

Supplementary information: Extensive epigenomic integration of the glucocorticoid response in primary human monocytes and in vitro derived macrophages

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Buffycoat	RNA-seq	H3K4me1-ChIP	H3K4me3-ChIP	H3K27ac-ChIP	GR-ChIP
bc9	MoMf_TA				
bc10			MoMf_TA		
bc11		MoMf_TA		MoMf_TA	Mf_TA
bc12	MoMf_TA			Mo_TA	Mo_TA
bc16				Mf_TA	
bc19		MoMf_TA	MoMf_TA		
bc27	MoMf_TA				
bc28	MoMf_TA				
bc33					Mo_TA
bc45					Mf_TA
bc52	Mo_TA-DEX 100nM				
bc53	Mo_TA-DEX 100nM				

Table S1 Healthy adult blood donation buffy coats and the assays performed on them.

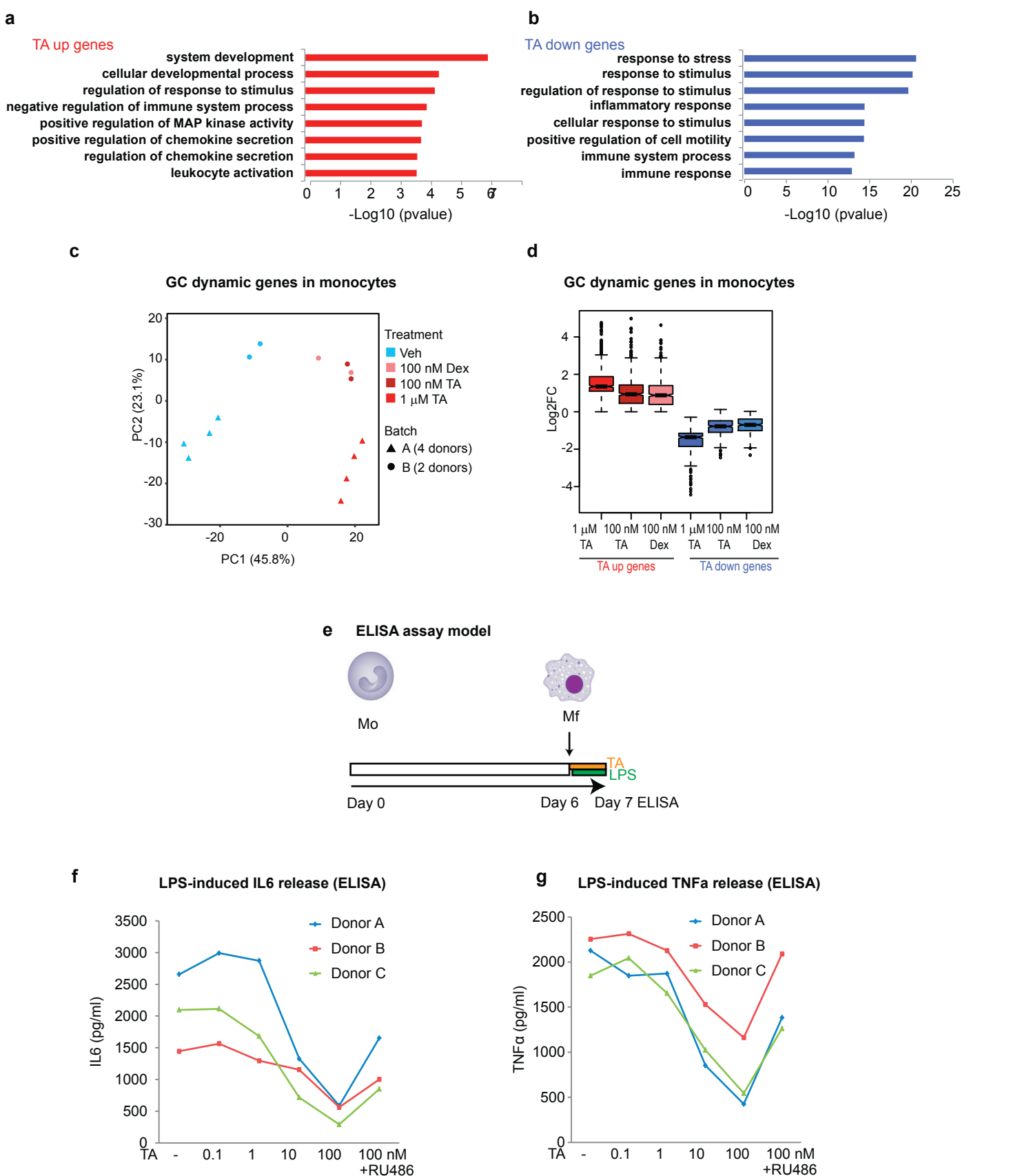


Figure S1

(a) Gene ontology analysis of the 629 TA up-regulated genes displayed on Fig. 1c-e.

