

***Atg7*-dependent canonical autophagy regulates the degradation of aquaporin 2 in prolonged hypokalemia**

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Supplementary Information

Supplementary Table S1. Effect of AQP2-specific deficiency of *Atg7* on kidney weight and biochemical parameters.

		<i>Atg7^{fl/fl}</i>		<i>Atg7^{ΔPC}</i>	
		Control	K ⁺ -depleted	Control	K ⁺ -depleted
	Body weight (g)	22.5 ± 0.5	19.5 ± 1.26**	24.5 ± 1.63**	23.3 ± 1.31 ⁺
Weight	Kidney weight (g)	0.16 ± 0.01	0.15 ± 0.02	0.17 ± 0.02	0.19 ± 0.02**/+
	KW/BW (%)	100 ± 8.43	115.15 ± 10.10*	108.65 ± 1.38	123.88 ± 8.05*/#
Blood	Plasma Na ⁺ (mmol/L)	150.2 ± 1.8	147.4 ± 2.1	150.2 ± 2.0	153.2 ± 2.1 ^{*/+/#}
	Plasma K ⁺ (mmol/L)	3.89 ± 0.32	2.97 ± 0.26**	4.27 ± 0.30	2.28 ± 0.25**/+/#
	Plasma Cl ⁻ (mmol/L)	112.8 ± 2.4	113.3 ± 1.5	114.2 ± 3.8	119.5 ± 4.1 ^{*/+/#}
	BUN (mg/dL)	27.2 ± 5.9	30.8 ± 4.49	27.0 ± 2.65	47.00 ± 7.79 ^{*/+/#}
	Urine volume (ml)	0.56 ± 0.37	2.48 ± 1.36**	2.37 ± 0.67**	3.02 ± 0.81 ^{**/+/#}
	Osmolality (mosmol/kgH ₂ O)	2244.1 ± 421	1168.1 ± 418**	1525.7 ± 401**	974 ± 185 ^{**/+/#}
Urine	Urine Na ⁺ (mmol/L)	70.7 ± 18.0	38.1 ± 16.8*	52.1 ± 20.0*	26.9 ± 6.2 ^{*/+/#}
	Urine K ⁺ (mmol/L)	137.6 ± 16.3	13.7 ± 6.0**	105.2 ± 32.2**	15.3 ± 3.7 ^{**/#}
	Urine Cl ⁻ (mmol/L)	132.1 ± 31.1	61.3 ± 24.9*	95.9 ± 33.2*	67.9 ± 18.0 ^{*/+/#}

t-test vs. *Atg7^{fl/fl}* Control * *P* < 0.05, ** *P* < 0.001

t-test vs. *Atg7^{fl/fl}* K⁺-depleted + *P* < 0.05, ++ *P* < 0.001

t-test vs. *Atg7^{ΔPC}* Control # *P* < 0.05, ## *P* < 0.001

All values are the means ± SEM.

