

A micropatterning platform for quantifying interaction kinetics between the T cell receptor and an intracellular binding protein

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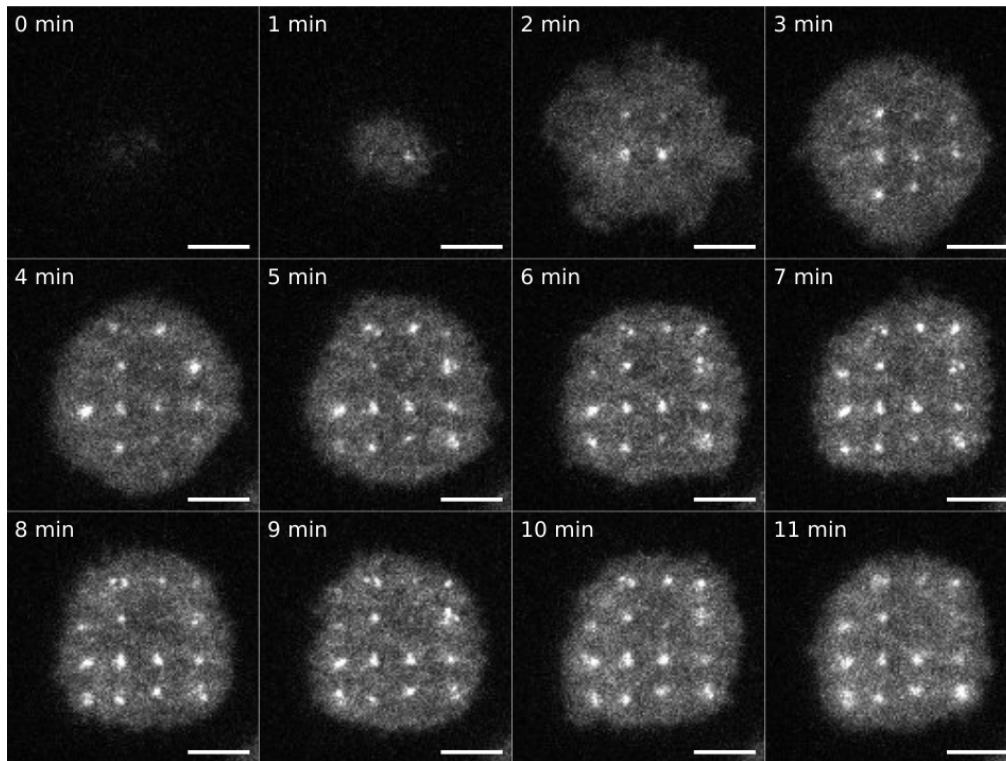


Figure S1. Time course of ZAP70-GFP recruitment to OKT3 sites. Cells were seeded onto a micropatterned OKT3 surface at $t = 0$ min. Cell attachment and formation of ZAP70-GFP patterns over time is shown for a representative cell. Scale bar is 5 μm .

