1	Supplementary Information for
2	
3	Pareto Optimality Explanation of the Glycolytic Alternatives in Nature
4	
5	Chiam Yu Ng, Lin Wang, Anupam Chowdhury, and Costas D. Maranas
6	
7	Corresponding author: Costas D. Maranas
8	Email: costas@psu.edu
9	
10	
11	This PDF file includes:
12	
13	Supplementary Information Text
14	Figs. S1 to S8
15	Tables S1 to S2
16	References for SI reference citations
17	
18	Other supplementary materials for this manuscript include the following:
19	
20	Supplementary Data Files S1 to S6

## 21 Supplementary Information Text

# 22 Integer cut constraints and the runtime reduction of optStoic.

Integer cut constraints are introduced in the optStoic formulation to exhaustively identify alternate optimal pathways that satisfy the design equation. Herein, we define k as the number of iteration for running optStoic algorithm ( $k \in \{1, 2, ..., \kappa\}$ ).

26

The formulation of optStoic in the method section (see main text) was reformulated as a combination of linear relations by introducing two non-negative real number (or integer) variables for each  $v_i$  as followed:

$$v_{j} = v_{j}^{f} - v_{j}^{r}, \quad \forall j \in J$$
  
where  $v_{j} \in \mathbb{Z}$ ,  $v_{j}^{f} \in \mathbb{Z}_{\geq 0}$  and  $v_{j}^{r} \in \mathbb{Z}_{\geq 0}$   
or  $(v_{j} \in \mathbb{R}, v_{j}^{f} \in \mathbb{R}_{\geq 0} \text{ and } v_{j}^{r} \in \mathbb{R}_{\geq 0})$   
 $|v_{j}| = v_{j}^{f} + v_{j}^{r}$ 

30

31 To this end, binary variables  $y_j^f$  and  $y_j^r$  are defined as followed:

 $y_j^f = \begin{cases} 1, \text{ if reaction } j \text{ carries non-zero flux in the forward direction } (v_j^f > 0) \\ 0, \text{ otherwise} \end{cases}$ (1)  $y_j^r = \begin{cases} 1, \text{ if reaction } j \text{ carries non-zero flux in the reverse direction } (v_j^r > 0) \\ 0, \text{ otherwise} \end{cases}$ (2)

32 Likewise,  $y_j^{f^k}$  and  $y_j^{r^k}$  are binary variables associated with the solution from the *k*-th 33 iteration. At iteration  $k = \kappa$ , the following constraints are added to the modified optStoic 34 formulation:

$$\sum_{j \in J \mid y_j^{f^k} + y_j^{r^k} = 1} (1 - y_j^f - y_j^r) + \sum_{j \in J \mid y_j^{f^k} + y_j^{r^k} = 0} (y_j^f + y_j^r) \ge 1 \qquad k = 1, 2, \dots, \kappa - 1 \quad (3)$$

$$y_j^f + y_j^r \le 1 \tag{4}$$

$$y_j^f \varepsilon \le v_j^f \le y_j^f M \tag{5}$$

$$y_i^r \varepsilon \le v_i^r \le y_i^r M \tag{6}$$

35 Constraint 3 is the integer cut constraint that ensures that at least one of reaction *j* that was identified in the previous iteration k is inactive in the current iteration. Constraint 4 36 37 enforces that only one of the binary variables (corresponding to the flux directions) for 38 each reaction *j* is active. Finally, constraints 5 and 6 restrict the flux (in forward or reverse 39 direction) to be strictly positive whenever the corresponding binary variable is active. The 40 parameter  $\varepsilon$  is a user-defined small positive real number. The MILP problems were solved 41 using the CPLEX v.12.6.1 solver accessed through the GAMS (v24.4.1) modeling system 42 and Gurobi Optimizer v6.5.1 using Python 2.7.

