

## Supplemental Information

### Multiomic Profiling Identifies

### *cis*-Regulatory Networks Underlying Human

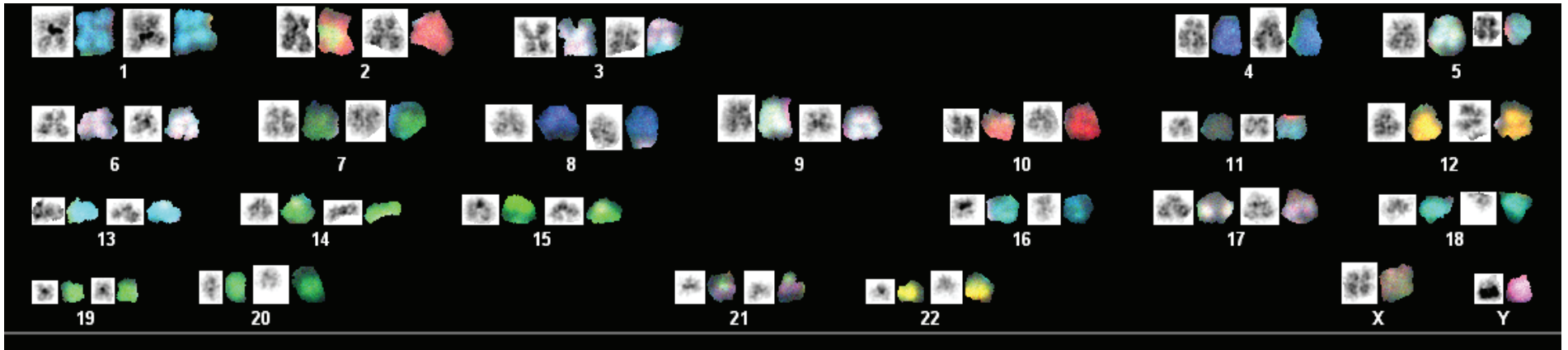
### Pancreatic $\beta$ Cell Identity and Function

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SKY EndoC-βH1 Metaphase S2.1

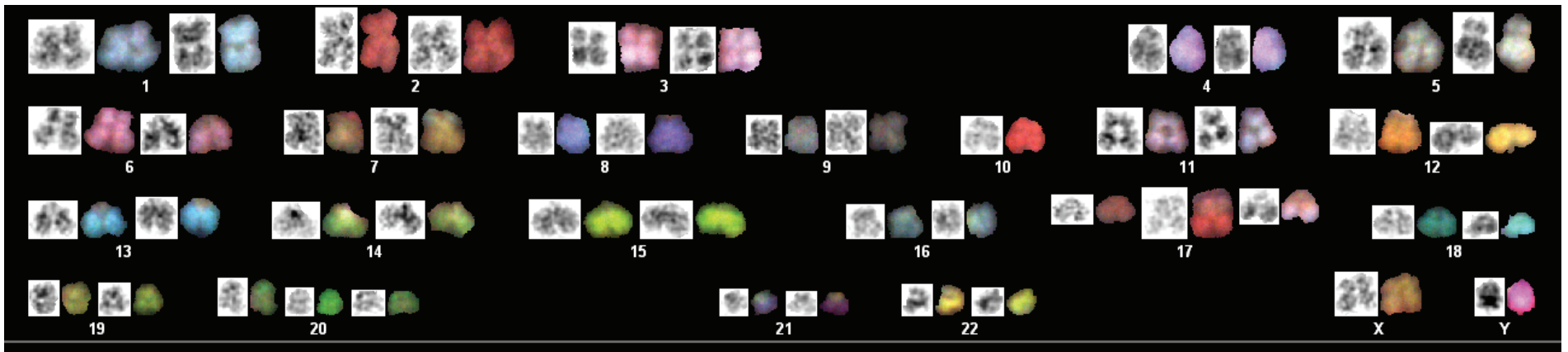
46,XY,der(17)t(10;17),der(17)t(3;17),der(17)t(3;17)