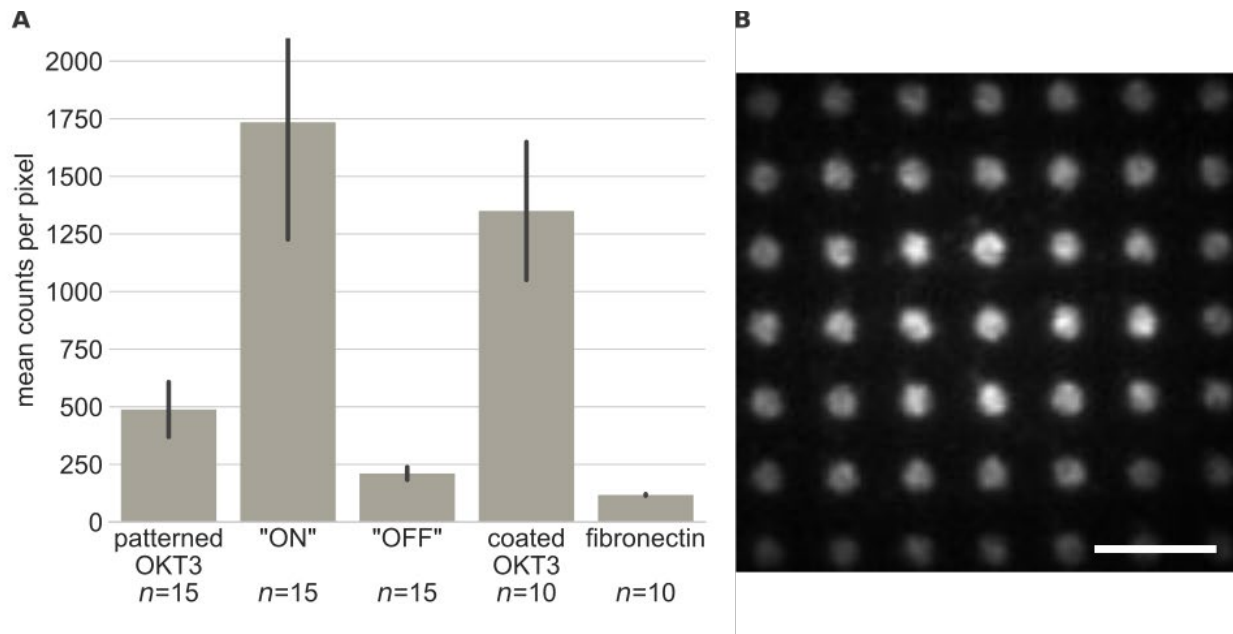


Figure S2. Relative OKT3 antibody densities on patterned and homogeneously coated OKT3 surfaces. (A) OKT3 antibodies were labeled with Zenon™ Alexa Fluor™ 647 Mouse IgG2a Fab fragments. Relative antibody densities are given by the fluorescence intensities. Data are from three independent experiments (average \pm SEM); the number of measured areas is indicated as *n*. **(B)** Image of a patterned OKT3 surface labeled with Zenon™ Fab fragments. Scale bar is 5 μ m.

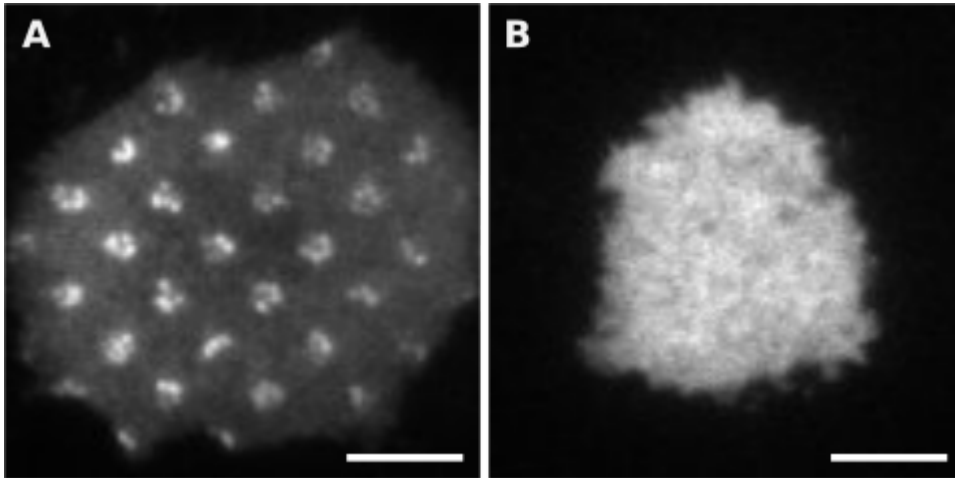


Fig S3: Specific ZAP70-GFP recruitment to aCD3 “ON” areas. Cells expressing ZAP70-GFP were seeded onto micropatterned surfaces featuring antibodies against **(A)** the CD3 ϵ epitope MEM-57 and **(B)** an HA-tag and allowed to adhere for 10 minutes before an image of the ZAP70-GFP distribution was recorded.