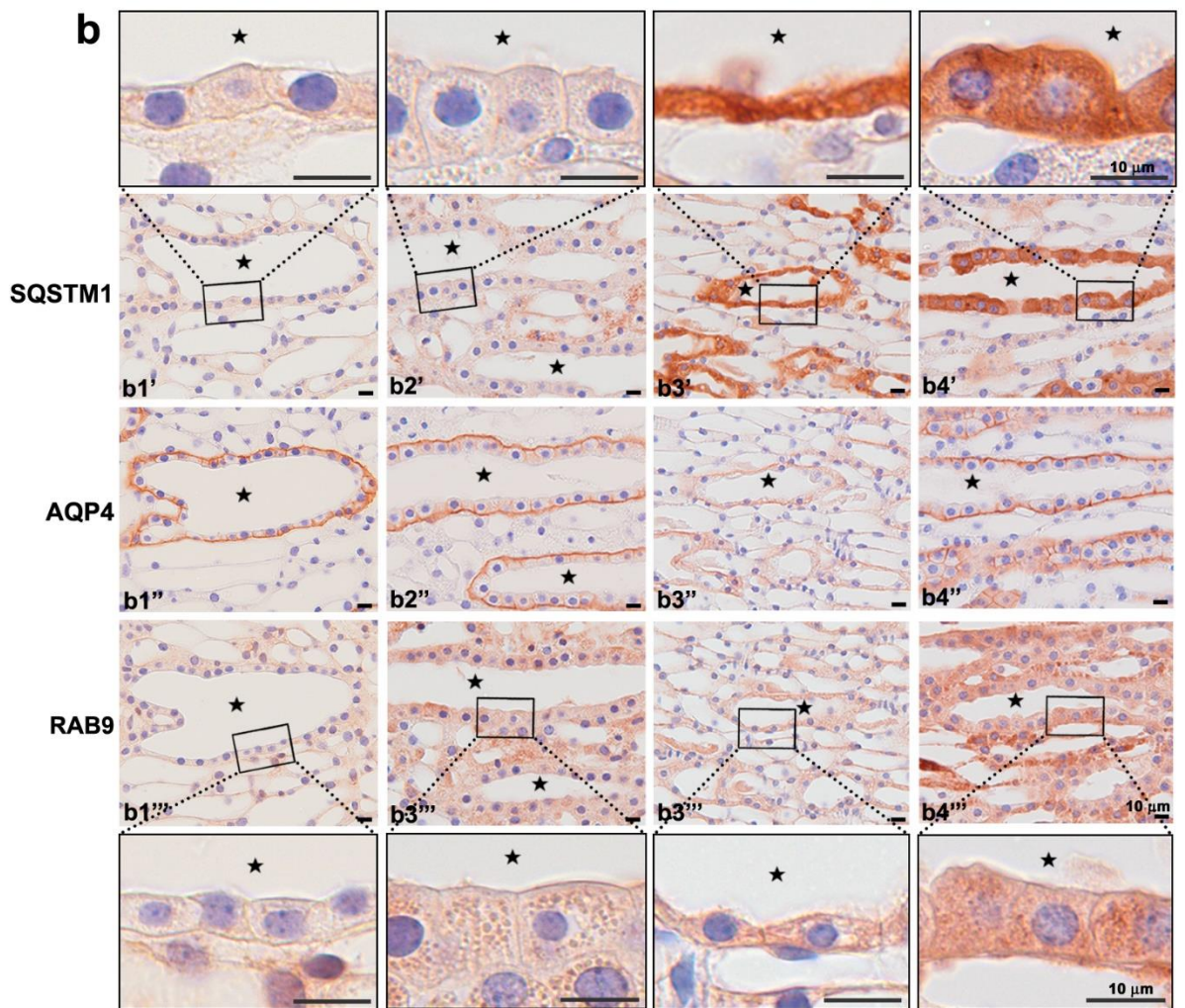