Common event to multiple metaphases



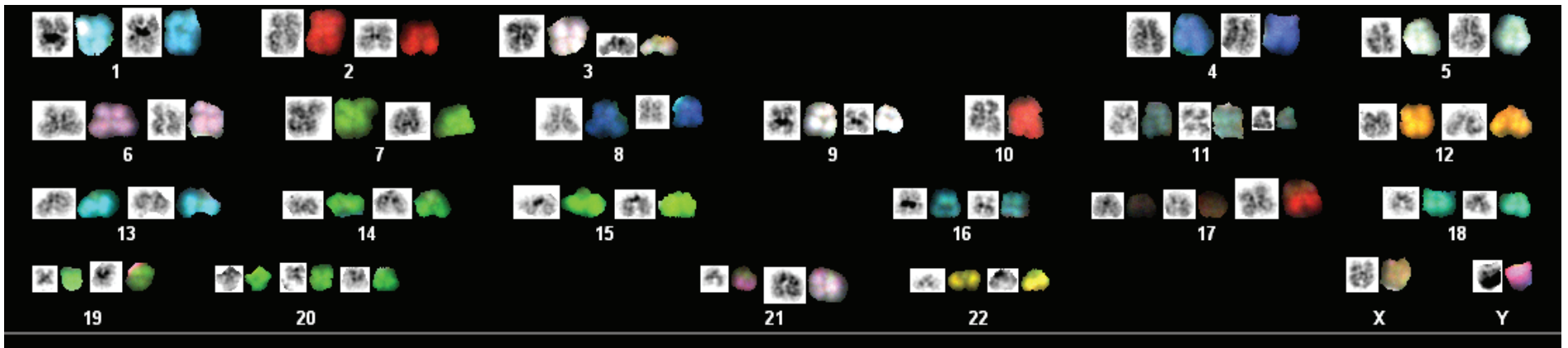
SKY EndoC-βH1 Metaphase 7

47,XY,t(7;18),-10,t(10;17),t(3;17),der(17)t(3;17),+20

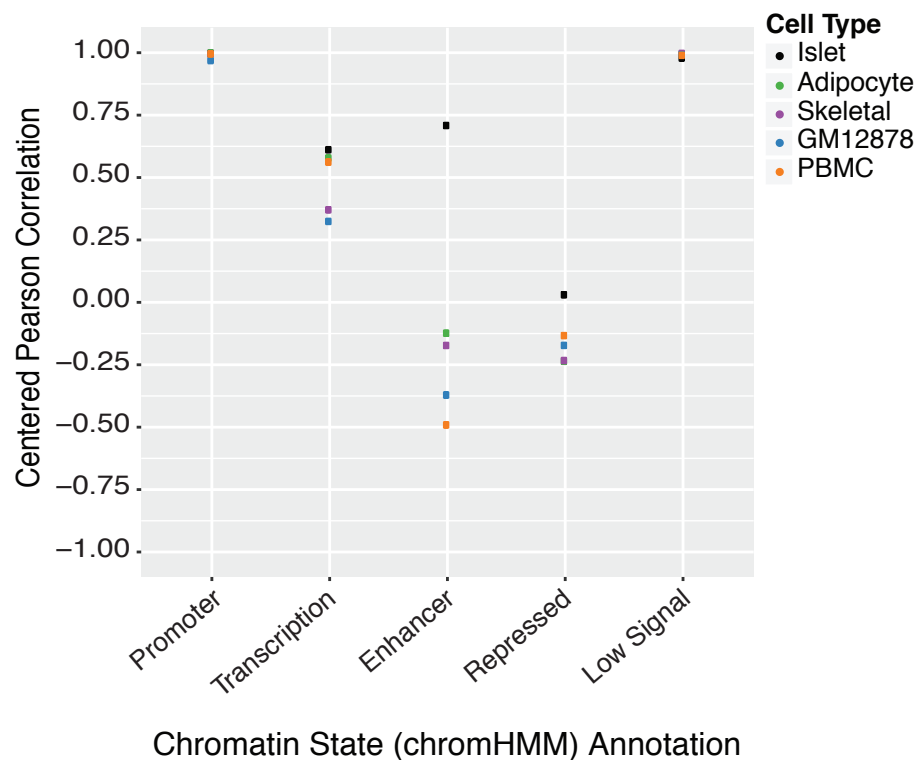
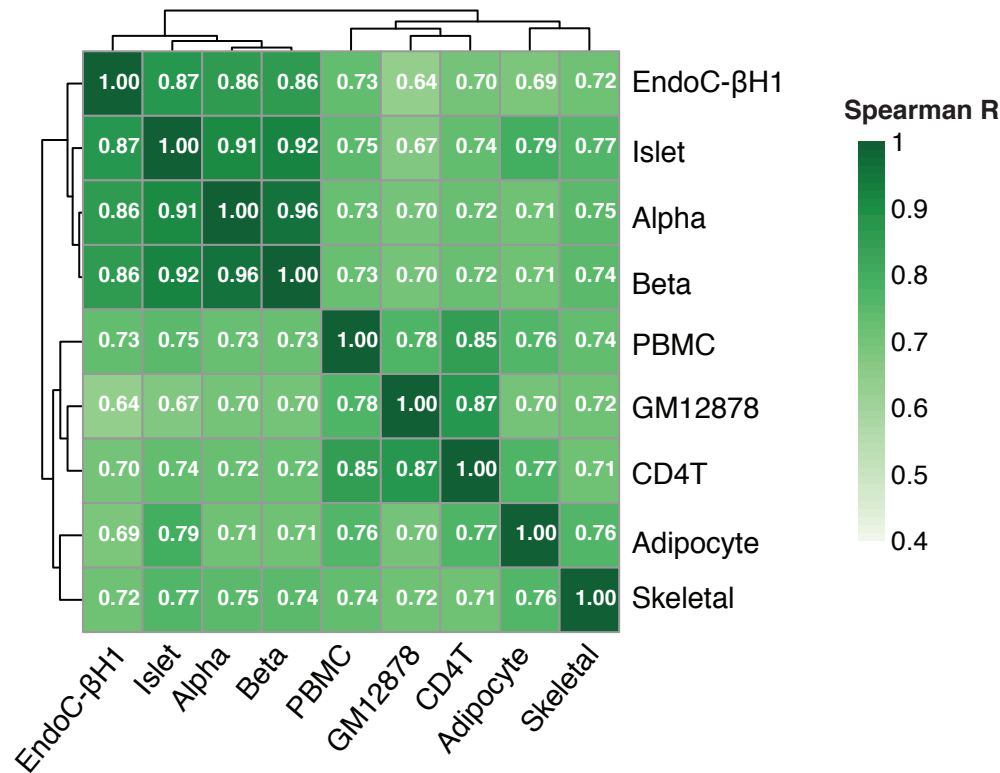
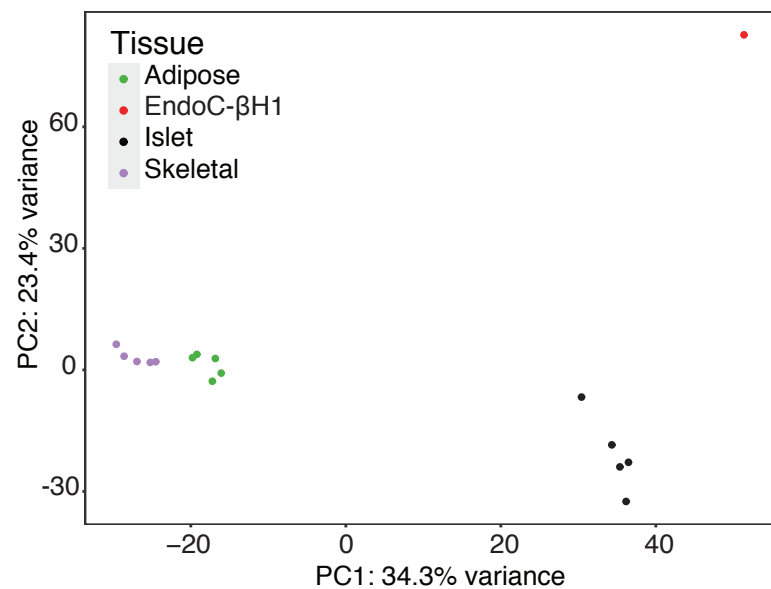
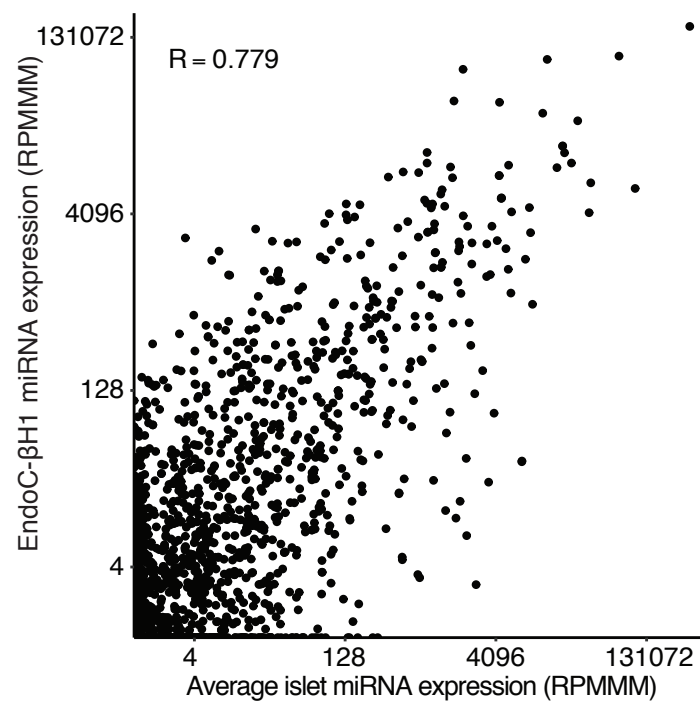


SKY EndoC-βH1 Metaphase S2.7

48,XY,t(3;21),-10,+der(17)t(10;17),+20



**Figure S1, related to Figure 1: Representative spectral karyotypes (SKY) of EndoC-βH1 cells at metaphase.** Specific metaphases shown are S2.1 (top), 7 (middle), and S2.7 (bottom). Common structural or numerical chromosomal aberrations that are evident in multiple metaphases are indicated in green.