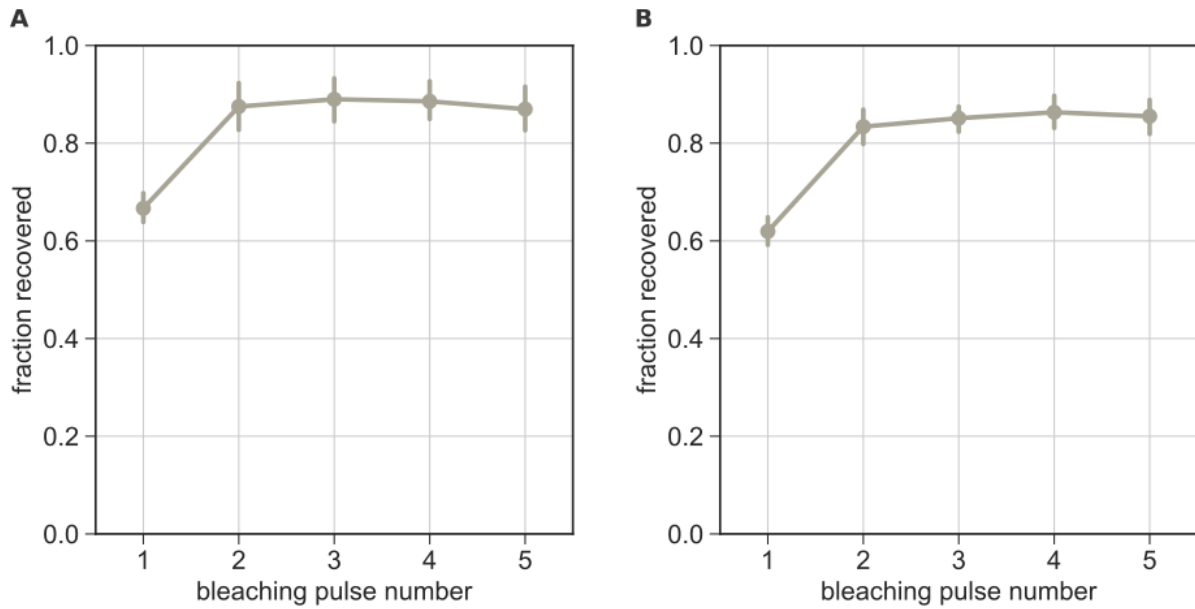


Figure S4. Recovering fraction of Zap70-GFP after repetitive bleach pulses. Images were recorded 1s after the end of a 500ms bleach pulse and normalized to the corresponding pre-bleach image. Data are from two independent experiments with 50 **(A)** and 21 **(B)** different cells (average + 95% confidence interval). The short time interval between bleach pulses does not allow for the recovery of the slower fraction of ZAP70-GFP. The recovering fraction increases with successive FRAP experiments to level out at ~88% **(A)** and ~85% **(B)**, indicating that the depletion of the cytosolic pool of ZAP70-GFP caused by the bleach pulse amounts to ~12% and ~15% at 22°C and 37°C, respectively.

