

SUPPLEMENTARY MATERIAL

Cationic Intrinsically Disordered Antimicrobial Peptides (CIDAMPs) Represent a New Paradigm of Innate Defense with a Potential for Novel Anti-Infectives

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Supplementary Figure 1

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340      350      360      370      380      400      410      420      430      440      450      460
YSYSRGHYES GSGQTSGFQO HESSSGQSSG YSKHSGSGGR SSSGQHGST SGQASSSGQH GQHEASRHS SGRGQHSSGS QSPGHQRG SSGQSPSSG QHGTGFRSS SSGPYVSGG
SRGHYES GSGQTSGFQO HESSSGQSSG Y
SRGHYES GSGQTSGFQO HESSSGQSSG
ES GSGQTSGFQO HESSSGQSSG Y
HS SGRGQHSSGS QSPGHQRG SSGQSPSSG QHGTGFRSS SSGPY
HS SGRGQHSSGS QSPGHQRG SSGQSPSSG QHGTGFRSS SSGP
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GRGQHSSGS QSPGHQRG SSGQSPSSG QHGTGFR
GRGQHSSGS QSPGHQRG SSGQSPSSG QHGTGFR
SSGS QSPGHQRG SSGQSPSSG QHGTGFR
GS QSPGHQRG SSGQSPSSG QHGTGFR

500      510      520      530      540      550      560      570
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SQ SSRGERQSS AGSSSSYGQH GSGSRQSLGR S HSGSGQS PPSRGRHES GSFQSSSYGP HG
GERQSS AGSSSSYGQH GSGSRQSLGR S
GERQSS AGSSSSYGQH GSGSRQSL

680      690      700      710      720      730      740      750      760      770      780
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GQHS GSGWSSNGP HGSVSGQSSG FGHKSGSGQS SG
SSSNGP HGSVSGQSSG FGHKSGSGQS SGYSOHGSGS SHSSGY SQSSRSE QHGSSGLSS SYGQHSGSH QSSGHRQSS GSHGSPSRV
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SSSNGP HGSVSGQSSG FGHKSGSGQS SG
SSNGP HGSVSGQSSG FGHKSGSGQS SGYS
SSNGP HGSVSGQSSG FGHKSGSGQS SGY
SNP HGSVSGQSSG FGHKSGSGQS SGY
NGP HGSVSGQSSG FGHKSGSGQS SGY
SSG FGHKSGSGQS SGYSOHGSGS SHSSGY

920      930      940      950      960      970
SGSGRSSSSG RHGSGSQSS GFGHKSSSQ SSGYTOHGSG SGHSSSYEQH GSRSGQSSRS
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SSSSG RHGSGSQSS GFGHKSSSQ SSGYTOHG
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SSSSG RHGSGSQSS GFGHKSSSQ SSGYI
SSSSG RHGSGSQSS GFGHKSSSQ SSGY
SSG RHGSGSQSS GFGHKSSSQ SSGYTOH

980      990      1000      1010      1020      1030      1040      1050      1060      1070      1080
RCHGSSSGSS SSYGQHGSG RQSLGHQHG SSGSQSPSPS RGRHSGSGO SSSYGPVRSQ SGWSSSRGPY ESGSGHSSGL GHRESRQSS SGYQHGSSS GHSSTHQHG
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G SSGSQSPSPS RGRHSGSGO SSSYGPY
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QSPS YGRHSGSGR SSSGQHGSG LGESSGFHH ESSSQSSS
QSPS YGRHSGSGR SSSGQHGSG LGESSGFHH E
QSPS YGRHSGSGR SSSGQHGSG LGE
GRHSGSGR SSSGQHGSG LGESSGFHH ESSSQSSY
GRHSGSGR SSSGQHGSG LGESSGFHH ESSSQSSS
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CRHSGSGR SSSGQHGSG LGESSGFHH ESSSQS
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GRHSGSGR SSSGQHGSG LGESSGFHH ESS
GRHSGSGR SSSGQHGSG LGESSGFHH ES
GRHSGSGR SSSGQHGSG LGESSGFHH E
RHSGSGR SSSGQHGSG LGESSGFHH ESSSQSSS
HGSGR SSSGQHGSG LGESSGFHH ESSSQSSS
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SSSGQHGSG LGESSGFHH ESSSQS
SSSGQHGSG LGESSGFHH ESSSQ
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SSSGQHGSG LGESSGFHH
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GHGSG LGESSGFHH ESSSQSSS
HSG LGESSGFHH ESSSQSSS

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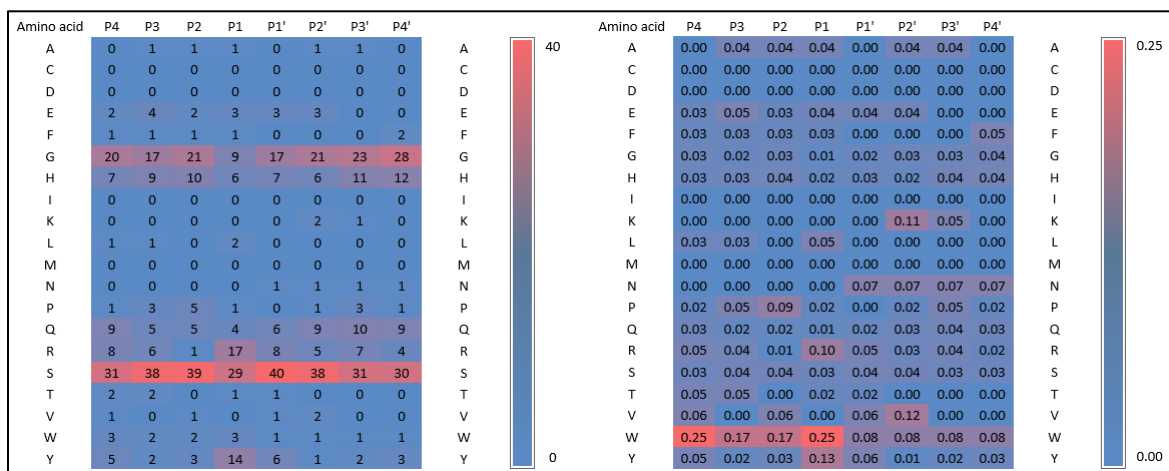
Supplementary Figure 1, continued

1250	1260	1270	1280	1290	1300	
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GRHGS	GLGHSSSHGQ	HGSSGGRSSS	RGPYE			
GRHGS	GLGHSSSHGQ	HGSSGGRSSS	RGPY			
HGS	GLGHSSSHGQ	HGSSGGRSSS	RGPYE			
HGS	GLGHSSSHGQ	HGSSGGRSSS	RGPY			
GS	GLGHSSSHGQ	HGSSGGRSSS	RGPY			
	GLGHSSSHGQ	HGSSGGRSSS	RGPY			
			ESRSGH	SSVFGQHESG	SGHSSAY	
			ESRSGH	SSVFGQHESG	SGHSSA	
			ESRSGH	SSVFGQHESG	SGHS	
			ESRSGH	SSVFGQHESG	SGH	
			SGH	SSVFGQHESG	SGHSSAY	
			SGH	SSVFGQHESG	SGHSSA	
1360	1370	1380	1390	1400	1410	
<u>RHRGSGQSP</u>	<u>SRGRHGSQSG</u>	<u>HSSSYGQHGS</u>	<u>GSQSSSSGR</u>	<u>HGSSGQSSG</u>	<u>FGHHSSSAQ</u>	
SP	SRGRHGSQSG	HSSSYGQHGS	GSQSS			
	RGRHGSQSG	HSSSYGQHGS	GSQSS			
	GRHGSQSG	HSSSYGQHGS	GSQSSSSGR	HGSSGQSSG	FGHHES	
	GRHGSQSG	HSSSYGQHGS	GSQSSSSSSGR	HGSSGQSSG	FGHHE	
	GRHGSQSG	HSSSYGQHGS	GSQSSSSSSGR	HG		
	GRHGSQSG	HSSSYGQHGS	GSQSSSSSSGR			
	GRHGSQSG	HSSSYGQHGS	GSQSS			
	GRHGSQSG	HSSSYGQHGS	GSQSS			
	HGSSGQ	HSSSYGQHGS	GSQSSSSSSGR	HGSSGQSSG	FGHHES	
	HGSSGQ	HSSSYGQHGS	GSQSSSSSSGR	HGSSGQSSG	FGHHES	
	HGSSGQ	HSSSYGQHGS	GSQSSSSSSGR	HGSSGQSSG	FGHHE	
	HGSSGQ	HSSSYGQHGS	GSQSS			
		QGHGS	GSQSSSSSSGR	HGSSGQSSG	FGHHES	
		QGHGS	GSQSSSSSSGR	HGSSGQSSG	FGHHES	
		QGHGS	GSQSSSSSSGR	HGSSGQSSG	FGHHES	
		QGHGS	GSQSSSSSSGR	HGSSGQSSG	FGHHE	
		QGHGS	GSQSSSSSSGR	HGSSGQSSG	FGHH	
		GS	GSQSSSSSSGR	HGSSGQSSG	FGHHE	
			SSSSGR	HGSSGQSSG	FGHHES	
			SSSSGR	HGSSGQSSG	FGHHES	
			SGR	HGSSGQSSG	FGHHES	
			GR	HGSSGQSSG	FGHHES	
				HGSSGQSSG	FGHHES	
1440	1450	1460	1470	1480	1490	1500
<u>GSRSGQSSRG</u>	<u>ERHGSSSGSS</u>	<u>SSYGQHGS</u>	<u>RQSLGHGQHG</u>	<u>SGSGQSPSPS</u>	<u>RGRHGSQSSQ</u>	<u>SSSYSPYGS</u>
G	ERHGSSSGSS	SSYGQHGS	RQSL			
	HGSSSSGSS	SSYGQHGS	RQSLGHGQHG	SGSGQSPSPS		
	HGSSSSGSS	SSYGQHGS	RQSL			
			QSLGHGQHG	SGSGQSPSPS	RGRHGSQSSQ	SSSY
			GHGQHG	SGSGQSPSPS	RGRHGSQSSQ	SSSYSPY
1610	1620	1630	1640	1650	1660	1670
<u>RGSQSSRQSPS</u>	<u>YGRHGSQSGR</u>	<u>SSSSGQHGS</u>	<u>LGESSGFGRH</u>	<u>ESSSGQSSSY</u>	<u>SGHGSQSSGHS</u>	<u>SGYGQHGSS</u>
QSPS	YGRHGSQSGR	SSSSGQHGS	LGESSGFGRH	ESSSGQSSS		
QSPS	YGRHGSQSGR	SSSSGQHGS	LGESSGFGRH	E		
QSPS	YGRHGSQSGR	SSSSGQHGS	LGESSGFGRH	ESSSGQSSSY		
	GRHGSQSGR	SSSSGQHGS	LGESSGFGRH	ESSSGQSSS		
	GRHGSQSGR	SSSSGQHGS	LGESSGFGRH	ESSSGQSSS		
	GRHGSQSGR	SSSSGQHGS	LGESSGFGRH	ESSSGQSS		
	GRHGSQSGR	SSSSGQHGS	LGESSGFGRH	ESSSG		
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	GSGSGR	SSSSGQHGS	LGESSGFGRH	ESSSGQSSS		
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		SSSSGQHGS	LGESSGFGRH	ESSSGQSS		
		SSSSGQHGS	LGESSGFGRH	ESSSGQ		
		SSSSGQHGS	LGESSGFGRH	ESSSG		
		SSSSGQHGS	LGESSGFGRH	ESSS		
		SSSSGQHGS	LGESSGFGRH	ESS		
		SSSSGQHGS	LGESSGFGRH	ES		
		SSSSGQHGS	LGESSGFGRH	E		
		SSSGQHGSS	LGESSGFGRH	ESSSGQSSS		
		SGQHGSS	LGESSGFGRH	ESSSGQSSS		
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Supplementary Figure 1, continued

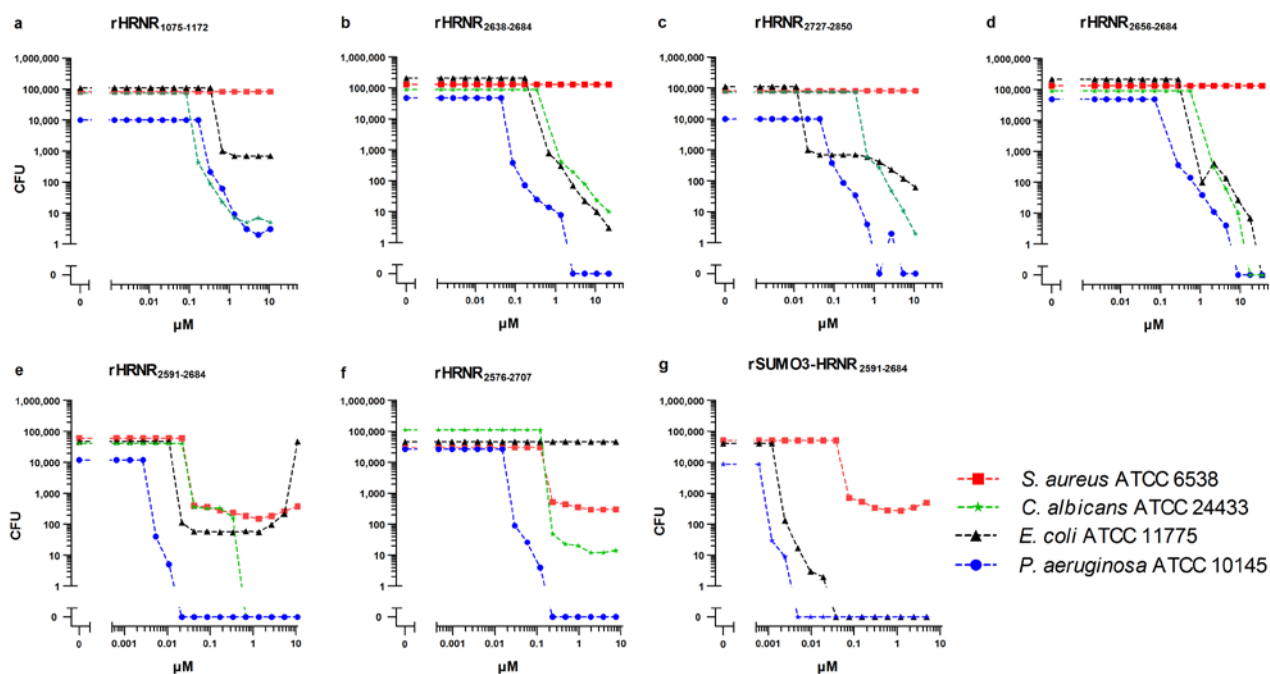
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	GRHGS	GLGHSSSHGQ	HGSGSGRSGS	RGPYE					
	GRHGS	GLGHSSSHGQ	HGSGSGRSGS	RGPY					
	HGS	GLGHSSSHGQ	HGSGSGRSGS	RGPYE					
	HGS	GLGHSSSHGQ	HGSGSGRSGS	RGPY					
	GS	GLGHSSSHGQ	HGSGSGRSGS	RGPY					
		GLGHSSSHGQ	HGSGSGRSGS	RGPY					
				ESRSGH	SSVFGQHESG	SGHSSAY			
				ESRSGH	SSVFGQHESG	SGHSSA			
				ESRSGH	SSVFGQHESG	SGHS			
				ESRSGH	SSVFGQHESG	SGH			
				SGH	SSVFGQHESG	SGHSSAY			
				SGH	SSVFGQHESG	SGHSSA			
1810	1820	1830	1840	1850	1860	1870	1880		
<u>GOSSSHGQHG</u>	<u>SGSSQSSSYG</u>	<u>QOGSGSGQSP</u>	<u>SRGRHGSGSG</u>	<u>HSSSYGQHGS</u>	<u>GSGWSSSSGR</u>	<u>HGSGSGQSSG</u>	<u>FGHHSSSWD</u>		
	G	QOGSGSGQSP	SRGRHGSGSG	HSSSYGQHGS	GSGWSSSSGR				
	G	QOGSGSGQSP	SRGRHGSGSG	HSSSYGQHGS	GSGWS				
		SGSGQSP	SRGRHGSGSG	HSSSYGQHGS	GSGWSSSSGR				
		SGSGQSP	SRGRHGSGSG	HSSSYGQHGS	GSGWSS				
		SP	SRGRHGSGSG	HSSSYGQHGS	GSGWS				
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			GRHGSGSG	HSSSYGQHGS	GSGWSSSSGR	HGSGSGQSSG	FGHHE		
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			GRHGSGSG	HSSSYGQHGS	GSGWS				
			GRHGSGSG	HSSSYGQHGS	GSGW				
			HGSGSG	HSSSYGQHGS	GSGWSSSSGR	HGSGSGQSSG	FGHHESS		
			HGSGSG	HSSSYGQHGS	GSGWSSSSGR	HGSGSGQSSG	FGHHE		
			HGSGSG	HSSSYGQHGS	GSGWSSSSGR	HGSGSGQSSG	FGHHE		
			HGSGSG	HSSSYGQHGS	GSGW				
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				QHGGS	GSGWSSSSGR	HGSGSGQSSG	FGHHE		
				QHGGS	GSGWSSSSGR	HGSGSGQSSG	FGHHE		
				QHGGS	GSGWSSSSGR	HGSGSGQSSG	FGHHE		
				GS	GSGWSSSSGR	HGSGSGQSSG	FGHHE		
					SSSSGR	HGSGSGQSSG	FGHHESSWQ		
					SSSSGR	HGSGSGQSSG	FGHHESSW		
					SGR	HGSGSGQSSG	FGHHESSW		
					GR	HGSGSGQSSG	FGHHESSWQ		
						HGSGSGQSSG	FGHHESSW		
1890	1900	1910	1920	1930	1940	1950	1960	1970	
<u>SSGYDQHGSG</u>	<u>SGHSSSYDQH</u>	<u>GRSRGSSRG</u>	<u>EDHGSSSGS</u>	<u>SSYGQHGSGS</u>	<u>ROSLGHGQHG</u>	<u>SGSGQSPSPS</u>	<u>RGRHGSGSGQ</u>	<u>SSSYGPGSG</u>	
			HGSSSGSS	SSYGQHGSGS	ROSLGHGQHG	SGSGQSPSPS			
			HGSSSGSS	SSYGQHGSGS	RQSL				
					QSLGHGQHG	SGSGQSPSPS	RGRHGSGSGQ	SSSY	
					GHGQHG	SGSGQSPSPS	RGRHGSGSGQ	SSSYGPY	
					GHGQHG	SGSGQSPSPS	RGRHGSGSGQ	SSSYGP	
					HGQHG	SGSGQSPSPS	RGRHGSGSGQ	SSSYGPY	
					HG	SGSGQSPSPS	RGRHGSGSGQ	SSSYGPY	
					G	SGSGQSPSPS	RGRHGSGSGQ	SSSYGPY	
						SGSGQSPSPS	RGRHGSGSGQ	SSSYGPY	
						SGSGQSPSPS	RGRHGSGSGQ	SSSYGP	
1980	1990	2000	2010	2020					
<u>SGWSSSRGPY</u>	<u>ESGSGHSSGL</u>	<u>GHRESRSGQS</u>	<u>SGYGQHGSSS</u>	<u>GHSSTHGQHG</u>					
	RGPY	ESGSGHSSGL	GHRESR						
		ESGSGHSSGL	GHRESRSGQS	SGYGQHGSSS	GHSSTHGQHG				
2220	2230	2240							
<u>RGPYESRSGH</u>	<u>SSVFGQHESG</u>	<u>SGHSSAYSCH</u>							
	ESRSGH	SSVFGQHESG	SGHSSAY						
	ESRSGH	SSVFGQHESG	SGHSSA						
	ESRSGH	SSVFGQHESG	SGHS						
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Supplementary Figure 2