(b) As (a) for the 451 down-regulated genes displayed on Fig. 1c-e.

(c) Principal component analysis of the eight samples derived from four donors' Mo exposed to 1 μ M TA (triangles) displayed in Fig. 1b and of two additional donor's Mo samples exposed to vehicle (0.1% DMSO), 100 nM TA or 100 nM dexamethasone (circles). Symbol colour and shading indicate conditions.

(d) Box plot representation of the log₂ fold-change for TA-dynamic gene in TA (1 and 0.1 μ M) or dex (0.1 μ M).

(e) Time line of LPS stimulation of Mo-derived Mf for the experiments displayed in panels (f) and (g).

TA was added 1 hour before LPS. The Elisa measurement was performed 24 h later.

(f) ELISA results against IL6 as a function of the indicated escalating TA doses in three donor's Mo-derived Mf. RU486 was used at a concentration of 1 μ M.

(g) As (f) but for TNF alpha release.

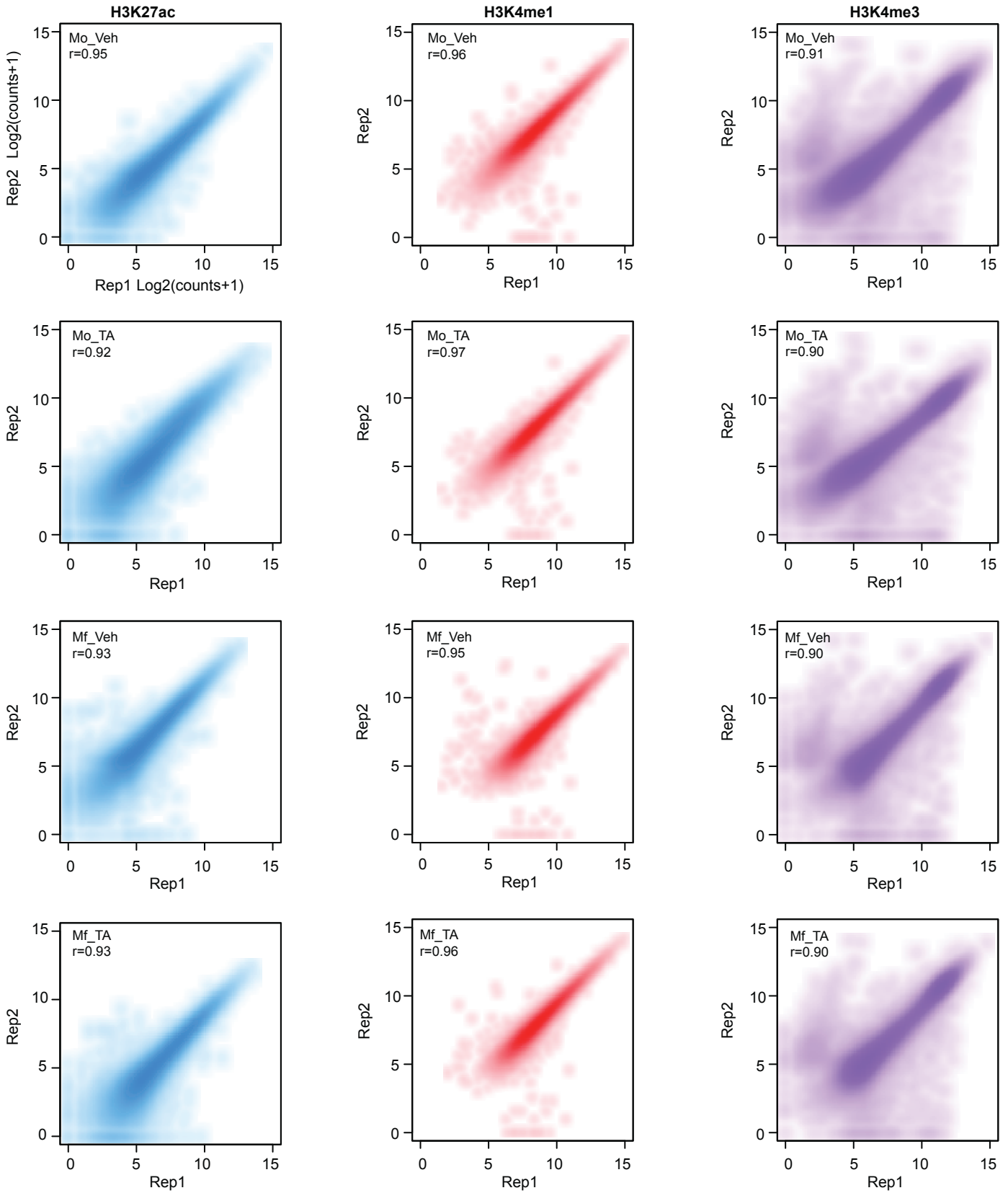


Figure S2

Pairwise comparisons of the post-translationally modified histone H3 ChIP-seq experiments performed with two healthy donors' cells. Note the Pearson correlation coefficients which are always above 0.9.