**A** Similarity of cell type chromatin states to EndoC- $\beta$ H1**B** Correlation of EndoC- $\beta$ H1 and other tissue RNA-seq profiles**C** PCA of EndoC- $\beta$ H1 and other tissue miRNA profiles**D** Correlation of EndoC- $\beta$ H1 and Islet miRNA profiles

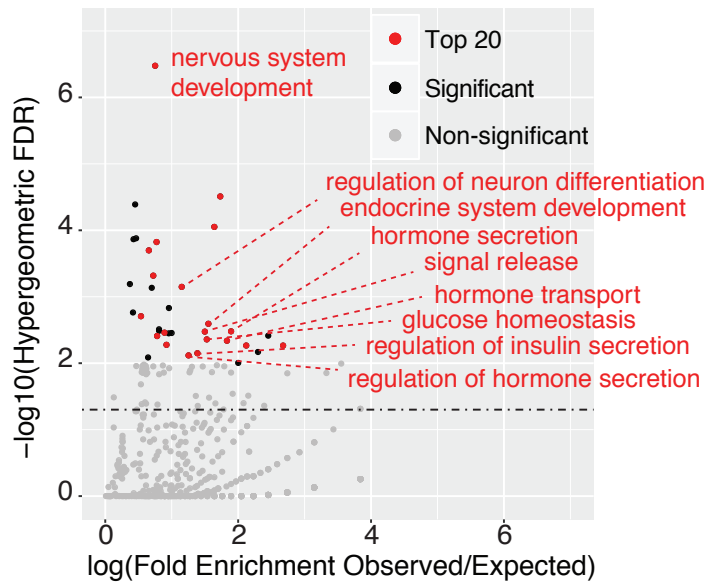
**Figure S2, related to Figure 2: Comparison of EndoC-βH1 and islet epigenomic and transcriptomic profiles.**

(A) Similarity (centered Pearson correlation) of human islet, adipocyte, skeletal muscle, GM12878, and PBMC chromatin state annotations (ChromHMM) to those of EndoC-βH1 at EndoC-βH1 OCRs. The similarity matrix was calculated using the “simil” function within the proxy version 0.4 R package (Meyer and Buchta, 2018).

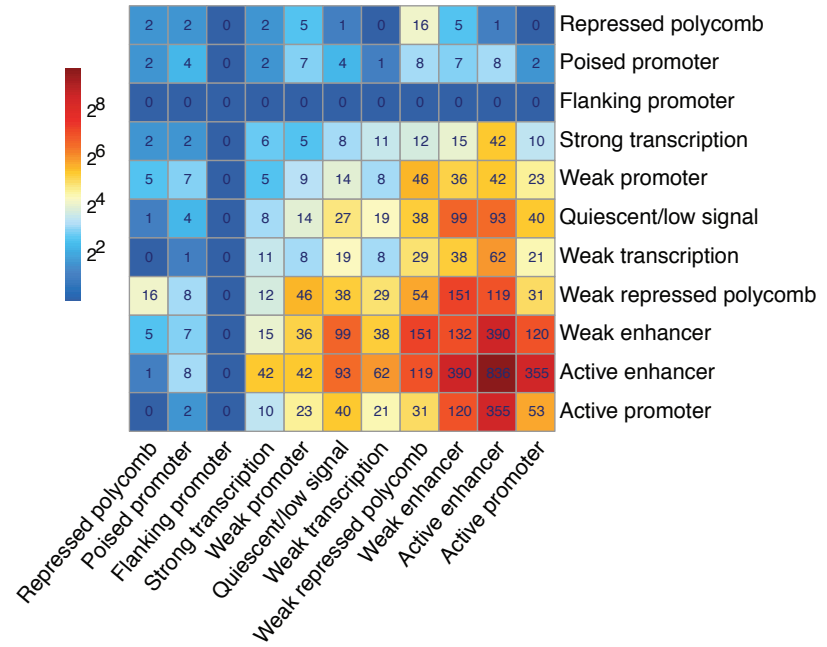
(B) Spearman correlation between EndoC-βH1 RNA-seq and the corresponding RNA-seq from islets, sorted alpha or beta cells (Ackermann et al., 2016), and other cell types and tissues. PBMC=peripheral blood mononuclear cells, GM12878 = lymphoblastoid cell line, CD4T = CD4+ T immune cell, skeletal = skeletal muscle, Alpha = primary islet alpha cells, Beta = primary islet beta cells.

(C) Principal component analysis (PCA) of miRNA-seq profiles from EndoC-βH1 and 5 representative human islet, skeletal muscle, and adipose tissue samples. (D) Scatter plot illustrating the resemblance of miRNA expression levels between EndoC-βH1 and human islets. RPMMM = reads per million mapped miRNA, R = Pearson R.