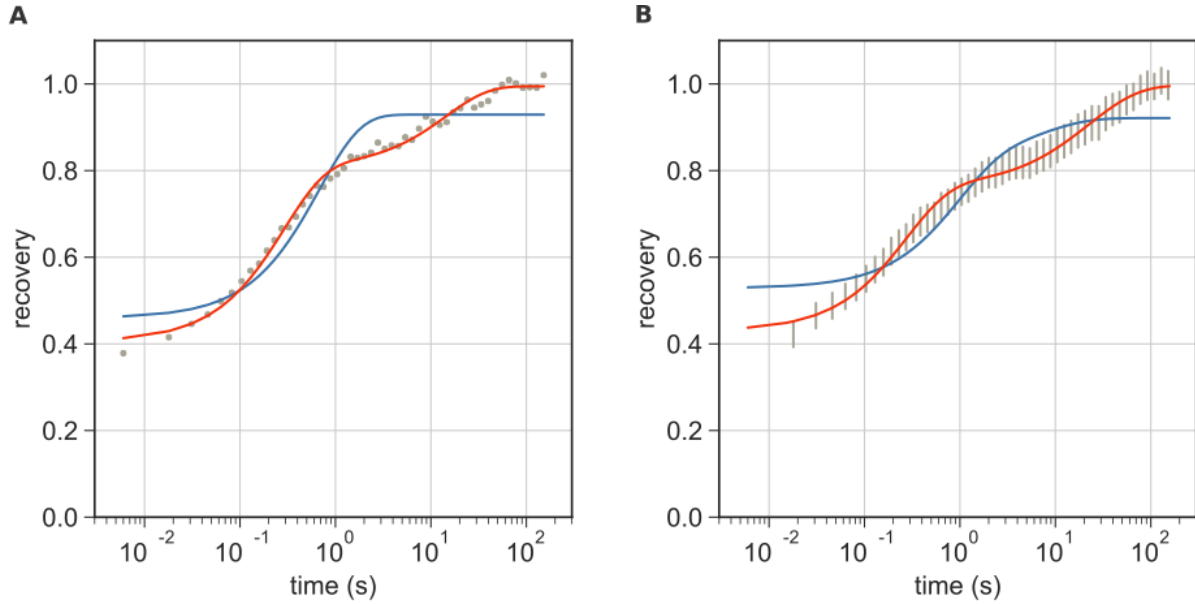


Figure S5. FRAP in cells expressing cytosolic GFP on a fibronectin-coated surface. (A) A representative FRAP curve of cytosolic GFP measured in a cell on a fibronectin-coated surface. A mono-exponential fit (Eq. 1) is shown in blue. A bi-exponential fit (Eq. 2, shown in red) yielded $\tau_1 = 0.29\text{s}$, $\tau_2 = 14.46\text{s}$, $f_m = 0.99$, $b = 0.40$ and $c = 0.68$. **(B)** Averaged FRAP curves of cytosolic GFP measured in cells on a fibronectin-coated surface. Data are from three independent experiments with 29 different cells. The 95% confidence interval is shown. Mean values \pm SE from cells expressing cytosolic GFP fitted individually with Eq. 2 were: $\tau_1 = 0.28 \pm 0.02\text{s}$, $\tau_2 = 25.4 \pm 2.4\text{s}$, $f_m = 1.00 \pm 0.01$, $b = 0.43 \pm 0.01$ and $c = 0.59 \pm 0.02$. The average of all fit curves is shown in red.

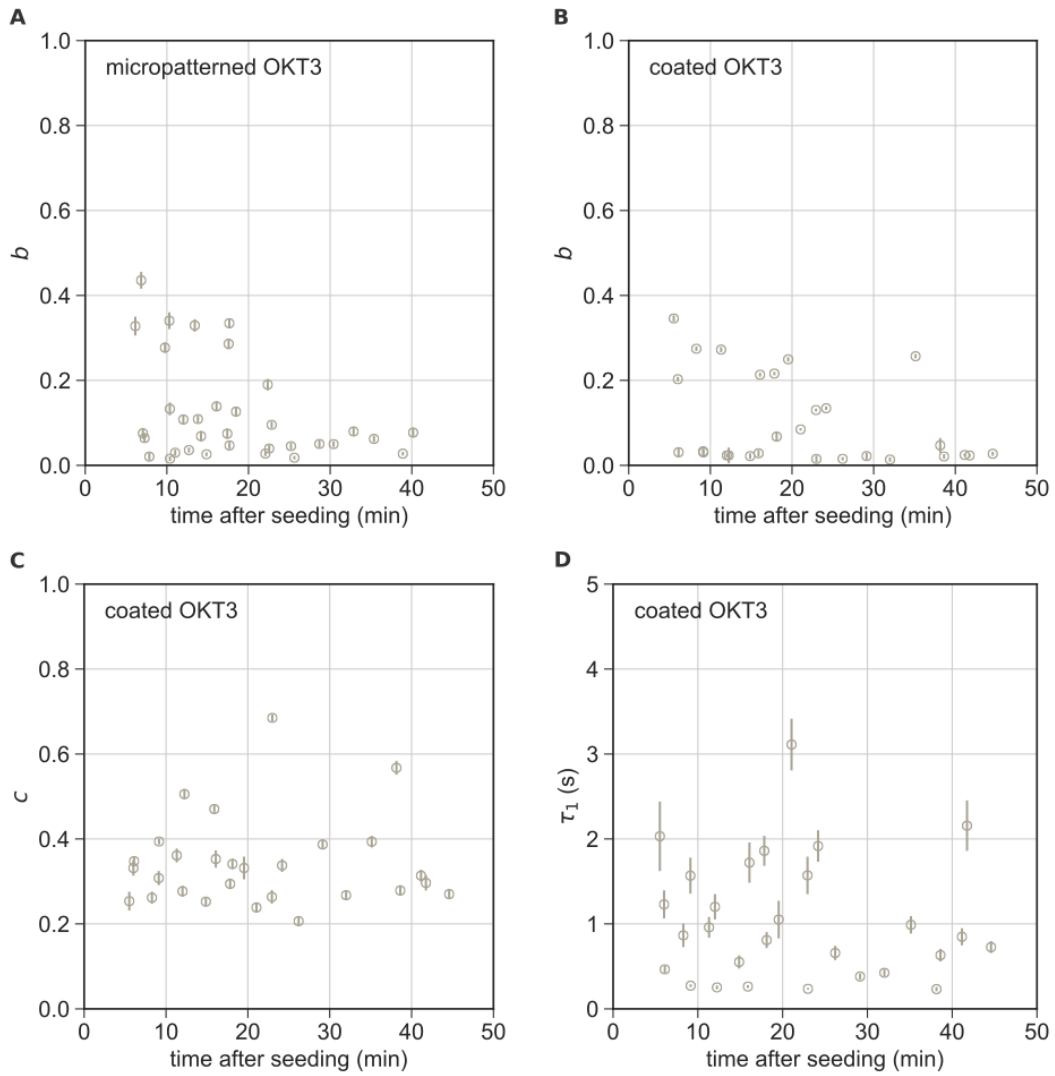


Figure S6. Comparison of FRAP parameters for cells on micropatterned and coated OKT3 surfaces at 22°C. The fitting parameters b , c and τ_1 are plotted over time after seeding for cells on micropatterned (N=34, 3 independent experiments) **(A)** and homogeneously coated (N=28, 4 independent experiments) **(B-D)** OKT3 surfaces. Error bars show the SE of the fit.

