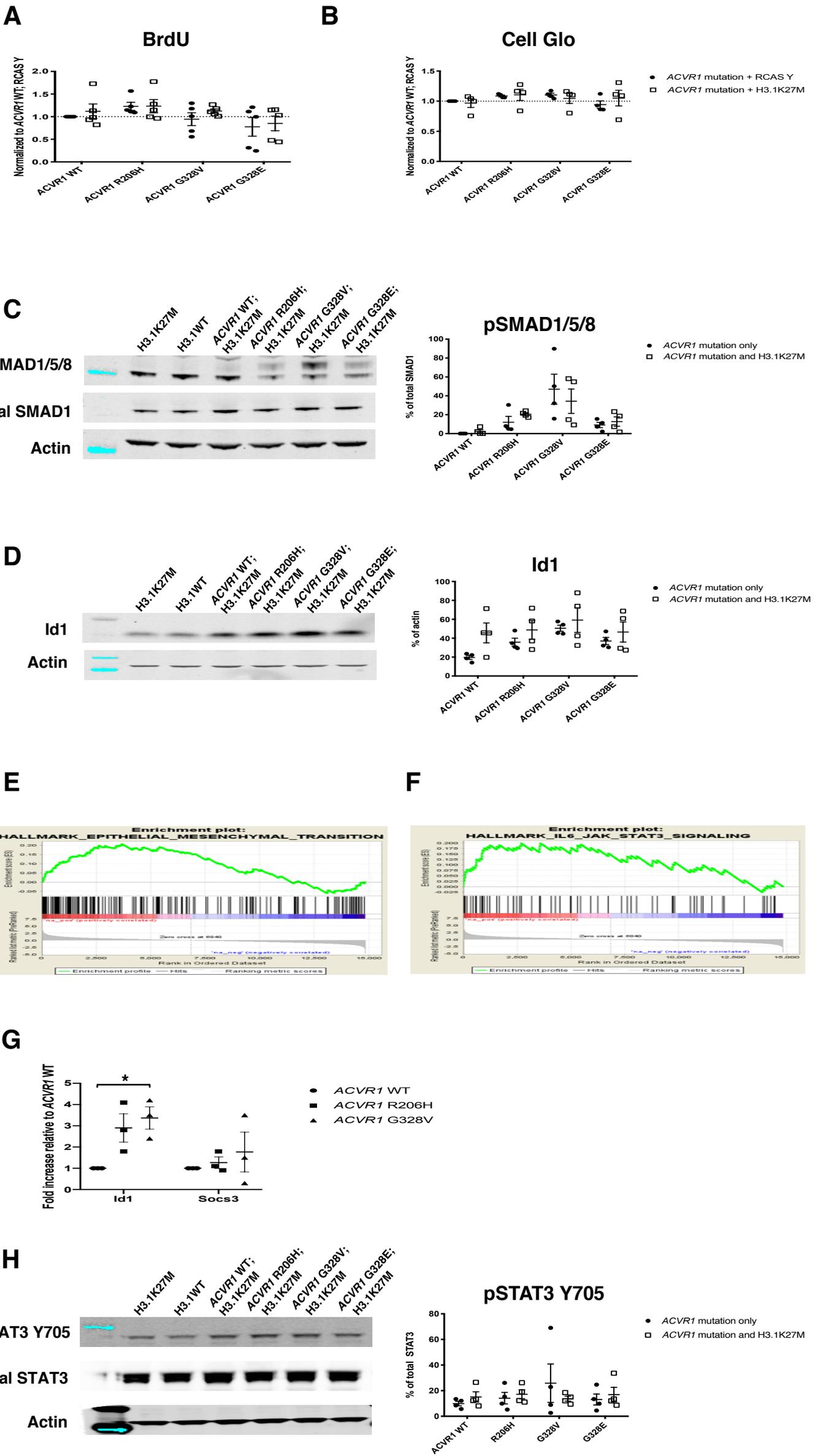


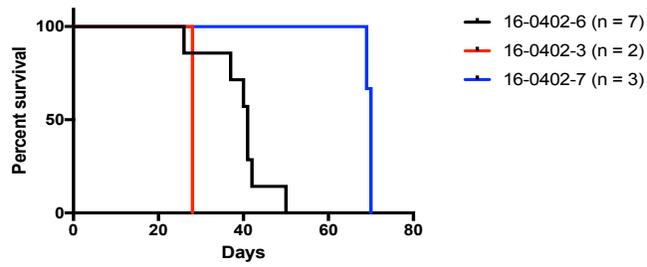
Supplementary Information

***ACVR1* R206H cooperates with H3.1K27M in promoting Diffuse Intrinsic Pontine Glioma pathogenesis**

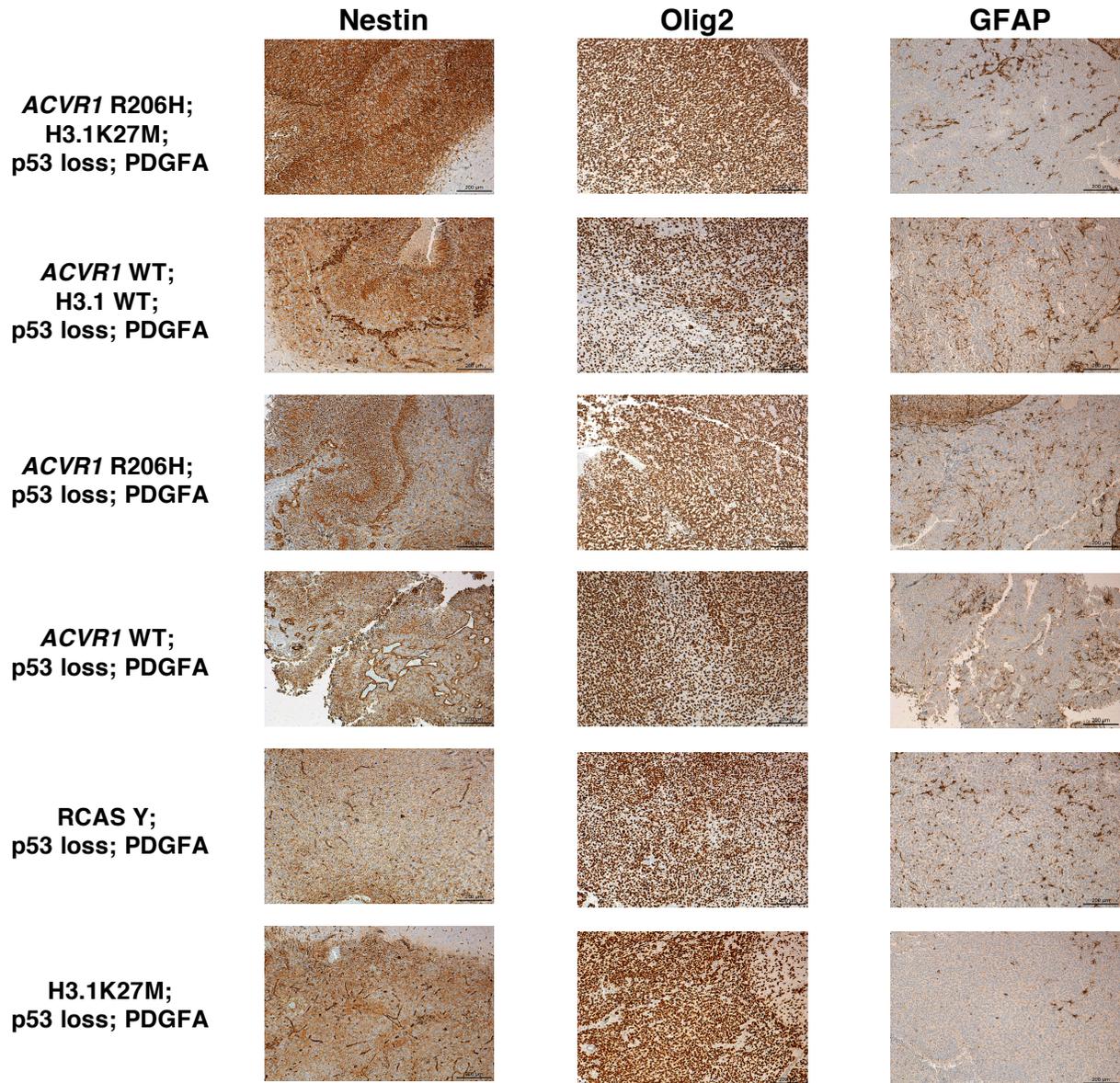
Hoeman et al.



Supplementary Figure 1. ACVR1 mutations and H3.1K27M do not have an additive effect on proliferation, cell survival, BMP, or STAT3 signaling *in vitro*. Brainstem progenitor cells isolated from postnatal day 3 Ntv-a;p53^{fl/fl} pups were cultured as neurospheres and infected with RCAS viruses *in vitro* as indicated. **(a and b)** Proliferation **(a)** and cell viability **(b)** of neurospheres infected with ACVR1 and H3.1K27M mutation viruses (white squares) compared to control neurospheres infected with ACVR1 mutations and RCAS Y (black circles), (n = 4). **(c and d)** Western blot analysis of pSMAD1/5/8 **(c)** and Id1 **(d)** of neurospheres infected with ACVR1 and H3.1K27M mutation viruses (white squares) compared to neurospheres infected with ACVR1 mutations only (black circles, represented in Fig. 1c and d), (n = 4). **(e)** GSEA analysis of Epithelial to Mesenchymal Transition genes as identified by RNA-Seq analysis (see Supplementary Data 1) in neurospheres infected with ACVR1 WT or ACVR1 R206H. Also see Supplementary Data 2. **(f)** GSEA analysis of IL6_JAK_STAT3_signaling pathway as identified by RNA-Seq analysis. Also see Supplementary Data 2. **(g)** qRT-PCR analysis of Id1 and Socs3 in ACVR1 WT, ACVR1 R206H, or ACVR1 G328V infected neurospheres (n = 3), * p = 0.05. **(h)** Western blot analysis of pSTAT3 Y705 of neurospheres infected with ACVR1 and H3.1K27M mutation viruses (white squares) compared to neurospheres infected with ACVR1 mutations only (black circles, represented in Fig. 1i), (n = 4). All data are represented as mean with SEM and analyzed using paired t tests.

