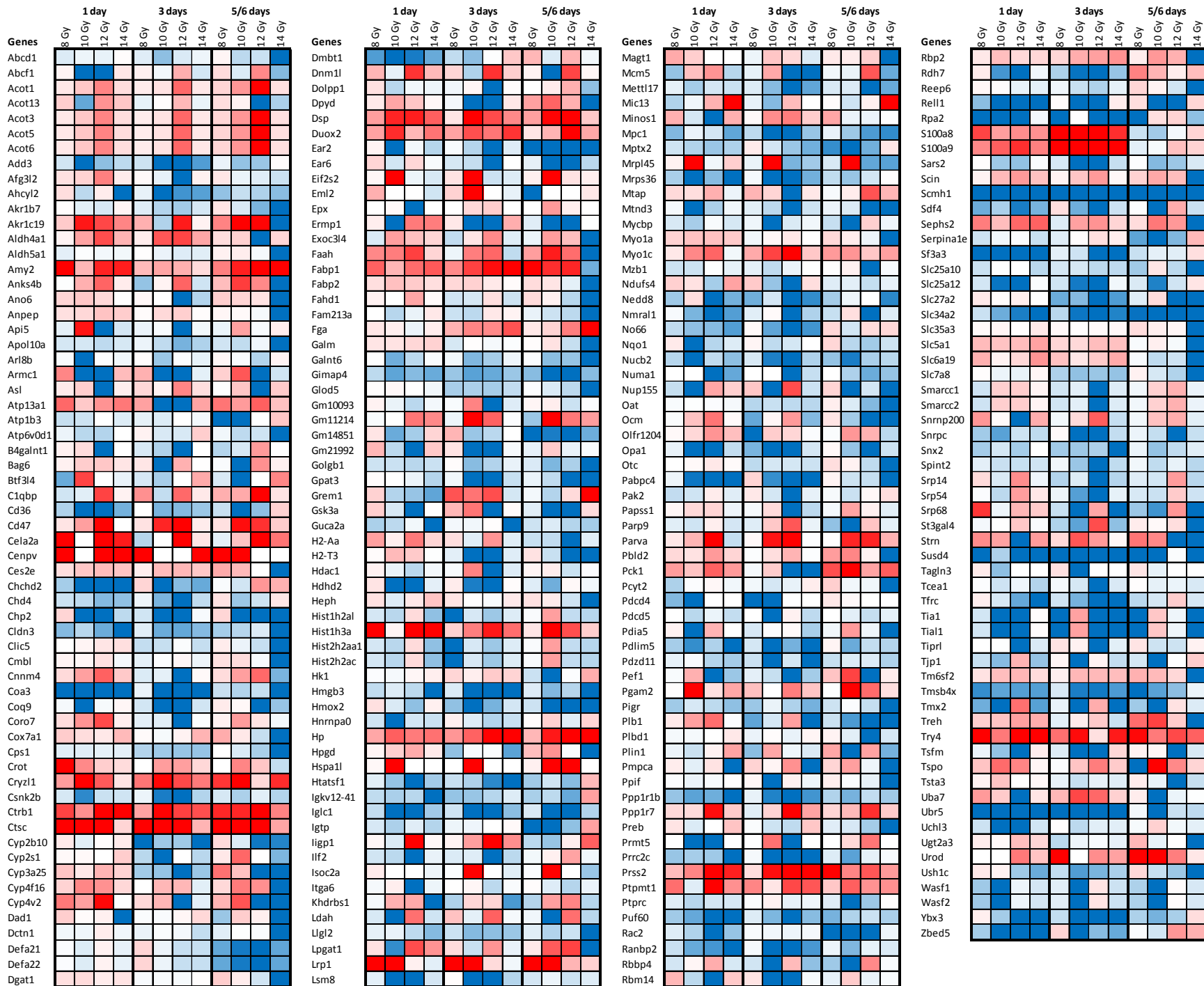
Supplemental Figure 8. Venn diagram of differential protein expression showing the effect of dose on day 1 after irradiation.

Supplemental Figure 9. Venn diagram of differential protein expression showing the effect of dose on day 3 after irradiation.

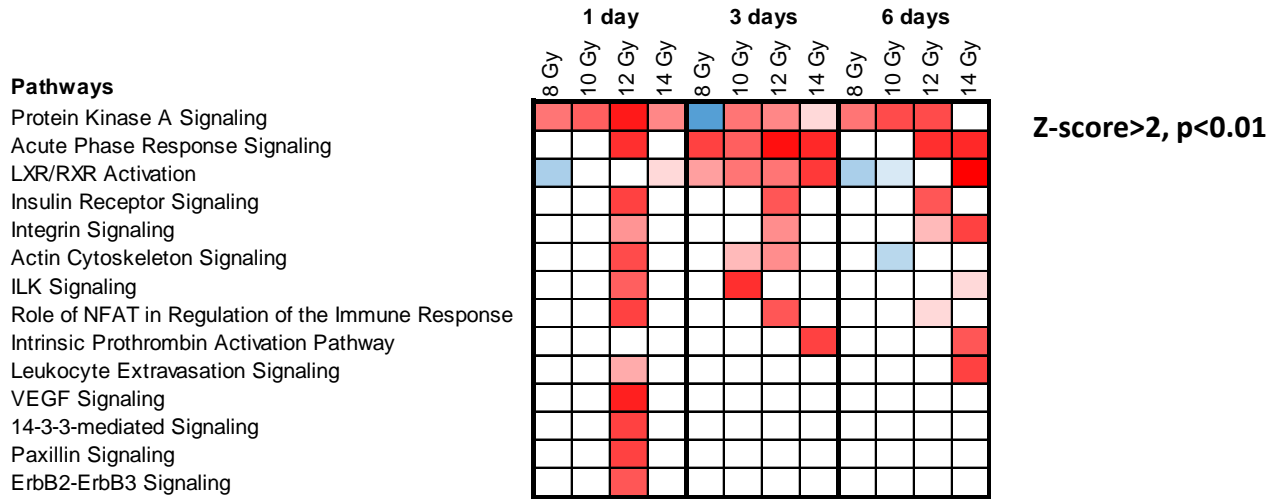
Supplemental Figure 10. Venn diagram of differential protein expression showing the effect of dose on day 6 after irradiation.

