

Supplementary Material

NFAT1 and NFAT2 differentially regulate CTL differentiation upon acute viral infection

Tianhao Xu¹, Ashleigh Keller¹, Gustavo J. Martinez^{1,*}

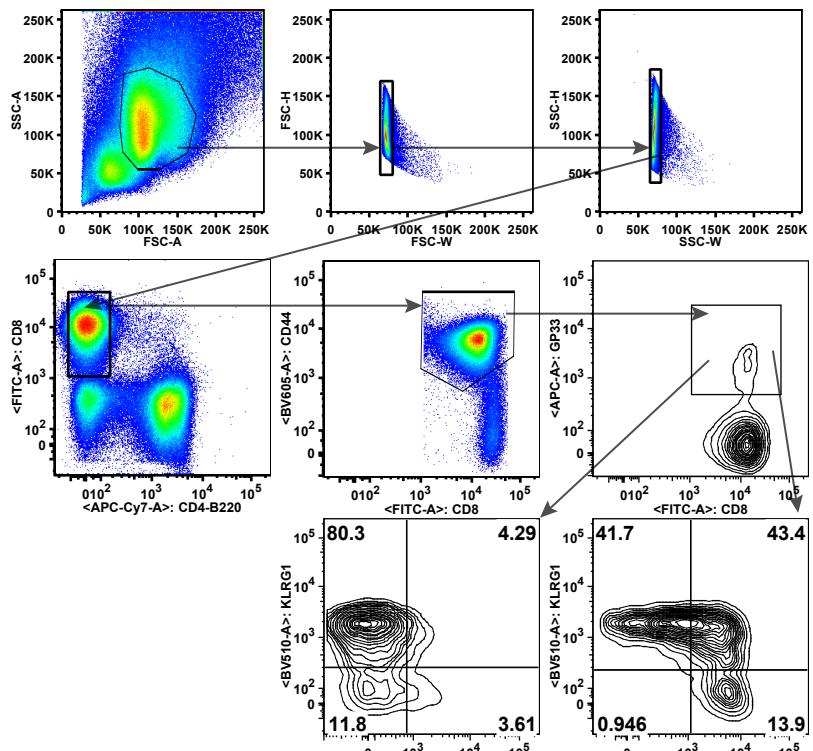
¹Department of Microbiology and Immunology, Chicago Medical School, Rosalind Franklin University, 3333 Green Bay Road, North Chicago, IL 60064, USA.

*Correspondence:

Dr. Gustavo Martinez

Gustavo.martinez@rosalindfranklin.edu

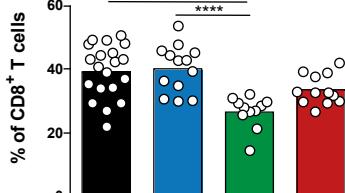
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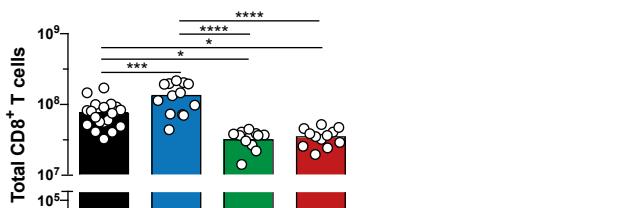
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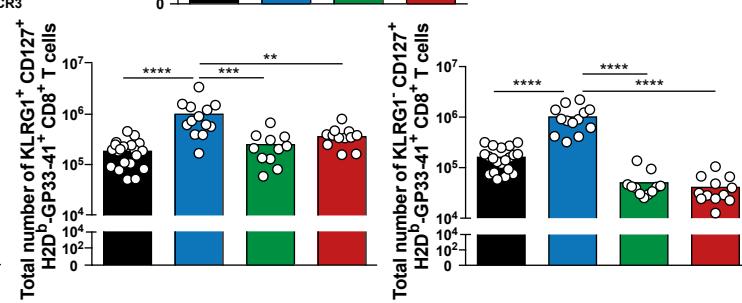
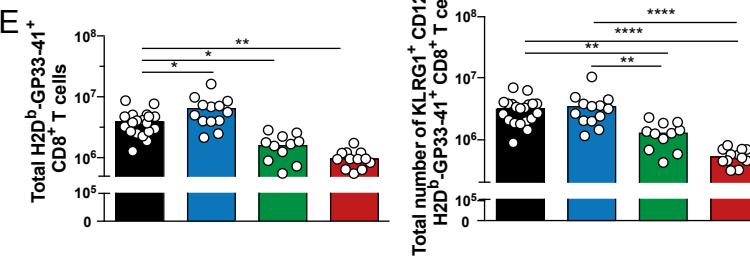
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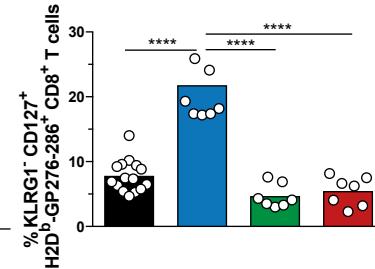
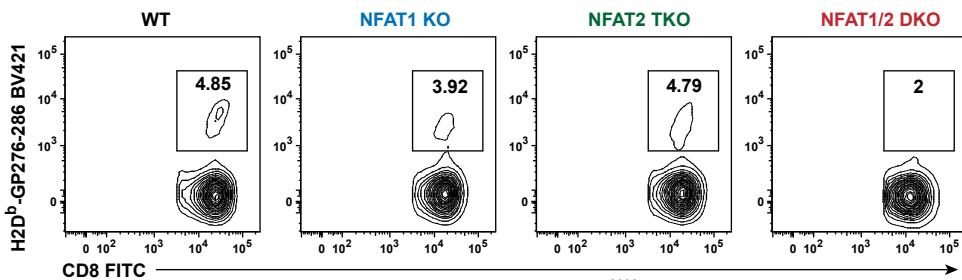
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E



F



G

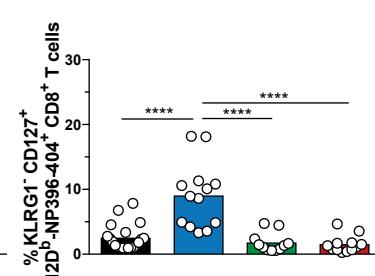
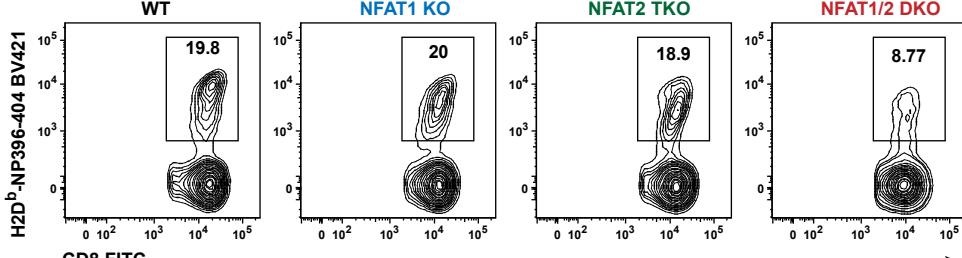
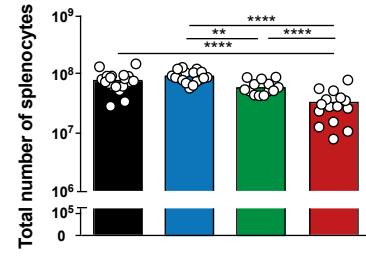
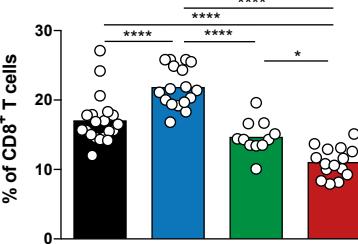