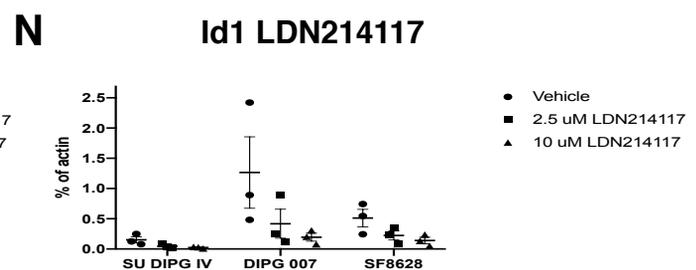
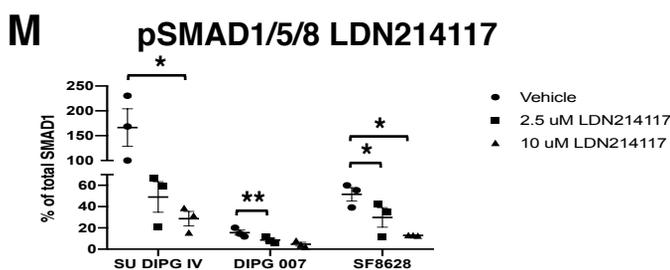
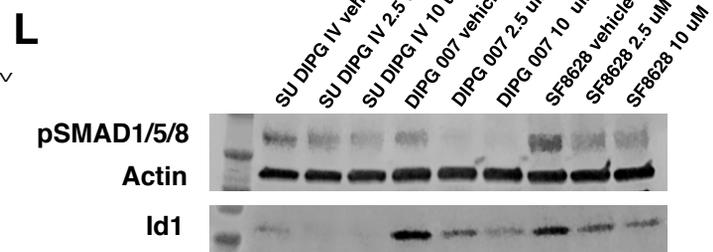
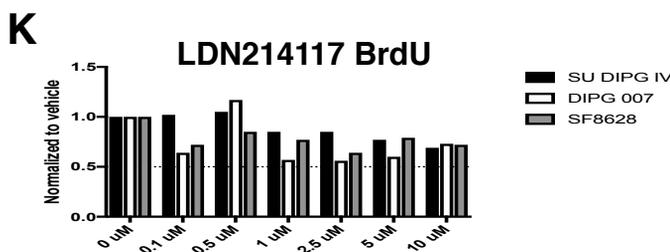
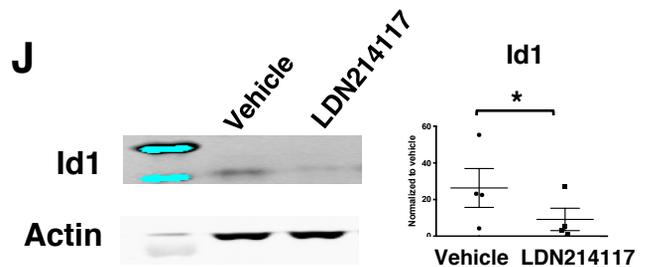
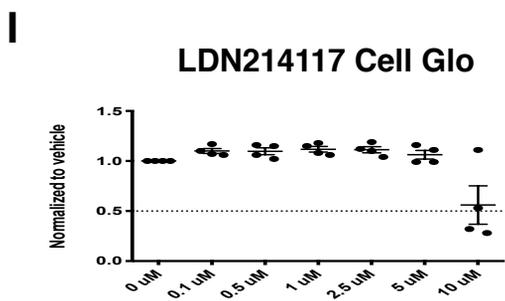
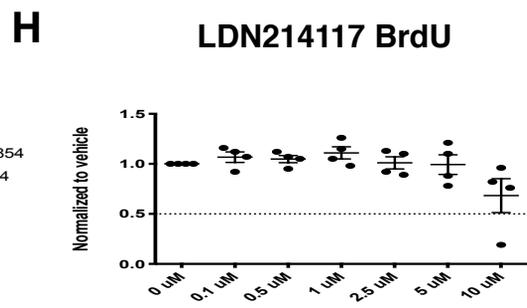
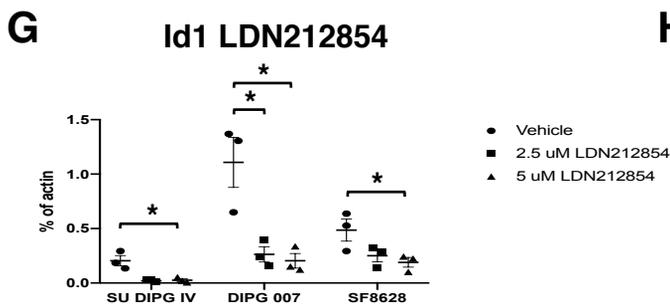
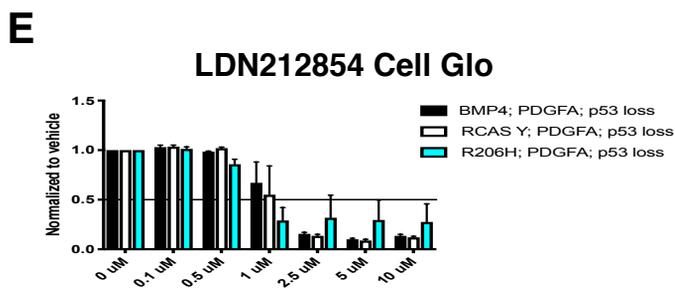
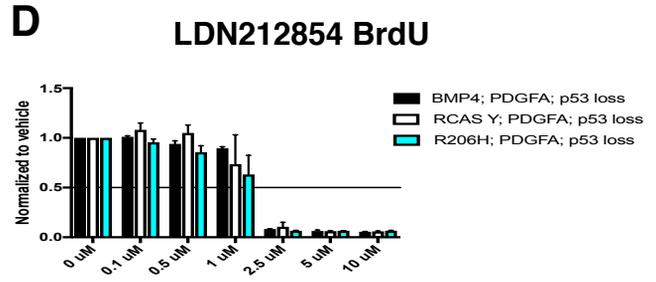
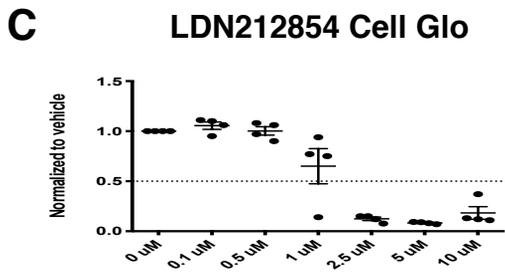
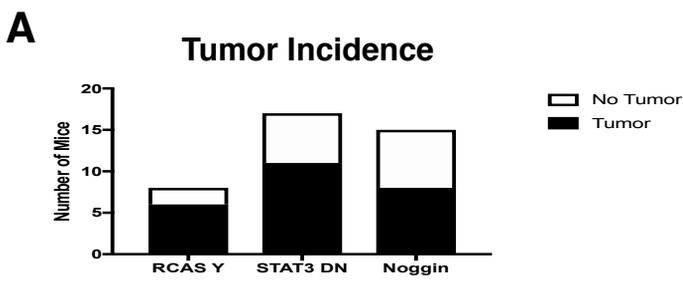