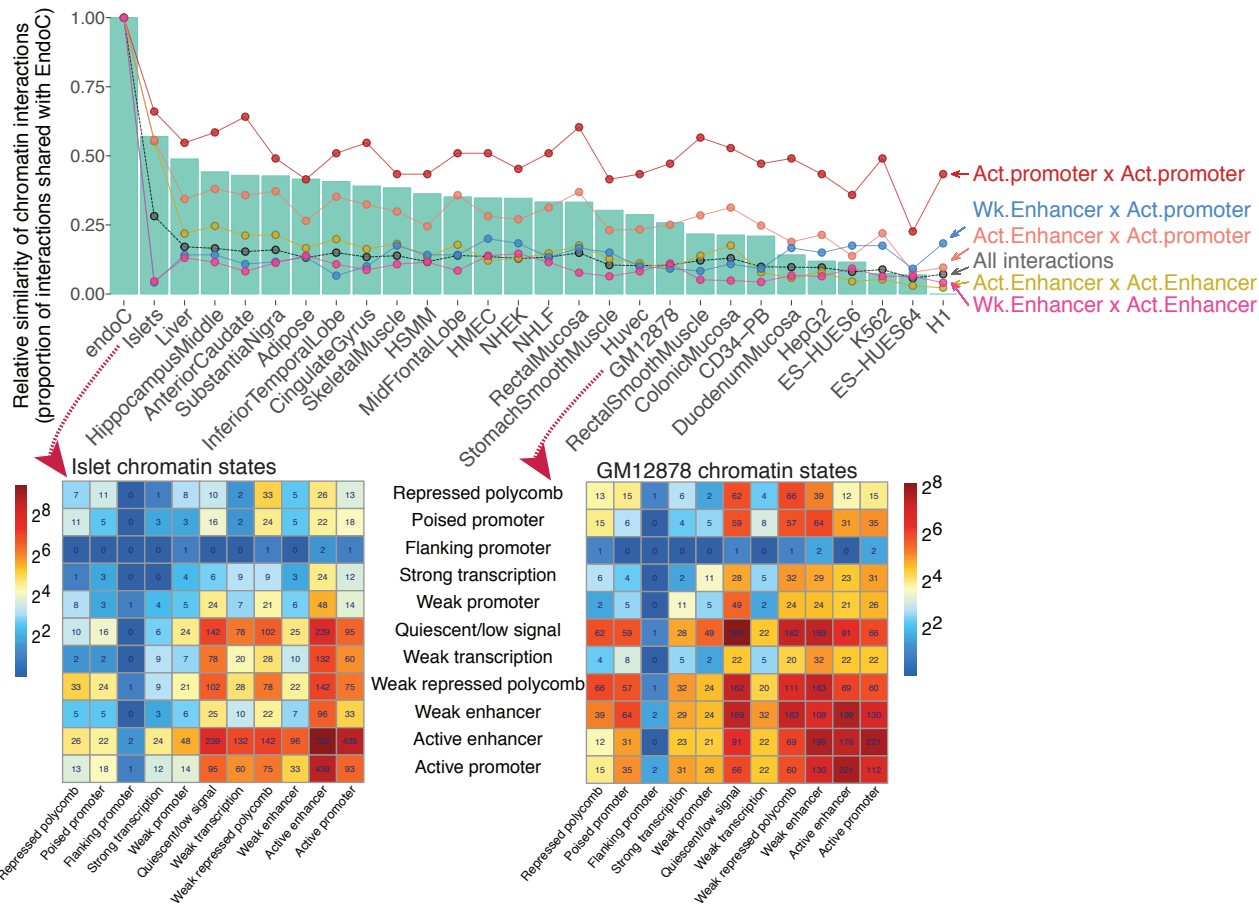
### A Pathway analysis of genes adjacent to EndoC-βH1 anchors



### B EndoC-βH1 Hi-C loop locations



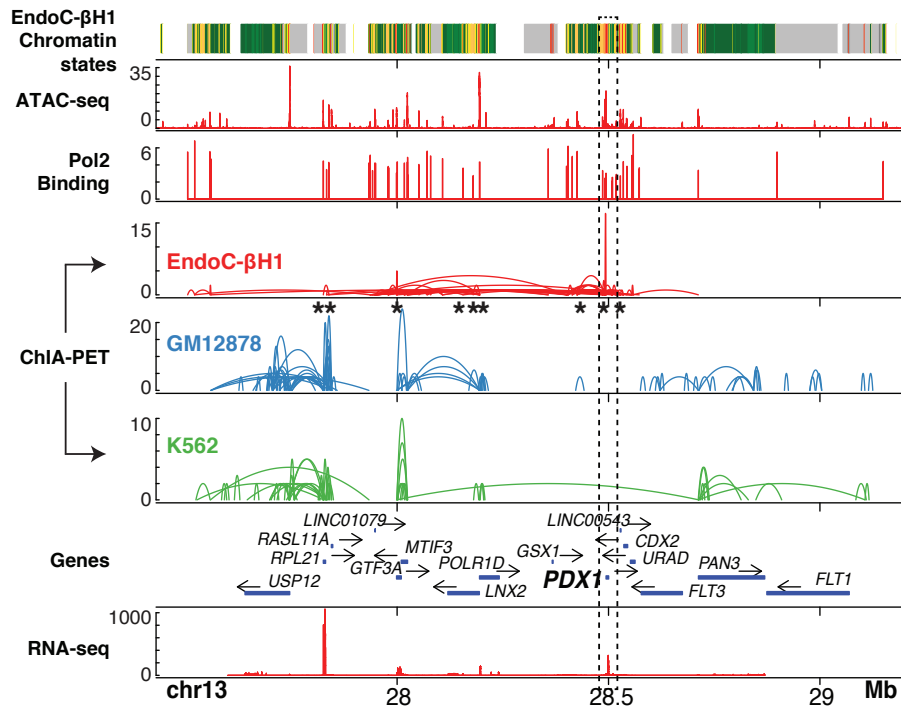
### C Specificity of EndoC-βH1 Hi-C looping locations