Figure S1. Defective CTL differentiation in different antigen-specific cells upon NFATs deficiency on day 8 post LCMV Arm infection. (A) General gating strategy used in FACS analysis. (B-D) Total splenocytes, percentage of CD8⁺ T cells, and total number of CD8⁺ T cells combined form three biological replicates. (E) Absolute number of antigen-specific H2D^b-gp33-41⁺ CD8⁺ T cells and each gated KLRG1 and CD127 sub-populations form three individual experiments. (F) Top panel shows a representative contour plot displaying the frequency of H2D^b-GP276-286⁺ CD8⁺ T cells within the different mice; bottom panel shows the total cell number of H2D^b-GP276-286⁺ CD8⁺ T cells per spleen and frequency of KLRG1 and CD127 sub-populations within these antigen-specific cells. (G) Top panel shows a representative contour plot displaying the frequency H2D^b-NP396-404⁺ CD8⁺ T cells within the different mice; bottom panel shows the total cell number of H2D^b-NP396-404⁺ CD8⁺ T cells per spleen and frequency of KLRG1 and CD127 sub-populations within these antigen-specific cells. Statistical analysis was done using non-paired One-Way ANOVA followed by Tukey's multiple comparisons. *: p ≤ 0.05, **: p ≤ 0.01, ***: p ≤ 0.001, ****: p ≤ 0.0001.

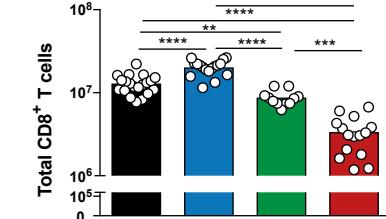
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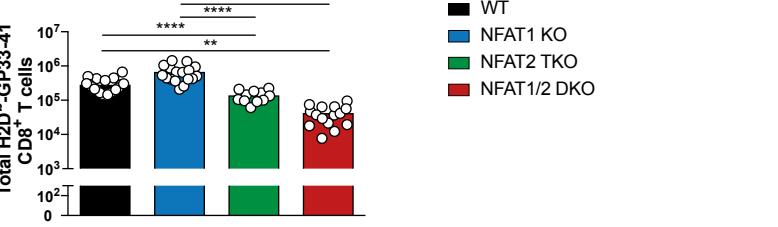
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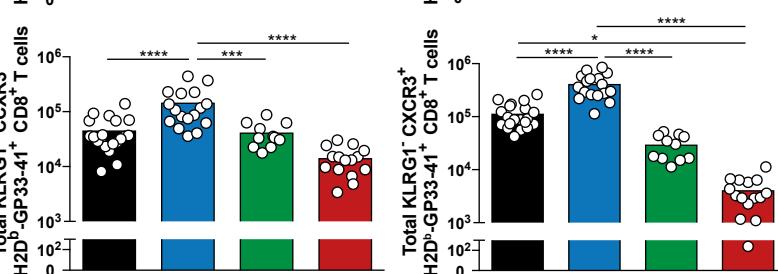
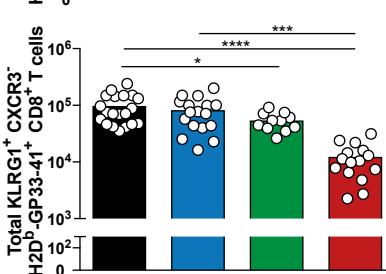
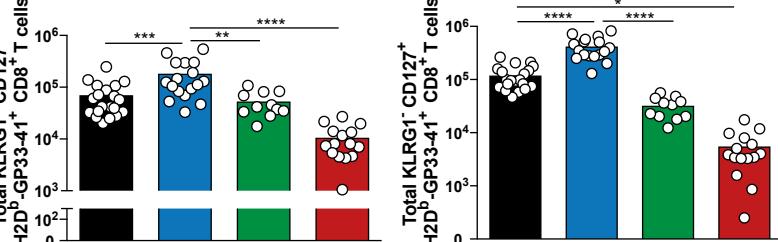
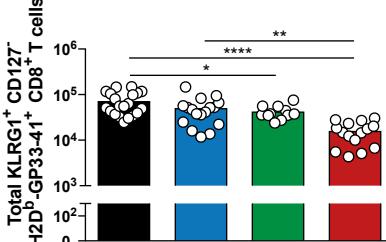
C