Supplemental Figure 11. Retinoid quantification in small intestine (jejunum) after TBI radiation. Radiation doses and times after radiation dose are notated. (a.) RA, (b.) ROL, (c.) RE. Retinoid levels were measured using n=5 per condition except for control (0) which is an average of n=15 un-irradiated control mice euthanized at day -6 (n=5), day 0 (n=5), or day 6 (n=5). The statistical analysis relied on the unpaired Student's *t*-test between groups and the Data is mean \pm standard deviation, * $p < 0.05$ using student's *t*-test between groups as compared to control (0). Data shown as a function of time after dose at a given radiation dose. Data shown as a function of dose on a given day after dose is shown in **Supplementary Fig. 12**. RA=retinoic acid, ROL = retinol, RE=total retinyl esters

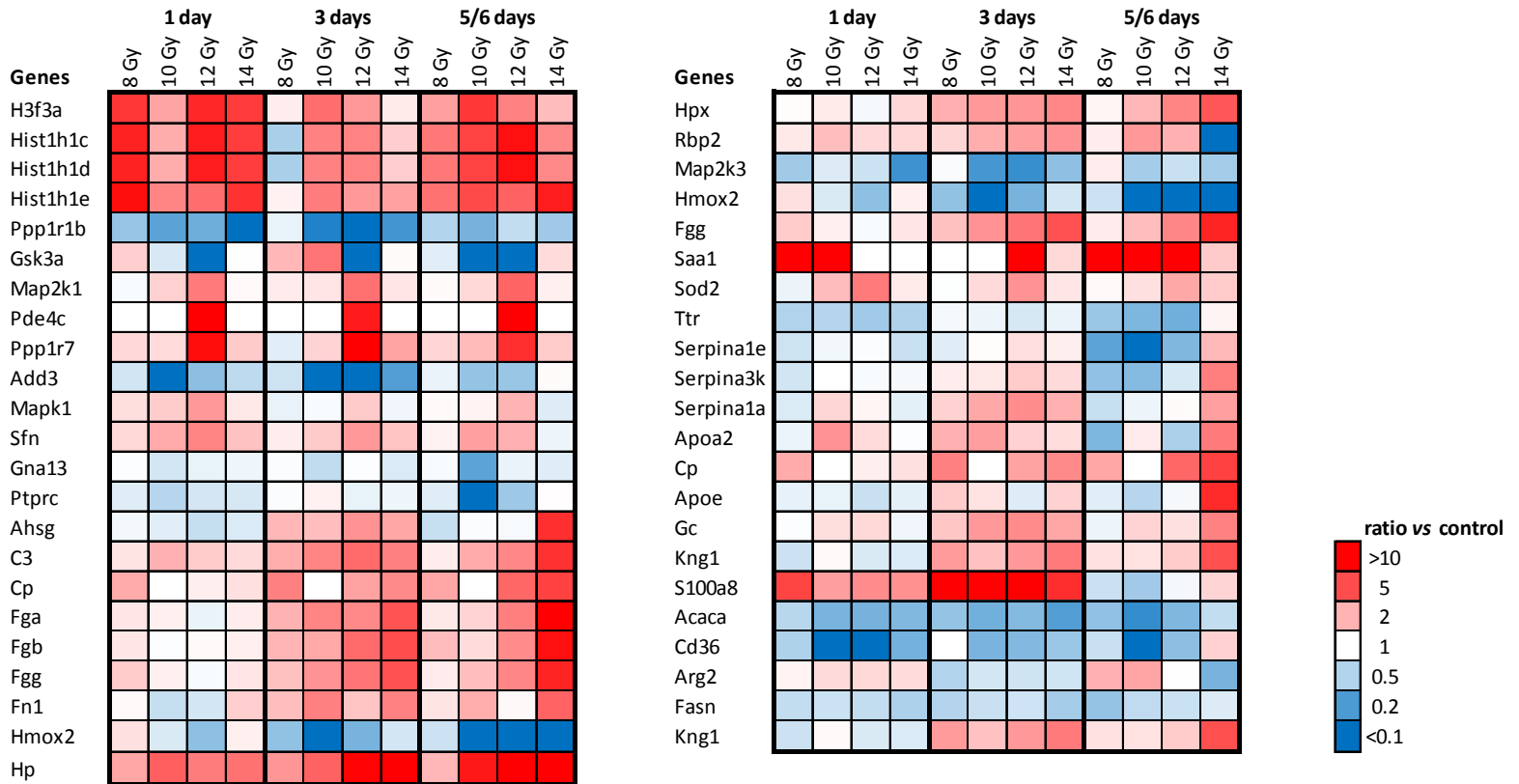
Supplemental Figure 12. Retinoid quantification in small intestine (jejunum) after TBI radiation. Radiation doses and times after radiation dose are notated. (a.) RA, (b.) ROL, (c.) RE. Retinoid levels were measured using n=5 per condition except for control (0) which is an average of n=15 un-irradiated control mice euthanized at day -6 (n=5), day 0 (n=5), or day 6 (n=5). The statistical analysis relied on the unpaired Student's *t*-test between groups and the Data is mean \pm standard deviation, * $p < 0.05$ using student's *t*-test between groups as compared to control (0). Data shown as a function of dose on a given day after dose. Data shown as a function of time after dose at a given radiation dose is shown in **Supplemental Fig. 11**. RA=retinoic acid, ROL = retinol, RE=total retinyl esters