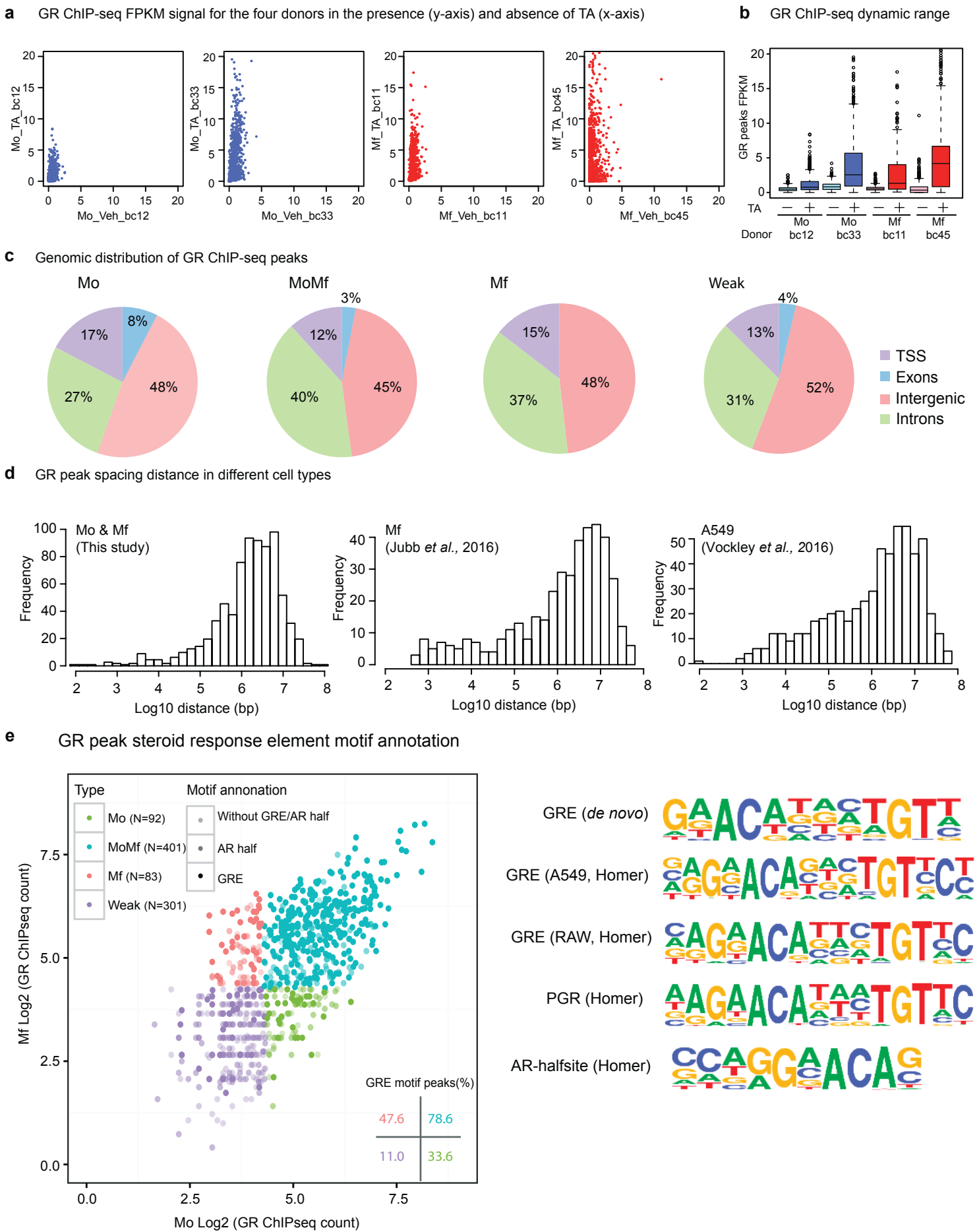


Figure S3

(a) Scatter plots of the four GR ChIP-seq experiment whereby the ChIP signal in the absence and presence of TA are plotted against each other for each donor sample pair. (b) The dynamic range of the four GR ChIP-seq experiments is indicated by the distributions of signal rendered as box plots. (c) Pie chart depiction of the proportions of GR-peaks in exons, introns, intergenic regions and transcription start sites for the four sets of GR peaks depicted in (e). (d) Distribution of intervals between GR peaks in three independent data sets. Since 23 chromosomes are considered, the 877 GR peaks (Fig. 3b) yield 854 intervals. The intervals between GR ChIP-seq peaks detected in bone marrow derived human macrophages (BMDM)¹ and the top 599 GR ChIP-seq peaks of A549 cells² are shown for comparison. (e) GR signal intensity in Mo and Mf at the 877 high confidence GR ChIP-seq peaks defines four sets of GR peaks (MoMf, Mo, Mf and 'Weak'). Overlap of each peak with bona fide AR half-sites and GREs is indicated by shading. The inset indicates percentages of peaks that harbour a GRE. GRE, PGR and AR half-site sequence logos obtained from the Homer database and used to scan DHS associated with H3K27ac-marked regions are depicted next to panel e.