D



E



■ WT
■ NFAT1 KO
■ NFAT2 TKO
■ NFAT1/2 DKO

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*

Figure S2. Defective CD8⁺ T cell differentiation upon deficiency of NFAT family members is maintained during the memory phase. Total splenocytes (**A**), percentage and total numbers of CD8⁺ T cells (**B-C**), total number of tetramer H2D^b-GP33-41⁺ CTLs (**D**), each gated KLRG1 and CD127 or CXCR3 sub-population within these antigen-specific cells from three individual experiments (**E**). Statistical analysis was done using non-paired One-Way ANOVA followed by Tukey's multiple comparisons. *: p ≤ 0.05, **: p ≤ 0.01, ***: p ≤ 0.001, ****: p ≤ 0.0001.

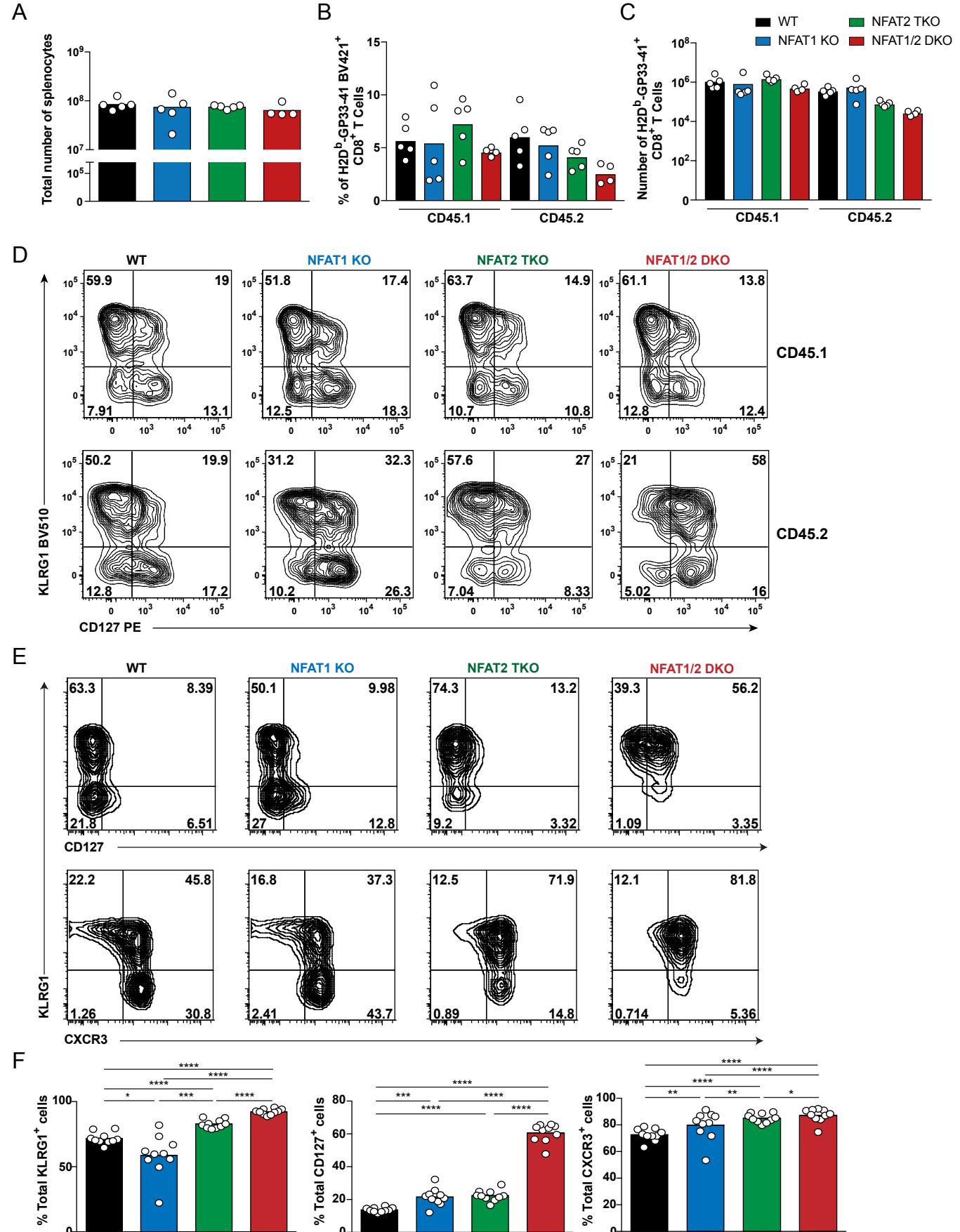


Figure S3. NFAT members regulate CD8⁺ T cell differentiation in a cell-intrinsic manner.

(A) Spleen cell counts. (B-C) Frequency and total number of H2D^b-GP33-41⁺ CD8⁺ T cells. (D) Representative contour plot displaying the expression of KLRG1 and CD127 in H2D^b-GP33-41⁺ recipient (CD45.1) or donor (CD45.2) CD8⁺ T cells from mix bone marrow chimera. (E) Representative plot showing the expression of KLRG1, CD127 and CXCR3 on adoptively-transferred P14 CD8⁺ T cells. (F) The combined frequency of total KLRG1⁺, CD127⁺ or CXCR3⁺ adoptively-transferred P14 CD8⁺ T cells from two independent experiments. Non-paired One-Way ANOVA followed by Tukey's multiple comparisons were used in (A-D), and student's T test was used in (F). *: p ≤ 0.05, **: p ≤ 0.01, ***: p ≤ 0.001, ****: p ≤ 0.0001.