43

The runtime of the modified optStoic algorithm depends on the size of the search space(i.e. database size). Therefore, blocked reactions (i.e., reactions incapable of carrying flux)

were identified first upon imposing the bounds on exchange fluxes (see constraint 4 in main
text) and excluded from the search space before running the algorithm. We also observed
that the runtime of optStoic significantly increases when more integer cuts are added. This

48 in at the function of option significantly increases when more integer cuts are added. This
 49 is caused by a large number of integer variables introduced in the second term of constraint
 50 2 to a bit of the second term of constraint

- 50 3 at each iteration k:
- 51

$$\sum_{i \in J \mid y_j^{f^k} + y_j^{r^k} \neq 1} (y_j^f + y_j^r)$$

 $\sum_{i \in I \setminus I_{exchange}} |v_j| = z^*$ 

52 Upon removal of the blocked reactions, there are still over 3,000 reactions exist in the set 53  $\{j \in J | y_j^{f^k} + y_j^{r^k} = 0\}$ . To solve this issue, we imposed an additional constraint on the 54 objective function as followed:

55

56

57

and used the following integer cut to prevent the same pathway from being identified for the same objective value  $z^*$ .

60 
$$\sum_{j \in J \mid y_j^{f^k} + y_j^{r^k} = 1} (1 - y_j^f - y_j^r) \ge 1, \quad \forall k = 1, 2, 3, ...$$

61 We run the modified optStoic algorithm in parallel for each fixed objective value  $z^*$  to 62 further reduce the total runtime.

# 63 Assessing the thermodynamic feasibility of a pathway.

64 The thermodynamic feasibility of each pathway under physiological concentration ranges65 are assessed using the max-min driving force (MDF) formulation (1).

66 Step 1: The  $\Delta_r G'^{\circ}$  for each reaction involved in a pathway  $(j \in J_{path})$  is estimated using 67 the Component Contribution method (2) at pH 7, 25°C and ionic strength of 0.1 M (3, 4).

68 Step 2: The MDF problem is solved for each pathway, which minimizes the maximum

69  $\Delta_r G'_j$  of a pathway by optimizing over the concentrations of all metabolites in the pathway. 70 The optimization formulation is given by:

$$\max_{c_i} \min_j \{-\Delta_r G_j'\} \qquad (MDF)$$
(7)

subject to 
$$\Delta_r G'_j = \Delta_r G'^\circ + RT \sum_{\substack{i \in I_{rest}}} S^T_{ij} \ln c_i, \quad \forall j \in J_{path}$$
 (8)

$$\ln c_i^{min} \le \ln c_i \le \ln c_i^{max}, \quad \forall i \in I_{path}$$
(9)

$$\ln r_p^{\min} \le \ln r_p \le \ln r_p^{\max} , \quad \forall \, p \in \mathbf{P}$$
(10)

71

where  $I_{path}$  is the set of all metabolites and  $J_{path}$  is the set of all reactions in a pathway,  $c_i$ is the concentration of metabolite *i*, *R* is the gas constant, *T* is the temperature,  $r_p$  is the concentration ratio for an ordered pair of metabolites p (e.g.,  $p = (ATP, ADP), r_{(ATP,ADP)} = c_{ATP}/c_{ADP}$ ), and **P** is a set of metabolite pairs (e.g.,  $P \in \{(ATP, ADP), (NADPH, NADP^+), (NADH, NAD^+)\}$ ). Note that the *S* matrix here refers to

77 the stoichiometric matrix of the pathway with  $S \in \mathbb{R}^{|I_{path}| \times |J_{path}|}$ .