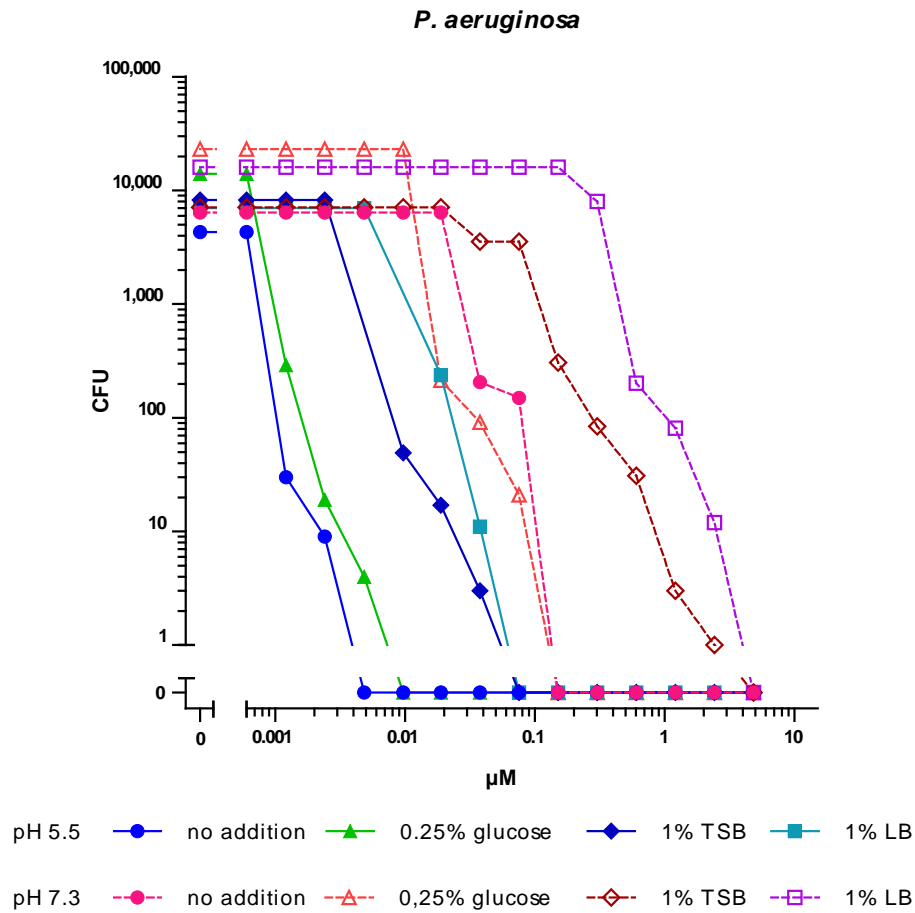
Supplementary Figure 2 | Analysis of the sequences surrounding the cleavage sites (P1 position) of all HRNR-peptides identified in stratum corneum. The heat maps show (left panel) the number of residues identified in position P4-P1 and P1'-P4', respectively, and (right panel) the values of these residues normalized vs. the total number of the particular amino acid in the HRNR canonical sequence. Annotation of P4-P1 and P1'-P4' according to ref.⁹³.

Supplementary Figure 3



Supplementary Figure 3 | HRNR fragments are potent broad-spectrum antimicrobial polypeptides. Sensitivity of *P. aeruginosa* ATCC 10145, *E. coli* ATCC 11775, *S. aureus* ATCC 6538 and *C. albicans* ATCC 24433 towards **a**, rHRNR₁₀₇₅₋₁₁₇₂; **b**, rHRNR₂₆₃₈₋₂₆₈₄; **c**, rHRNR₂₇₂₇₋₂₈₅₀; **d**, rHRNR₂₆₅₆₋₂₆₈₄; **e**, rHRNR₂₅₉₁₋₂₆₈₄; **f**, rHRNR₂₅₇₆₋₂₇₀₇; and **g**, rSUMO3-HRNR₂₅₉₁₋₂₆₈₄ has been tested in a CFU assay system at pH 5.5 in 10 mM NaP/1% TSB (**a-d**) or in 10 mM NaP/0.25% glucose (**e-g**). Note *E. coli* ATCC 11775 missing sensitivity towards rHRNR₂₅₇₆₋₂₇₀₇ at these conditions (**f**), and an “antibiotic paradox” (decrease of sensitivity at increasing peptide concentrations) towards rHRNR₂₅₉₁₋₂₆₈₄ (**e**). AA-sequences of the HRNR-polypeptides are shown in (Supplementary Table 1). Results shown are representatives (n=2) (**a-d, f, g**) and (n=3) (**e**).

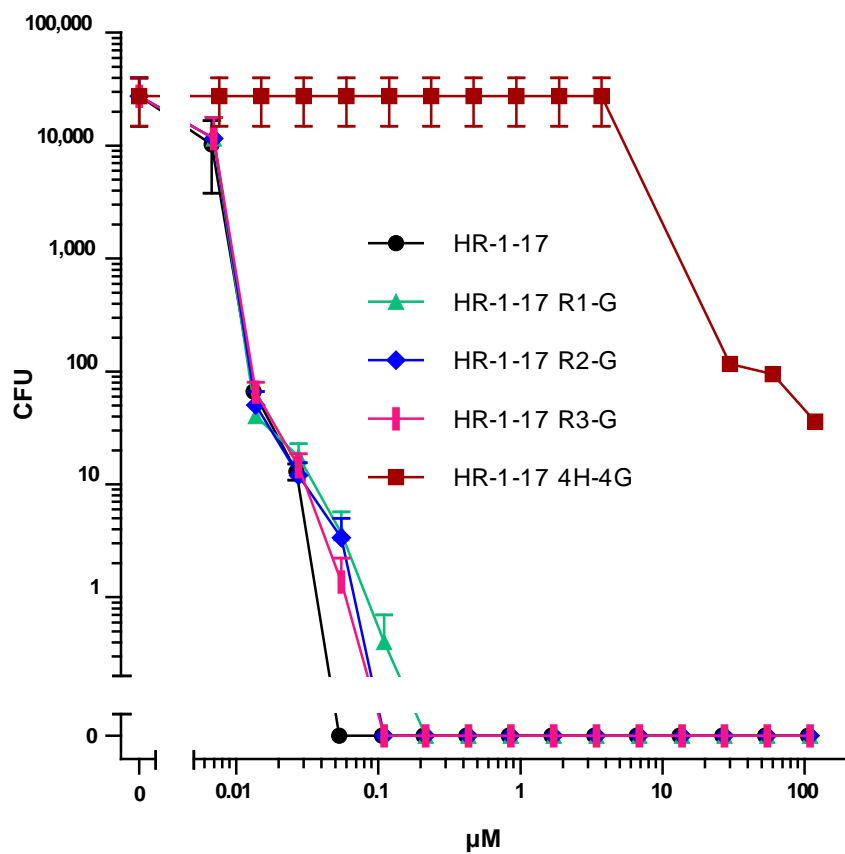
Supplementary Figure 4



Supplementary Figure 4 | *P. aeruginosa*-cidal activity of SUMO3-HRNR₂₅₉₁₋₂₆₈₄ is pH-dependent and is inhibited by “Tryptic Soybean Broth, TSB” and “Luria Broth, LB”.

Recombinant SUMO3-HRNR₂₅₉₁₋₂₆₈₄ was tested at the indicated conditions in 10 mM NaP/0.25% glucose or 1% TSB, 1% LB or no addition. Note a shift of the dose-response curve by both, increased pH and addition of nutrients like TSB or LB. All data shown are representatives (n=2).

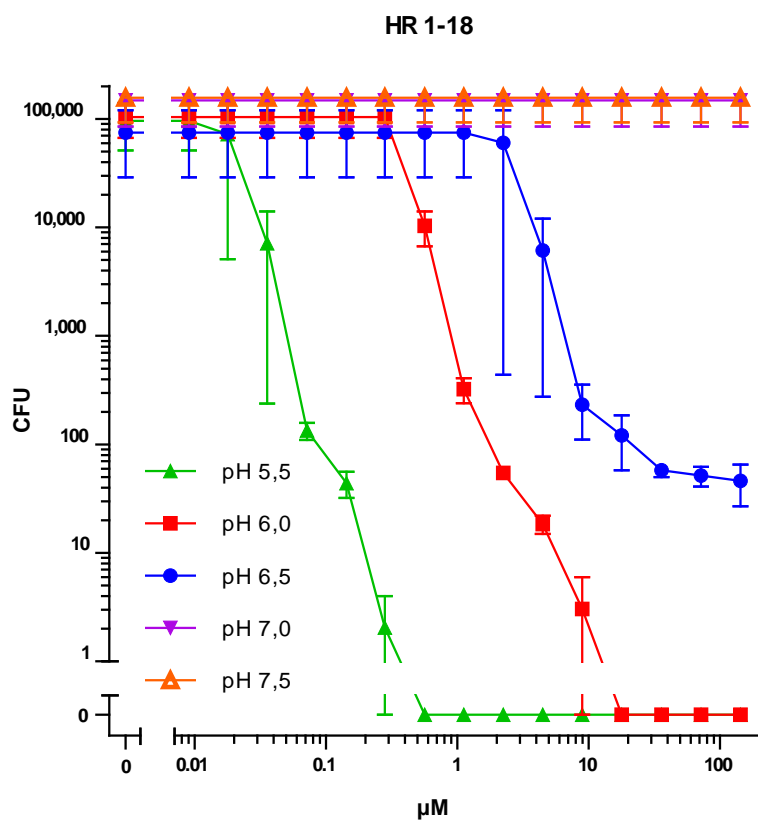
Supplementary Figure 5:



Supplementary Figure 5 | *P. aeruginosa* ATCC 10145-cidal activity of HR 1-17 mutants.

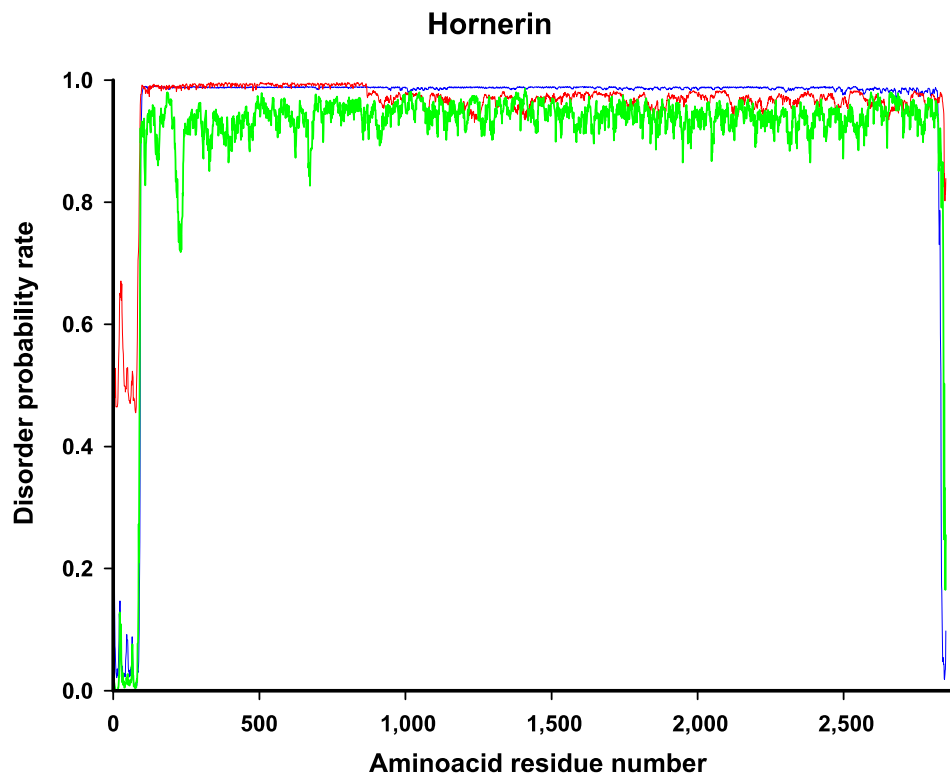
Pseudomonas aeruginosa ATCC 10145-cidal activity in 10 mM NaP/0.25% glucose/pH 5.5, was examined for four mutants of HR1-17 with replacement of either Arg or His by Gly (AA-sequences are shown in Supplementary Table 4). Data represent the mean \pm s.e.m. (n=3).

Supplementary Figure 6:



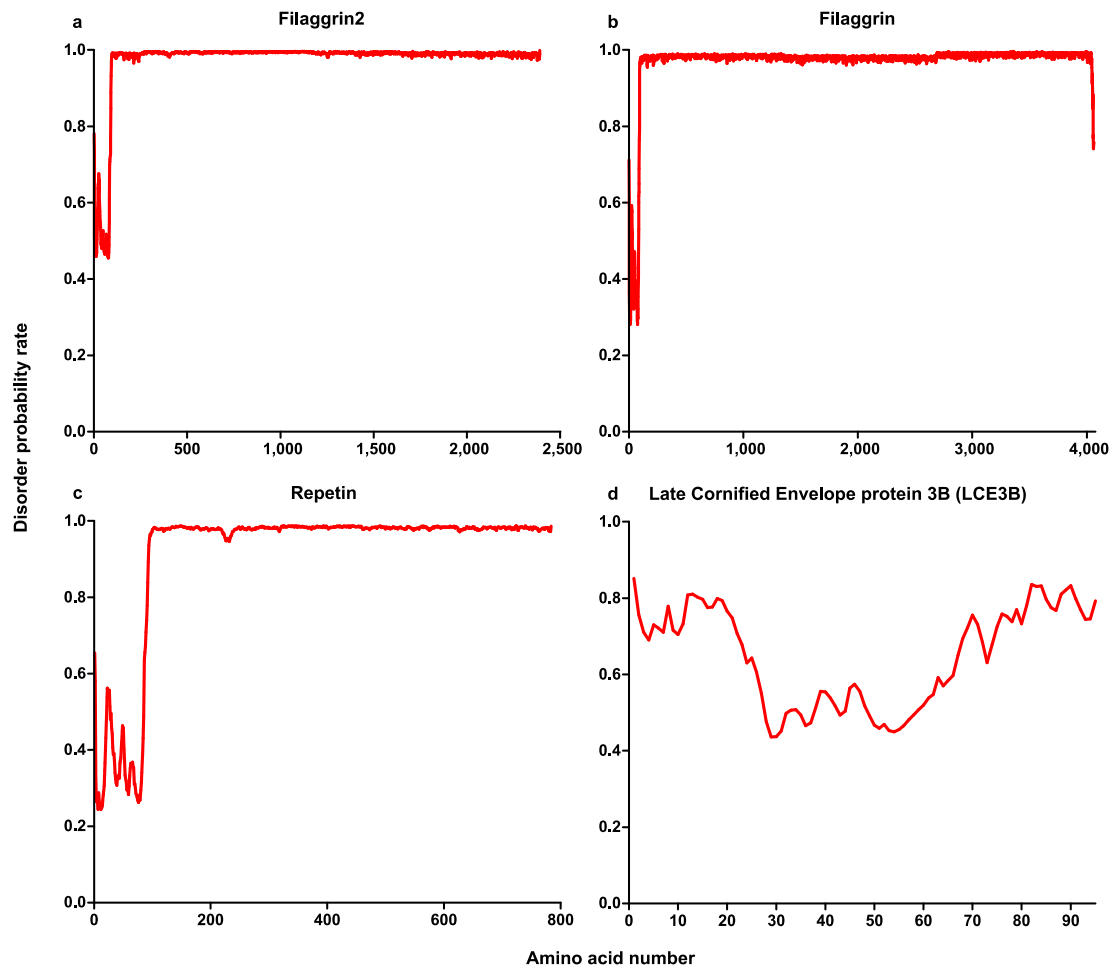
Supplementary Figure 6 | *P. aeruginosa*-cidal activity of HR1-18 depends on pH. *P. aeruginosa* ATCC 10145-cidal activity of HR1-18 was examined in 10 mM NaP/ 0.25% glucose/pH 5.5-7.5. Data shown represent the mean \pm s.e.m. (n=3).

Supplementary Figure 7:



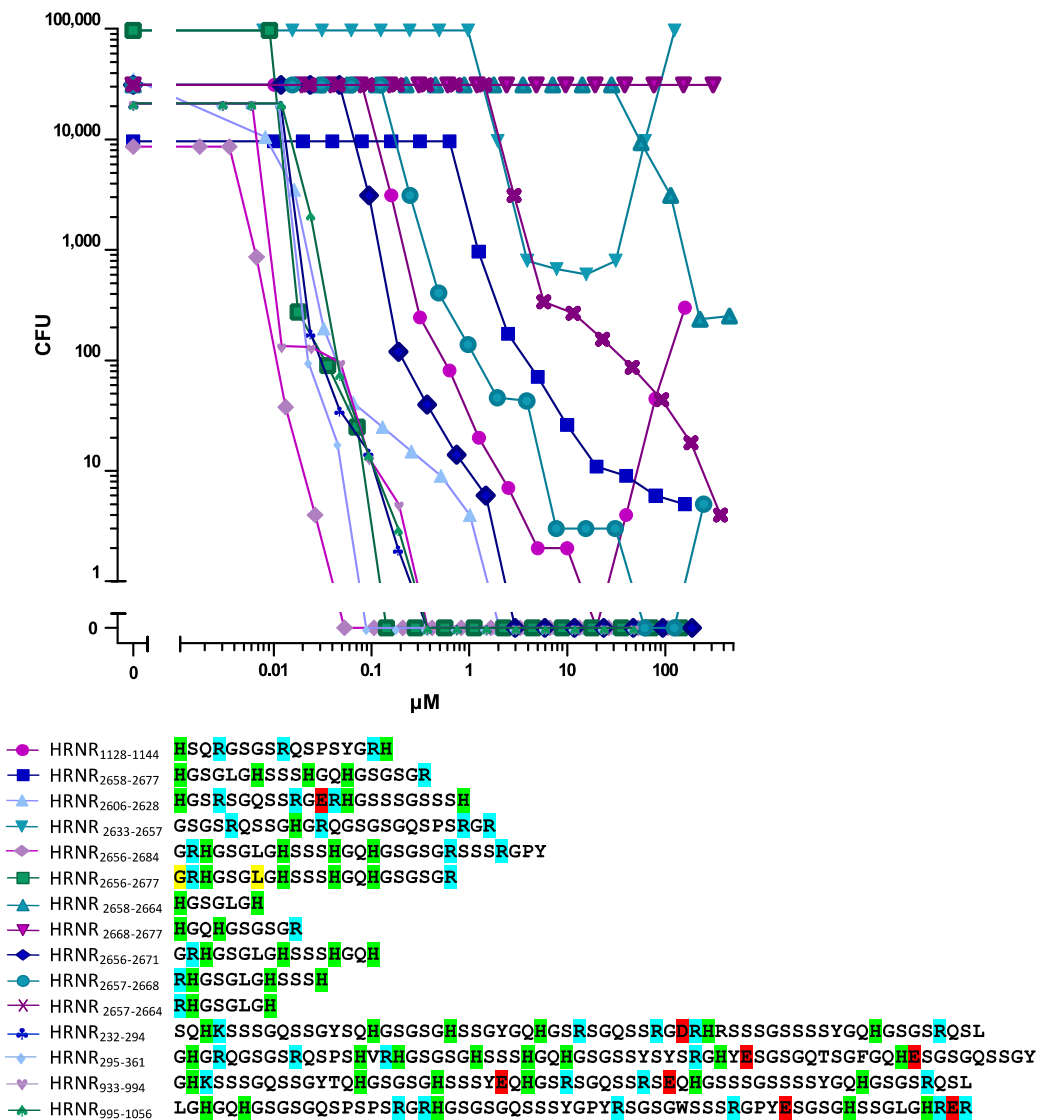
Supplementary Figure 7 | Evaluation of the intrinsic disorder predisposition in HRNR by three computational tools. The amino acid position (abscissa) is shown as a function of the disorder probability rate. Results of the PreDisorder program⁹⁴ are given in blue, of the DisoClust program⁹⁵ given in red and results of the DISOPRED2 program⁹⁶ appear in green. There is a clear difference between the N-terminal ordered EF-hand-like-domain (AA 1-100), the area of repetitive Gly/Ser-rich units detected as a large intrinsically disordered region and the small C-terminal ordered region.