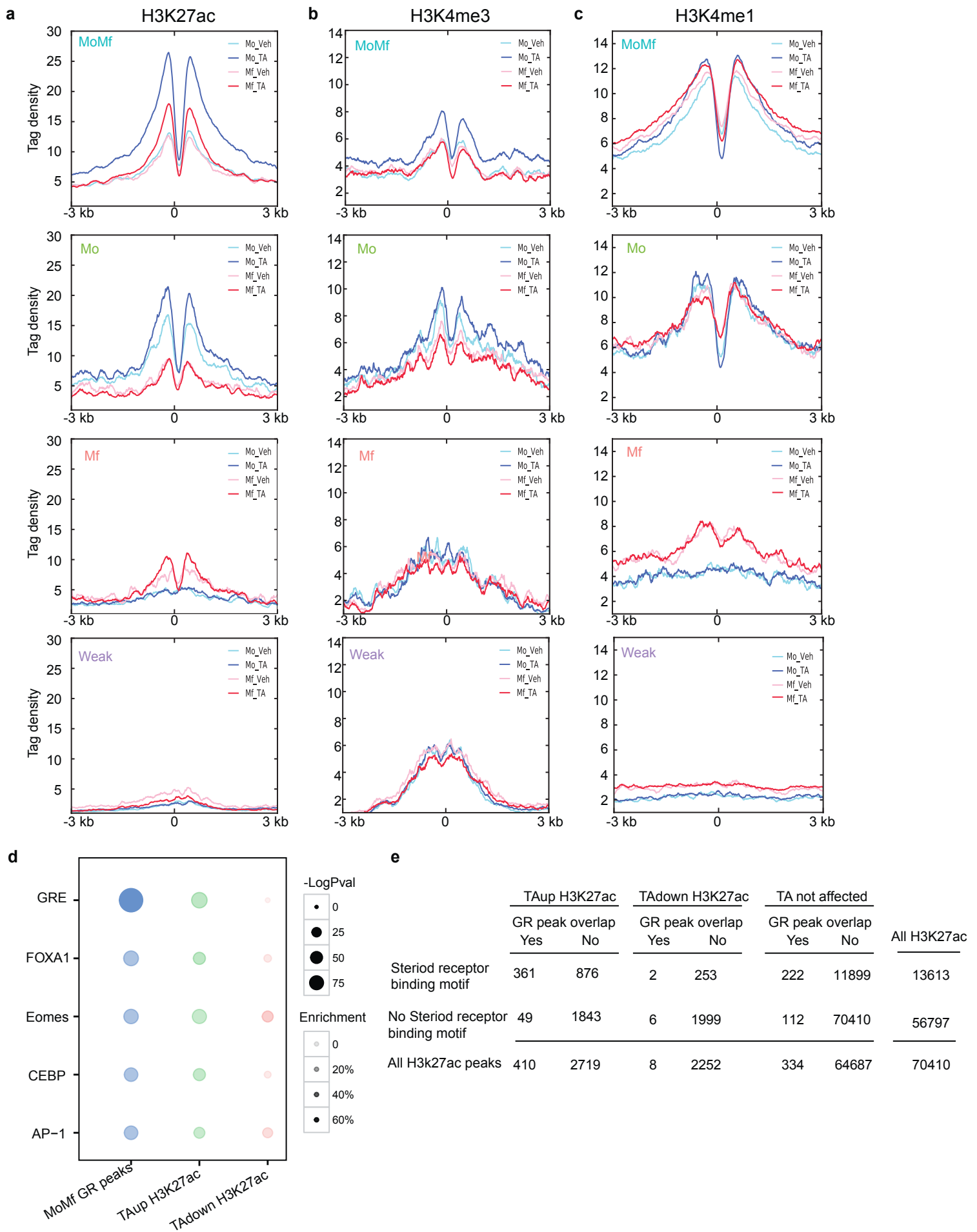


Figure S4

(a) H3K27ac ChIP-seq signal intensity in 6 kb windows centred on the four types of GR binding sites displayed in Figure 3c. (b) and (c) as (a) but for H3K4me3 and H3K4me1, respectively. (d) Statistical (P-value) enrichment (% above background) rendition of the indicated TF motif instances enriched in DHS bearing any one of the 401 MoMf GR peaks depicted on Fig. 3C in DHS associated with up- and down-regulated H3K27ac regions displayed on Fig. 2B. Note the depletion of lineage determining CEBP- and GR-associated motifs in TA-down-regulated H3K27ac DHS. (e) Contingency table of the occurrence of any of the full steroid receptor motifs (displayed on Fig. S3e) in H3K27ac-associated DHS that gain, lose or are not affected by TA for peaks that overlap one of the 877 GR-binding sites (Fig. 3c) or not, and arranged in two rows as a function of the presence or absence of steroid receptor motif instances.