Supplementary Figure 2. *ACVR1* R206H and H3.1K27M are able to generate secondary tumors *in vivo*. Survival curve of Ntv-a;p53^{fl/fl} mice that were implanted with neurospheres derived from *ACVR1* R206H, PDGFA, H3.1K27M, and Cre primary tumors (n = 3 biologically different neurosphere lines).



Supplementary Figure 3. Nestin, Olig2, and GFAP expression are not changed by *ACVR1*

R206H. Representative IHC images of Nestin, Olig2, and GFAP from various PDGFA tumors. 10x magnification, scale bar = 200 μ M.



Supplementary Figure 4. ACVR1 inhibitor LDN214117 does not inhibit proliferation or cell

viability *in vitro*. (a) Tumor incidence of mice that were injected with RCAS-PDGFA, RCAS-Cre,

RCAS-ACVR1 R206H, and RCAS-Y (6/8 tumors = 75%) or RCAS-STAT3 DN (11/17 tumors = 65%)

or RCAS-Noggin (8/15 tumors = 53%), Fisher's exact test. (b) Tumor grade from mice injected in

(a), Fisher's exact test. (c) Tumor derived neurospheres from Nestin tv-a; p53^{fl/fl} mice injected

with RCAS-ACVR1 R206H, RCAS-H3.1K27M, RCAS-Cre, and RCAS-PDGFA were treated ACVR1

inhibitor LDN212854 for 24 hours and assessed for cell viability (n = 4). (d and e) All Ntv-a;p53^{fl/fl}

mice were injected with RCAS-PDGFA and RCAS-Cre along with additional RCAS viruses as

indicated. Proliferation (d) and cell viability (e) of neurospheres incubated with LDN212854 for

24 hours (BMP4 lines n = 2, RCAS Y lines n = 2, and ACVR1 R206H lines n = 4). (f and g) Western

blot quantification of pSMAD1/5/8 (f) and Id1 (g) of human lines treated with LDN212854 from

Fig. 6h (n = 3 independent experiments), * p < 0.05, paired t test. (h-j) Neurospheres were

derived from primary tumors of RCAS-ACVR1 R206H, RCAS-H3.1K27M, RCAS-Cre, and RCAS-

PDGFA injected Nestin tv-a;p53^{fl/fl} mice. Proliferation (h) and cell viability (i) of neurospheres

treated with LDN214117 for 24 hours (n = 4). (j) Western blot analysis of Id1 expression of

LDN214117 treated neurospheres versus vehicle neurospheres (n = 4), * p < 0.05, paired t test.