- 78 79 Constraint 8 relates the Gibbs free energy of reaction  $(\Delta_r G_i)$  with the standard Gibbs free energy of reaction  $(\Delta_r G'^{\circ}_i)$  and the mass action ratio. The concentrations of all metabolites 80 are allowed to vary between 1  $\mu$ M ( $c_i^{min}$ ) and 100 mM ( $c_i^{max}$ ) in constraint 9. 81 82 Concentration ratios of common cofactor pairs (e.g., NADPH/NADP<sup>+</sup>, NADH/NAD<sup>+</sup> and 83 ATP/ADP) play an important role in a cell as they determine the driving force of a large 84 number of biosynthesis reactions (5). The concentration ratios of energy and redox 85 cofactors are therefore allowed to vary within the maximum and minimum values found in 86 the literature (6-8) and the Bionumbers database (9) in constraint 10. Constraint 9 is 87 optional depending on the case study. Herein, we assumed that the designed pathway 88 operates at steady-state and within a single compartment of a cell at a temperature (T) of 89 25°C, an ionic strength of 0.1 M and pH 7.0. The pathway with a positive objective 90 function (MDF) indicates that it is thermodynamically infeasible within the given 91 physiological concentration (and ratio) ranges is omitted from the subsequent step. 92 Importantly, the objective function of the enzyme cost minimization problem is convex 93 only when all  $\Delta_r G'_i < 0$  in a pathway. The MDF problem is solved using Gurobi Optimizer 94 v6.5.1 solver and Python script modified from the Component Contribution Python 95 package (2).
- 96 We have previously found that when imposing the metabolite concentration ranges 97 (constraint 9) strictly to experimentally measured metabolite levels, the MDF formulation 98 is often over-constrained and may become infeasible due to several factors. They include: 99 (i) measurement errors of the absolute intracellular metabolite concentrations, (ii) 100 estimation errors of Gibbs free energy from group-contribution based approaches, (iii) the 101 MDF formulation assumes that metabolite concentrations are homogenous (i.e., 102 compartmentalization of metabolites and potential occurrence of substrate-channeling are 103 ignored), and (iv) MDF analysis is performed with a pathway and not on the entire metabolic network. In addition, when we added the uncertainties in  $\Delta_r G_j^{\prime \circ}$  to the MDF 104 formulation (i.e., allowing the  $\Delta_r G'_j$ ° of each reaction to vary between a range given by 105  $\Delta_r G_i^{\prime \circ} \pm SE_i$  in constraint 8), we found that MDF analysis will identify the more optimistic 106 solution given the degree of freedom to have a more negative  $\Delta_r G_j^{\prime \circ}$ . Since, we are 107 comparing between pathways, the relative contribution of the errors of  $\Delta_r G_i^{\prime \circ}$  estimation 108 109 has a lesser contribution towards the overall analysis and would not affect our conclusions. 110 Therefore, MDF analysis of the pathways is studied within a larger physiological 111 metabolite concentration ranges, and the errors of Gibbs free energy are not considered in 112 the study.

### 113 Minimization of protein cost.

114 The minimal enzyme demand in units of mg protein/mmol glucose/h for each one of the 115 thermodynamically feasible pathways is then estimated based on the enzyme cost 116 minimization (ECM) method (10, 11). The formulation is as followed:

minimize 
$$PC = \frac{1}{v_{EX\_glc}} \sum_{j} M_{E,j} \lambda_{E,j}$$
 (ECM) (11)

subject to  $\lambda_{E,i}$ 

$$= \frac{v_j}{k_{cat,j}^+} \left(1 - \exp\left(\frac{\Delta_r G_j'}{RT}\right)\right)^{-1} \left(1 + \prod_{i \in I_{re,j}} \left(\frac{K_{M,ij}}{c_i}\right)^{q_{ij}^+}\right), \forall j$$
(12)

$$\Delta_r G'_j = \Delta_r G'^{\circ}_j + RT \ln \frac{\prod_{i \in I_{\mathbf{pr},j}} C_i^{q_{ij}}}{\prod_{i \in I_{\mathbf{re},j}} C_i^{q_{ij}^+}}, \quad \forall j \in J_{path}$$
(13)

$$\Delta_r G'_j < 0, \qquad \forall \, j \in J_{path} \tag{14}$$

$$c_i^{min} \le c_i \le c_i^{max}, \quad \forall i \in I_{path}$$
(15)

$$r_p^{\min} \le r_p \le r_p^{\max}, \quad \forall \ p \in \mathbf{P}$$
 (16)