Fig. S5

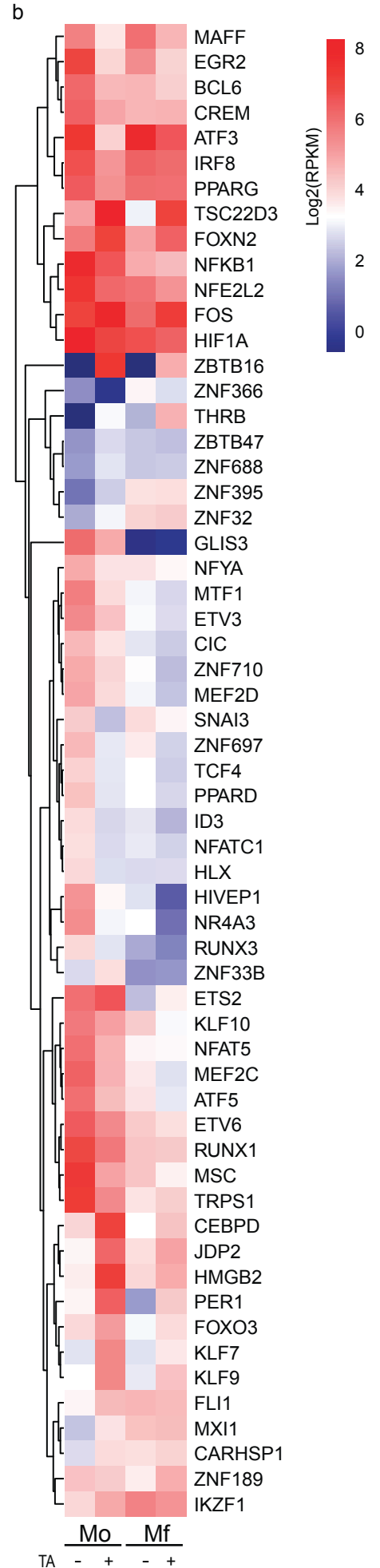
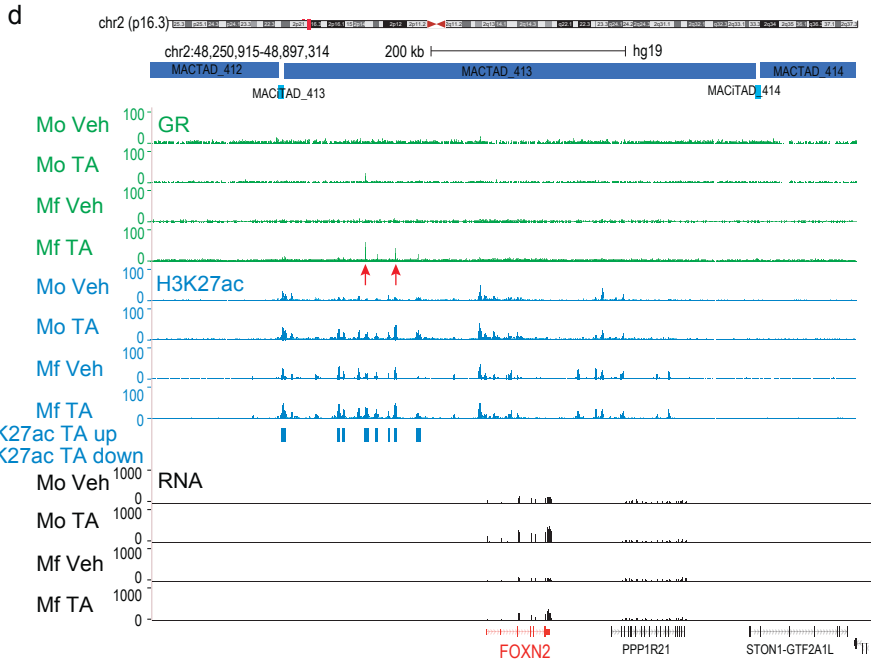