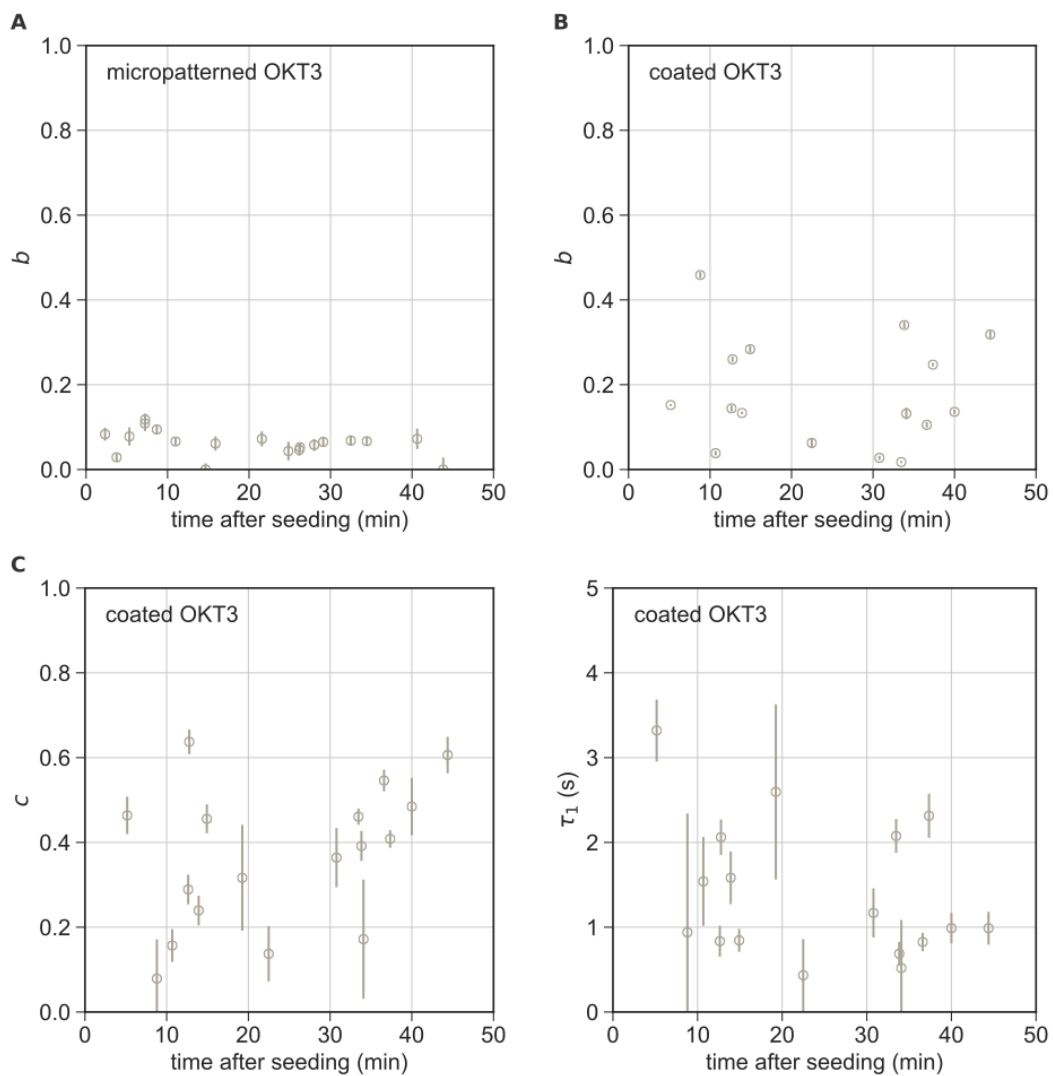


Figure S7. Comparison of FRAP parameters for cells on micropatterned and coated OKT3 surfaces at 37°C. The fitting parameters b , c and τ_1 are plotted over time after seeding for cells on micropatterned (N=19, 2 independent experiments) **(A)** and homogeneously coated (N=17, 2 independent experiments) **(B-D)** OKT3 surfaces. Error bars show the SE of the fit.