Supplementary Figure 8:



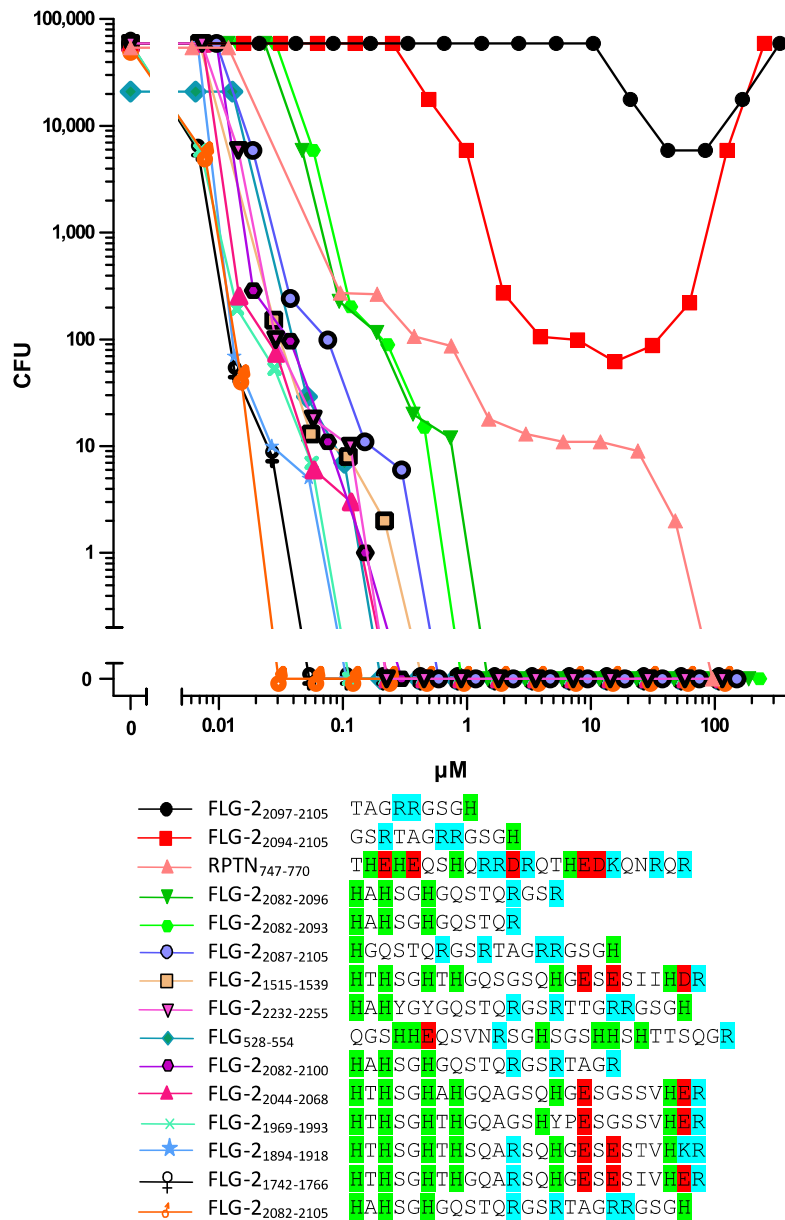
Supplementary Figure 8 | Evaluation of the intrinsic disorder predisposition FLG-2, FLG, RPT and LCE3B by the DisoClust program. a: FLG-2; b: FLG; c: RPT; d: LCE3B. The amino acid position (abscissa) is shown as a function of the disorder probability rate. Results of the DisoClust program⁹⁵ are given. In the three “S100-fused-type-proteins FLG2, FLG and RPT there is a clear difference between the N-terminal ordered EF-hand-like-domain (AA 1-100) and the area rich in repetitive disorder promoting AA. LCE3B reveals, possibly due to its high content of Cys and His, a disorder probability between 50 - 70 % in the relevant stretch of AA 56 to AA 78 in LCE3B₅₆₋₆₈. Notably, this peptide is investigated for antimicrobial activity in its thiol-form at pH 5.5, where His is protonated. This is not taken into account by the DisoClust program and may further increase the disorder probability rate.

Supplementary Figure 9:



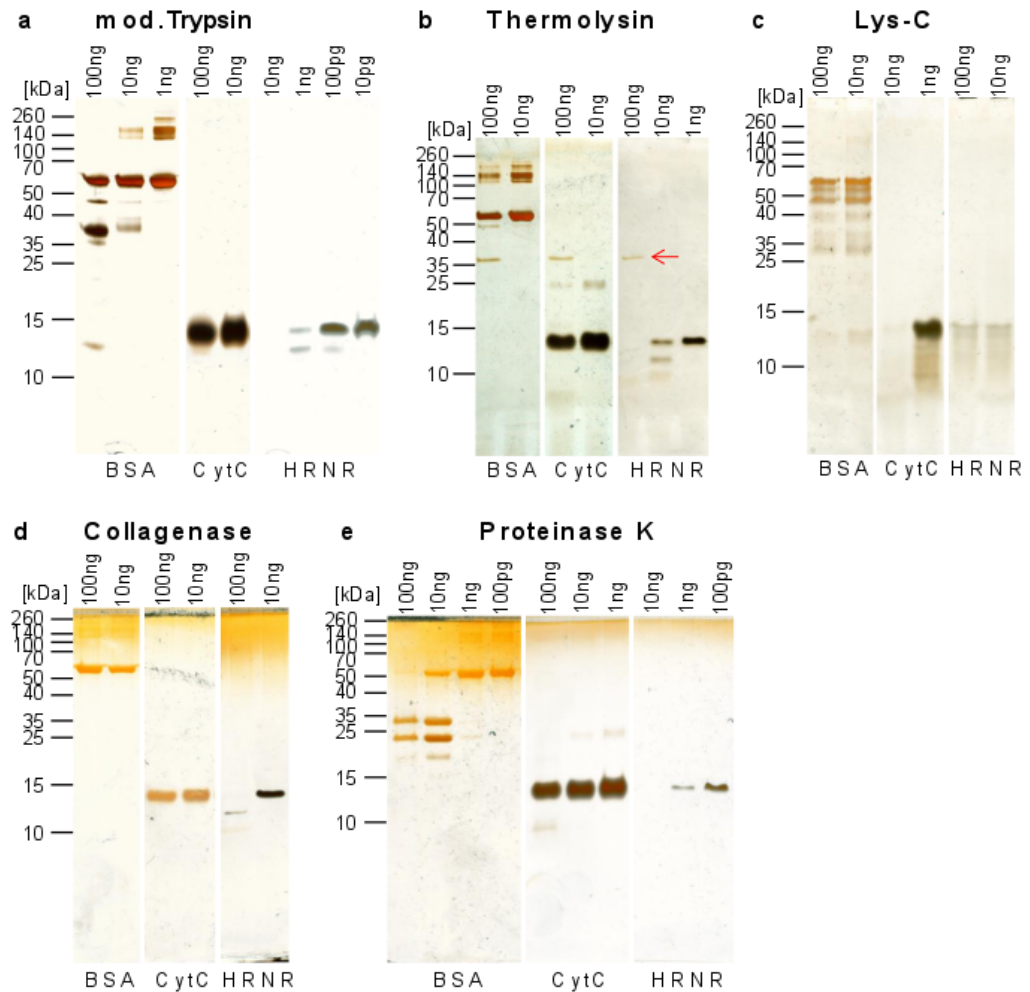
Supplementary Figure 9 | *P. aeruginosa*-cidal activity of selected HRNR peptides. Sensitivity of *P. aeruginosa* ATCC 10145 towards various peptides was tested in 10 mM NaP/0.25% glucose/pH 5.5. In the peptide's amino acid sequences residues cationic at pH 7, cationic only at pH 5.5 and anionic at pH 5.5 are colored blue, green and red, respectively. Antimicrobial paradox¹⁶ (increased bacterial growth with increasing CIDAMP-concentrations) was observed for HRNR₁₁₂₈₋₁₁₄₄ and HRNR₂₆₃₃₋₂₆₅₇. Representative dose-response curves of selected HRNR-peptides are shown (for the number of experiments see Supplementary Table 6).

Supplementary Figure 10:



Supplementary Figure 10 | *P. aeruginosa*-cidal activity of selected FLG-2-, FLG- and RPTN-derived CIDAMPs. Sensitivity of *P. aeruginosa* ATCC 10145 was tested in 10 mM NaP/ 0.25% glucose/pH 5.5. Residues cationic at pH 7 (R, K), cationic only at acidic pH (H) and anionic (E, D) are colored blue, green and red, respectively. Antimicrobial paradox¹⁶ (increased bacterial growth with increasing CIDAMP-concentrations) was observed for FLG-2₂₀₉₄₋₂₁₀₅ and FLG-2₂₀₉₇₋₂₁₀₅. Representative dose-response curves of selected FLG-2-; FLG- and RPTN-peptides are shown (for the number of experiments see Supplementary Table 7).

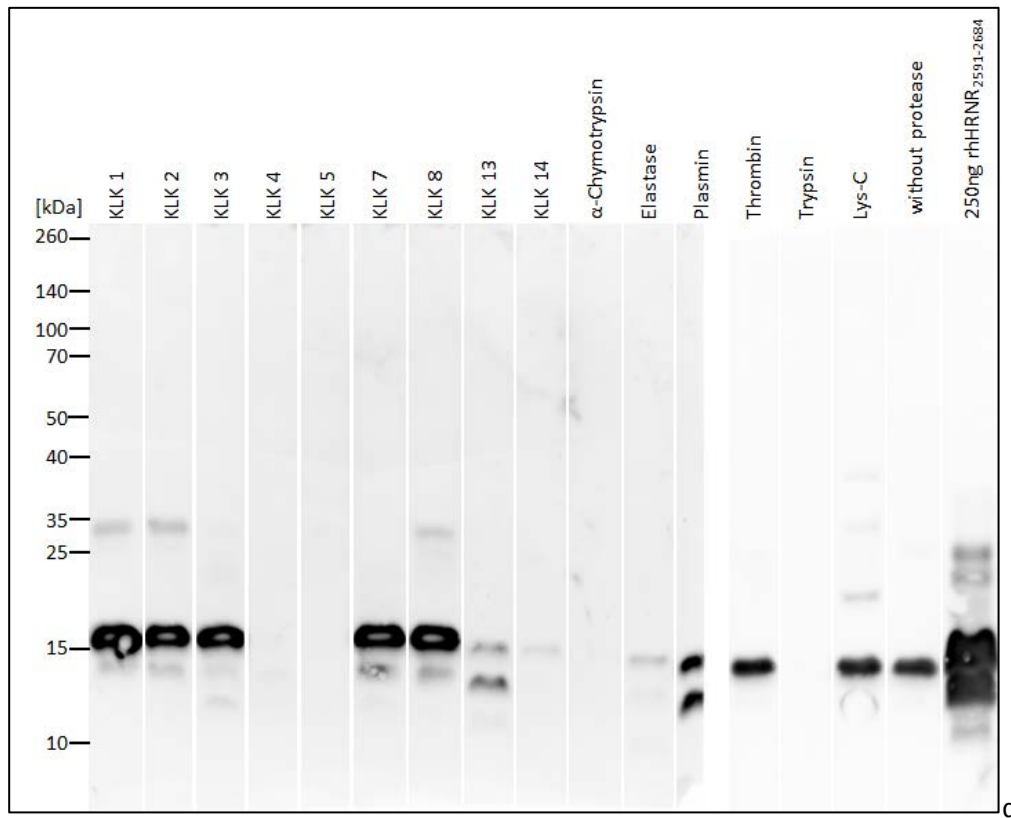
Supplementary Figure 12:



Supplementary Figure 12 | Protease sensitivity of rHRNR₂₅₉₁₋₂₆₈₄, BSA and cytochrome C.

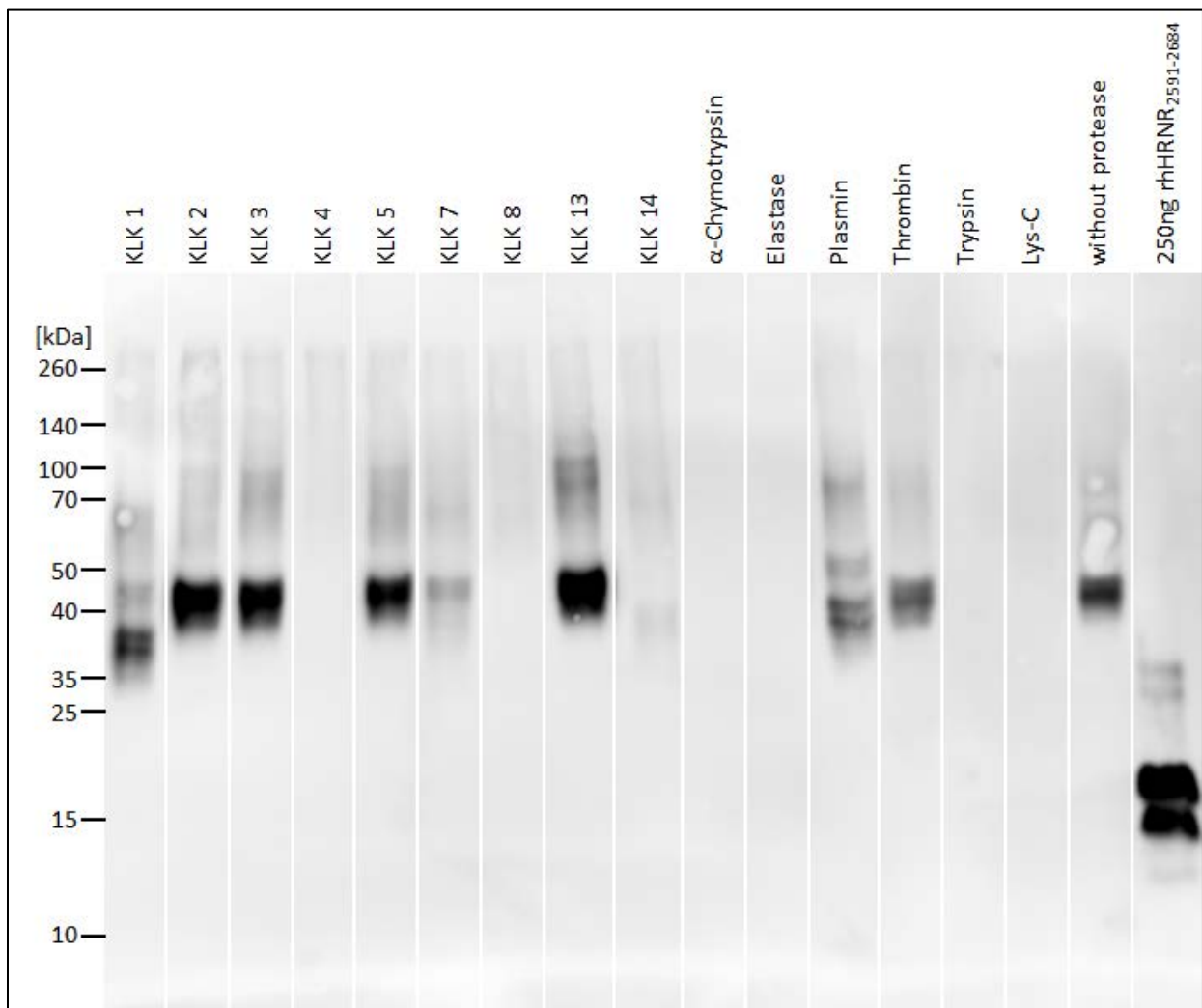
1 µg each of bovine serum albumin (BSA), cytochrome (Cyt) C or rHRNR₂₅₉₁₋₂₆₈₄ (HRNR, a different HRNR-fragment as used in Fig. 8) were incubated with the proteases indicated above in protease-specific buffers for 1 h at 37 °C. Thereafter samples were subjected to a SDS-PAGE analysis and proteins were detected by silver staining. **a**, modified (methylated) serine proteinase trypsin; **b**, the metalloproteinase thermolysin (red arrow); **c**, the serine proteinase Lys C; **d**, the metalloproteinase collagenase; and **e**, proteinase K. Note the sensitivity of rHRNR₂₅₉₁₋₂₆₈₄ towards digestion by Lys C and collagenase, albeit higher concentrations, although predicted cleavage sites for these enzymes are absent in rHRNR₂₅₉₁₋₂₆₈₄ (for AA-sequence see Supplementary Table 1). Data shown are representatives (n=3).