a

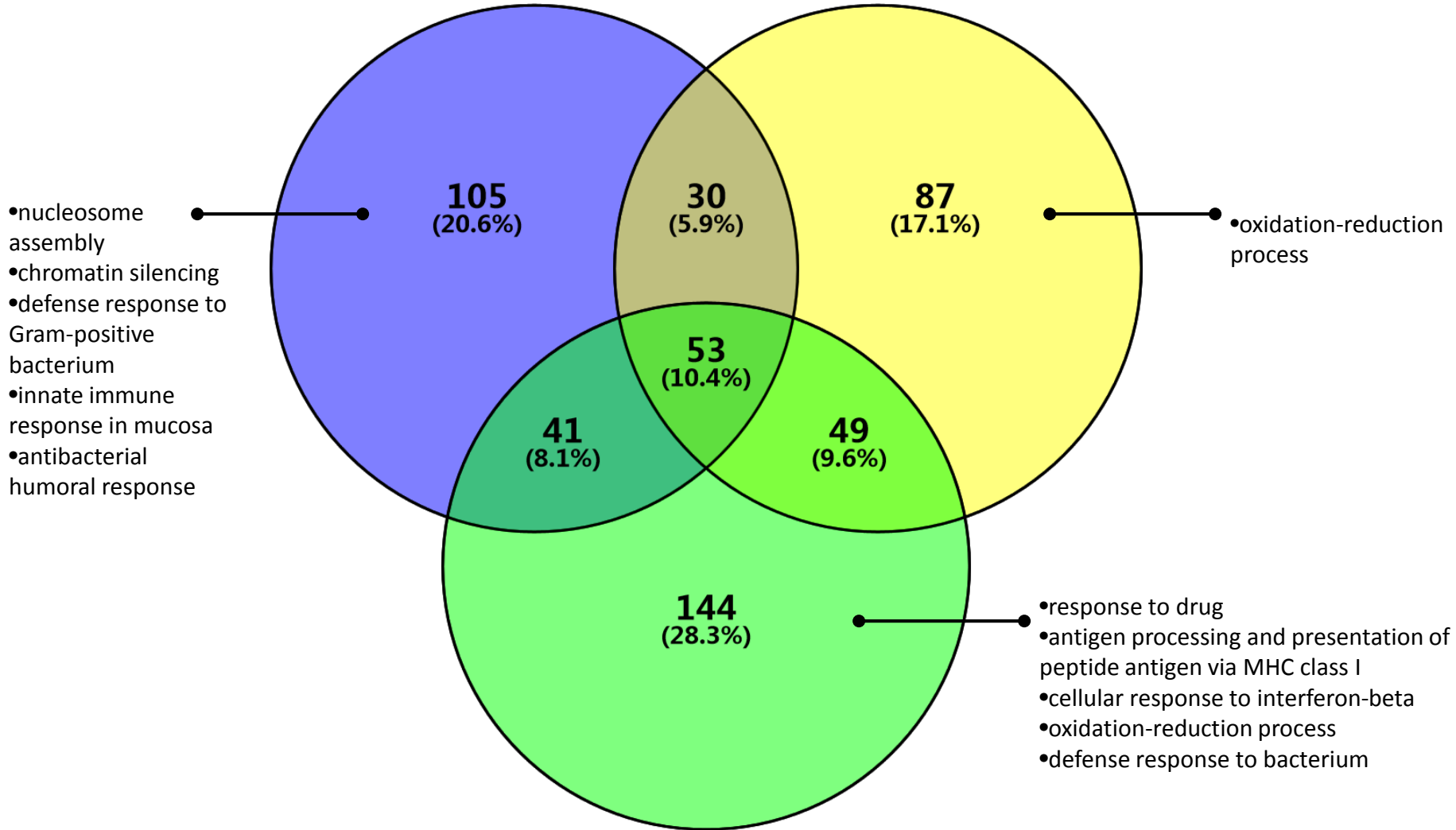


b



8 Gy 1 Day

8 Gy 3 Days

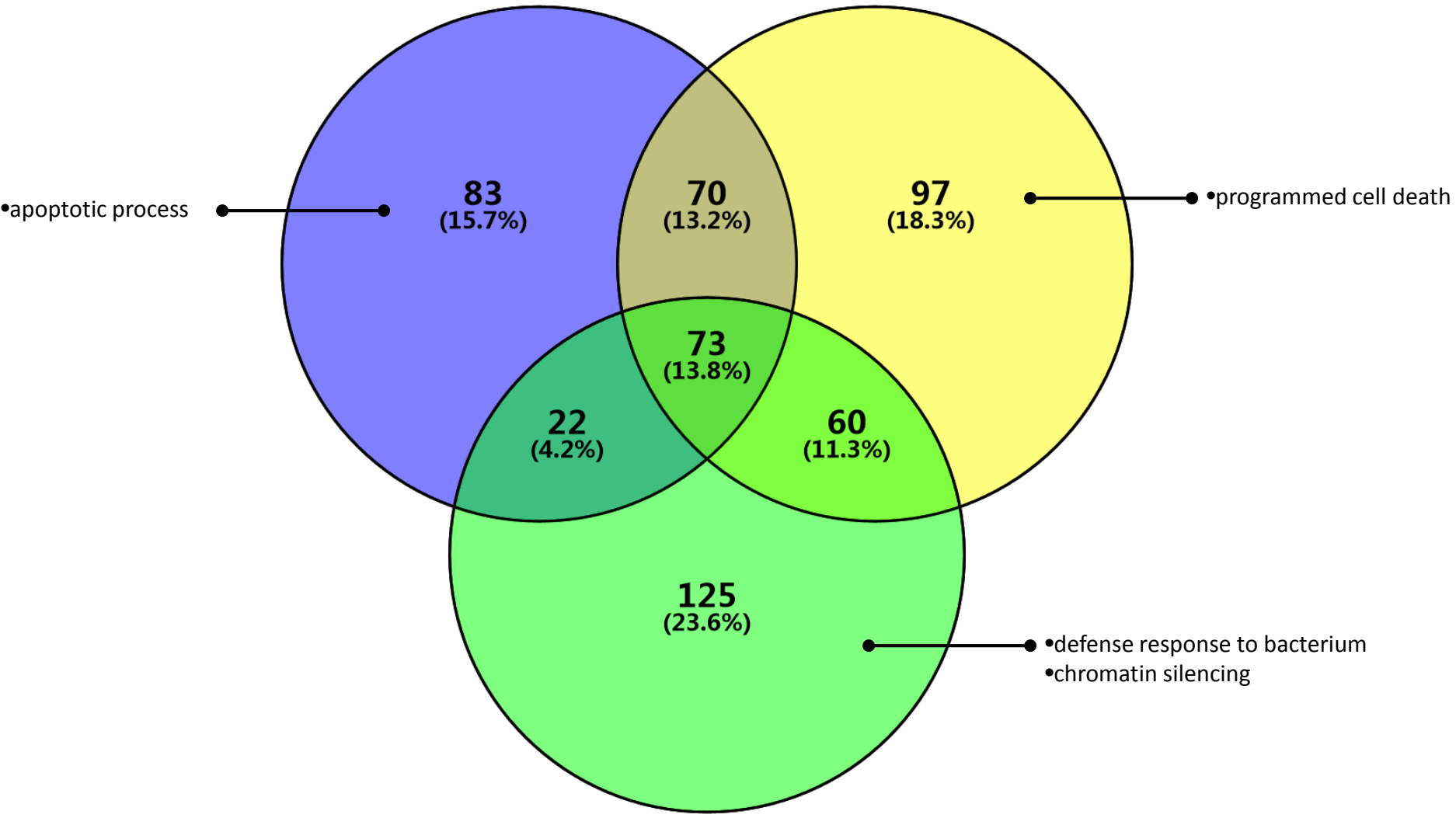


8 Gy 6 Days

10 Gy 1 Day