117

where  $M_{E,j}$  is the molecular weight of enzyme per active site for reaction j,  $v_{EX_{alc}}$  is the 118 glucose uptake flux (mmol Glucose/h),  $\lambda_{E,j}$  is the enzyme level for reaction j,  $v_j$  is the flux 119 through reaction j,  $k_{cat,j}^+$  is the turnover number of the reaction in the forward direction, 120  $I_{re,j}$  is the set of reactants in reaction j,  $I_{pr,j}$  is the set of products in reaction j, the set of all 121 122 metabolites in the pathway  $I_{path}$  is the union of  $I_{re,j}$  and  $I_{pr,j}$ ,  $K_{M,ij}$  is the Michaelis-Menten constant of the enzyme for reaction j towards metabolite i,  $q_{ij}^+$  and  $q_{ij}^-$  is the stoichiometric 123 coefficient of metabolite *i* in reaction *j*.  $q_{ij}^+ > 0$  if metabolite *i* is a reactant in reaction *j* 124 and  $q_{ij}^+ = 0$  otherwise, whereas  $q_{ij}^- > 0$  if metabolite *i* is a product in reaction *j* and  $q_{ij}^- = 0$ 125 126 0 otherwise. Note that in the preprocessing step, all the reactions are re-arranged such that 127 flux  $v_i$  through each of them is strictly positive.

128

129 The objective function (equation 11) involves the minimization of the sum of the enzymatic 130 cost (µg Protein/ mmol Glucose/ h) for each reaction in the pathway normalized by the 131 glucose uptake rate. Constraint 12 defines the enzyme level for a reaction *j* as a function 132 derived from the reversible Michaelis-Menten kinetic equation (10). Constraint 13 is 133 equivalent to constraint 8 recasted using concentrations. Constraint 14 ensures that all 134 reactions have a negative change in free energy and prevents division by zero in equation 135 12. Constraints 15 and 16 impose the bounds on the concentration ranges and concentration 136 ratio ranges. The above formulation can be simplified by substituting the concentration 137 variable  $c_i$  with logarithmic concentrations  $x_i = \ln C_i$  and thus converting the product term 138 into a summation.

139

140 According to Flamholz *et al.* (10), the enzyme cost minimization (ECM) formulation can 141 be rewritten by substituting  $x_i = \ln c_i$  as followed: 142

minimize 
$$PC = \frac{1}{v_{EX\_glc}} \sum_{j} M_{E,j} \lambda_{E,j}$$
 (ECM) (17)

subject to 
$$\lambda_{E,j} = \frac{v_j}{k_{cat,j}^+} \frac{1 + \exp\left(\sum_{i \in I_{re,j}} q_{ij}^+ (\ln K_{M,ij} - x_i)\right)}{1 - \exp\left(\sum_{i \in I_{path}} S_{ij}^T x_i + \frac{\Delta_r G_j^{(0)}}{RT}\right)}, \forall j \in J_{path}$$
 (18)

$$\Delta_r G'_j = \Delta_r G'_j + R \cdot T \cdot \sum_{i \in I_{path}} S^T_{ij} x_i, \qquad \forall j \in J_{path}$$
(19)

$$\Delta_r G'_j \le 0 - \varepsilon, \qquad \forall \, j \in J_{path} \tag{20}$$

$$\ln c_i^{min} \le x_i \le \ln c_i^{max}, \quad \forall i \in I_{path}$$
(21)

$$\ln r_p^{\min} \le \ln r_p \le \ln r_p^{\max}, \quad \forall \ p \in \mathbf{P}$$
(22)

143 Using log-concentration simplifies the formulation as the product term in equations 12 and 144 13 can be replaced by the summation term in equations 18 and 19, respectively. Note that 145 we have set  $\varepsilon$  to a very small number (i.e., 1e<sup>-6</sup>) to ensure that the denominator of  $\lambda_{e,j}$  does 146 not become zero. We increase  $\varepsilon$  stepwise by 10-fold up to 0.1 if the optimization failed to 147 converge at a lower  $\varepsilon$  value. If the optimization still fails to terminate successfully at  $\varepsilon =$ 148 0.1, we exclude the pathway from the final solution.