Supplementary Figure 13:



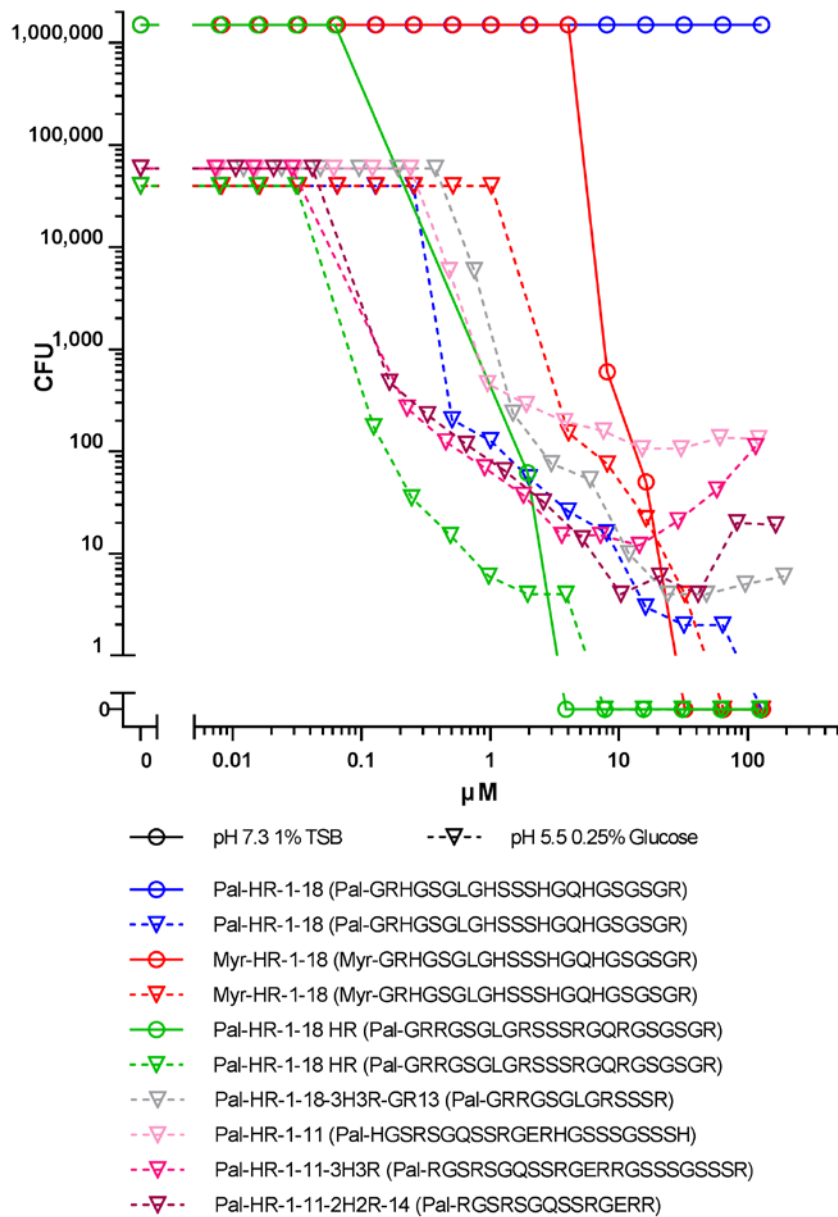
Supplementary Figure 13 | KLK4, KLK5, KLK13, KLK14 and trypsin, but not KLK1, KLK7, KLK8 and LysC digest rHRNR₂₅₉₁₋₂₆₈₄. 500 ng rHRNR₂₅₉₁₋₂₆₈₄ each were incubated with 0.7 nM of the respective, activity-checked protease in buffer (100 mM Tris, pH 7.5, 150 mM NaCl, 5 mM EDTA, 0.05 Tween 20) for 15 h. For SDS-PAGE analyses 250 ng of rHRNR₂₅₉₁₋₂₆₈₄ were used and then analyzed by rHRNR₂₅₉₁₋₂₆₈₄-Westernblot. Lane 16 (“without protease”) contains a control sample treated in the same manner, however in the absence of a protease. In lane 17, 250 ng of an rHRNR₂₅₉₁₋₂₆₈₄-pool (for AA-sequence see Supplementary Table 1) have been directly applied. Note the presence of multiple bands, which originate from oligomeric HRNR-complexes¹². Data shown are representatives (n=3).

Supplementary Figure 14:



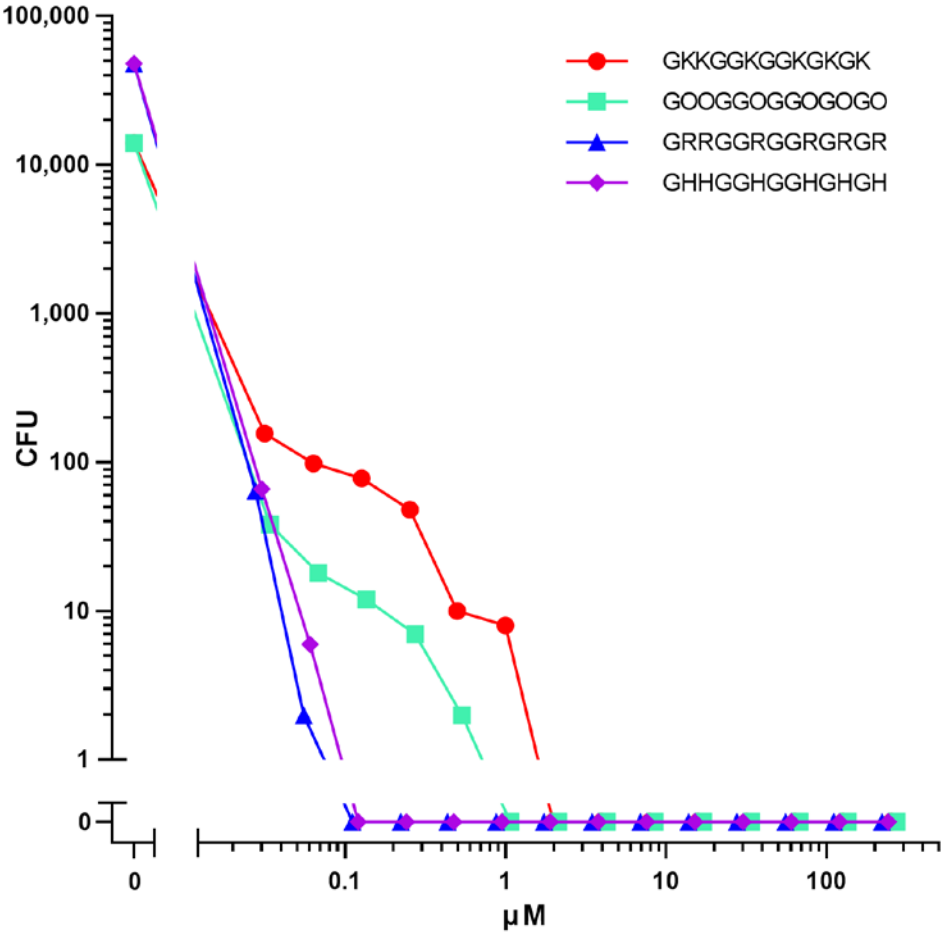
Supplementary Figure 14 | KLK1, KLK4, KLK7, KLK8, KLK14, trypsin and LysC, but not KLK13 digest PAD1-deiminated rHRNR₂₅₉₁₋₂₆₈₄. 500 ng PAD1-deiminated rHRNR₂₅₉₁₋₂₆₈₄ each were incubated for 15 h with 0.7 nM of the respective, activity-tested protease in buffer (100 mM Tris pH 7.5, 150 mM NaCl, 5 mM EDTA, 0.05% Tween 20). For SDS-PAGE analyses 250 ng of the protein were used and then analyzed by rHRNR₂₅₉₁₋₂₆₈₄-Westernblot. Lane 16 ("without protease") contains a control sample (PAD1-deiminated rHRNR₂₅₉₁₋₂₆₈₄), treated in the same manner, however in the absence of a protease. In lane 17, 250 ng of an rHRNR₂₅₉₁₋₂₆₈₄-pool have been directly applied. Note a marked decrease of PAD1-deiminated rHRNR₂₅₉₁₋₂₆₈₄ electromobility (lane 16) relative to rHRNR₂₅₉₁₋₂₆₈₄ (lane 17). Data shown are representatives (n=3).

Supplementary Figure 15:



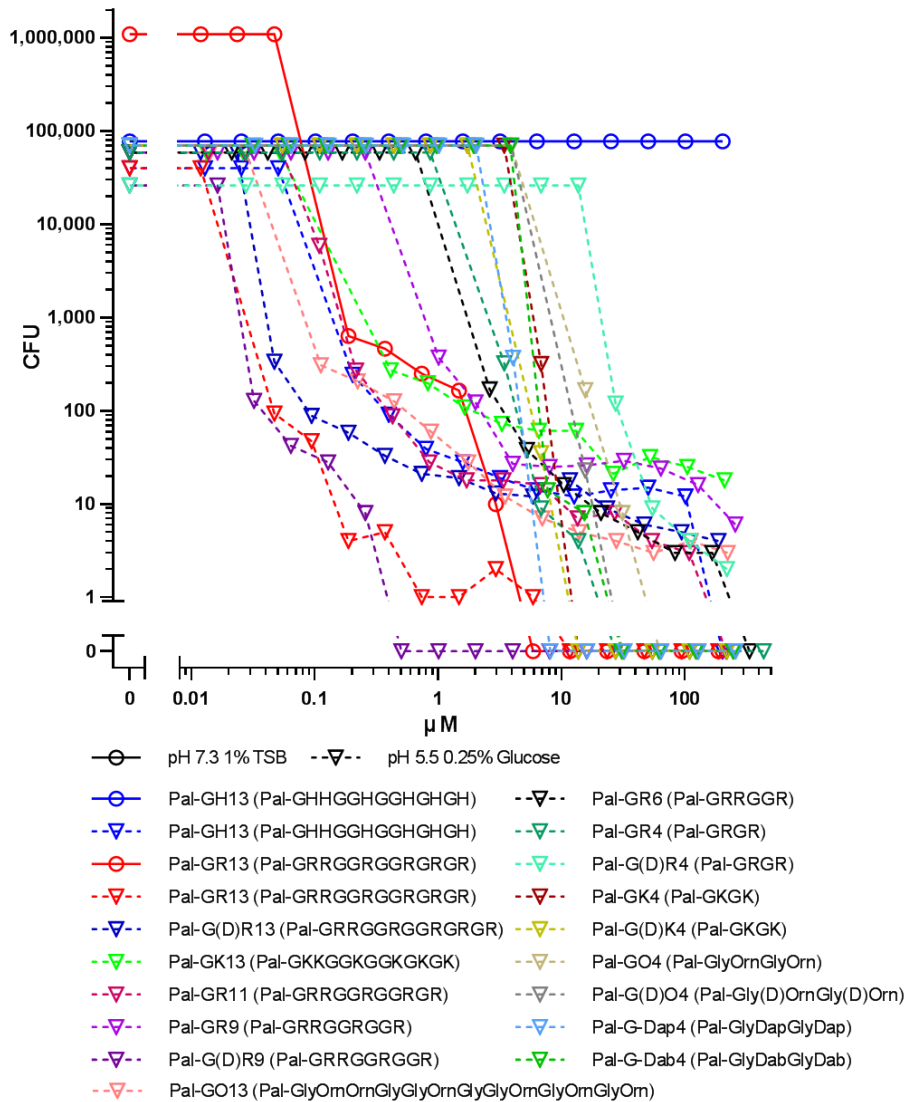
Supplementary Figure 15 | Palmitoylation and myristoylation of CIDAMPs improves *S. aureus*-cidal activity of HR1-18, HR1-11 and its Arg-mutants. *S. aureus* ATCC 6538-cidal activity, assayed either in 10 mM NaP, pH 5.5, containing 0.25% glucose, or in 10 mM NaP, pH 7.3, containing 1 % TSB, was examined in a CFU assay system. Representatives are shown (for the number of experiments see Table 5).

Supplementary Fig. 16



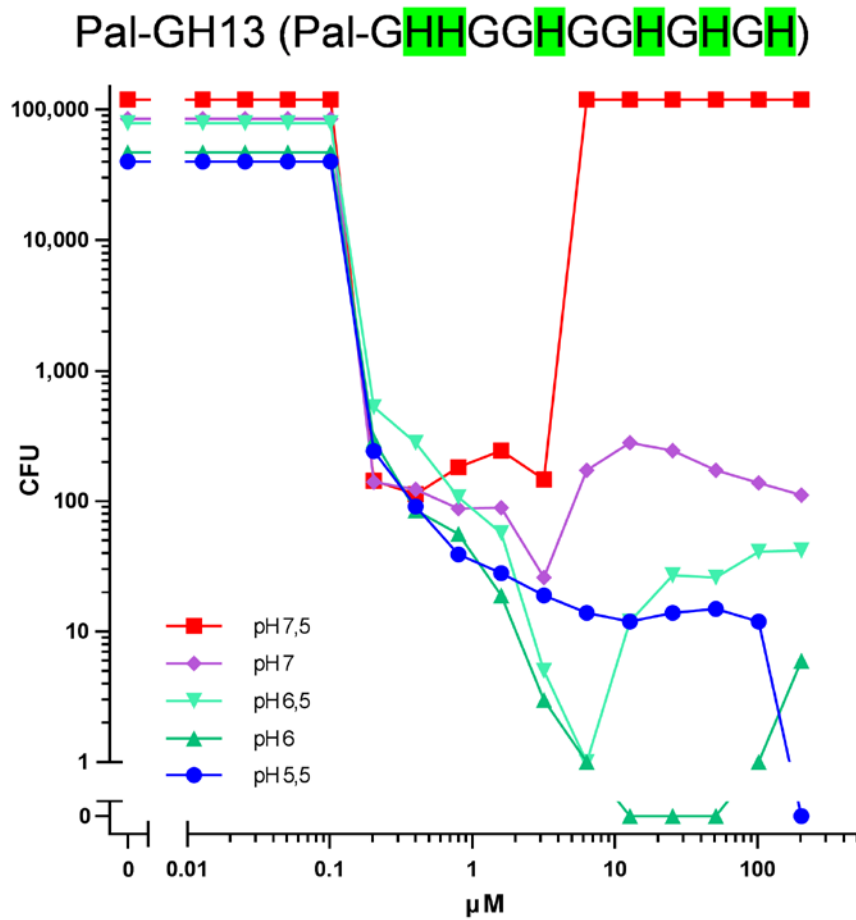
Supplementary Fig. 16 | *P. aeruginosa*-bactericidal activity of Gly-rich 13-mer CIDAMPs containing Lys, Orn, Arg or His. *P. aeruginosa* ATCC 10145-bactericidal activity was examined in a CFU assay system in 10 mM NaP, pH 5.5, containing 0.25% glucose. Data shown are representatives (n = 2).

Supplementary Figure 17



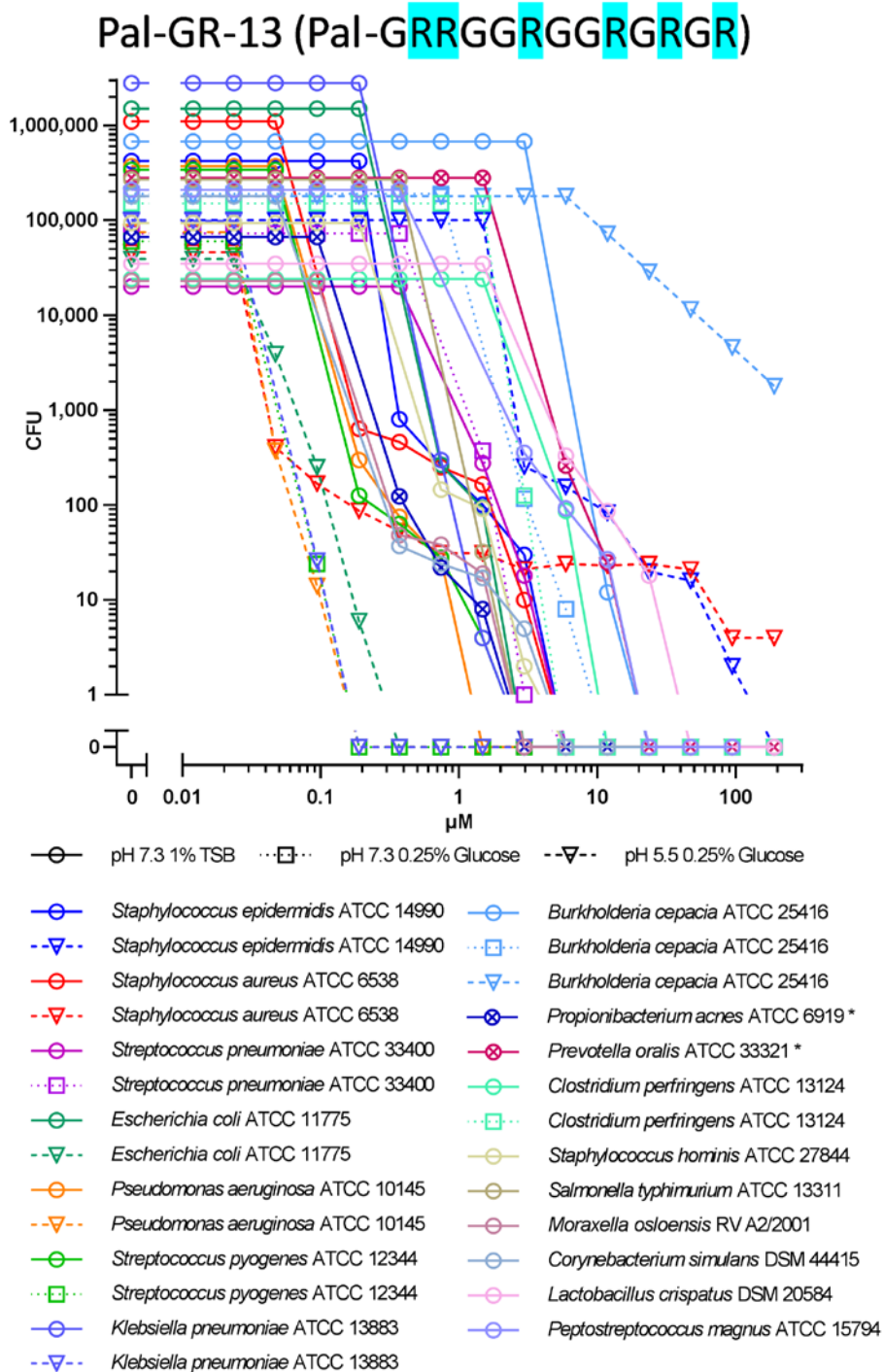
Supplementary Figure 17 | *S. aureus*-cidal activity of palmitoylated CIDAMPs depends on the peptide chain length and positively charged AAs. *S. aureus* ATCC 6538-cidal activity of the lipopeptides was analyzed in a CFU assay system, in 10 mM NaP, pH 5.5, containing 0.25% glucose (solid lines), or in 10 mM NaP, pH 7.3, containing 1 % TSB (dashed lines). Note the loss of sensitivity when Pal-GH13 is tested in TSB-containing medium at pH 7.3. Dab: L-2,4-Diaminobutyric acid, Dap:L-2,3-Diaminopropionic acid. Representatives are shown (for the number of experiments see Table 2).

Supplementary Figure 18



Supplementary Figure 18 | pH-dependency analysis of Pal-GH13-*S. aureus*-cidal activity reveals an “antibiotic paradox”. *S. aureus* ATCC 6538-cidal activity of Pal-GH13 was analyzed in a CFU assay system in 10 mM NaP (pH 5.5; 6.0; 6.5; 7.0; 7.5) containing 0.25% glucose. Note that an increase of the pH towards pH 7.5 causes an “antibiotic paradox”¹⁶, which is characterized by an increase of CFU with increasing lipopeptide concentrations. The result of a single experiment is shown.