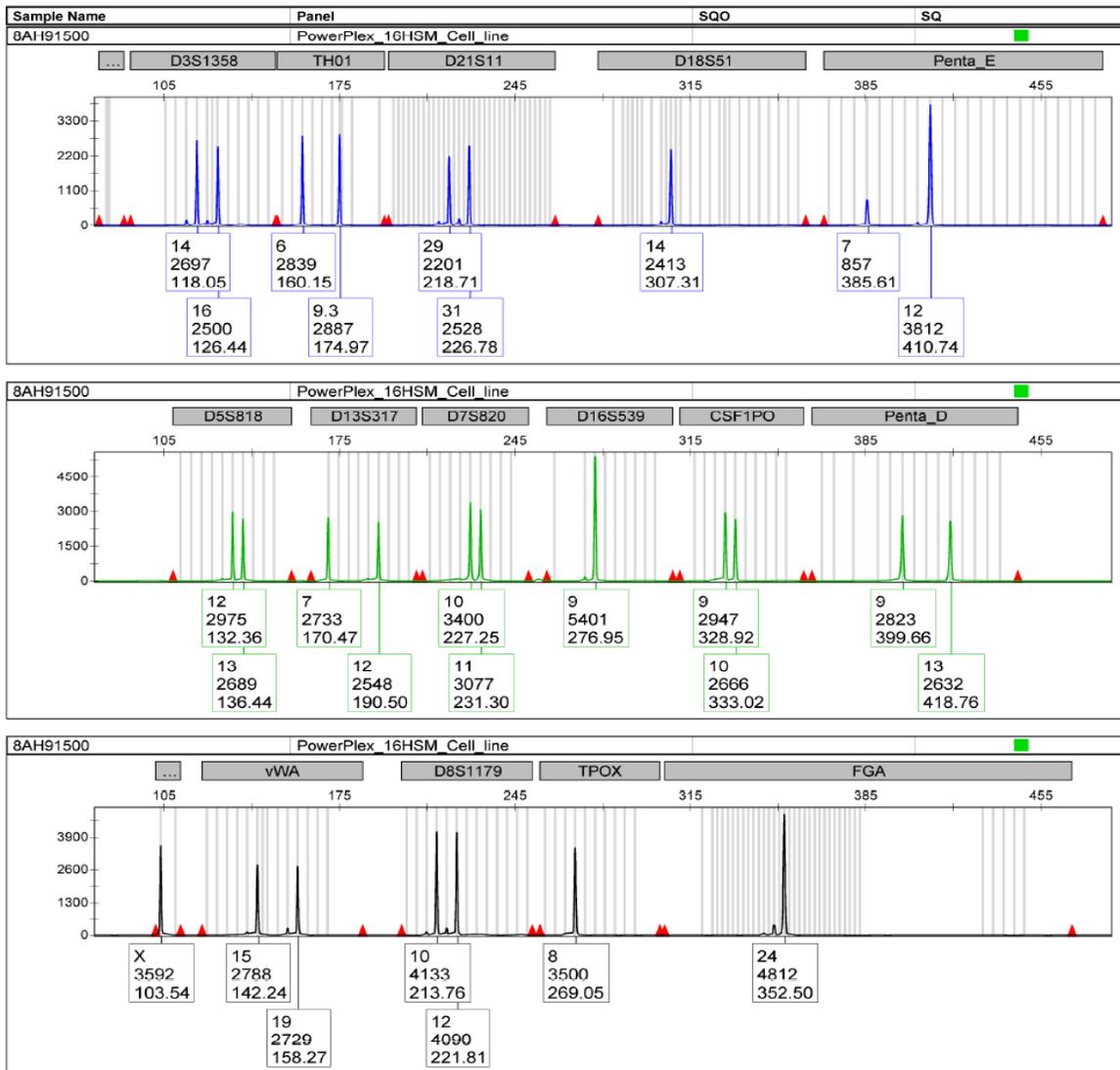
(k-n) ACVR1 mutant and wildtype human DIPG lines were treated with ACVR1 inhibitor

LDN214117 for 24 hours. (k) Proliferation of LDN214117 treated human lines (n = 3

independent experiments, triplicate wells). (l-n) Western blot analysis and quantification of