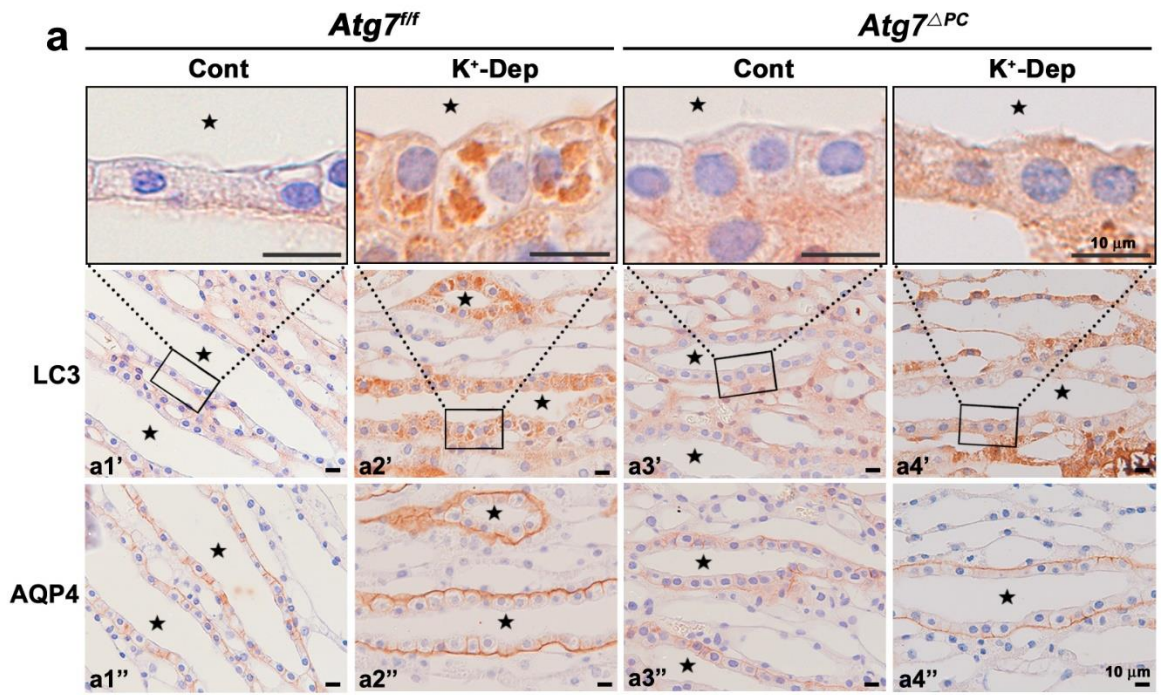