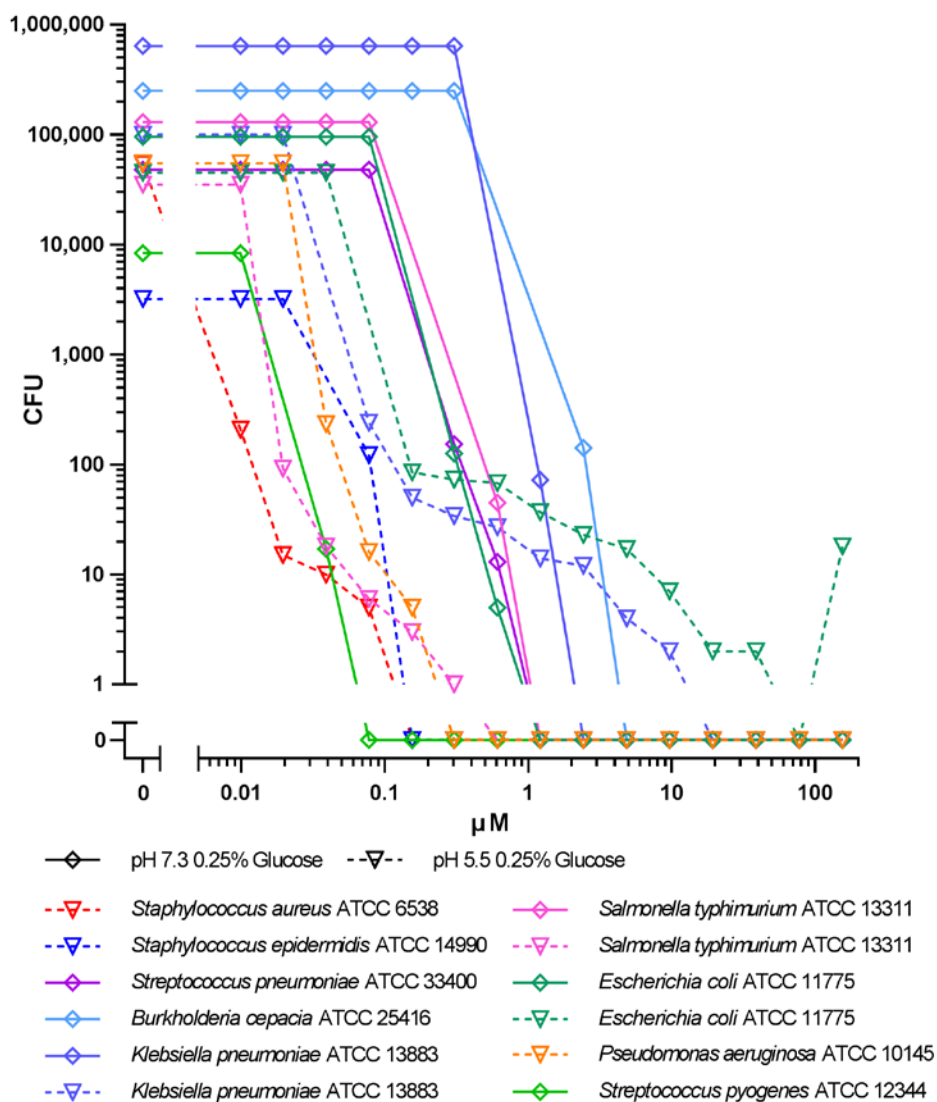
Supplementary Figure 19



Supplementary Figure 19: Antimicrobial activity spectrum of Pal-GR13. Sensitivity of selected bacteria towards Pal-GR13 was tested in a CFU assay system with 10 mM NaP containing either 0.25% glucose or 1% TSB as test medium, at pH 5.5 or pH 7.3. Representatives are shown (for the number of experiments see Table 7).

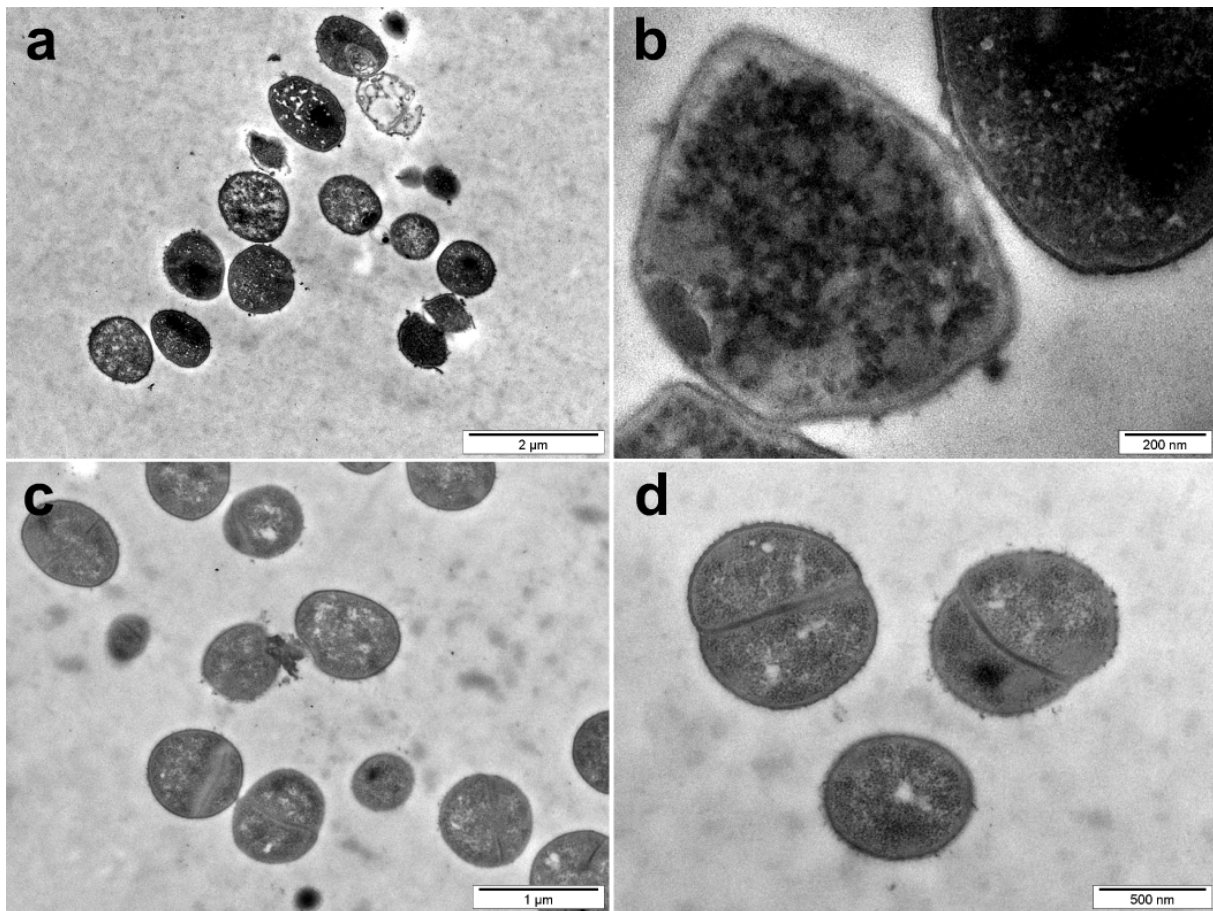
Supplementary Figure 20

Palmitoyl-LCE-3B₅₆₋₆₈ (Pal-SHHRCRSHRCRR)



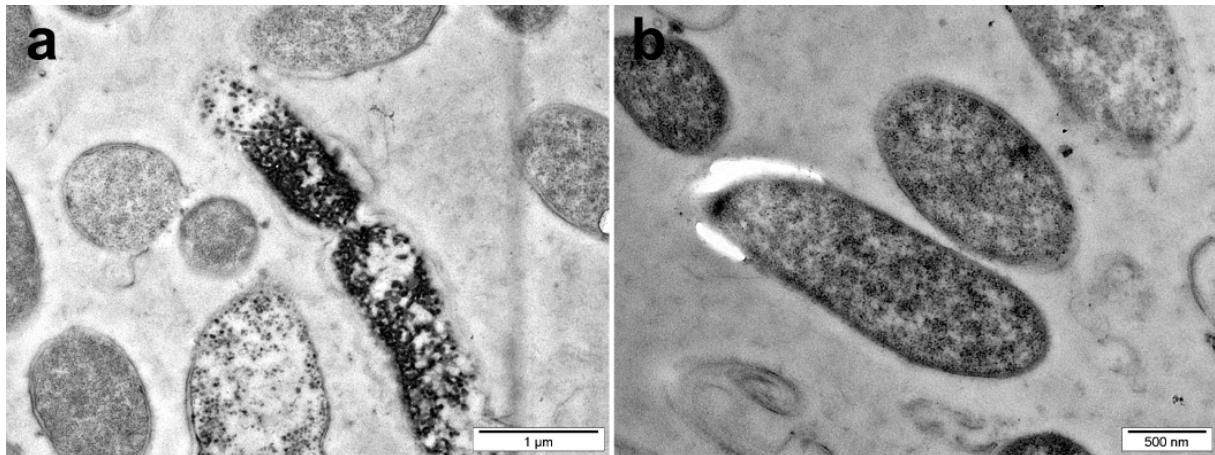
Supplementary Figure 20: Antimicrobial activity spectrum of Pal-LCE-3B₅₆₋₆₈. Sensitivity of bacteria towards Pal-LCE-3B₅₆₋₆₈ was tested in a CFU assay system in 10 mM NaP, containing 0.25% glucose and, depending on the microbe, at pH 5.5, 6.0 or pH 7.3, as indicated in Supplementary Table 7. Representatives are shown (for the number of experiments see Table 8).

Supplementary Figure 21



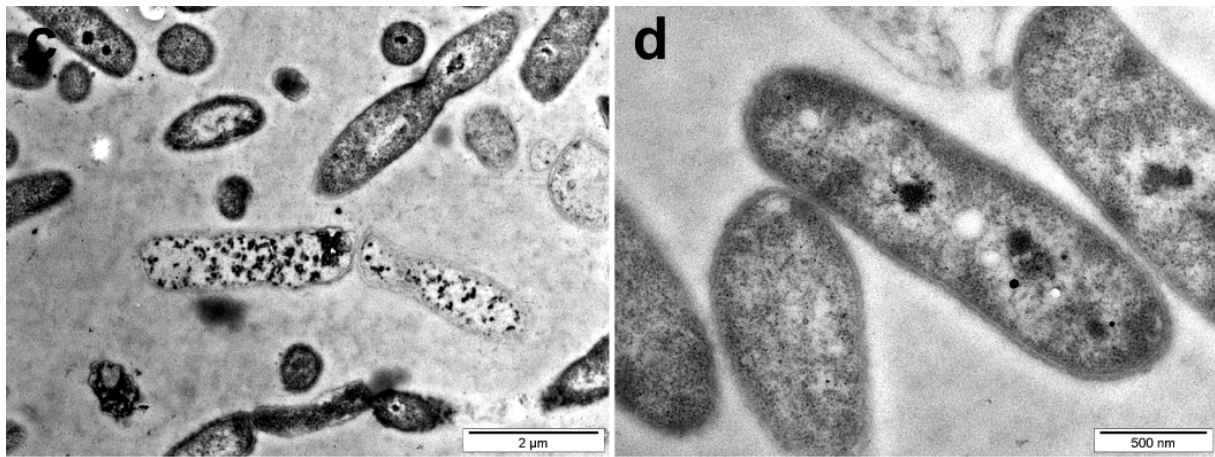
Supplementary Figure 21| TEM of LCE3B₅₆₋₆₈-treated *S. aureus*. 10^7 *S. aureus* ATCC 6538, suspended in 150μl 10 mM NaP, pH 5.5, containing 0.25% glucose, were treated with LCE3B₅₆₋₆₈ (SHHRCCRS^{SHRCRR}, 125 μg/ml) for 90 min at 37°C and then analyzed by transmission electron microscopy (TEM)(**a, b**). *S. aureus* ATCC 6538, incubated with 10 mM NaP, pH 5.5, containing 0.25% glucose, for 90 min at 37°C, served as control (**c, d**). Note the condensation of electron-dense cytoplasmic material in LCE3B₅₆₋₆₈-treated bacteria (**a, b**). Images are representative of two independent experiments, sampling on average 10 images.

Supplementary Figure 22 a, b



Supplementary Figure 22 a, b | TEM of Pal-GR13-treated *E. coli*. 10^7 *E. coli* ATCC 11775, suspended in 150µl 10 mM NaP, pH 7.3, containing 1% TSB, were treated with Pal-GR13 (Pal-GRRGGRGGRRGR, 375 µg/ml) for 60 min at ambient temperature and then analyzed by transmission electron microscopy (TEM) (a). *E. coli* ATCC 11775, incubated with 10 mM NaP, pH 7.3, containing 1% TSB for 60 min at ambient temperature, served as control (b). Note condensation of electron-dense cytoplasmic material in Pal-GR13-treated bacteria (a). Images are representative of two independent experiments, sampling on average 10 images.

Supplementary Figure 22c, d

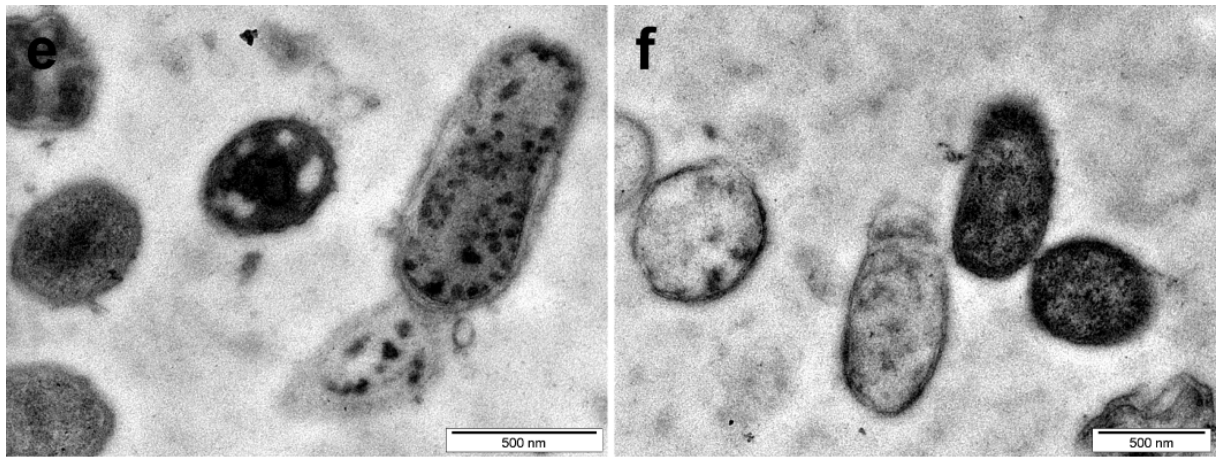


Supplementary Figure 22c, d | TEM of Pal-GR13-treated *Burkholderia cepacia*.

10^7 *Burkholderia cepacia* ATCC 25416, suspended in 150 μ l 10 mM NaP, pH 7.3, containing 1% TSB, were treated with Pal-GR13 (Pal-GRRGGRRGGRRGR, 375 μ g/ml) for 60 min at ambient temperature and then analyzed by transmission electron microscopy (TEM) (a).

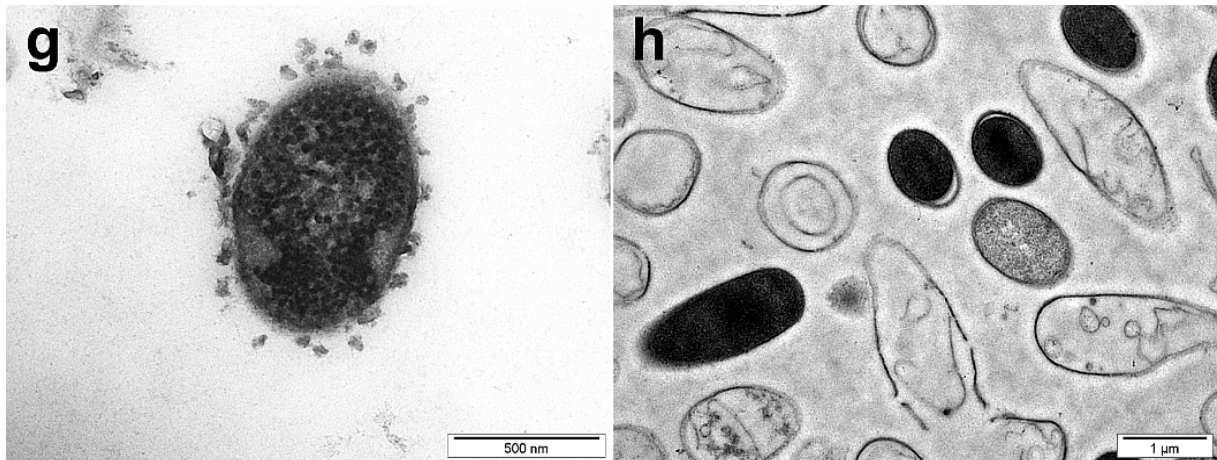
Burkholderia cepacia ATCC 25416, incubated with 10 mM NaP, pH 7.3, containing 1% TSB for 60 min at ambient temperature, served as control (b). Note condensation of electron-dense cytoplasmic material in Pal-GR13-treated bacteria (a). Images are representative of two independent experiments, sampling on average 10 images.

Supplementary Figure 22e, f



Supplementary Figure 22e, f | TEM of Pal-GR13-treated *Prevotella oralis*. 10^7 *Prevotella oralis* ATCC 33321, suspended in 150 μ l 10 mM NaP, pH 7.3, containing 1% TSB, were treated with Pal-GR13 (Pal-GRRGGRGGRGRGR, 375 μ g/ml) for 60 min at ambient temperature and aerobic conditions and then analyzed by transmission electron microscopy (TEM)(a). *Prevotella oralis* ATCC 33321, incubated with 10 mM NaP, pH 7.3, containing 1% TSB for 60 min at ambient temperature and aerobic conditions, served as control (b). Note condensation of electron-dense cytoplasmic material in Pal-GR13-treated bacteria (a). Images are representative of two independent experiments, sampling on average 10 images.