149

150 The optimal concentrations of metabolites obtained from the MDF problem are used as the 151 initial condition for the ECM problem, which is then solved using the sequential least 152 squares quadratic programming method (Python SciPy package). Due to the lack of 153 experimentally measured kinetic parameters, we assumed generic values ( $M_E =$ 154  $40 \ kDa, k_{cat} = 79 \ s^{-1} \ and \ K_M = 200 \ \mu M$ ) (12) for all kinetic parameters as was carried 155 out in the original study (10). This implies that all enzymes were treated as equally fast in 156 every pathway. The allowable metabolite concentration ranges are identical to that of the 157 MDF analysis.

158





Fig. S1. Pathways shown here perform the overall conversion defined below the panels. 161 162 (A) Design A is a pathway with disjoint subnetworks that generate cofactors such as ATP. 163 This design is obtained using the previous optStoic formulation. (B) The  $S_{int}$  matrix, which contains only internal reactions, was processed by removing rows containing cofactors. 164 The basis of the null space of the resulting  $S_{red}$  matrix is then obtained  $(null(S_{red}) =$ 165  $N_{red}$ ). Each row of the  $N_{red}$  matrix is an internal cycle that results in no net non-cofactor 166 metabolite production. The loop law is imposed as  $N_{red}^T G = 0$ , which implies that flux 167 could traverse only through one of the directions in a loop. Two cases are shown here for 168 169 the loop involving reaction R1 (D-Fructose-1,6-phosphate +  $H_2O \rightarrow D$ -Fructose-6phosphate +  $P_i$ ) and R2 (D-Fructose-6-phosphate + ATP  $\rightarrow$  D-Fructose-1,6-phosphate + 170 ADP). In case (ii) (a), when reaction R1 is active ( $v_{R1} > 0$ ), then reaction R2 can carry 171 only zero flux or flux in the same direction with R1. (C) After adding the loop law 172 173 constraints, we found that ATP and redox generation occurs only on the main carbon 174 transfer pathway.





Fig. S2. The distribution of the glycolytic pathway alternatives based on (A) total flux through a pathway, and (B) the number of reactions in a pathway. Note that the total flux through a pathway and the number of reactions are calculated without accounting for the exchange reactions. The colors represent the ATP yield per glucose (mol ATP/mol glucose) generated by a pathway at a fixed glucose uptake flux. Red dashed lines indicate the mean values, whereas blue dashed lines denote the median values.

- 183
- 184
- 185



**Fig. S3.** Distribution of absolute metabolite concentrations across different organisms (8). The fraction of metabolites that are within 1  $\mu$ M and 100 mM are 97.1%, 97.7% and 97.5% for mammalian cells, yeast and *E. coli*, respectively. The fraction of metabolites that fall within 1  $\mu$ M and 10 mM are 94.1%, 90.9% and 94.2% for mammalian cells, yeast and *E. coli*, respectively.



Fig. S4. The ATP yield versus minimal protein cost scatter plot. Note that the Y-axis is
categorical. Jittering effect was applied to the plot to show the distribution more clearly.
Pathways are color-coded based on the type of redox cofactors produced: (Blue) 2 NADH,
(Green) 1 NADH and 1 NADPH, and (Red) 2 NADH.





202 203 Fig. S5. Robustness analysis of the effect of ATP and ADP concentrations on the top ten

204 1-ATP generating pathways. Note that the legends for the x and y-axes are the same for 205 all the ten pathways.



207

Fig. S6. Robustness analysis of the effect of ATP and ADP concentrations on the top fortytwo 2-ATP generating pathways. Note that the legends for the x and y-axes are the same for all the 42 pathways. In addition, the ranges of the axes and heat map scales are the same for each panel.



Fig. S7. Robust 1-ATP and 2-ATP generating glycolytic pathways within a broad range of ATP/ADP ratio. The pathway diagrams were generated using the pathway visualization tool described in the Method section. Pathway (A) is the 9th 1-ATP generating pathways

216 ranked by protein cost in Fig. S5. Pathways (B), (C), (D) and (E) are 36th to 39th 2-ATP

217 generating pathways ranked by protein cost in Fig. S6.