Figure S1. Light micrographs of the inner medulla of control (Cont) and K⁺-depleted (K⁺-Dep) *Atg7^{f/f}* and *Atg7^{ΔPC}* mouse kidneys illustrating immunolabeling of LC3 (**a1'-a4'**), SQSTM1 (**b1'-b4'**) and RAB9 (**b1'''-b4'''**). Inner medullary collecting ducts (IMCDs) (stars) were identified by immunolabeling on the basolateral plasma membrane for AQP4 using consecutive sections (**a1''-a4''**) and (**b1''-b4''**). **(a)** In the images, it can be observed that the intense immunoreactivity for LC3 in K⁺-Dep *Atg7^{f/f}* mice is markedly and restrictively decreased in the AQP4-positive IMCD cells in K⁺-Dep *Atg7^{ΔPC}* mice. **(b)** Strong immunoreactivity for SQSTM1 is observed restrictively in the AQP4-positive IMCD cells of *Atg7^{ΔPC}* mice. Note the prominent increase of RAB9 in K⁺-Dep *Atg7^{ΔPC}* mice. Boxes in **(a, b)** are higher magnification of the areas indicated by the rectangles in upper or lower panels.

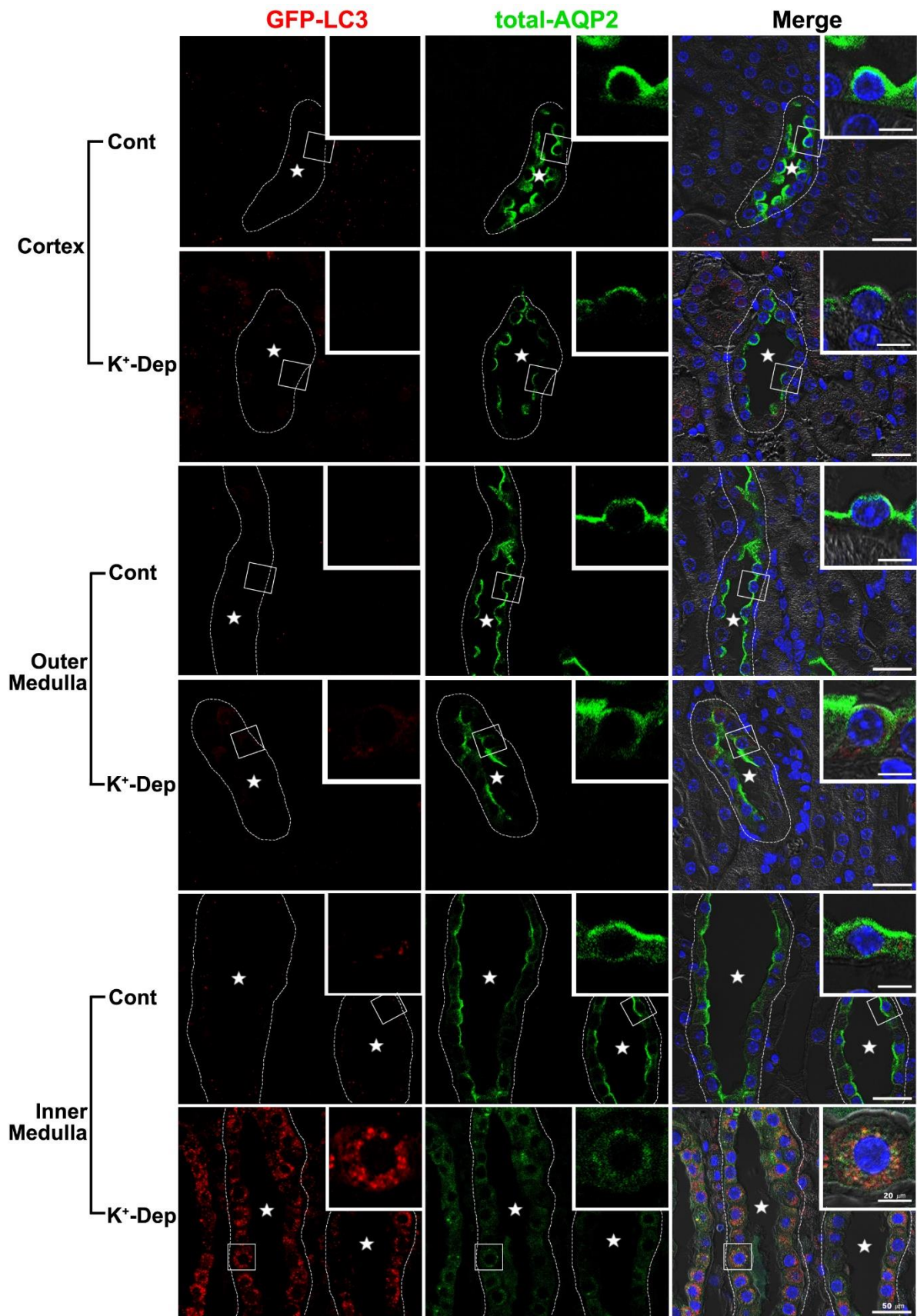


Figure S2. Confocal micrographs of the cortex, outer and inner medulla of control (Cont) and

K⁺-depleted (K⁺-Dep) GFP-LC3 transgenic mice illustrating double labeling with GFP-LC3 (red) and total-AQP2 (green). After potassium depletion for 2 weeks, GFP-LC3-positive dots are markedly accumulated in the inner medullary collecting duct (IMCD) cells, but a few in both cortical and outer medullary collecting duct cells. Note that total-AQP2 is colocalized with these GFP-LC3-positive puncta in the IMCD cells. Stars indicate the lumen of collecting ducts.