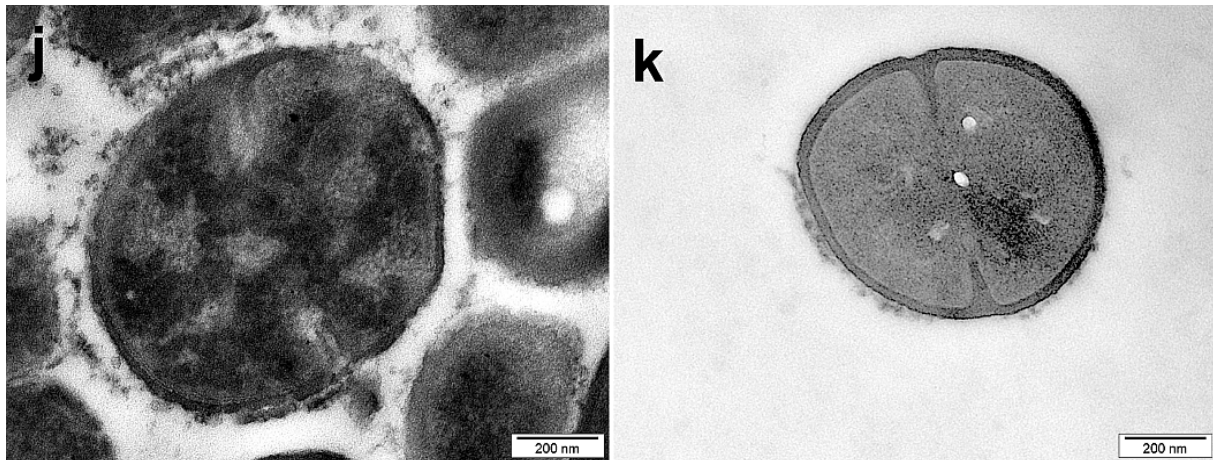
Supplementary Figure 22g, h



Supplementary Figure 22g, h | TEM of Pal-GR13-treated *Clostridium perfringens*.

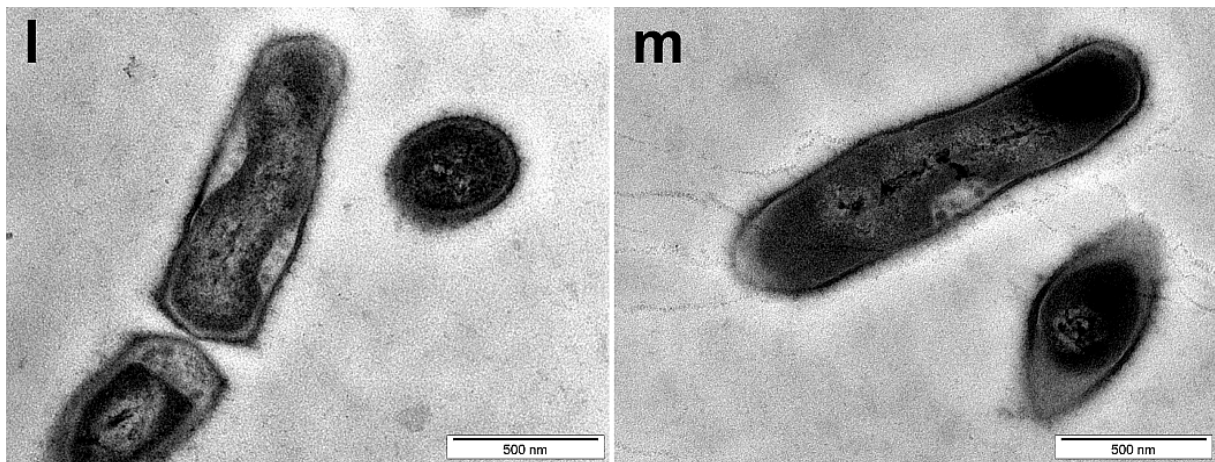
10^7 *Clostridium perfringens* ATCC 13124, suspended in 150 μ l 10 mM NaP, pH 7.3, containing 1% TSB, were treated with Pal-GR13 (Pal-GRRGGRRGRGR, 375 μ g/ml) for 60 min at ambient temperature and aerobic conditions and then analyzed by transmission electron microscopy (TEM)(a). *Clostridium perfringens* ATCC 13124, incubated with 10 mM NaP, pH 7.3, containing 1% TSB for 60 min at ambient temperature and aerobic conditions, served as control (b). Note condensation of electron-dense cytoplasmic material and liberation of mesosome-like vesicles in Pal-GR13-treated bacteria (a). Images are representative of two independent experiments, sampling on average 10 images.

Supplementary Figure 22j, k



Supplementary Figure 22j, k: TEM of Pal-GR13-treated *Staphylococcus epidermidis*. 10^7 *Staphylococcus epidermidis* ATCC 14990, suspended in 150 μ l 10 mM NaP, pH 7.3, containing 1% TSB, were treated with Pal-GR13 (Pal-GRRGGRRGGRRGR, 375 μ g/ml) for 60 min at ambient temperature and then analyzed by transmission electron microscopy (TEM)(**a**). *Staphylococcus epidermidis* ATCC 14990, incubated with 10 mM NaP, pH 7.3, containing 1% TSB for 60 min at ambient temperature, served as control (**b**). Note condensation of electron-dense cytoplasmic material and liberation of mesosome-like vesicles in Pal-GR13-treated bacteria (**a**). Images are representative of two independent experiments, sampling on average 10 images.

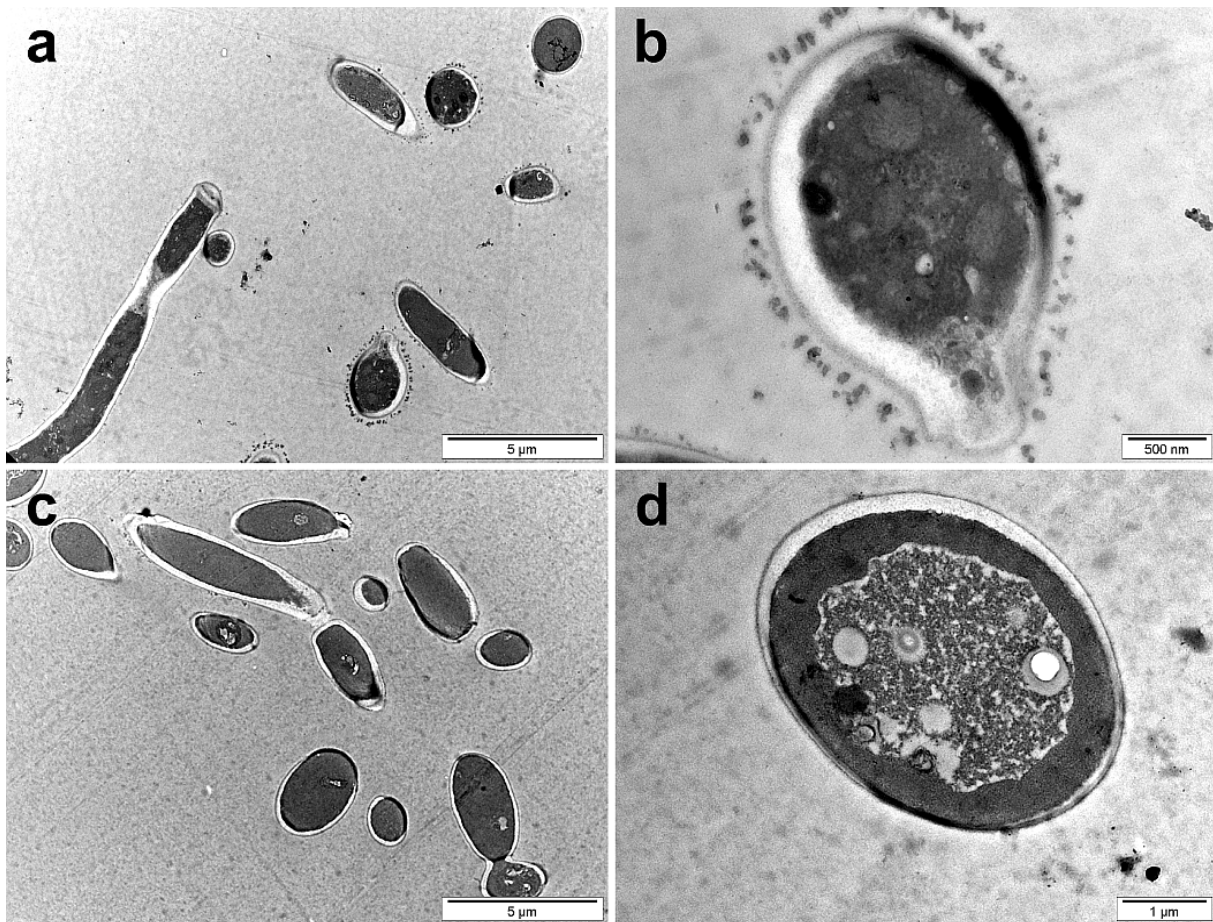
Supplementary Figure 22 I, m



Supplementary Figure 22 I, m | TEM of Pal-GR13-treated *Propionibacterium acnes*. 10^7

Propionibacterium acnes ATCC 6919, suspended in 150 μ l 10 mM NaP, pH 7.3, containing 1% TSB, were treated with Pal-GR13 (Pal-GRRGGRGGRGRGR, 375 μ g/ml) for 60 min at ambient temperature and aerobic conditions and then analyzed by transmission electron microscopy (TEM)(a). *Propionibacterium acnes* ATCC 6919, incubated with 10 mM NaP, pH 7.3, containing 1% TSB for 60 min at ambient temperature, served as control (b). Note condensation of electron-dense cytoplasmic material in Pal-GR13-treated bacteria (a). Images are representative of two independent experiments, sampling on average 10 images.

Supplementary Figure 23



Supplementary Figure 23 | TEM of Pal-GR13-treated *C. albicans*. 10^7 *C. albicans* ATCC 24433, suspended in 150 μ l 10 mM NaP, pH 7.3, containing 1% TSB, were treated with Pal-GR13 (Pal-GRRGGRRGGRRGR, 375 μ g/ml) for 60 min at ambient temperature and then analyzed by transmission electron microscopy (TEM) (a, b). *C. albicans* ATCC 24433, incubated with 10 mM NaP, pH 7.3, containing 1% TSB for 60 min at ambient temperature, served as control (c, d). Note nuclear blebs and chromatin margination and condensation in the nucleus in Pal-GR13-treated bacteria (a, b). Images are representative of two independent experiments, sampling on average 10 images.

Supplementary Tables

Supplementary Table 1 | Amino acid sequences of recombinant HRNR fragments used in this study.

Name	Amino Acid Sequence
rHRNR ₁₀₇₅₋₁₁₇₂	T H G Q H G S T S G Q S S S C G Q H G A S S G Q S S S H G Q H G S G S S Q S S G Y G R Q G S G S G Q S P G H G R G S G S R Q S P S Y G R H G S G S G R S S S S G Q H G S G L G E S S G F G H H E S
rHRNR ₂₅₉₁₋₂₆₈₄	S Q H G S G S G H S S G Y G Q H G S R S G Q S S R G E R H G S S S G S S S H Y G Q H G S G S R Q S S G H G R Q G S G S G Q S P S R G R H G S G Y G H S S S H G Q H G S G S G R S S S R G P Y
rHRNR ₂₆₃₈₋₂₆₈₄	Q S S G H G R Q G S G S G Q S P S R G R H G S G Y G H S S S H G Q H G S G S G R S S S R G P Y
rHRNR ₂₆₅₆₋₂₆₈₄	G R H G S G Y G H S S S H G Q H G S G S G R S S S R G P Y
rHRNR ₂₅₉₁₋₂₆₄₄	S Q H G S G S G H S S G Y G Q H G S R S G Q S S R G E R H G S S S G S S S H Y G Q H G S G S R Q S S G H G R
rHRNR ₂₅₇₆₋₂₇₀₇	G F G H H E S S S G Q S S S Y Q H G S G S G H S S G Y G Q H G S R S G Q S S R G E R H G S S S G S S S H Y G Q H G S G S R Q S S G H G R Q G S G S G Q S P S R G R H G S G Y G H S S S H G Q H G S G S R S S R G P Y E S R S G H S S V F G Q H E S G S G H S S A Y
rHRNR ₂₇₂₇₋₂₈₅₀	S G Q S T F D Q E G S S T G Q S S S Y G H R G S G S S Q S S G Y G R H G A G S G Q S L S H G R H G S G S G Q S S S Y G Q H G S G S G Q S S G Y S Q H G S G S G Q D G Y S Y C K G G S N H D G G S S G S Y F L S F P S T S P Y E Y V Q E Q R C Y F Y Q
SUMO3-His-tag	M G H H H H H H G G M S E E K P K E G V K T E N D H I N L K V A G Q D G S V V Q F K I K R H T P L S K L M K A Y C E R Q G L S M R Q R F R F D G Q P I N E T D T P A Q L E M E D E D T I D V F Q Q Q T G G

Residues cationic at pH 7, cationic only at acidic pH and anionic at pH >5 are colored blue, green and magenta, respectively.

Supplementary Table 2 | Tryptic soybean broth (TSB) inhibits *P. aeruginosa* sensitivity towards HRNR₂₆₅₆₋₂₆₇₇ (HR 1-18).

AA-sequence	Net charge*	Name	LD90 (µg/ml)	LD100 (µg/ml)	Condition**
GRHGSGLGHSSSGQHGSGR	+6	HR 1-18, HRNR ₂₆₅₆₋₂₆₇₇	0.038	0.15	0% TSB
GRHGSGLGHSSSGQHGSGR	+6	HR 1-18, HRNR ₂₆₅₆₋₂₆₇₇	1.18	9.38	1% TSB
GRHGSGLGHSSSGQHGSGR	+6	HR 1-18, HRNR ₂₆₅₆₋₂₆₇₇	37.5	>300	3% TSB
GRHGSGLGHSSSGQHGSGR	+6	HR 1-18, HRNR ₂₆₅₆₋₂₆₇₇	>300	>300	10% TSB

*At pH 5.5. **Sensitivity of *P. aeruginosa* ATCC 10145 was examined in 10 mM NaP/0.25%

glucose/pH 5.5. Residues cationic at pH 7 and cationic only at acidic pH are colored blue and green, respectively. Representatives are shown (n=2).

Supplementary Table 3 | *P. aeruginosa*-cidal activity of HR 1-18 and its variants.

AA-sequence	Net charge*	Name	n	LD90 (µg/ml)	LD100 (µg/ml)	Condition***
GRHGSGLGHSSSHGQHSGSGR	+6	HR 1-18, HRNR ₂₆₅₆₋₂₆₇₇	7	0.038	0.15	pH5.5
LRHGSGLGHSSSHGQHSGSGR	+6	HR 1-18-Leu	2	4.7	>300	pH5.5
GRHGSGLGHSSSHGQHSGSGR	+5	HR 1-18,L-G/Q-G	2	0.038	0.15	pH5.5
GRHGGGCGHGGCGGHGGGGR	+6	HR 1-18,L-G/Q-G/6S-6G	2	0.038	0.15	pH5.5
GRHGGGCGHGGCGGHGGGGR	+6	HR 1-18,G+H	2	<0.019	0.075	pH5.5
GRHGGGCGHGGCGGHGGGGR	0	HR 1-18,G+H	2	>300	>300	pH7.3
GRRGSGLGRHSSSHGGRGSGSGR	+6	HR 1-18,4H-4R	2	0.038	0.15	pH5.5
GRRGGGCGHGGCGGHGGGGR	+6	HR 1-18,G+R	2	<0.019	0.075	pH5.5
GRRGSGLGRHSSSHGGRGSGSGR	+6	HR 1-18,4H-4R/L-G	2	0.038	0.15	pH5.5
SRRSSLSRHSSSHGGRGSGSGR	+6	HR 1-18,4H-4R/9G-9S	2	0.15	1.18	pH5.5
Biot-GRHGSGLGHSSSHGQHSGSGR	+6	Biotin-HR1-18	2	>300/1.18**	>300	pH5.5

*At pH 5.5. **Antimicrobial paradox¹⁶. ***Sensitivity of *P. aeruginosa* ATCC 10145 was examined in 10 mM NaP/0.25% glucose/pH 5.5 or pH 7.3. In AA-sequences, residues cationic at pH 7 and cationic only at acidic pH are colored blue and green, respectively. AA replaced by Gly, Arg, Ser or Leu or biotinylated are colored magenta. Note the increase of the LD in a Leu-mutant of HR1-18 (colored yellow) and the loss of antimicrobial activity in a Gly- and His-containing HR1-18 mutant at neutral pH (LD values colored blue and red, respectively). n: number of experiments.

Supplementary Table 4 | *P. aeruginosa*-killing by HR1-17 mutants.

AA-sequence	Name	Net charge*	LD90 (µg/ml)	LD100 (µg/ml)
GRHGSGLGHSSSHGQHGSQSGR ^{SSSR} GPY	HR 1-17, HRNR ₂₆₅₆₋₂₆₈₄	+7	0.038	0.15
GRHGSGLGHSSSHGQHGSQSGR ^{SSSR} GPY	HR 1-17R1-G, HRNR ₂₆₅₆₋₂₆₈₄ R2657G	+6	0.038	0.3
GRHGSGLGHSSSHGQHGSQSGR ^{SSSR} GPY	HR 1-17R2-G, HRNR ₂₆₅₆₋₂₆₈₄ R2677G	+6	0.038	0.15
GRHGSGLGHSSSHGQHGSQSGR ^{SSSR} GPY	HR 1-17R3-G, HRNR ₂₆₅₆₋₂₆₈₄ R2681G	+6	0.038	0.15-0.3
GRHGSGLGHSSSHGQHGSQSGR ^{SSSR} GPY	HR 1-174H-4G, HRNR ₂₆₅₆₋₂₆₈₄ all H vs G	+3	75	>300

*At pH 5.5. Residues cationic at pH 7 and cationic only at acidic pH are colored blue and green, respectively. Amino acids replaced in wild type HR1-17 by Gly are colored magenta. *P. aeruginosa* ATCC 10145-cidal activity was examined in 10 mM NaP/0.25% glucose/pH 5.5. All data shown are representatives (n=2).

Supplementary Table 5 | *P. aeruginosa*-cidal activity of HR1-18 does not depend on its AA-sequence and chirality.

AA-sequence	Net charge*	Name	LD90 (µg/ml)	LD100 (µg/ml)
GRHGSGLGHSSSHGQHGSGSGR	+6	HR 1-18, HRNR ₂₆₅₆₋₂₆₇₇	0.075	0.59
GR _D H _D GS _D GL _D GH _D S _D S _D S _D H _D GQ _D H _D GS _D GS _D GR _D	+6	(D)HR 1-18, HRNR ₂₆₅₆₋₂₆₇₇	0.038	0.3
RGS _D SG _D HG _D SS _D SH _D GL _D SG _D H _D RG	+6	rev-HR 1-18	<0.019	0.3
SLSSGHGSGHGQRGGHRS _D SG _D	+6	scr-HR 1-18	<0.019	0.15

*At pH 5.5. Residues cationic at pH 7 and cationic only at acidic pH are colored blue and green, respectively. *P. aeruginosa* ATCC 10145-cidal activity was examined in 10 mM NaP/0.25% glucose/pH 5.5. Data are representatives (n=3).

Supplementary Table 6 | HRNR peptides and its mutants are *P. aeruginosa*-cidal CIDAMPs.