Fig. S8. (A-D) All four variants of the semi-phosphorylative ED pathway and (E) the NADPH-dependent EMP pathway described in the text. The pathway diagrams were generated using the pathway visualization tool described in the Method section. (F) The tradeoff plot of the minimal protein cost and the ATP yield of all glycolytic pathway variants as shown in Figure 3 (B). In addition to the ED (pink star) and the EMP pathways (red star), the semi-phosphorylative ED pathway variants (A-D) are represented as lightblue squares, whereas the NADPH-dependent EMP pathway is shown in the yellow square.

<b>KEGG ID</b>	Description	KEGG ID	Description
C00001	H2O	C00112	CDP
C00002	ATP	C00131	dATP
C00003	NAD+	C00138	Reduced ferredoxin
C00004	NADH	C00139	Oxidized ferredoxin
C00005	NADPH	C00144	GMP
C00006	NADP+	C00206	dADP
C00007	Oxygen	C00286	dGTP
C00008	ADP	C00360	dAMP
C00009	Orthophosphate	C00361	dGDP
C00010	СоА	C00362	dGMP
C00011	CO2	C00363	dTDP
C00013	Diphosphate	C00364	dTMP
C00015	UDP	C00365	dUMP
C00016	FAD	C00390	Ubiquinol
C00020	AMP	C00399	Ubiquinone
C00035	GDP	C00458	dCTP
C00044	GTP	C00459	dTTP
C00055	СМР	C00460	dUTP
C00063	СТР	C01352	FADH2
C00075	UTP		
C00080	H+		
C00081	ITP		
C00104	IDP		
C00105	UMP		

Table S1. Cofactors that were removed from the S matrix when generating the internal stoichiometric matrix ( $S^*$ ).

Table S2A. The number of pathways that are thermodynamically feasible (MDF < 0) at physiological concentration ranges and ratio.

Conditions	# of 1-	# of 2-	# of 3-	# of 4-	# of 5-
	ATP	ATP	ATP	ATP	ATP
	Pathways	Pathways	Pathways	Pathways	Pathways
optStoic	5,739	3,430	1,873	659	215
condition (i)	4,550	2,891	1,542	466	165
condition (ii)	2,549	1,099	281	4	0
condition (iii)	2,525	1,098	281	4	0
condition (iv)	1,824	778	173	2	0
condition (v)	538	105	0	0	0
condition (vi)	2,558	1,099	281	4	0

i. All metabolites are allowed to vary between 1 µM and 100 mM.

235	ii.	Same with (i), except that ATP and ADP concentrations are bounded based on
236		Park <i>et al.</i> (8) (i. e., 1.66 mM $\leq C_{ATP} \leq 11.4$ mM; 0.429 mM $\leq C_{ADP} \leq$
237		0.715 mM).

238 iii. Same with (ii), except that the concentration range of CO<sub>2</sub> was bounded based on 239 Park *et al.* (8) (i.e., 50  $\mu$ M  $\leq C_{CO_2} \leq 10$  mM).

iv. All metabolites are allowed to vary between 1  $\mu$ M and 100 mM except CO<sub>2</sub>. The range of CO<sub>2</sub> was obtained from Park *et al.* (8) (i.e., 50  $\mu$ M  $\leq C_{CO_2} \leq 10$  mM). The ratio ranges for different cofactor pairs were imposed as followed: 0.2  $\leq$  $C_{ATP} \leq 20, 0.2 \leq C_{NADPH} \leq 100, 0.0005 \leq C_{NADH} \leq 0.5$  have d on data collected

243 
$$\frac{C_{APP}}{C_{ADP}} \le 20, \ 0.2 \le \frac{C_{NADP}}{C_{NADP}} \le 100, \ 0.0005 \le \frac{C_{NAD}}{C_{NAD}} \le 0.5 \text{ based on data collected}$$
244 from Bionumbers (9) and literature (6-8).

245 v. All metabolites other than CO<sub>2</sub> are allowed to vary between 1 
$$\mu$$
M and 10 mM.