Refseq Genes

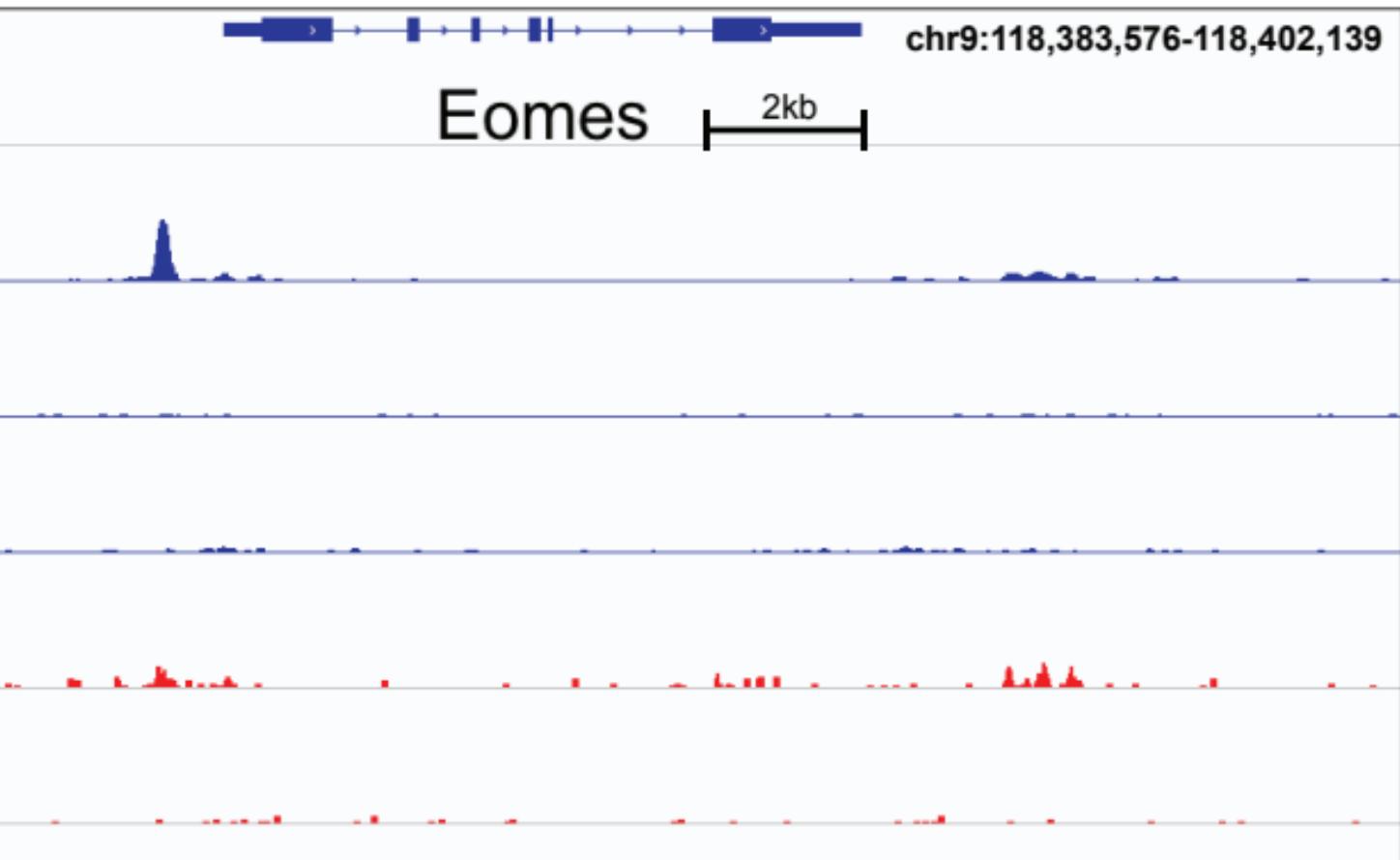