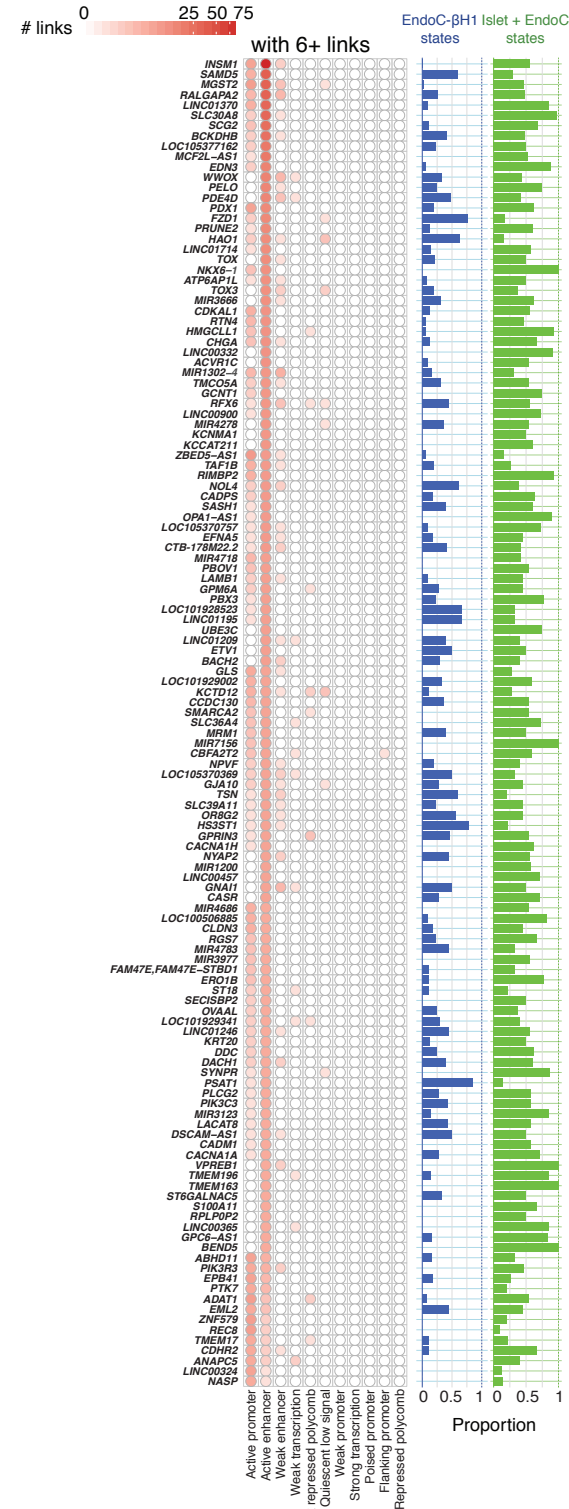
**Figure S3, related to Figure 3: Enrichment analysis and annotation of EndoC-βH1 Hi-C loops.**

(A) Biological process gene ontology terms enriched in genes adjacent to EndoC-βH1-specific Hi-C anchors. Enrichment analysis was performed with GREAT (McLean et al., 2010) using the single gene whose transcription start site (TSS) was nearest to each anchor. Results with an adjusted (Benjamini & Hochberg) hypergeometric p-value < 0.05 were regarded as statistically significant. (B) Frequency of EndoC-βH1 Hi-C loop anchors and their corresponding chromatin state (ChromHMM) annotations. (C) (Top) Bar plot depicting the average similarity of chromatin state (ChromHMM) annotations for each cell type at EndoC-βH1 Hi-C loop anchor positions. Overlaid line plots highlight the relative similarity for various chromatin state interactions (e.g. active promoter vs. active promoter). (Bottom) Select heat maps showing the frequency of human islet (left) and GM12878 (right) chromatin states at EndoC-βH1 loop anchors.