10 Gy 3 Days

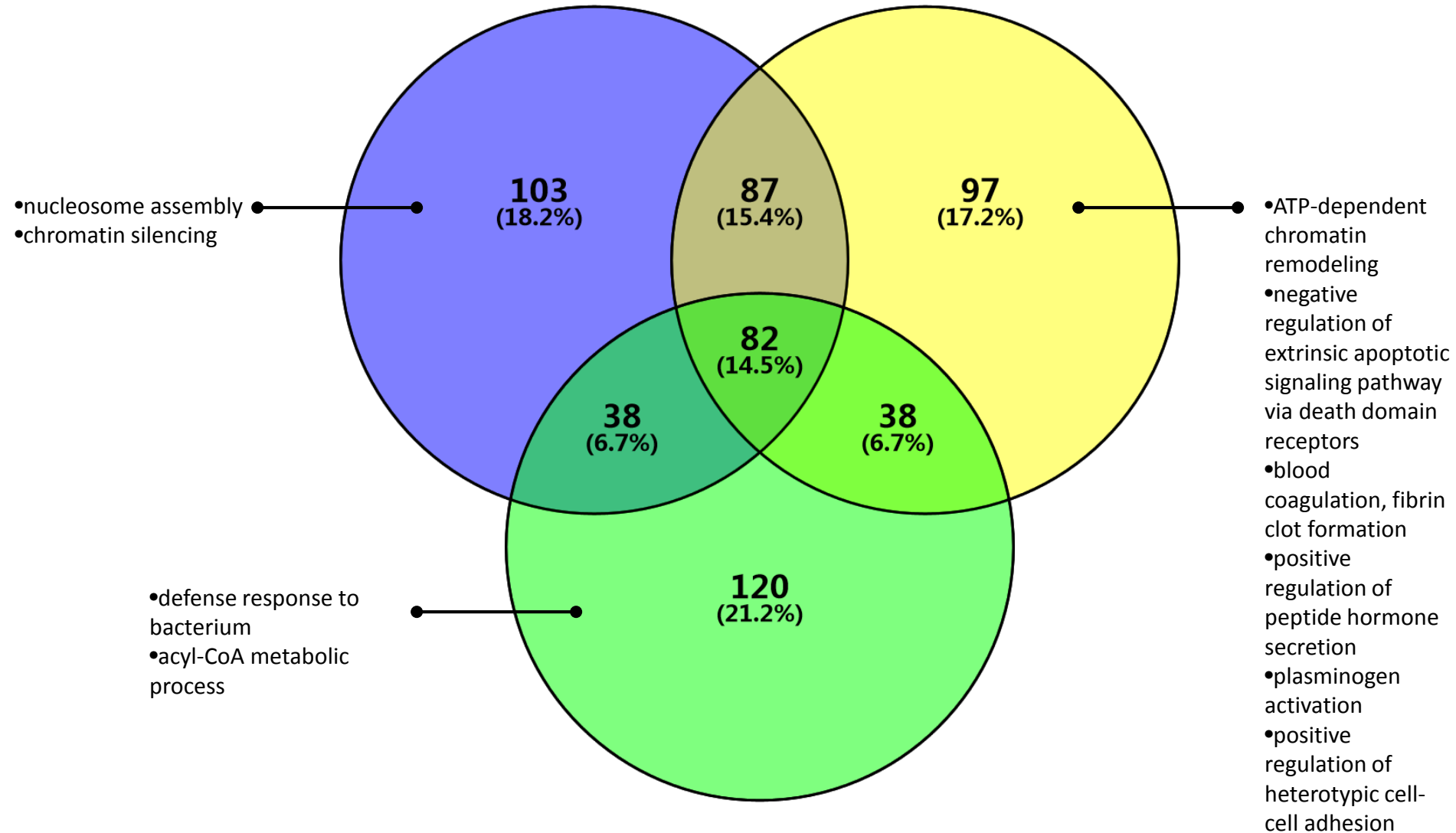
10 Gy 6 Days



12 Gy 1 Day

12 Gy 3 Days

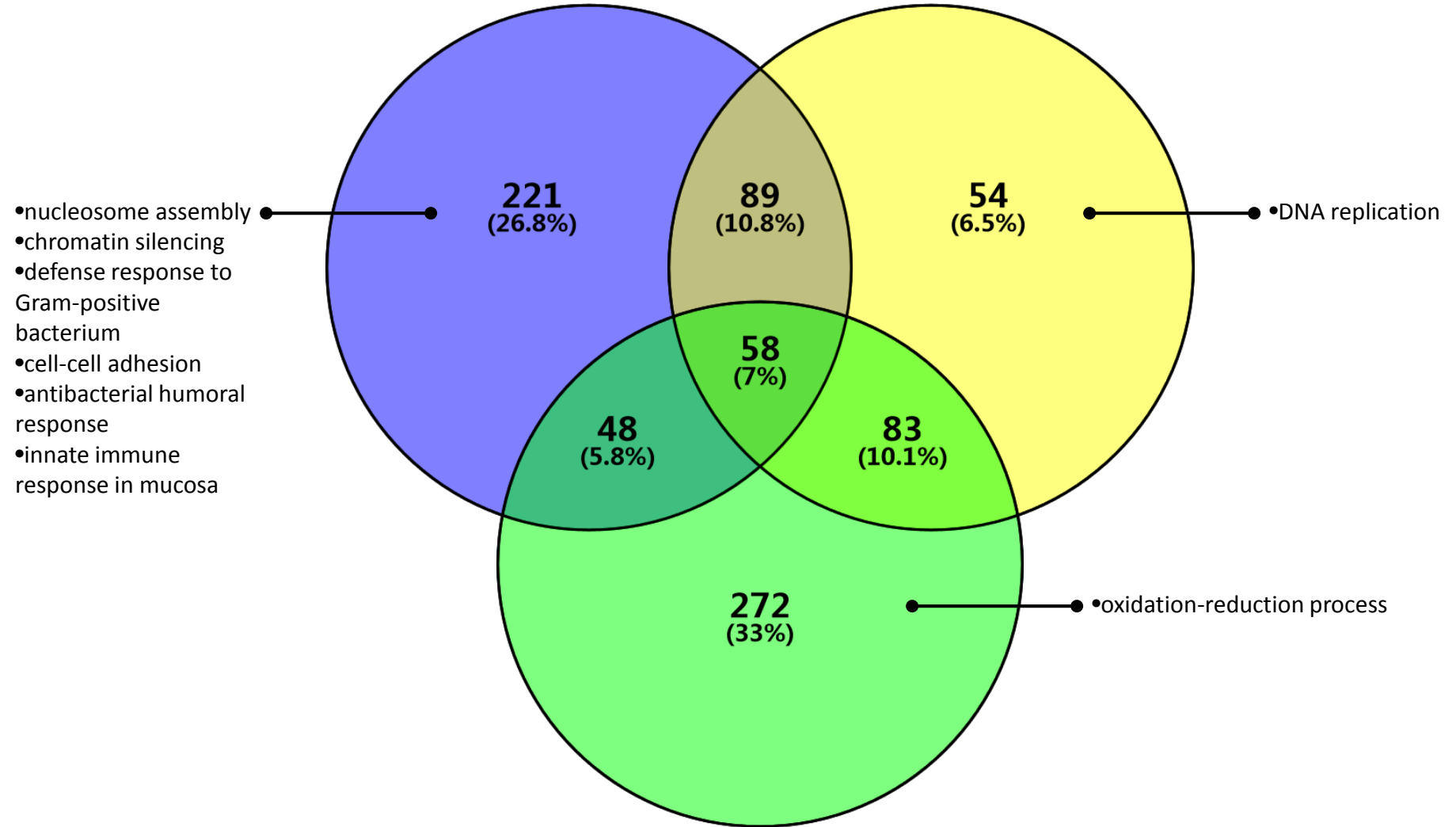
12 Gy 6 Days