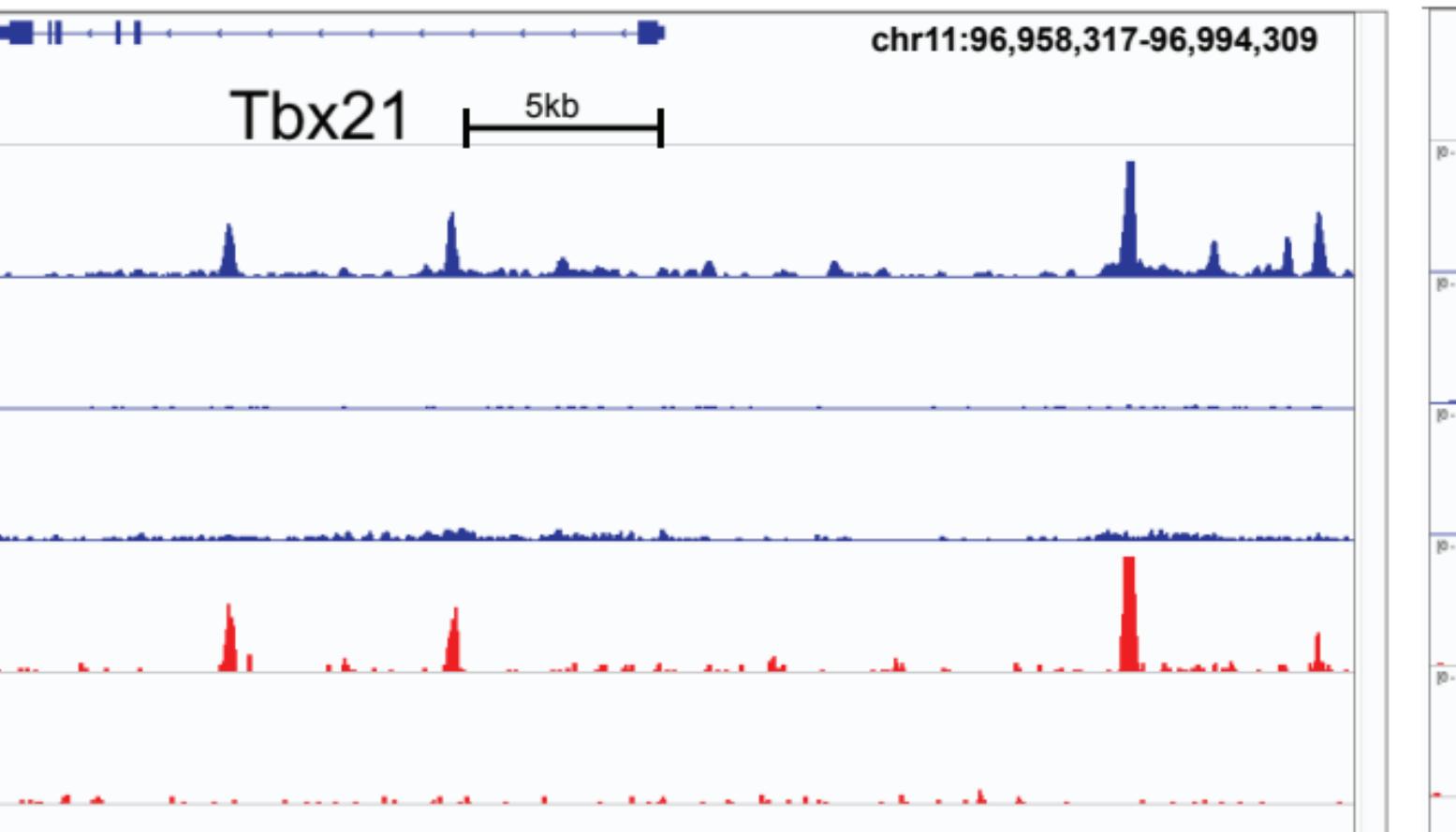