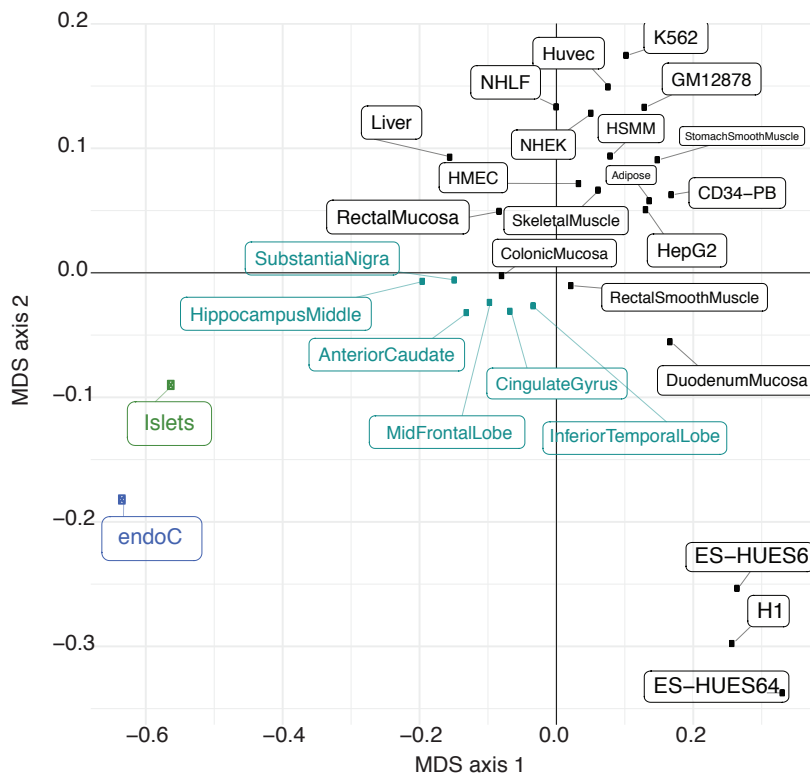
### A Connectivity at *PDX1* locus



### B Network connectivity of EndoC-βH1 anchor genes



### C MDS based on dissimilarity with EndoC-βH1 chromHMM states



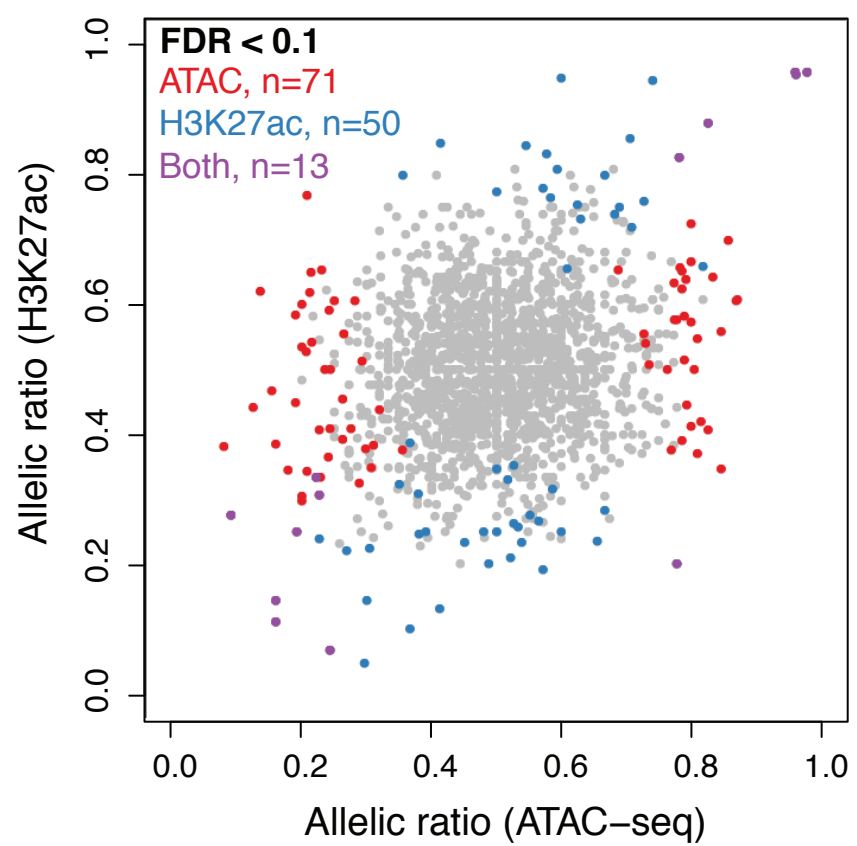


**Figure S4, related to Figure 4: Annotation and evaluation of EndoC-βH1 RNA Pol2 ChIA-PET interactions.**

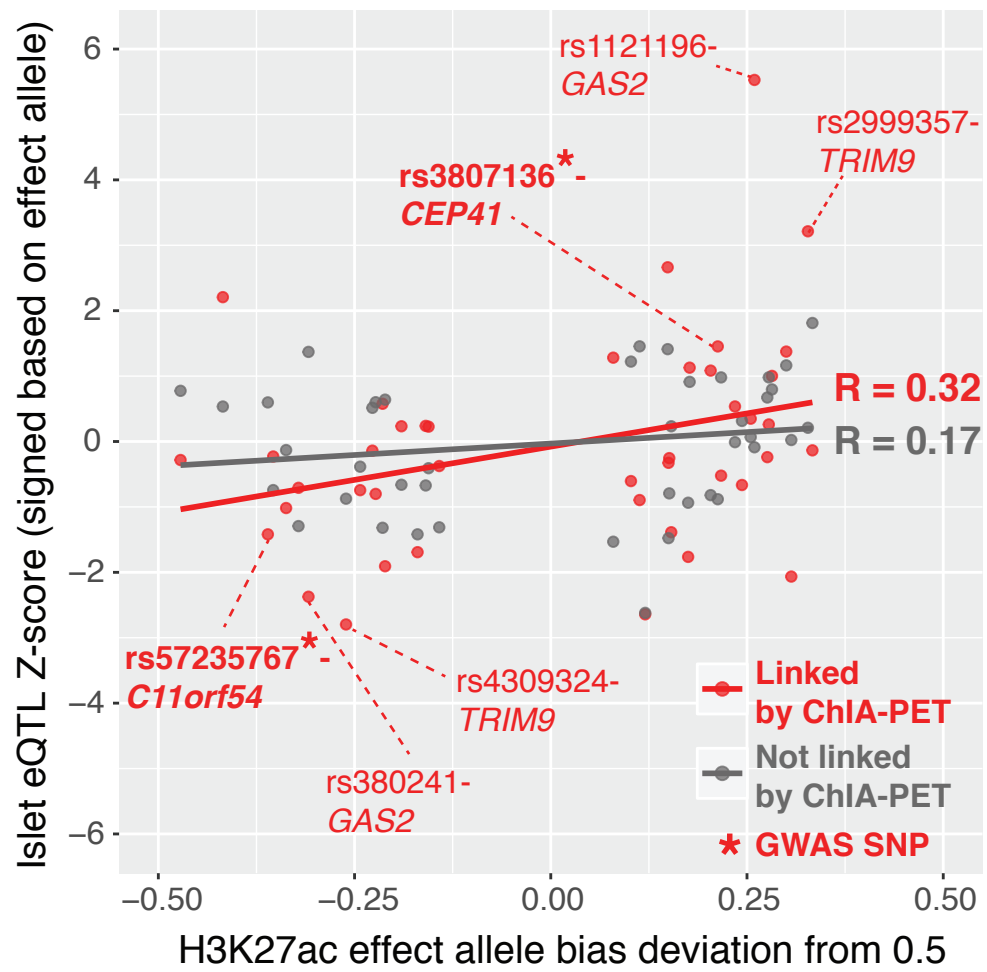
(A) Genome-wide view of RNA polymerase 2-mediated (Pol2 ChIA-PET) chromatin interactions around the PDX1 locus on chromosome 13 in EndoC-βH1. Asterisks under EndoC-βH1 (red) ChIA-PET interactions indicate interacting sites/anchors identified by targeted 4C-seq analyses in this locus in human islets (Pasquali et al., 2014). (B) Connectivity of EndoC-βH1 ChIA-PET interactions when considering nodes with 6 or more links to other regulatory elements. (Left) Circular plots depict the number of links that occur with corresponding regulatory elements (e.g., active promoter, active enhancer). (Right) bar plots illustrate the proportion of interacting nodes that exhibit the active regulatory element chromatin states exclusively in EndoC-βH1 (blue) or identical chromatin states in both EndoC-βH1 and islet (green). (C) Multidimensional scaling (MDS) plot based on