246 The range of CO<sub>2</sub> was obtained from Park *et al.* (8) (i.e., 50  $\mu$ M  $\leq$  C<sub>CO<sub>2</sub></sub>  $\leq$  10

248 
$$0.2 \le \frac{c_{ATP}}{c_{ADP}} \le 20, \, 0.2 \le \frac{c_{NADPH}}{c_{NADP}} \le 100, \, 0.0005 \le \frac{c_{NADH}}{c_{NAD}} \le 0.5.$$

249 vi. All metabolites are allowed to vary between 1  $\mu$ M and 100 mM. The ratio ranges

250 for different cofactor pairs were imposed as followed:  $1 \le \frac{C_{ATP}}{C_{ADP}} \le 10,000$ .

Table S2B. All the constraints used for the simulation of Table S2A.

Conditions	(i)	(ii)	(iii)	(iv)	(v)	(vi)
(A) $1 \mu M \leq C_i \leq 100 \text{ mM}$ for all	+	+	+	+		+
metabolite <i>i</i>						
(B) 1.66 mM $\leq C_{ATP} \leq 11.4$ mM;		+	+			
$0.429 \text{ mM} \le C_{ADP} \le 0.715 \text{ mM}$						
(C) 50 $\mu$ M $\leq C_{CO_2} \leq 10 \text{ mM}$			+	+	+	
(D) $0.2 \le \frac{c_{ATP}}{c_{ADP}} \le 20;$				+	+	
$0.2 \le \frac{C_{NADPH}}{C_{NADP}} \le 100;$						
$0.0005 \le \frac{C_{NADH}}{C_{NAD}} \le 0.5$						
(E) $1 \ \mu M \le C_i \le 10 \ mM$ , for all					+	
metabolite <i>i</i>						
(F) $1 \le \frac{c_{ATP}}{c_{ATP}} \le 10,000$						+

255	Refer	rences
256	1.	Noor E, et al. (2014) Pathway thermodynamics highlights kinetic obstacles in
257		central metabolism. PLoS computational biology 10(2):e1003483.
258	2.	Noor E, Haraldsdottir HS, Milo R, & Fleming RM (2013) Consistent estimation
259		of Gibbs energy using component contributions. PLoS computational biology
260		9(7):e1003098.
261	3.	Biemans-Oldehinkel E, Mahmood NA, & Poolman B (2006) A sensor for
262		intracellular ionic strength. Proceedings of the National Academy of Sciences of
263		the United States of America 103(28):10624-10629.
264	4.	Storey KB (2004) Functional Metabolism: Regulation and Adaptation (John
265		Wiley & Sons, Inc., Hoboken, NJ, USA) p 616.
266	5.	Zhang J, et al. (2015) Determination of the Cytosolic NADPH/NADP Ratio in
267		Saccharomyces cerevisiae using Shikimate Dehydrogenase as Sensor Reaction.
268		Scientific Reports 5:12846.
269	6.	Bennett BD, et al. (2009) Absolute metabolite concentrations and implied enzyme
270		active site occupancy in Escherichia coli. Nature chemical biology 5(8):593-599.
271	7.	Gerosa L, et al. (2015) Pseudo-transition Analysis Identifies the Key Regulators
272		of Dynamic Metabolic Adaptations from Steady-State Data. Cell Syst 1(4):270-
273		282.
274	8.	Park JO, et al. (2016) Metabolite concentrations, fluxes and free energies imply
275		efficient enzyme usage. Nature chemical biology.
276	9.	Milo R, Jorgensen P, Moran U, Weber G, & Springer M (2010) BioNumbersthe
277		database of key numbers in molecular and cell biology. Nucleic acids research
278		38(Database issue):D750-753.
279	10.	Flamholz A, Noor E, Bar-Even A, Liebermeister W, & Milo R (2013) Glycolytic
280		strategy as a tradeoff between energy yield and protein cost. Proceedings of the
281		National Academy of Sciences of the United States of America 110(24):10039-
282		10044.
283	11.	Noor E, et al. (2016) The Protein Cost of Metabolic Fluxes: Prediction from
284		Enzymatic Rate Laws and Cost Minimization. PLoS computational biology
285		12(11):e1005167.
286	12.	Bar-Even A, et al. (2011) The moderately efficient enzyme: evolutionary and
287		physicochemical trends shaping enzyme parameters. <i>Biochemistry</i> 50(21):4402-
288		4410.
289		