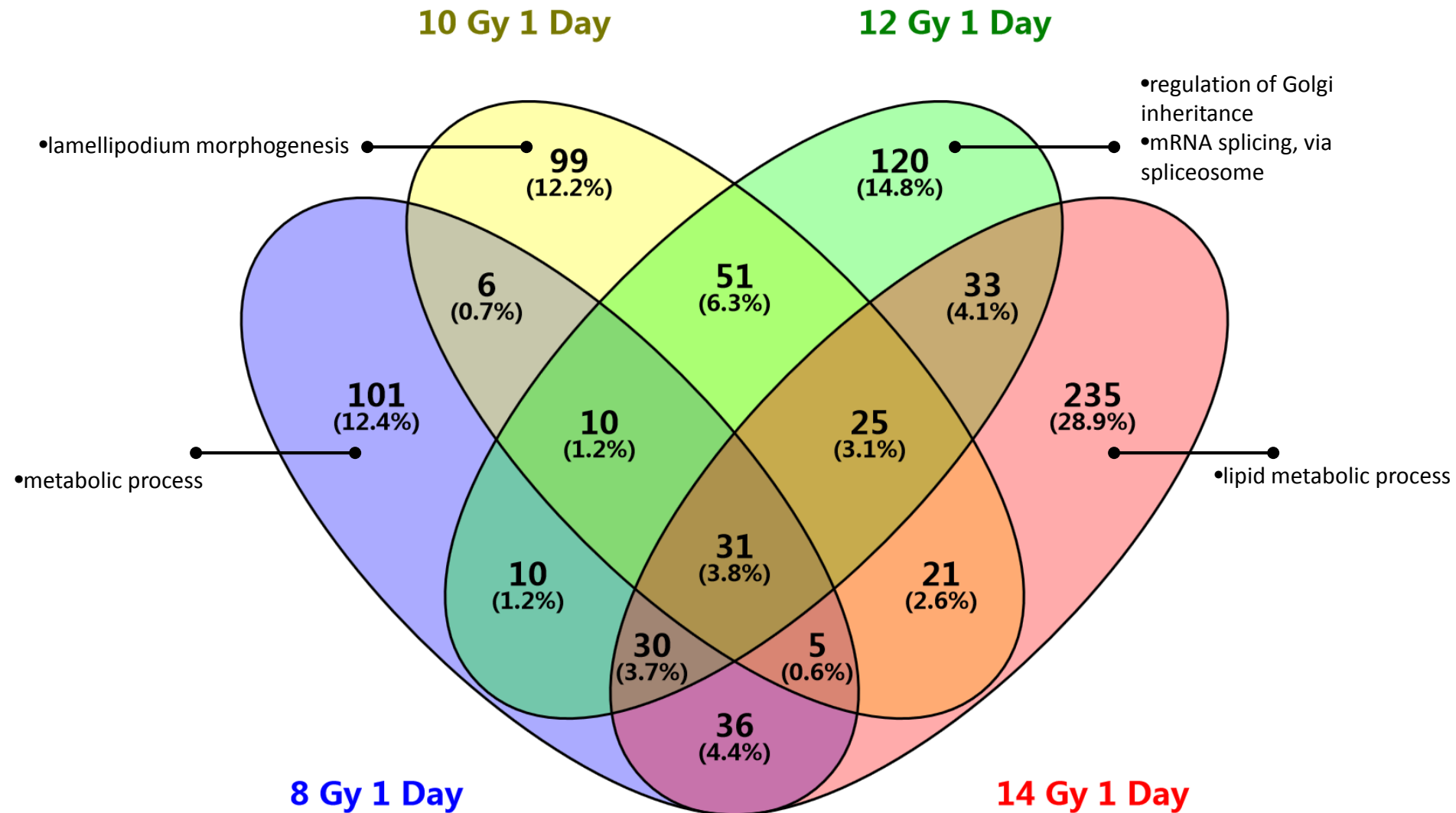


14 Gy 1 Day

14 Gy 3 Days

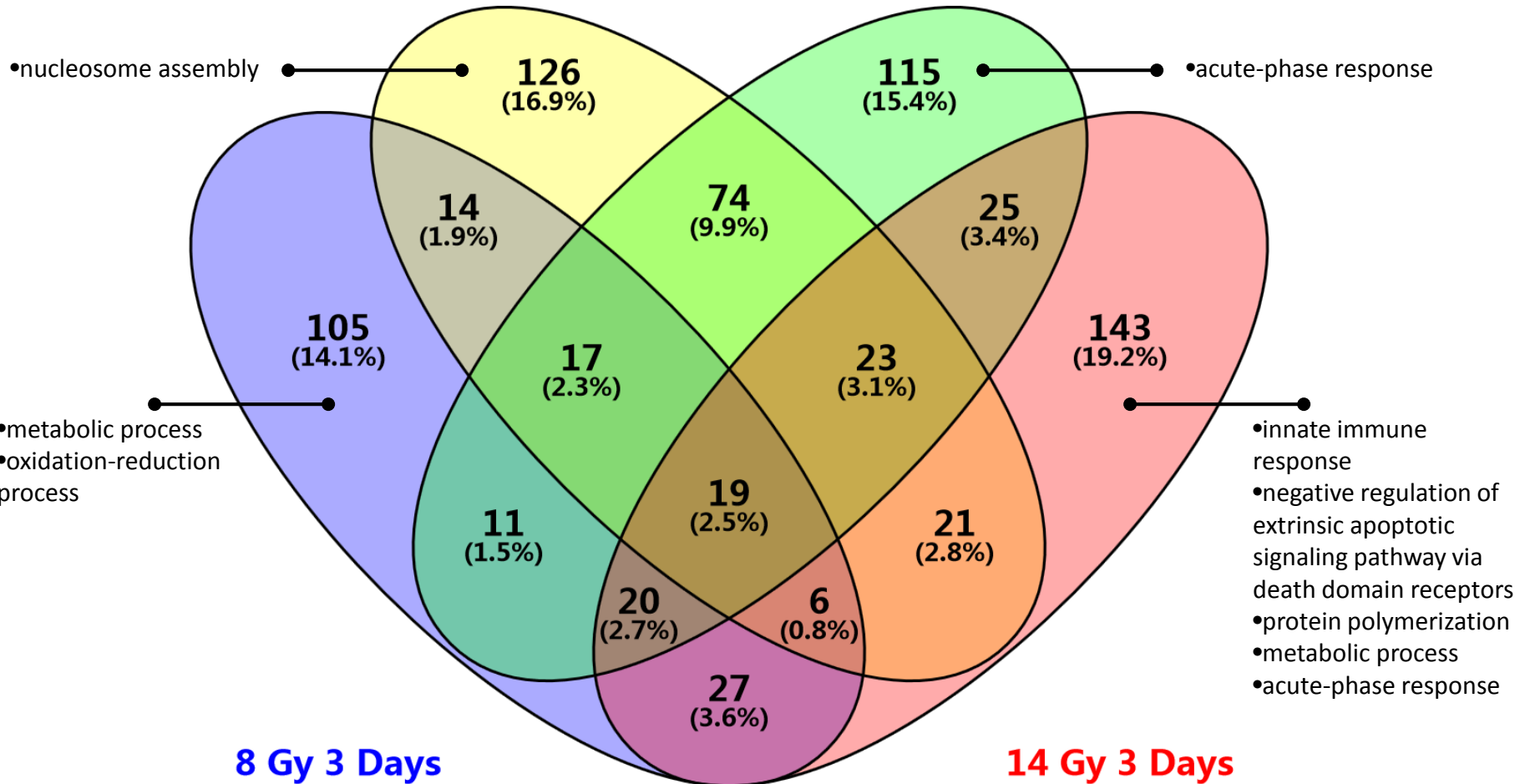
14 Gy 5 Days





10 Gy 3 Days

12 Gy 3 Days