AA-sequence	Net charge*	Name	n	LD90 (µg/ml)	LD100 (µg/ml)	Condition***
QSPSY	±0	HR 2-3, HRNR ₁₁₃₇₋₁₁₄₁	1	>300	>300	pH5.5, 1% TSB
SSSSR	+1	HR 1-9, HRNR ₂₆₇₈₋₂₆₈₁	1	>300	>300	pH5.5, 1% TSB
SGQSSR	+1	HR 1-2, HRNR ₂₆₁₀₋₂₆₁₅	1	>300	>300	pH5.5, 1% TSB
SSSRGPY	+1	HR 1-10, HRNR ₂₆₇₈₋₂₆₈₄	1	>300	>300	pH5.5, 1% TSB
QSPSYGR	+1	HR 2-6, HRNR ₁₁₃₇₋₁₁₄₃	1	>300	>300	pH5.5, 1% TSB
SGQSSRGR	+1	HR 1-3, HRNR ₂₆₁₀₋₂₆₁₈	1	>300	>300	pH5.5, 1% TSB
GSGSRQSPSY	+1	HR 2-2, HRNR ₁₁₃₂₋₁₁₄₁	1	>300	>300	pH5.5, 1% TSB
QGSGSQSPSR	+1	HR 1-6, HRNR ₂₆₄₅₋₂₆₅₅	1	>300	>300	pH5.5, 1% TSB
QSSGR	+2	HR 1-5, HRNR ₂₆₃₈₋₂₆₄₄	1	>300	>300	pH5.5, 1% TSB
SGSGSR	+2	HR 2-9, HRNR ₁₁₄₄₋₁₁₅₀	1	>300	>300	pH5.5, 1% TSB
GSGSRQSPSYGR	+2	HR 2-8, HRNR ₁₁₃₂₋₁₁₄₃	1	37.5	>300	pH5.5, 1% TSB
QGSGSQSPSRGR	+2	HR 1-7, HRNR ₂₆₄₅₋₂₆₅₇	1	>300	>300	pH5.5, 1% TSB
QSPSYGRSGSGSR	+3	HR 2-5, HRNR ₁₁₃₇₋₁₁₅₀	2	>300	>300	pH5.5, 1% TSB
GSGSRQSPSYGRQGSGSGR	+3	HR 2-4Q, HRNR ₁₁₃₂₋₁₁₅₀ H1144Q	1	>300	>300	pH5.5, 1% TSB
GSGSRQSSGRGRQGSGSGQ	+3	HR 1-16, HRNR ₂₆₃₂₋₂₆₅₁	1	>300	>300	pH5.5, 1% TSB
QGSGSQSPSRGRQSGSGRQSPSY	+3	HR 2-1, HRNR ₁₁₁₈₋₁₁₄₁	1	>300	>300	pH5.5, 1% TSB
GRSGSGSQSSSYSPYSGSGWSSSR	+3	HRNR ₂₄₂₂₋₂₄₄₇	2	18.75/150**	>300	pH5.5, 1% TSB
GRSGSGSQSSSYSPYSGSGWSSSRGPY	+3	HRNR ₁₉₅₂₋₁₉₈₀	2	18.75/150**	>300	pH5.5, 1% TSB
GRSGSGSQSSSYSPYSGSGWSSSRGPY	+3	HRNR ₂₄₂₂₋₂₄₅₀	2	2.35/>300**	>300	pH5.5, 1% TSB
RSGLGSSS	+4	HR 1-27, HRNR ₂₆₅₇₋₂₆₆₈	1	75-100*	1.18/300*	pH5.5, 0.25% glucose
GRSGGLGSSS	+4	HR 1-37, HRNR ₂₆₅₆₋₂₆₆₈	2	9.38	>300	pH5.5, 0.25% glucose
SSSGQSGSGSR	+4	HR 1-38, HRNR ₂₆₆₄₋₂₆₇₇	2	>300/9.38**	>300	pH5.5, 0.25% glucose
SRQSPSYGRSGSR	+4	HR 2-15, HRNR ₁₁₃₅₋₁₁₄₈ S1148R	1	75	>300	pH5.5, 1% TSB
RQSPSYGRSGSGSR	+4	HR 2-14, HRNR ₁₁₃₆₋₁₁₅₀	1	300	>300	pH5.5, 1% TSB
GSGSRQSPSYGRSGSGSR	+4	HR 2-4, HRNR ₁₁₃₂₋₁₁₅₀	4	>300	>300	pH5.5, 1% TSB
SRQSPSYGRSGSGSR	+4	HR 2-13, HRNR ₁₁₃₅₋₁₁₅₀	1	>300	>300	pH5.5, 1% TSB
GSRQSPSYGRSGSGSR	+4	HR 2-12, HRNR ₁₁₃₄₋₁₁₅₀	1	>300	>300	pH5.5, 1% TSB
GSGSRQSSYGRSGSGSR	+4	HR 2-4S, HRNR ₁₁₃₂₋₁₁₅₀ P1139S	2	150	>300	pH5.5, 1% TSB
GSGSRQSLYGRSGSGSR	+4	HR 2-4L, HRNR ₁₁₃₂₋₁₁₅₀ P1139L	1	75	>300	pH5.5, 1% TSB
SGSSSSSYGQSGSGSR	+4	HR 1-4, HRNR ₂₆₁₉₋₂₆₃₇	1	150	>300	pH5.5, 1% TSB
SGSGSGSSGYGQSGSR	+4	HR 1-1, HRNR ₂₅₉₁₋₂₆₀₉	1	>300	>300	pH5.5, 1% TSB
GRSGSGSQSSSYGPYSGSGWSSSRGPY	+4	HRNR ₁₀₁₃₋₁₀₄₁	2	1.18/75**	>300	pH5.5, 1% TSB
RSGLGSSS	+5	HR 1-26, HRNR ₂₆₅₇₋₂₆₇₁	2	0.3/150**	>300/2,35	pH5.5, 0.25% glucose
GRSGGLGSSSGQ	+5	HR 1-25, HRNR ₂₆₅₆₋₂₆₇₁	2	0.3	4.7	pH5.5, 0.25% glucose
SGRQSGSRQSPSYGR	+5	HR 2-16, HRNR ₁₁₂₈₋₁₁₄₄	2	0.59/300**	37.5	pH5.5, 0.25% glucose
GSGSRQSPSRGRSGSGSR	+5	HR 2-4R, HRNR ₁₁₃₂₋₁₁₅₀ Y1141R	2	37.5/300**	>300	pH5.5, 1% TSB
GSGSRQSPSRGRSGSGSR	+5	HR 2-4H, HRNR ₁₁₃₂₋₁₁₅₀ Y1141H	2	75	>300	pH5.5, 1% TSB
SGSLGSSSGQSGSGSR	+5	HR 1-8, HRNR ₂₆₅₈₋₂₆₇₇	1	18.75/>300**	>300	pH5.5, 1% TSB
RSGLGSSSGQSGSGSR	+5	HR 1-8, HRNR ₂₆₅₈₋₂₆₇₇	1	4.7	>300	pH5.5, 0.25% glucose
SGSRSGQSSRGRSGSSSS	+5	HR1-11, HRNR ₂₆₀₆₋₂₆₂₈	2	0.075	2.35	pH5.5, 0.25% glucose
GSGSRQSPSYGRSGSGSRSSSSGQ	+5	HR 2-11, HRNR ₁₁₃₂₋₁₁₅₇	2	37.5	>300	pH5.5, 1% TSB
GRSGSGSSSGQSGSGSR	+6	HRNR ₂₁₈₆₋₂₂₀₇	2	0.15/75**	4.7-9.37**	pH5.5, 0.25% glucose
GRSGGLGSSSGQSGSGSR	+6	HR 1-18, HRNR ₂₆₅₆₋₂₆₇₇	7	0.038	0.15	pH5.5, 0.25% glucose
LRSGGLGSSSGQSGSGSR	+6	HR 1-18-Leu,	2	4.7	>300	pH5.5, 0.25% glucose
LGSSSGQSGSGSRSSSRGPYPSRLG	+6	HR 1-36, HRNR ₂₆₆₃₋₂₆₉₀	2	0.15	2.35	pH5.5, 0.25% glucose
GRSGGLGSSSGQSGSGSRSSSRGPY	+7	HR 1-17, HRNR ₂₆₅₆₋₂₆₈₄	2	0.038	0.15	pH5.5, 0.25% glucose
GRSGSGSSGYTQSGSGSSSYGRSGSR	+8	HRNR ₉₃₃₋₉₉₄	2	0.3	2.35	pH5.5, 0.25% glucose
GRSGSGSRQSPSRGRSGSGSSSGQSGSR	+9	HRNR ₂₉₅₋₃₆₁	2	0.15	0.59	pH5.5, 0.25% glucose
GSSYSYRCHYSGSGTSGFGQSGSGSSSEY	+9	HRNR ₉₉₅₋₁₀₅₆	2	0.3	2.35	pH5.5, 0.25% glucose
LRSGGLGSSSGQSGSGSR	+10	HRNR ₂₃₂₋₂₉₄	2	0.3	2.35	pH5.5, 0.25% glucose

*At pH 5.5. **Antimicrobial paradox¹⁶. ***Sensitivity of *P. aeruginosa* ATCC 10145 was examined in 10 mM NaP. Residues cationic at pH 7, cationic only at acidic pH, anionic and mutated are colored blue, green, magenta and yellow, respectively. Note the marked increase of the LD value when a selected AA is replaced in defined HRNR-peptides (LD values colored in yellow). n: number of experiments.

Supplementary Table 7 | IDPRs of FLG-2, FLG and Repetin are *P. aeruginosa*-cidal CIDAMPs.

AA-sequence	Name	n	Net charge*	LD90 (µg/ml)	LD100 (µg/ml)
TAGRRGSG	FLG-2 ₂₀₉₇₋₂₁₀₅	1	+3	>300	>300
GSRTAGRRGSG	FLG-2 ₂₀₉₄₋₂₁₀₅	1	+4	>300/2.35**	>300
HAHSGHGQSTQR	FLG-2 ₂₀₈₂₋₂₀₉₃	2	+4	0.15	2.35
HTHSGHTHGQSGSQHGESESIIDR	FLG-2 ₁₅₁₅₋₁₅₃₉	2	+4	0.075	0.59
HAHSGHGQSTQRGSR	FLG-2 ₂₀₈₂₋₂₀₉₆	2	+5	0.15	1.18
HTHSGHTHGQAGSHYPESGSSVHER	FLG-2 ₁₉₆₉₋₁₉₉₃	2	+5	0.038	0.3
HTHSGHAHGQAGSQHGESGSSVHER	FLG-2 ₂₀₄₄₋₂₀₆₈	1	+5	0.038	0.3
HTHSGHTHGQARSQHGESISIVHER	FLG-2 ₁₇₄₂₋₁₇₆₆	2	+5	0.038	0.15
THHEQSHQRRDRQTHEDKQNRQR	RPTN ₇₄₇₋₇₇₀	2	+5	0.3	150
HGQSTQRGSRTAGRRGSG	FLG-2 ₂₀₈₇₋₂₁₀₅	2	+6	0.075	1.18
HAHSGHGQSTQRGSRTAGR	FLG-2 ₂₀₈₂₋₂₁₀₀	2	+6	0.038	0.3
HAHYGYGQSTQRGSRTTGRRGSG	FLG-2 ₂₂₃₂₋₂₂₅₅	2	+7	0.075	0.59
QGSHHHQSVNRSGHSGSHSHHTTSQGR	FLG ₅₂₈₋₅₅₄	2	+7	0.15	0.59
HTHSGHTHSQARSQHGESISTVHKR	FLG-2 ₁₈₉₄₋₁₉₁₈	2	+7	0.038	0.3
HAHSGHGQSTQRGSRTAGRRGSG	FLG-2 ₂₀₈₂₋₂₁₀₅	2	+8	< 0.038	0.075

*At pH 5.5. **Antimicrobial paradox¹⁶. Residues cationic at pH 7, cationic only at acidic pH and anionic are colored blue, green and magenta, respectively. Sensitivity of *P. aeruginosa* ATCC 10145 was tested in 10 mM NaP/0.25% glucose/pH5.5. n: number of experiments.

Supplementary Table 8 | CIDAMPs with *S. aureus* ATCC 6538-cidal activity

AA-sequence	Name	Net charge*	n	LD90 (µg/ml)	LD100 (µg/ml)	Condition**
GR ^{blue} GSGLG ^{blue} SSSS ^{green} GQ ^{blue} GSGSGR ^{blue}	HR 1-18, HRNR ₂₆₅₆₋₂₆₇₇	+6	2	>300	>300	pH5.5
LR ^{magenta} GSGLG ^{blue} SSSS ^{green} GQ ^{blue} GSGSGR ^{blue}	HR 1-18-Leu G2656Leu	+6	2	300	>300	pH5.5
GRR ^{blue} GSGLGR ^{blue} SSSR ^{green} GGRR ^{blue} GSGSGR ^{blue}	HR 1-18,4H-4R	+6	2	150	>300	pH5.5
GRR ^{blue} GSGLGR ^{blue} SSSR ^{green} GGRR ^{blue} GSGSGR ^{blue}	HR 1-18,4H-4R	+6	2	9.375	>300, max.killing: 98.8%	pH7.0
GRR ^{blue} GSGLGR ^{blue} SSSR ^{green} GGRR ^{blue} GSGSGR ^{blue}	HR 1-18,4H-4R/L-G	+6	1	>300	>300	pH5.5
GRR ^{blue} GG ^{blue} GR ^{blue} GGRR ^{blue} GGRR ^{blue} GGRR ^{blue}	HR 1-18,G+R	+6	1	>300	>300	pH5.5
SR ^{magenta} SS ^{green} SL ^{blue} SR ^{blue} SSSR ^{green} SSSR ^{green} SSSR ^{green}	HR 1-18,4H-4R/9G-9S	+6	1	150	>300	pH5.5
SR ^{magenta} SS ^{green} SL ^{blue} SR ^{blue} SSSR ^{green} SSSR ^{green} SSSR ^{green}	HR 1-18,4H-4R/9G-9S	+6	2	75	>300	pH7.2
GRR ^{blue} GGRR ^{blue} GGRR ^{blue}	GR13	+6	2	75	>300	pH5.5
GRR ^{blue} GGRR ^{blue} GGRR ^{blue}	GR13	+6	1	75	>300	pH7.0
SH ^{blue} RC ^{blue} CR ^{blue} SH ^{blue} RCRR ^{blue}	LCE-3B ₅₆₋₆₈	+8	2	0.075	>150, max.killing: 99.3%	pH5.5
SH ^{blue} RR ^{blue} FR ^{blue} SH ^{blue} QCRR ^{blue}	LCE-3C ₅₅₋₆₇	+8	2	2.35	>150, max.killing: 97.7%	pH5.5
SH ^{blue} RR ^{blue} FR ^{blue} SH ^{blue} QCRR ^{blue} QR ^{blue} SN ^{blue} SC ^{blue} DR ^{blue}	LCE-3C ₅₅₋₇₅	+9	2	0.59	>150, max.killing: 99.4%	pH5.5
SH ^{blue} RPRL ^{blue} F ^{blue} RRRR ^{blue}	LCE-2B ₇₄₋₈₆	+9	2	1.18	>150, max.killing: 97.4%	pH5.5

*At pH 5.5. ** *S. aureus* ATCC 6538 sensitivity was examined in 10 mM NaP/0.25% glucose/pH5.5, pH 7.0 or pH 7.2. Residues cationic at pH 7 and cationic only at acidic pH are colored blue and green, respectively. Amino acids in HR1-18 replaced by Gly, Arg, Ser or Leu are colored magenta. Note the higher antimicrobial potency of selected CIDAMPs at neutral pH (shaded) and the highest potency of the “Late-Cornified-Envelope (LCE) protein” fragment LCE-3B₅₆₋₆₈ at acidic pH (bold). n: number of experiments.

Supplementary Table 9 | *S. aureus* ATCC 6538 -bactericidal activity of S-palmitoylated HRNR-CIDAMPs.

AA-sequence	Name	n	Net charge	LD90 (µg/ml)	LD100 (µg/ml)	Condition***
GQHGSSSGHSSTHGQHGSTSGQSSSC-Pal*	HRNR ₅₉₈₋₆₂₃	2	+4	2.35	>150	pH5.5, 0.25% glucose
GQHGSSSGHSSTHGQHGASGQSSSC-Pal*	HRNR ₂₀₀₄₋₂₀₂₉	2	+4	0.59	>150/75**	pH5.5, 0.25% glucose
Pal-CGQHGATSGQSSSHGQHGSGSSQSSR	HRNR ₆₂₃₋₆₄₈	2	+4	2.35	>150	pH5.5, 0.25% glucose
Pal-CGQHGATSSGQSSSHGQHGSGSSQSSGYGR	HRNR ₁₀₈₉₋₁₁₁₇	2	+4	>150	>150	pH5.5, 0.25% glucose
GRHGSGSGQSSGFCHHESSSWQSSGC-Pal*	HRNR ₁₃₈₉₋₁₄₁₄	2	+4	1.18	18.8	pH5.5, 0.25% glucose
SGHSSVFGQHSGSGHSSAYSQHSGSGHFC-Pal*	HRNR ₁₇₄₈₋₁₇₇₈	2	+4	0.3	9.38	pH5.5, 0.25% glucose

*amide, **antimicrobial paradox¹⁶. ***Antimicrobial activity was examined in 10 mM NaP. Residues cationic at pH 7, cationic only at acidic pH, anionic and S-palmitoyl-Cys are colored blue, green, magenta and yellow, respectively. Hydrophobic residues are shaded grey. n: number of experiments.

Supplementary Table 10 | Sensitivity of *Corynebacterium simulans* and *P. aeruginosa* towards selected FLG- and HRNR-derived CIDAMPs

AA-sequence	Name	n	Net charge*	LD90 <i>C. simulans</i> (µg/ml)	LD100 <i>C. simulans</i> (µg/ml)	LD90 <i>P. aeruginosa</i> (µg/ml)	LD100 <i>P. aeruginosa</i> (µg/ml)
QGS ^H HE ^H QSVNR ^R SG ^H SGS ^H HS ^H HTTSQGR	FLG ₅₂₈₋₅₅₄	1/2	+7	>150	>150	0.15	0.59
^H TQNSSSGQAASS ^H EQAR	FLG ₆₈₄₋₇₀₁	1/1	+3	>150	>150	>300	>300
QSGTR ^H ETSSGGQAASS ^H EQAR	FLG ₁₆₅₂₋₁₆₇₄	1/1	+1	>150	>150	>300	>300
QSGT ^H HA ^H TNSSGGQAASS ^H EQAR	FLG ₂₃₀₁₋₂₃₂₃	1/1	+2	>150	>150	>300	300
QSGTR ^H ETQTSSGGQAASS ^H EQAR	FLG ₂₆₂₅₋₂₆₄₇	1/1	+3	>150	>150	>300	>300
QSGTR ^H EA ^H TSSGGQAASS ^H EQAR	FLG ₃₂₇₃₋₃₂₉₅	1/1	+2	>150	>150	>300	300
SG ^H KSSSGQSSGY ^Q HGSGSG ^H SSGYG ^Q HGSR ^R SGQSSR	HRNR ₂₃₂₋₂₉₄	2/1	+11	1.18	>150/37.5**	0.3	2.35
^G DR ^R HRSSSGSSSSSYG ^Q HGSGSRQSL							
^G HGRQSGSR ^R QSPS ^H VR ^H HGSGSG ^H SSS ^H GQ ^H HGSGSSYS	HRNR ₂₉₅₋₃₆₁	2/1	+10	0.59	75/9.375**	0.15	0.59
YSR ^G HY ^E SGSGQTSGFGQ ^H ESGSGQSSGY							
^G HKSSSGQSSGYT ^Q HGSGSG ^H SSSYE ^Q HGSR ^R SGQSSRS	HRNR ₉₃₃₋₉₉₄	1/1	+8	4.7	>150	0.3	2.35
^E QHSSSGSSSSSYG ^Q HGSGSRQSL							
LG ^H HGQ ^H HGSGSGQSPSPS ^R GR ^H HGSGSGQSSSYG ^P YRSGS	HRNR ₉₉₅₋₁₀₅₆	1/1	+8	4.7	>150	0.3	2.35
GWSSSR ^G PY ^E SGSG ^H SSSLG ^H RES ^R							

*at pH 5.5. **antimicrobial paradox¹⁶. Residues cationic at pH 7, cationic only at acidic pH and

anionic are colored blue, green and magenta, respectively. Sensitivity of *P. aeruginosa* ATCC 10145 towards the peptides was tested at pH 5.5 and towards *Corynebacterium simulans* DSM 44415 it was tested at pH 6.0 in 10 mM NaP/0.25% glucose. n: number of experients.