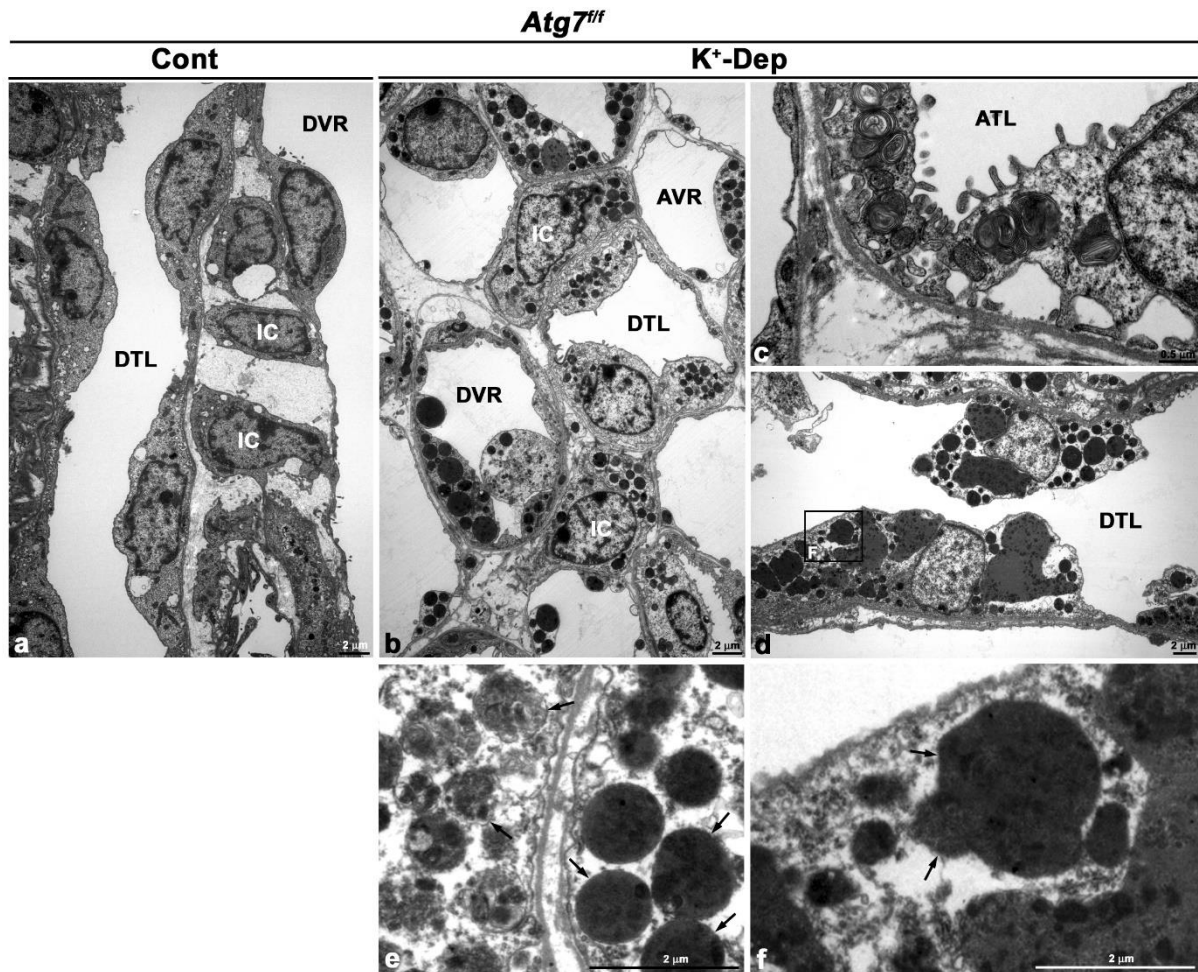


Figure S3. Transmission electron micrographs of the inner medulla of *Atg7^{f/f}* mouse kidneys. **(a)** On normal diet (Cont), no autophagic vacuoles are observed in the cytoplasm of interstitial cell (IC), descending thin limb (DTL), or descending vasa recta (DVR). **(b-f)** After K⁺-depletion (K⁺-Dep), the accumulation of numerous autophagic vacuoles containing multilamellar bodies or degradation materials can be observed in the IC, DTL, ascending thin limb (ATL), DVR, and ascending vasa recta (AVR). **(f)** Higher magnifications of the areas outlined in **(d)**. A single membrane (arrows) can be observed surrounding various autophagic vacuoles.