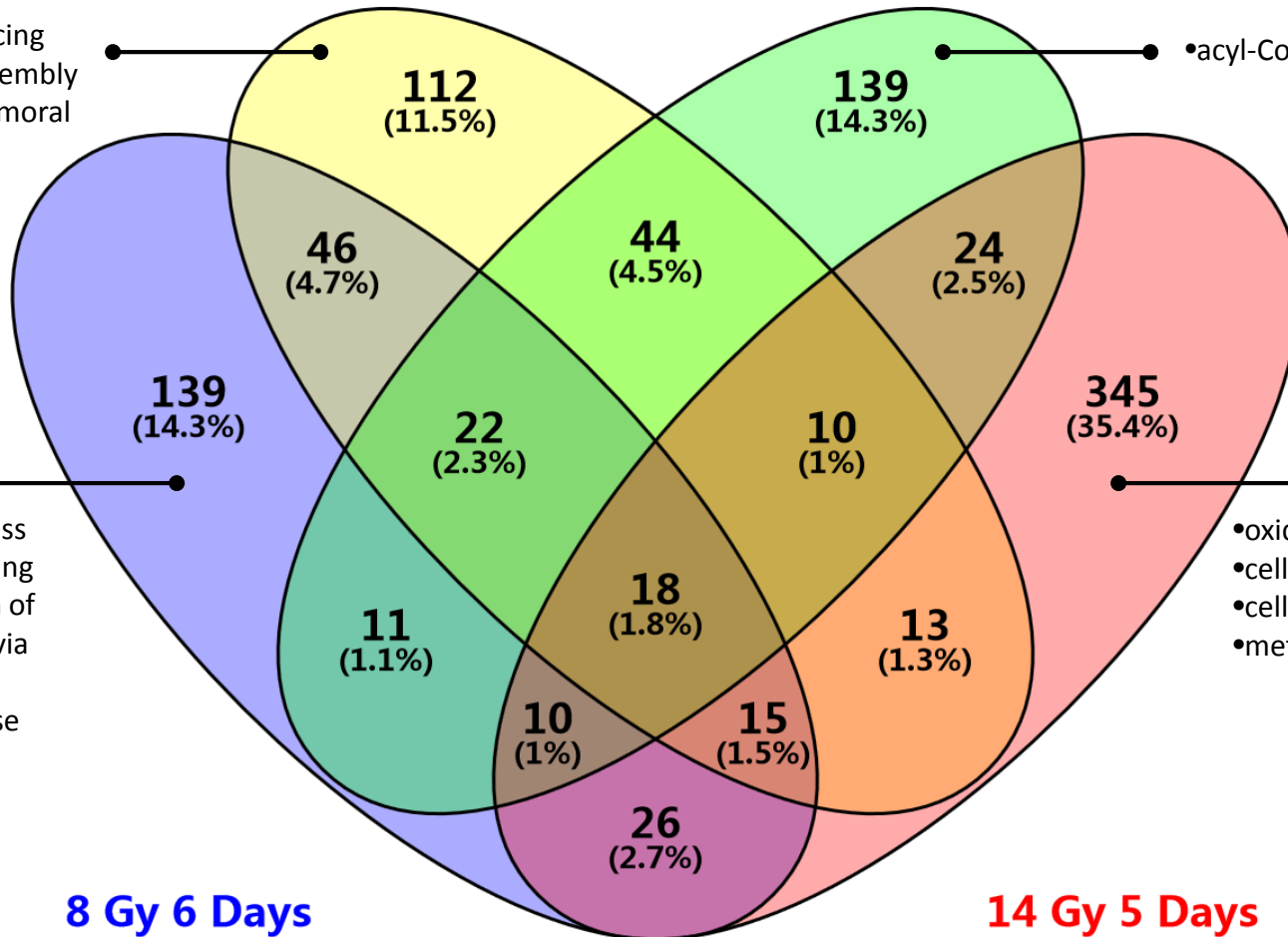


10 Gy 6 Days

12 Gy 6 Days

- chromatin silencing
- nucleosome assembly
- antibacterial humoral response

- acyl-CoA metabolic process



- metabolic process
- antigen processing and presentation of peptide antigen via MHC class I
- defense response