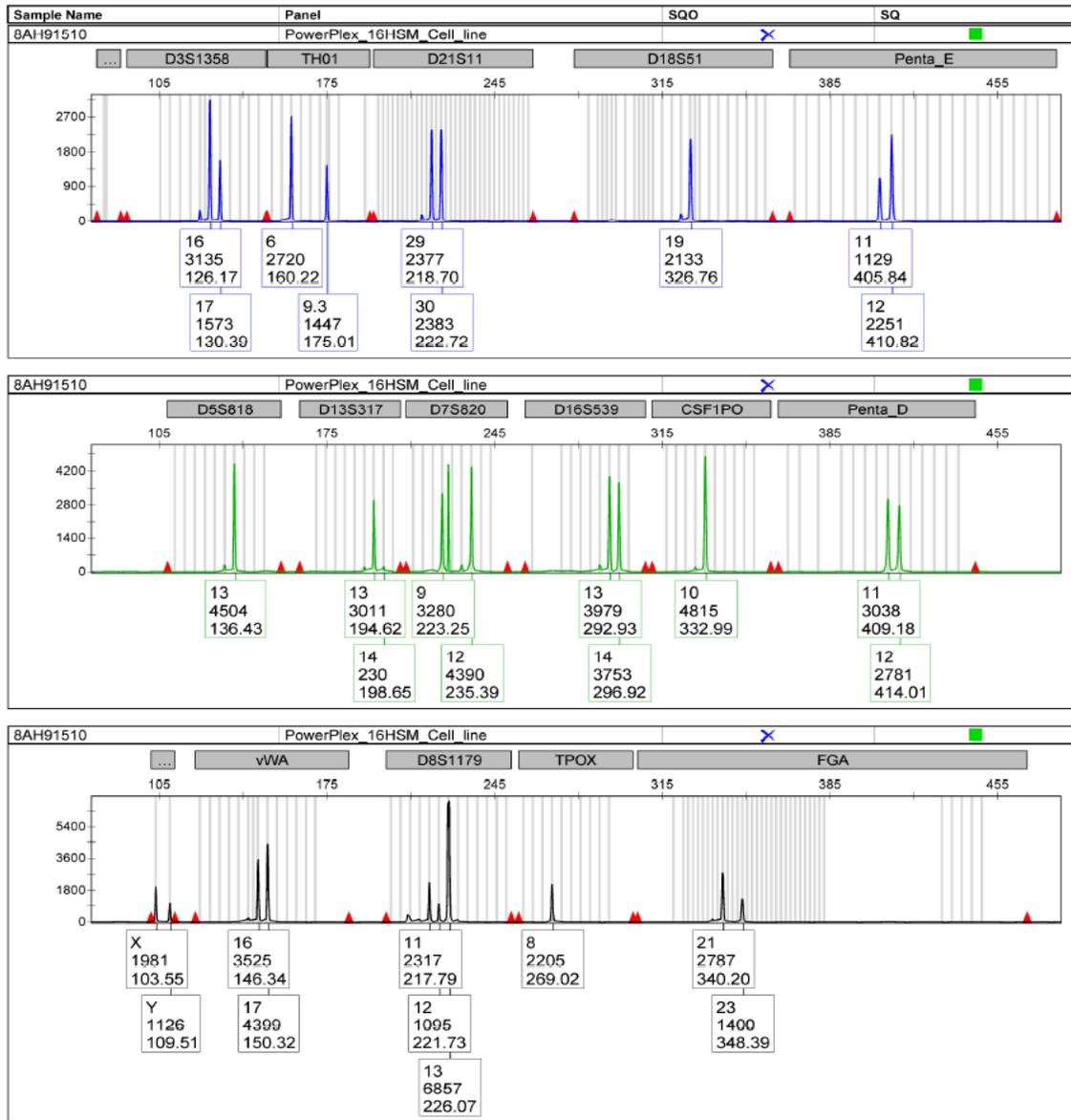
pSMAD1/5/8 (l and m) and Id1 (l and n) expression of LDN214117 treated human lines (n = 3

independent experiments). All data represented as mean with SEM. * p < 0.05, paired t test.