Figure S4. NFAT1 and NFAT2 bind to *Tbx21* and *Eomes* loci. The binding of NFAT1 and biotin Tagged NFAT2 on *Tbx21* and *Eomes* loci is depicted with igv_2.4.13 using published ChIP-seq data GSE64409 and GSE98726.

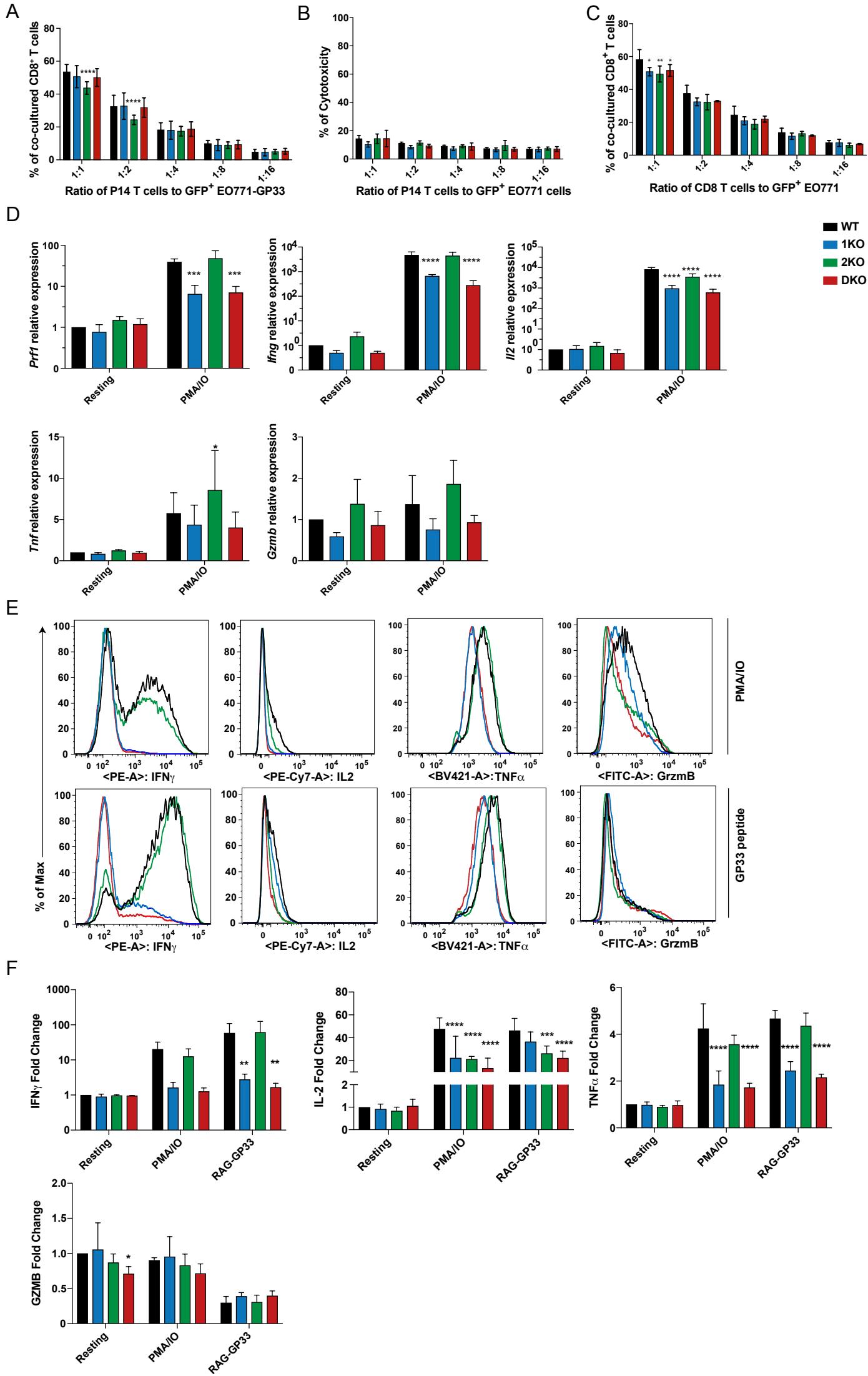


Figure S5. Defective cytokine production and cytotoxicity upon NFAT deficiency. (A) The mean percentage of CD8⁺ T cells incubated at different ratios with EO771 cells expressing GP33-41 from three independent experiments. (B-C) Mean cytotoxicity (B) or percentage of P14 CD8⁺ T cells (C) when incubated at different ratios with parental EO771 cells without GP33-41 antigen. (C) Cytotoxicity was measured by the frequency of GFP⁺ cells within the CD8⁺ population from three independent experiments. (D) Expression of effector genes in NFAT-deficient CTLs. P14 CD8⁺ T cells used for cytotoxicity assay were restimulated with PMA and ionomycin for 4hrs, after which RNA was extracted and expression of genes determined by real time RT-qPCR. *Rpl32* was used as control and the relative mRNA expression of different groups was determined by normalizing to WT resting condition (set as 1). (E-F) The representative and combined expression of IFN- γ , TNF- α , IL-2 and Granzyme B upon restimulation of *in vitro* activated P14 CD8⁺ T cells from the indicated genotypes. Cells were restimulated with PMA (10nM) and Ionomycin (1 μ M) or Rag^{-/-} splenocytes primed with GP33-41 peptide (0.2ug/ml), both in the presence of Brefeldin A, and intracellular staining performed after 4h of stimulation. The mean of expression fold change was calculated by normalizing to WT resting condition from three independent experiments. Two-Way ANOVA followed by Dunnett comparisons were performed to analysis within group statistical differences. *: p \leq 0.05, **: p \leq 0.01, ***: p \leq 0.001, ****: p \leq 0.0001.