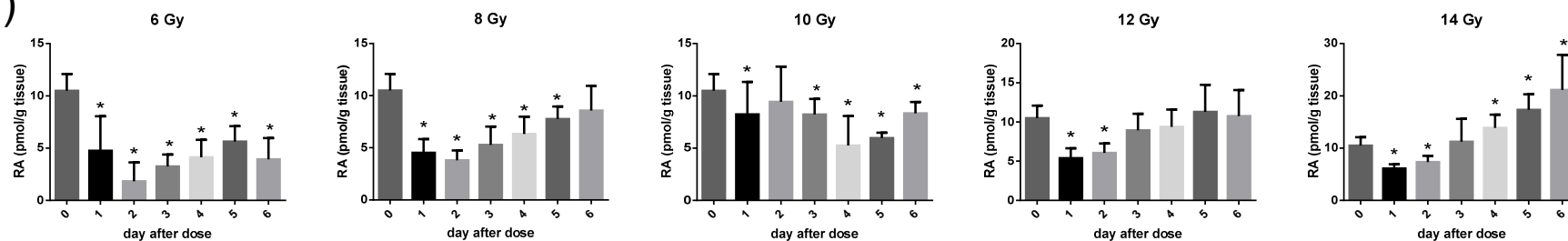
- oxidation-reduction process
- cell-matrix adhesion
- cell adhesion
- metabolic process

8 Gy 6 Days

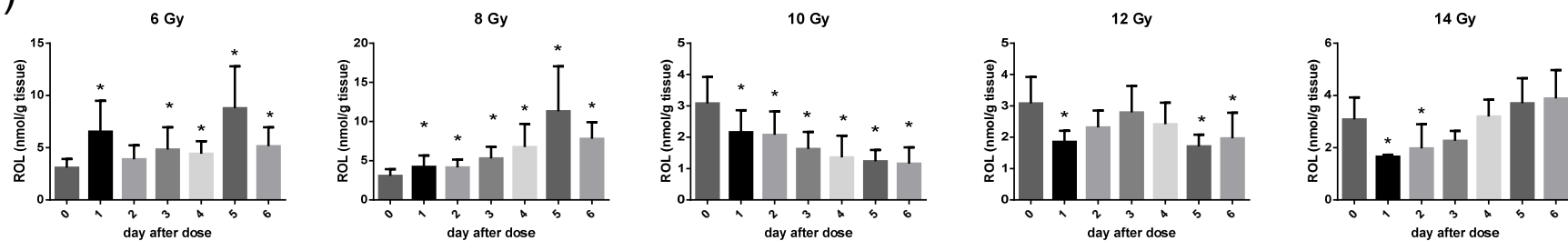
14 Gy 5 Days

SF11

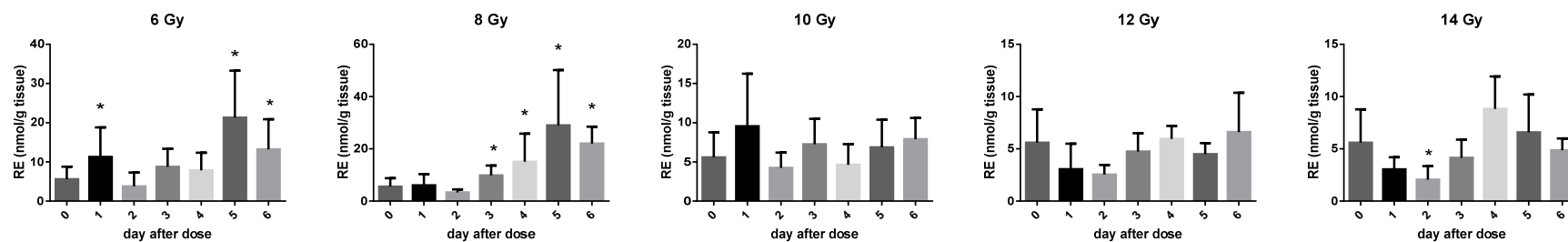
a.)



b.)

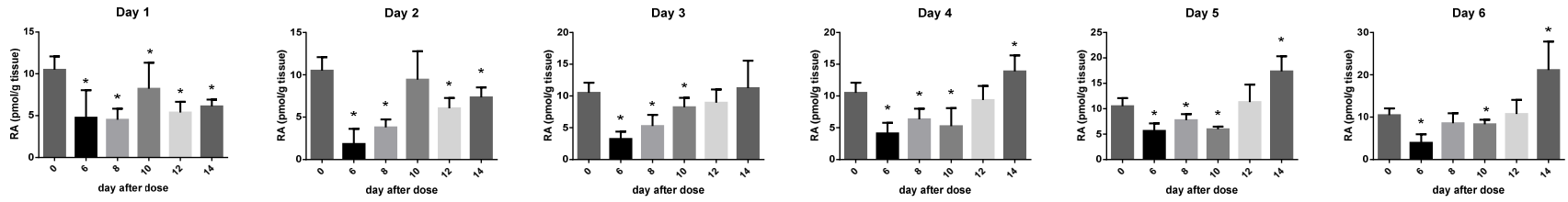


c.)

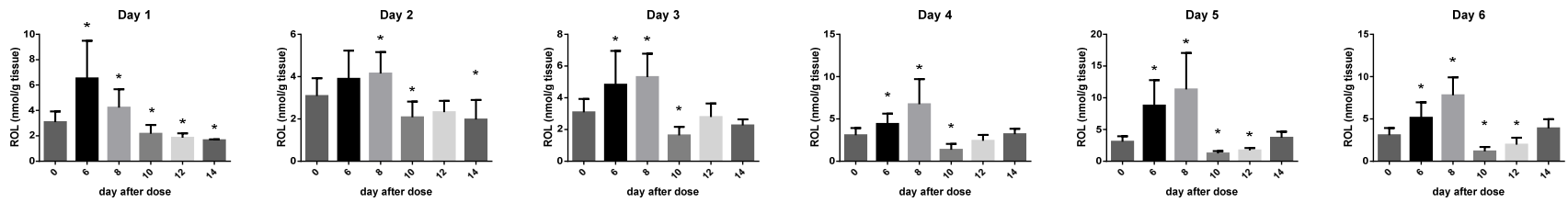


SF12

a.)



b.)



c.)