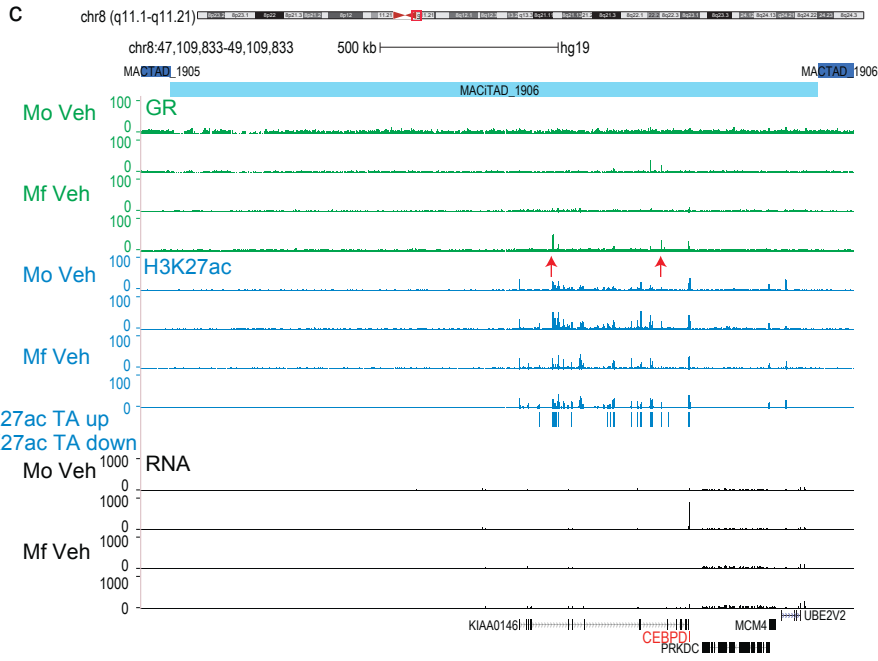
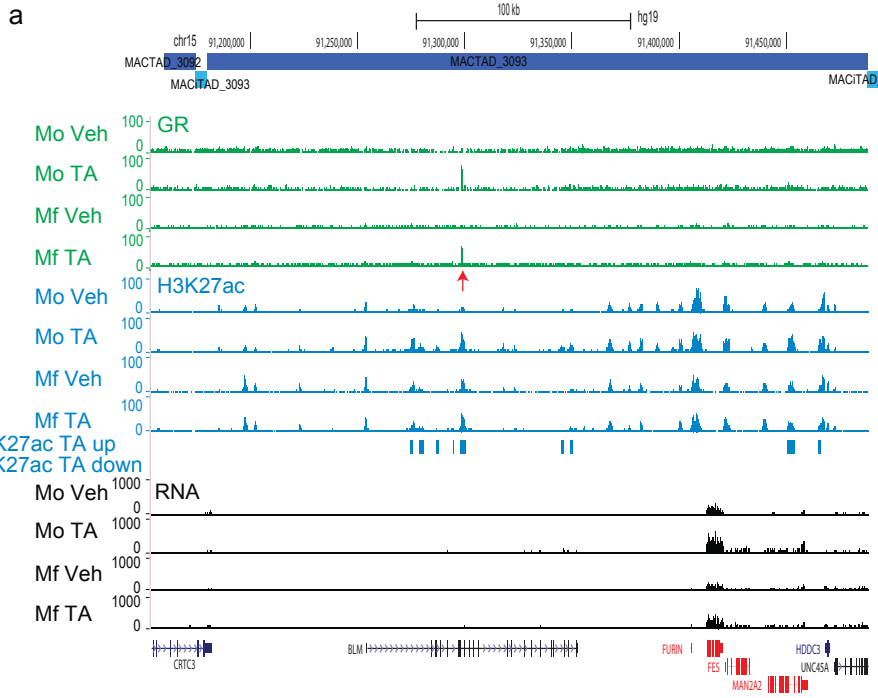