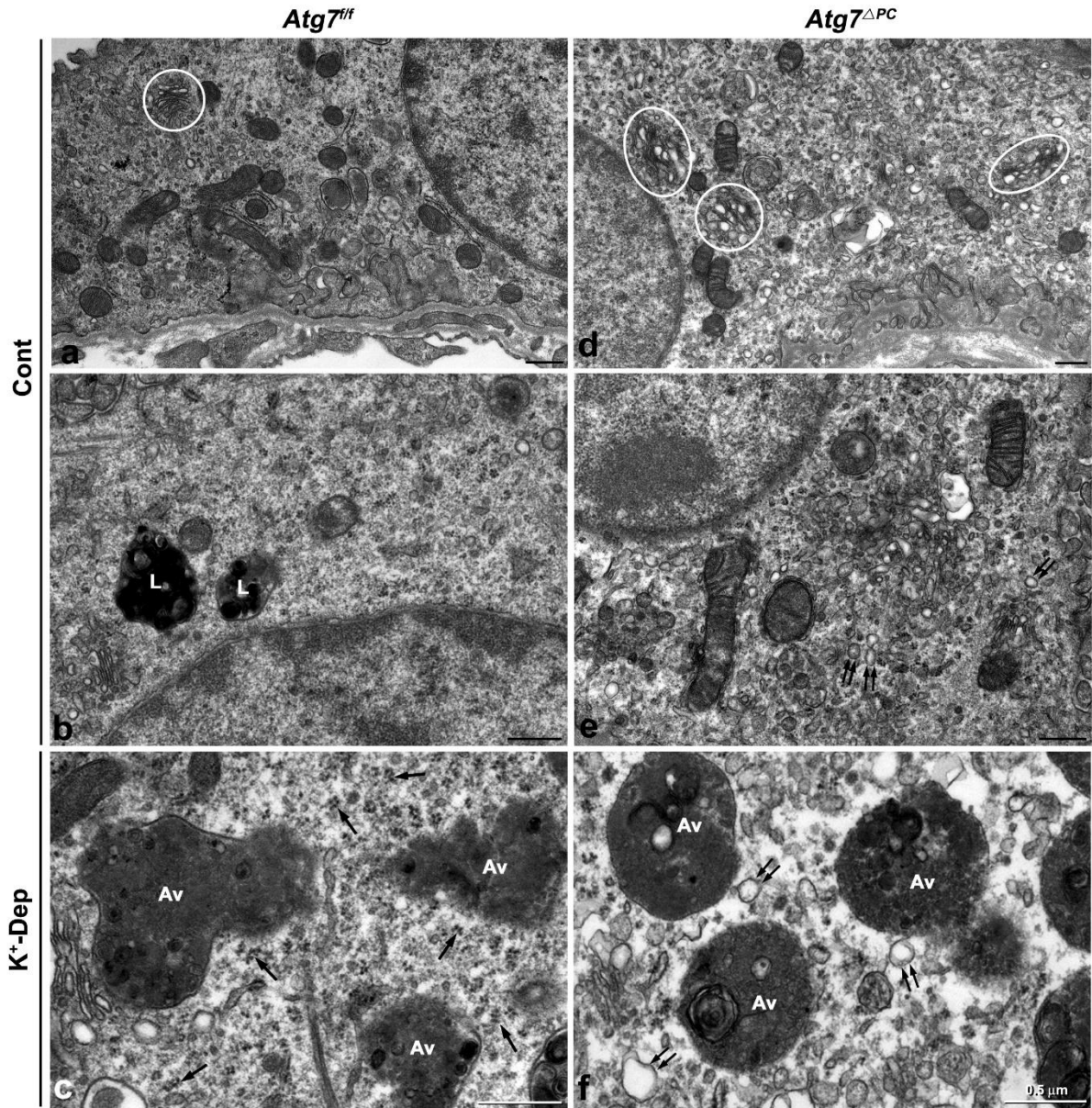


Figure S4. Transmission electron micrographs of inner medullary collecting duct (IMCD) cells. **(a-c)** In *Atg7^{fl/fl}* mice, polysomes (arrows) are markedly increased throughout the cytoplasm in IMCD cells, as are autophagic vacuoles (Av) after K⁺-depletion (K⁺-Dep). **(d-f)** In *Atg7^{ΔPC}* mice, in contrast, Golgi complexes (circles) and small tubuloalveolar shaped vesicles (double arrows) are markedly increased in both control (Cont) and K⁺-Dep groups.