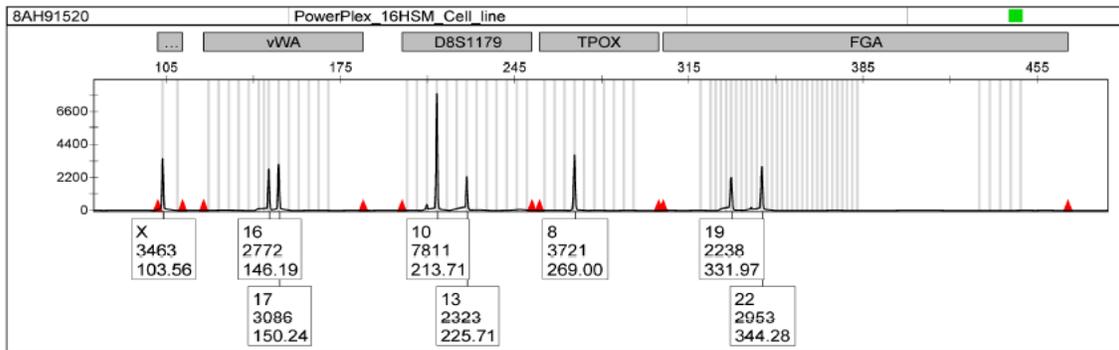
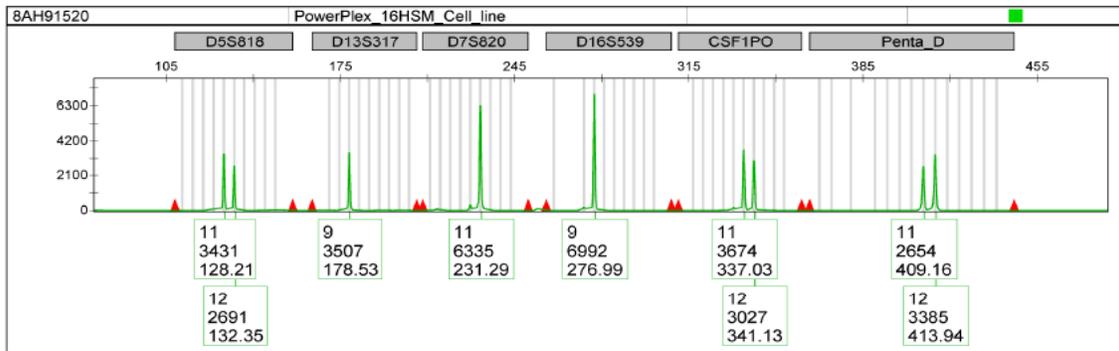
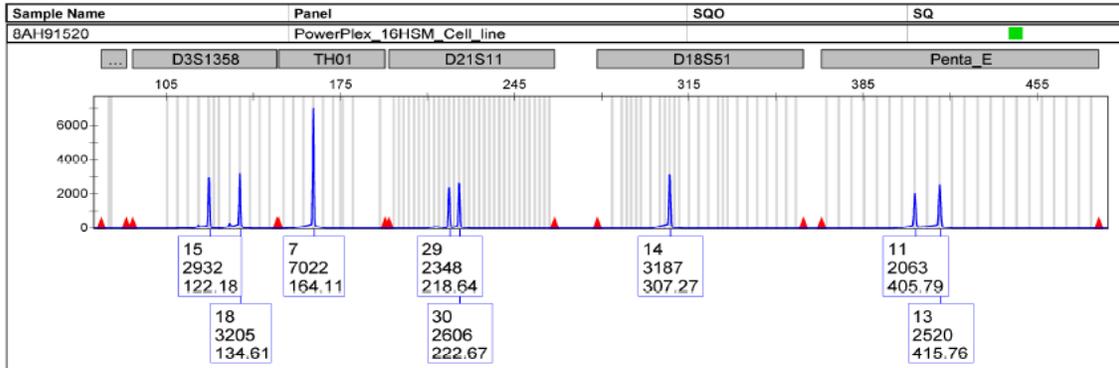


Supplementary Figure 5. Authentication of human DIPG cell line SU DIPG IV.

Electropherogram of STRs from human DIPG cell line SU DIPG IV. STR profiling results were obtained through PowerPlex 16HS + Mouse marker testing. Also see Supplementary Data 4.



Supplementary Figure 6. Authentication of human DIPG cell line DIPG 007. Electropherogram of STRs from human DIPG cell line DIPG 007. STR profiling results were obtained through PowerPlex 16HS + Mouse marker testing. Also see Supplementary Data 4.



Supplementary Figure 7. Authentication of human DIPG cell line SF8628. Electropherogram of STRs from human DIPG cell line SF8628. STR profiling results were obtained through PowerPlex 16HS + Mouse marker testing. Also see Supplementary Data 4.