Figure S5

(a) UCSC genome browser screenshot of chromosome 15q26.1 depicting MACTAD 3093 harbouring the TA-inducible genes *FURIN*, *FES* and *MAN2A2* genes and a single MoMf-GR ChIP-seq peak (red arrow) in the 7th intron of the *BLM* gene.

(b) Heat map clustered on the basis of expression patterns for TFs that change expression at least two-fold as a function of TA-exposure in Mo and/or Mf. The colour scale indicates the level of expression (log₂ RPKM).

(c) UCSC genome browser screenshot of chromosome 8q11.1-q11.21 depicting MAC(i)TAD 1906 which bears the TA-inducible *CEBPD* gene that codes for the bZIP CCAAT enhancer binding protein D transcription factor. TAD dimensions, GR ChIP-seq signal (red arrow), H3K27ac ChIP-seq signal, called dynamic H3K27ac regions and RNA-seq signal are displayed above the human genes. Note the presence of at least twenty TA-induced regions that largely coincide with non-coding RNA KIAA0146 and the *CEBPD* gene in a region that extends 385 kb downstream from *CEBPD*.

(d) As for (c) but chromosome 9q31.2-q31.3 depicting MACTAD 2146 which bears the TA-inducible gene *FOXN2*, multiple GR binding sites (red arrows) and many induced H3K27ac regions.

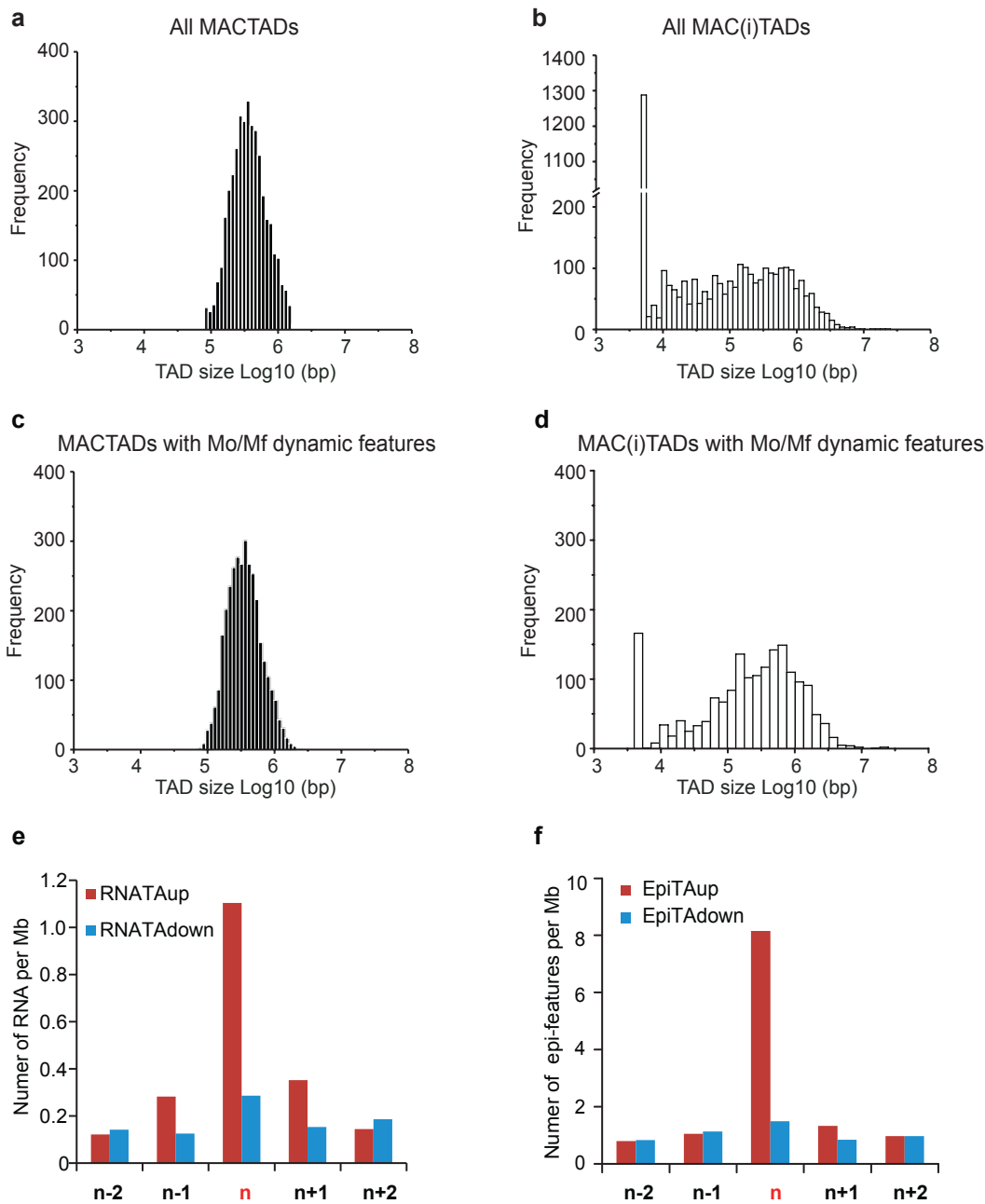


Figure S6

(a) Histogram of the size-distribution of all MACTADs (n=3,740). (b) Histogram of the size-distribution of all MAC(i)TADs (n=3,769). (c) Histogram of the size-distribution of all MACTADs that bear GR, dynamic genes and/or epigenetic features (n=3,318). (d) Histogram of the size-distribution of all MAC(i)TADs that bear GR, dynamic genes and/or epigenetic features. (n=1,750). (e) Density (features per Mb) of TA-up-, and TA-down-regulated transcripts in 249 GR peak-bearing TADs and the associated sets of four neighbouring TADs. (f) As (e) but for H3-bourne epigenetic features.