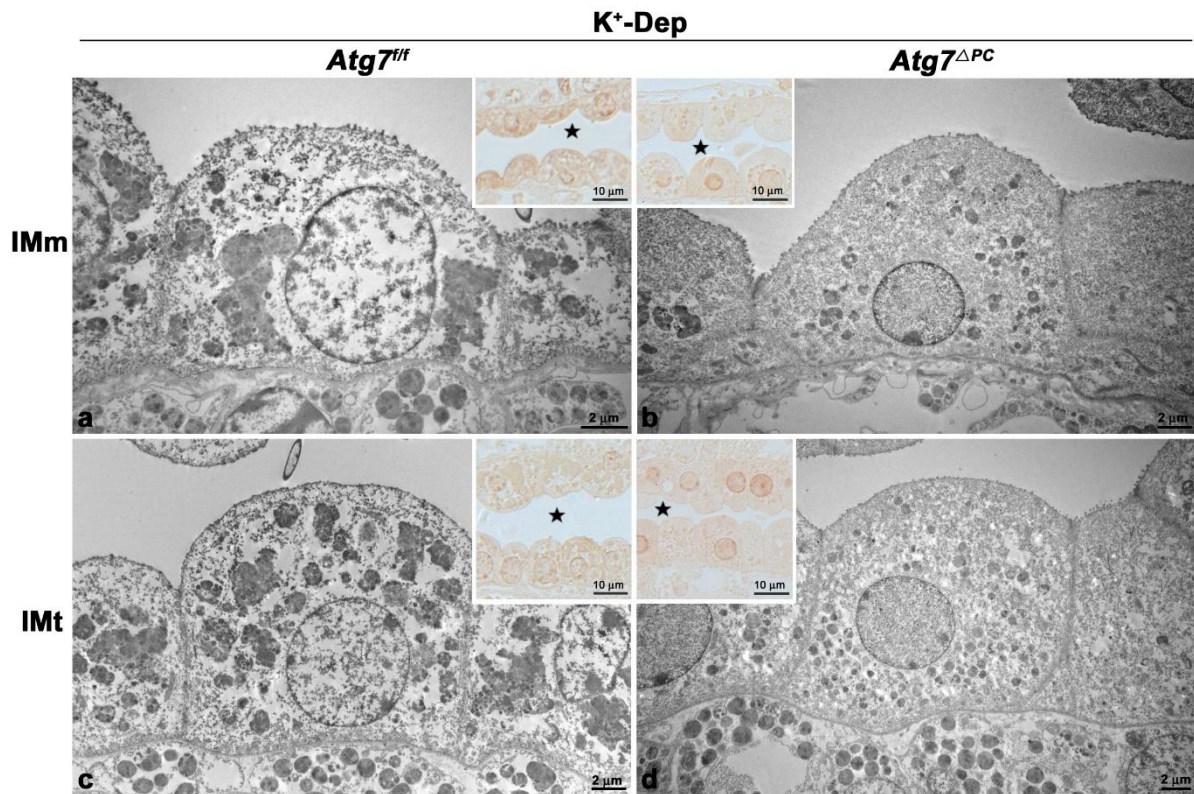


Figure S5. Immunoelectron micrographs of the middle (IMm). **(a, b)** and the terminal (IMt). **(c, d)** parts of the inner medulla of K⁺ depleted (K⁺-Dep) *Atg7^{f/f}*. **(a, c)** and *Atg7^{ΔPC}* **(b, d)** mice illustrating immunostaining for pS256-AQP2. The inserts are light micrographs of 1 μm-thick semi-thin sections of the same group. No immunoreactivity of pS256-AQP2 is observed in either the canonical autophagic vacuoles in K⁺-Dep *Atg7^{f/f}* mice **a, c** or the non-canonical autophagic vacuoles in K⁺-Dep *Atg7^{ΔPC}* mice **(b, d)**. Stars indicate the lumen of inner medullary collecting ducts **(a-d)**.

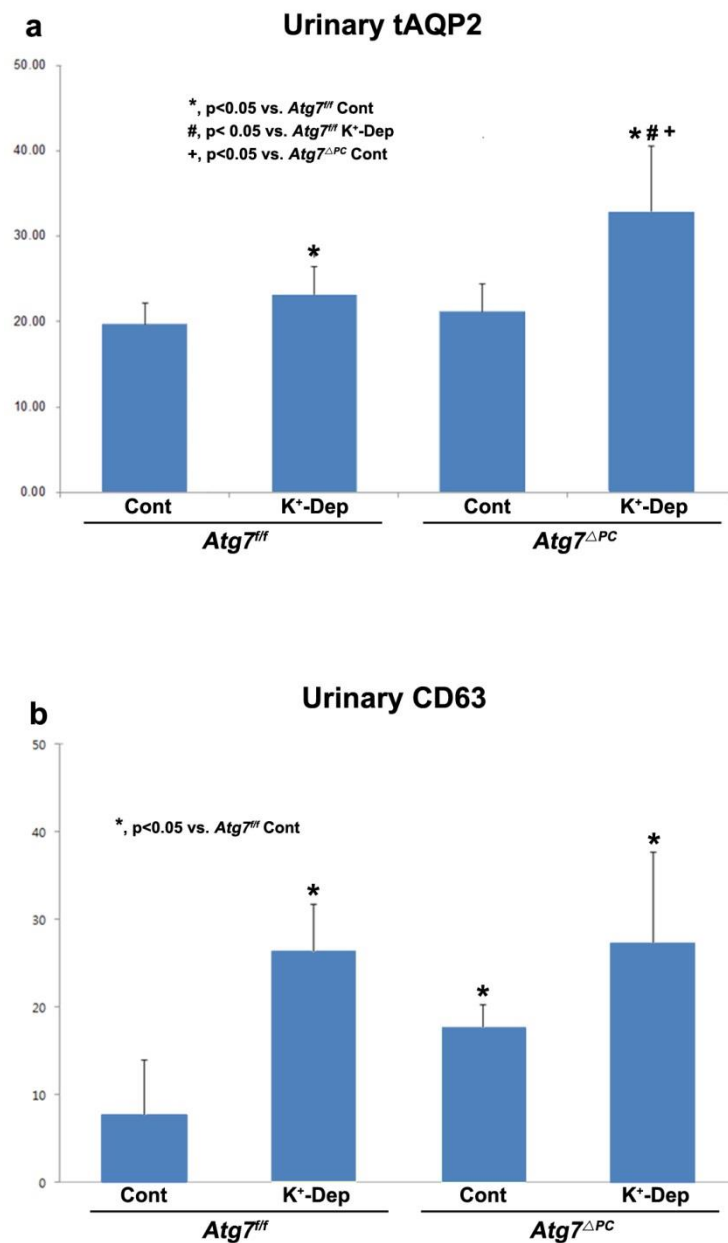


Figure S6. Enzyme-linked immunosorbent assay of urinary total-AQP2 (a) and CD63 (b). Note that urinary excretion of total-AQP2 and CD63 are significantly increased after K⁺ depletion in both *Atg7^{fl/fl}* mice and *Atg7^{ΔPC}* mice compare to those of control mice.