Supplementary Table 1 ACVR1 virus combinations and lesion incidence *in vivo*

Virus Combination	Lesion Incidence
ACVR1 R206H, H3.1K27M, Cre (Ntv-a;p53 ^{fl/fl})	4/10
ACVR1 R206H, H3.3K27M, Cre (Ntv-a;p53 ^{fl/fl})	0/10
ACVR1 R206H, H3.1K27M, Cre (Ntv-a;p53 ^{fl/fl} ;PTEN ^{fl/fl})	6/10
ACVR1 R206H, H3.1K27M, Cre, BMP4 (Ntv-a;p53 ^{fl/fl})	3/6
ACVR1 R206H, H3.1 K27M, Cre, BMP4 (Ntv-a;p53 ^{fl/fl} ;PTEN ^{fl/fl})	0/13
ACVR1 R206H, H3.1K27M, Cre, Activin A (Ntv-a;p53 ^{fl/fl})	0/5
ACVR1 R206H, H3.1K27M, Cre, STAT3 WT (Ntv-a; p53 ^{fl/fl})	0/14
ACVR1 R206H, H3.1K27M, Cre, STAT3 WT (Ntv-a;p53 ^{fl/fl} ;PTEN ^{fl/fl})	0/4
ACVR1 R206H, H3.1K27M, Cre (B6 Ntv-a;PTEN ^{fl})	0/12
ACVR1 G328V, H3.1K27M, Cre (Ntv-a;p53 ^{fl/fl})	4/13
ACVR1 G328V, H3.1K27M, Cre (Ntv-a;p53 ^{fl/fl} ;PTEN ^{fl/fl})	6/15
ACVR1 G328V, H3.1K27M, Cre, BMP4 (Ntv-a;p53 ^{fl/fl})	0/7
ACVR1 G328E, H3.1K27M, Cre (Ntv-a;p53 ^{fl/fl})	1/8
ACVR1 G328E, H3.1K27M, Cre (Ntv-a;p53 ^{fl/fl} ;PTEN ^{fl/fl})	0/5
ACVR1 R206H only (Ntv-a;p53 ^{fl/fl})	0/9
ACVR1 G328V only (Ntv-a;p53 ^{fl/fl})	0/7
ACVR1 G328E only (Ntv-a;p53 ^{fl/fl})	0/5
ACVR1 WT only (Ntv-a;p53 ^{fl/fl})	0/3
H3.1K27M only (Ntv-a;p53 ^{fl/fl})	0/4
ACVR1 R206H, H3.1K27M (Ntv-a;p53 ^{fl/fl})	0/4
ACVR1 R206H, Cre (Ntv-a;p53 ^{fl/fl})	0/6
H3.1K27M, Cre (Ntv-a;p53 ^{fl/fl})	1/7
H3.1K27M, Cre (Ntv-a;p53 ^{fl/fl} ;PTEN ^{fl/fl})	0/6
ACVR1 R206H, H3.1 WT, Cre Ntv-a;p53 ^{fl/fl})	0/6
ACVR1 WT, H3.1K27M, Cre (Ntv-a;p53 ^{fl/fl})	0/7

Supplementary Methods

Western blot antibodies

Antibody	Company
pSMAD1/5/8	Cell Signaling Technologies (#9511)
pSMAD1/5/8	Millipore (AB3848-I)
Total SMAD1	Cell Signaling Technologies (#9743)
Actin	Cell Signaling Technologies (#3700)
Actin (I-19)	Santa Cruz (sc-1616)
Id1	BioCheck (BCH-1/37-2)
Hes1	Cell Signaling Technologies (#11988)
CD31	Abcam (ab28364)
Phospho-Stat3 (Tyr705) (D3A7) XP	Cell Signaling Technologies (#9145)
Stat3 (D3Z2G)	Cell Signaling Technologies (#12640)

Antibodies for IF and IHC

Antibody	Company
Anti-phospho-histone H3	Millipore (#04-1093)
HA-probe (Y-11)	Santa Cruz (sc-805)
Nestin	BD Pharmigen (#556309)
Olig2	Millipore (AB9610)
GFAP	Dako (Z 0334)
CD44	BD Biosciences (#550538)
CD31	Abcam (ab28364)
pSTAT3 (Tyr705) (D3A7)	Cell Signaling Technologies (#9145)
Iba1	Wako (#019-19741)
Vimentin	Santa Cruz (sc-7557-R)
Anti-DDK	Origene (TA50011-100)

Primers for qRT-PCR

Gene	Forward (5'→3')	Reverse (5'→3')
Actin	TATTGGCAACGAGCGGTTCC	GGCATAGAGGTCTTTACGGATGTC
Id1	CCTAGCTGTTGCTGAAGGC	CTCCGACAGACCAAGTACCAC
Id2	ATGAAAGCCTTCAGTCCGGTG	AGCAGACTCATCGGGTCGT
Id3	CTGTCCGGAACGTAGCCTGG	GTGGTTCATGTCGTCCAAGAG
Id4	CCGCCCAACAAGAAAGTCAG	CCAGGATGTAGTCGATAACGTG
L1Cam	AAAGGTGCAAGGGTGACATTC	TCCCCACGTTCTGTAGGT
CD44	TCGATTTGAATGTAACCTGCCG	CAGTCCGGGAGATACTGTAGC
Snai2	TGGTCAAGAAACATTTCAACGCC	GGTGAGGATCTCTGGTTTTGGTA
Vimentin	CGTCCACACGCACCTACAG	GGGGGATGAGGAATAGAGGCT
TNC	ACGGCTACCACAGAAGCTG	ATGGCTGTTGTTGCTATGGCA
Sox10	AGGTTGCTGAACGAAAGTGAC	CCGAGGTTGGTACTTGTAGTCC