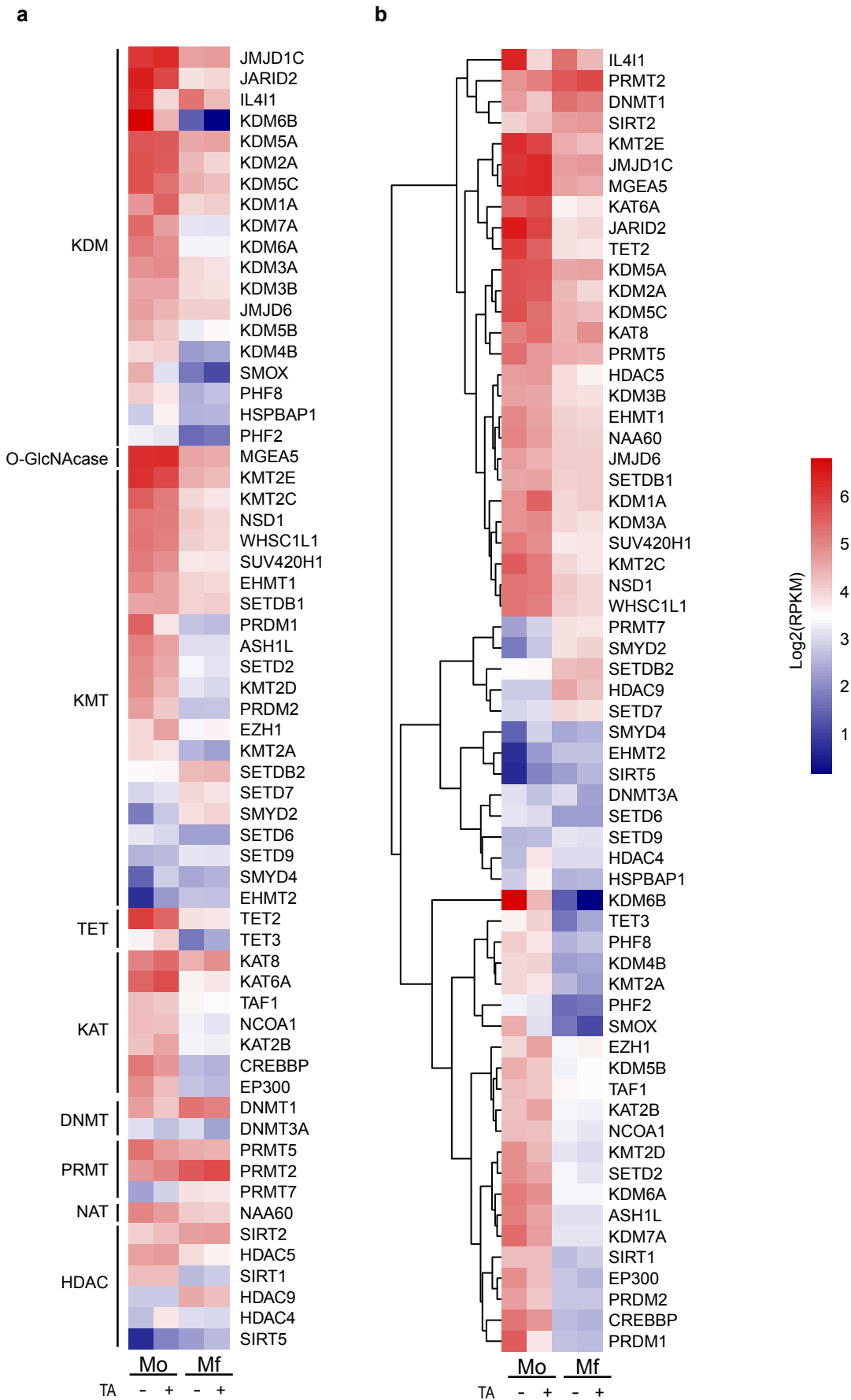


Figure S7

(a) Heat map clustered on the basis chromatin remodelling enzyme types ('writers' and 'erasers') that change expression at least two-fold as a function of Mo to Mf differentiation and/or TA-exposure. The colour scale indicates the level of expression (log₂ RPKM). (b) As (a), but clustered on expression dynamics rather than on enzyme type.

References

1. Jubb, A. W., Young, R. S., Hume, D. A. & Bickmore, W. A. Enhancer Turnover Is Associated with a Divergent Transcriptional Response to Glucocorticoid in Mouse and Human Macrophages. *J Immunol* **196**, 813-822, doi:10.4049/jimmunol.1502009 (2016).
2. Vockley, C. M. et al. Direct GR Binding Sites Potentiate Clusters of TF Binding across the Human Genome. *Cell* **166**, 1269-1281 e1219, doi:10.1016/j.cell.2016.07.049 (2016).