pairwise Chi-square distances of vectors of proportions of chromatin states in EndoC-βH1, islets, and additional Epigenomics Roadmap cell and tissue types at EndoC-βH1 defined ChIA-PET interacting nodes.

# A Correlation of cis-regulatory element allelic imbalance

N = 1734 SNPs, R = 0.2



# B Cis-regulatory element imbalance at human islet eQTL SNP-gene pairs



**Figure S5, related to Figure 5: Assessment of cis-regulatory element allelic imbalance in EndoC-βH1 and islets.**

(A) Correlation of cis-regulatory element allelic imbalance for SNPs with heterozygous genotypes in EndoC-βH1. For a given heterozygous SNP ( $n = 1,734$  total with 20X coverage in EndoC-βH1 ATAC-seq and H3K27ac data), allelic imbalance ratios were calculated and plotted. Colored points signify SNPs with significant allelic imbalance ( $FDR < 10\%$ ) in ATAC-seq (red), H3K27ac (blue), or both (purple) datasets. (B) Correlation of human islet eQTL SNP-gene pair direction of effect (z-score) from (Varshney et al., 2017) and their corresponding H3K27ac effect allele bias deviation from 0.5 in EndoC-βH1. Red points indicate eQTL SNP-gene pairs that are also linked by an EndoC-βH1 ChIA-PET interaction. Asterisks indicate eQTL SNPs that are also diabetes-associated GWAS SNPs.