Supplementary Table 11 | Palmitoylated or myristoylated CIDAMPs containing Arg, His, Lys, Dab or Dap as cationic AAs are bactericidal for *P. aeruginosa* ATCC 10145

AA-sequence	Name	n	LD100 (µg/mL)	LD90 (µg/mL)	Condition
Pal-GRHGSGLGHSSSHGQHGSQSGSR	Pal-HR 1-18, Pal-HRNR ₂₆₅₆₋₂₆₇₇	2	0.59	0.075	pH5.5, 0.25% glucose
Myr-GRHGSGLGHSSSHGQHGSQSGSR	Myr-HR 1-18, MyrHRNR ₂₆₅₆₋₂₆₇₇	1	0.3	0.075	pH5.5, 0.25% glucose
Pal-GRRGSGGLGRSSSRGQRGSGSGSR	Pal-HR 1-18-HR	2	0.59	0.038	pH5.5, 0.25% glucose
Pal-GRRGSGGLGRSSSRGQRGSGSGSR	Pal-HR 1-18-HR	1	4.7	1.18	pH7.3, 1% TSB
Pal-HGSRSGQSSRGERHGSSSGSSSH	HR 1-11, HRNR ₂₆₀₆₋₂₆₂₈	2	0.3	0.075	pH5.5, 0.25% glucose
Pal-RGSRSGQSSRGERRRGSSSSGSSSR	Pal-HR 1-11-3H3R	2	0.15	0.038	pH5.5, 0.25% glucose
Pal-RGSRSGQSSRGERRR	Pal-HR 1-11-2H2R-14	1	0.15	0.038	pH5.5, 0.25% glucose
Pal-GHHGGHGGHGHGH	Pal-GH13	1	0.3	0.075	pH5.5, 0.25% glucose
Pal-GRRGGRGGRGRGR	Pal-GR13	1	0.3	0.038	pH5.5, 0.25% glucose
Pal-GR ₀ R ₀ GGR ₀ GGR ₀ GR ₀ GR ₀	(D)-Pal-GR13	1	0.3	0.075	pH5.5, 0.25% glucose
Pal-GRRGGRGGRGR	Pal-GR11	2	0.3	0.075	pH5.5, 0.25% glucose
Pal-GRRGGRGGR	Pal-GR9	1	0.59	0.038	pH5.5, 0.25% glucose
Pal-GR ₀ R ₀ GGR ₀ GGR ₀	(D)-Pal-GR9	1	0.59	0.038	pH5.5, 0.25% glucose
Pal-GRRGGR	Pal-GR6	1	4.7	1.18	pH5.5, 0.25% glucose
Pal-GRGR	Pal-GR4	2	9.375	2.35	pH5.5, 0.25% glucose
Pal-GR ₀ GR ₀	Pal-(D)-GR4	1	150	18.75	pH5.5, 0.25% glucose
Pal-GKKGKGGKGGKGGKGGK	Pal-K-GR13	1	0.15	<0.019	pH5.5, 0.25% glucose
Pal-GOOGGOGGGOGGOGO	Pal-Orn-GR13	1	0.3	0.075	pH5.5, 0.25% glucose
Pal-GKGGK	Pal-GK4	2	9.375	2.38	pH5.5, 0.25% glucose
Pal-GK ₀ GK ₀	Pal-(D)-GK4	1	4.7	1.18	pH5.5, 0.25% glucose
Pal-GOGO	Pal-GO4	1	18.75	2.35	pH5.5, 0.25% glucose
Pal-GO ₀ GO ₀	Pal-(D)-GO4	1	9.38	1.18	pH5.5, 0.25% glucose
Pal-GlyDabGlyDab	Pal-G-Dab4	1	9.38	2.35	pH5.5, 0.25% glucose
Pal-GlyDapGlyDap	Pal-G-Dap4	2	9.38	2.35	pH5.5, 0.25% glucose
Pal-GRRGSGGLGRSSSR	Pal-HR 1-18-3H3R-GR13	2	4.7	0.59	pH5.5, 0.25% glucose

Residues cationic at pH 7, cationic only at acidic pH and anionic at pH 7 are colored blue, green and magenta, respectively. Dab: L-2,4-Diaminobutyric acid, Dap:L-2,3-Diaminopropionic acid. n: number of experiments.

Supplementary Table 12 | Late cornified envelope protein (LCE)-peptide fragments and its palmitoylated derivatives are potent and efficient *S. aureus*-cidal antimicrobials.

AA-sequence	Name	Net Charge*	LD100 (µg/mL)	LD90 (µg/mL)	Condition
SHHRCCRS ^{SH} RCRR	LCE-3B ₅₆₋₆₈	+8	>150, max.killing: 99.3%	0.075	pH5.5, 0.25% glucose
SHHRHF ^{RS} HQCRR	LCE-3C ₅₅₋₆₇	+8	>150, max.killing: 97.7%	2.35	pH5.5, 0.25% glucose
SHHRHF ^{RS} HQCRRQR ^{SN} SC ^{DR}	LCE-3C ₅₅₋₇₅	+9	>150, max.killing: 99.4%	0.59	pH5.5, 0.25% glucose
Pal-SHHRCCRS ^{SH} RCRR	Pal- LCE-3B ₅₆₋₆₈	+8	0.15	<0.019	pH5.5, 0.25% glucose
Pal-SHHRHF ^{RS} HQCRR	Pal- LCE-3C ₅₅₋₆₇	+8	0.3	0.038	pH5.5, 0.25% glucose
Pal-SHHRHF ^{RS} HQCRRQR ^{SN} SC ^{DR}	Pal- LCE-3C ₅₅₋₇₅	+9	0.59	0.15	pH5.5, 0.25% glucose

Residues cationic at pH 7, cationic only at acidic pH and anionic are colored blue, green and magenta, respectively. *: at pH 5.5. Representatives are shown (n=2).

Supplementary Table 13 | Antimicrobial activity spectrum of Palmitoyl-HR1-18.*

Microorganism	n	LD100 (µg/mL)	LD90 (µg/mL)	Conditions
<i>Burkholderia cepacia</i> ATCC 25416	2	>300	75	pH 7.3, 0.25% glucose
<i>Burkholderia cepacia</i> ATCC 25416	2	>300	>300	pH 5.5, 0.25% glucose
<i>Klebsiella pneumoniae</i> ATCC 13883	1	1.18	0.3	pH 5.5, 0.25% glucose
<i>Prevotella oralis</i> ATCC 33321	1	>300	37.5	pH 6.5, 0.25 % glucose
<i>Prevotella oralis</i> ATCC 33321	2	150	18.75	pH 7.3, 1 % TSB
<i>Pseudomonas aeruginosa</i> ATCC10145	2	0.59	0.075	pH 5.5, 0.25% glucose
<i>Clostridium perfringens</i> ATCC 13124	1	37.5	18.75	pH 7.3, 1 % TSB
<i>Clostridium perfringens</i> ATCC 13124	1	75	37.5	pH 5.5, 0.25% glucose
<i>Clostridium perfringens</i> ATCC 13124	1	18.75	4.7	pH 7.3, 0.25% glucose
<i>Propionibacterium acnes</i> ATCC 6919	1	0.59	0.3	pH 5.5, 0.25% glucose
<i>Propionibacterium acnes</i> ATCC 6919	1	4.7	1.18	pH 7.3, 1 % TSB
<i>Propionibacterium acnes</i> ATCC 6919	1	2.35	0.59	pH 7.3, 0.25% glucose
<i>Staphylococcus aureus</i> ATCC 6538	2	37.5	1.18	pH 5.5, 0.25% glucose
<i>Staphylococcus aureus</i> ATCC 6538	1	>300	>300	pH 7.3, 1 % TSB
<i>Staphylococcus epidermidis</i> ATCC 14990	1	>300	>300	pH 7.3, 1 % TSB
<i>Streptococcus pneumoniae</i> ATCC 33400	2	9.37	2.35	pH 6.5, 0.25% glucose
<i>Streptococcus pneumoniae</i> ATCC 33400	1	37.5	9.38	pH 7.3, 1 % TSB
<i>Streptococcus pneumoniae</i> ATCC 33400	1	4.7	2.35	pH 7.3, 0.25% glucose
<i>Streptococcus pyogenes</i> ATCC 12344	2	0.59	0.15	pH 6.0, 0.25% glucose
<i>Streptococcus pyogenes</i> ATCC 12344	1	9.38	9.38	pH 7.3, 1 % TSB
<i>Streptococcus pyogenes</i> ATCC 12344	1	0.3	0.15	pH 7.3, 0.25% glucose

*AA-sequence: Pal-GRHGSGGLGHSSSHGQHGSGSGR. Sensitivity of bacteria towards Pal-HR1-18 was tested in a CFU assay system in 10 mM NaP, depending on the microbe, at pH 5.5, 6.5 or pH 7.3, which contained 0.25% glucose and 1% TSB, respectively. n: number of experiments.

Supplementary Table 14 | Antimicrobial activity spectrum of Palmitoyl-HR1-18HR.*

Microorganism	n	LD100 (µg/mL)	LD90 (µg/mL)	Conditions
<i>Burkholderia cepacia</i> ATCC 25416	2	150	75	pH 7.3, 1 % TSB
<i>Burkholderia cepacia</i> ATCC 25416	2	300	75	pH 7.3, 0.25% glucose
<i>Prevotella oralis</i> ATCC 33321	1	75	37.5	pH 7.3, 1 % TSB
<i>Pseudomonas aeruginosa</i> ATCC10145	1	4.7	1.18	pH 7.3, 1 % TSB
<i>Pseudomonas aeruginosa</i> ATCC10145	2	0.59	0.038	pH 5.5, 0.25% glucose
<i>Clostridium perfringens</i> ATCC 13124	1	9.375	4.7	pH 7.3, 0.25% glucose
<i>Propionibacterium acnes</i> ATCC 6919	1	4.7	1.18	pH 7.3, 1 % TSB
<i>Staphylococcus aureus</i> ATCC 6538	2	4.7	0.3	pH 5.5, 0.25% glucose
<i>Staphylococcus aureus</i> ATCC 6538	1	9.38	1.18	pH 7.3, 1 % TSB
<i>Streptococcus pneumoniae</i> ATCC 33400	1	9.38	9.38	pH 7.3, 0.25 % glucose
<i>Streptococcus pyogenes</i> ATCC 12344	1	0.3	0.15	pH 7.3, 0.25 % glucose

* AA-sequence: Pal-GRRGSLGRSSSRGQRGSGSGR. Sensitivity of bacteria towards Pal-HR1-18HR was tested in a CFU assay system in 10 mM NaP, depending on the microbe, which contained 0.25% glucose and 1% TSB, respectively, at pH 5.5 or pH 7.3. n: number of experiments.

Supplementary Table 15 | Results of a CIDAMP-MIC*-screening at CO-ADD

Sample Name	CO-ADD Sample ID	GN_001	GN_003	GN_004	GN_042	GP_020	FW_001	FW_002	MA_007
		<i>E. coli</i> ATCC 29222 FDA control	<i>K. pneumoniae</i> ATCC 70593 DSMZ	<i>A. baumannii</i> ATCC 19606 Type strain	<i>P. aeruginosa</i> ATCC 27853 Type strain	<i>S. aureus</i> ATCC 43300 MRSA	<i>C. albicans</i> ATCC 90028 CSLI Ref	<i>C. neoformans</i> ATCC 20811 H99	HEK293 ATCC CRL-1073
		MIC (µg/mL)							
		CGC (µg/mL)							
LCE-2B74-86	C0137532	>32 (14.8%) >32 (15.9%)	>32 (32.1%) >32 (31.8%)	>32 (17.5%) >32 (38.7%)	>32 (4.8%) >32 (10.9%)	>32 (45.1%) >32 (29.0%)	>32 (4.0%) >32 (-3.2%)	>32 (27.2%) >32 (7.0%)	>32
LCE-3B56-68	C0137533	>32 (22.5%) >32 (17.5%)	>32 (25.7%) >32 (24.1%)	>32 (18.4%) >32 (38.6%)	>32 (1.6%) >32 (11.6%)	>32 (48.1%) >32 (39.5%)	>32 (0.8%) >32 (0.1%)	8 (95.9%) 8 (104.2%)	>32
LCE-3C56-67	C0137534	>32 (15.0%) >32 (12.6%)	>32 (23.0%) >32 (19.3%)	>32 (9.6%) >32 (36.9%)	>32 (9.9%) >32 (11.3%)	>32 (47.4%) >32 (36.9%)	>32 (6.2%) >32 (5.7%)	8 (97.8%) 8 (102.3%)	>32
LCE-3C56-75	C0137535	>32 (15.5%) >32 (11.7%)	>32 (25.7%) >32 (20.5%)	>32 (7.4%) >32 (17.6%)	>32 (-0.4%) >32 (4.9%)	>32 (41.5%) >32 (25.2%)	>32 (10.0%) >32 (7.8%)	4 (105.8%) 4 (105.3%)	>32
Pal-HR1-18-HR	C0137536	4 (95.0%) 8 (96.0%)	32 (82.9%) 32 (91.1%)	4 (97.0%) 1 (98.3%)	16 (97.5%) 32 (95.9%)	4 (84.0%) 8 (83.4%)	16 (99.0%) 16 (98.9%)	1 (105.0%) 1 (106.1%)	>32
Pal-HR1-11-2H2R-14	C0137537	8 (57.8%) 16 (95.8%)	>32 (32.7%) >32 (36.7%)	8 (98.3%) 4 (98.2%)	32 (81.8%) 32 (93.4%)	16 (95.0%) 8 (91.1%)	>32 (36.3%) >32 (31.7%)	4 (107.3%) 4 (106.8%)	>32
Pal-GR13	C0137538	4 (96.9%) 8 (95.1%)	32 (91.9%) 16 (92.3%)	2 (96.0%) 4 (97.0%)	8 (96.3%) 8 (97.1%)	0.5 (96.6%) 0.25 (99.4%)	4 (99.4%) 4 (99.1%)	0.25 (109.2%) 0.25 (109.6%)	>32
Pal-GR9	C0137539	>32 (28.9%) >32 (36.6%)	>32 (25.4%) >32 (21.3%)	>32 (16.9%) >32 (28.1%)	>32 (13.4%) >32 (12.9%)	32 (84.0%) 32 (100.3%)	>32 (5.7%) >32 (2.7%)	>32 (-0.9%) >32 (10.3%)	>32
Pal-GR4	C0137540	>32 (14.2%) >32 (11.2%)	>32 (19.2%) >32 (23.1%)	32 (82.8%) 32 (79.6%)	>32 (5.5%) >32 (11.4%)	8 (97.4%) 8 (98.1%)	32 (79.8%) 8 (85.0%)	2 (103.9%) 2 (106.3%)	>32
Pal-G-Dap4	C0137541	>32 (15.7%) >32 (13.3%)	>32 (15.6%) >32 (11.4%)	>32 (15.5%) >32 (25.9%)	>32 (9.5%) >32 (10.6%)	8 (96.2%) 8 (99.0%)	8 (97.2%) 16 (94.3%)	1 (106.0%) 1 (105.9%)	>32
Pal-CE-3C55-67	C0137542	>32 (15.0%) >32 (17.3%)	>32 (29.9%) >32 (31.2%)	>32 (15.2%) >32 (25.9%)	>32 (10.1%) >32 (10.8%)	>32 (71.3%) >32 (59.0%)	16 (93.9%) 10 (93.6%)	2 (103.9%) 1 (107.8%)	>32
Pal-CE-3B56-68	C0137543	>32 (17.3%) >32 (18.9%)	>32 (30.1%) >32 (23.6%)	>32 (11.5%) >32 (30.3%)	>32 (10.5%) >32 (12.5%)	16 (96.9%) 16 (97.6%)	1 (95.4%) 2 (98.3%)	0.25 (105.7%) 0.25 (106.1%)	>32
S-Pal-HRNR1748-1778	C0137544	>32 (13.0%) >32 (11.6%)	>32 (17.2%) >32 (22.9%)	>32 (12.7%) >32 (21.6%)	>32 (5.2%) >32 (12.2%)	>32 (59.0%) >32 (32.0%)	>32 (16.4%) >32 (4.1%)	4 (108.0%) 4 (104.7%)	>32
Pal-HR1-18-GR13	C0137546	32 (95.8%) >32 (40.2%)	>32 (19.3%) >32 (13.9%)	16 (99.5%) 16 (99.5%)	>32 (16.8%) >32 (20.8%)	16 (100.2%) 32 (78.1%)	>32 (8.5%) >32 (-0.8%)	16 (104.6%) 32 (101.0%)	>32
Pal-HR1-18	C0142188	>32 (9.8%) >32 (15.6%)	>32 (11.5%) >32 (10.5%)	32 (91.0%) 32 (98.8%)	>32 (7.3%) >32 (1.2%)	>32 (-2.7%) >32 (7.5%)	>32 (20.4%) >32 (23.6%)	>32 (16.3%) 32 (102.7%)	>32 (15.0%) >32 (49.9%)
Myr-HR1-18	C0142189	>32 (11.6%) >32 (16.0%)	>32 (4.6%) >32 (8.1%)	>32 (4.1%) >32 (7.1%)	>32 (6.6%) >32 (3.8%)	>32 (0.3%) >32 (1.9%)	>32 (9.1%) >32 (0.0%)	>32 (-11.3%) >32 (9.2%)	>32 (9.8%) >32 (60.9%)
HR1-11 Palm	C0142190	>32 (11.4%) >32 (8.3%)	>32 (1.3%) >32 (5.6%)	>32 (3.1%) >32 (6.5%)	>32 (7.4%) >32 (5.4%)	>32 (-0.2%) >32 (3.6%)	>32 (15.3%) >32 (4.4%)	>32 (-0.8%) >32 (17.3%)	>32 (24.1%) >32 (29.2%)
PalHR1-11-3H3R	C0142191	32 (96.2%) 32 (95.6%)	>32 (5.6%) >32 (-0.9%)	8 (99.4%) 8 (102.2%)	>32 (11.9%) >32 (9.9%)	>32 (15.1%) >32 (18.6%)	32 (75.1%) >32 (19.7%)	32 (114.1%) 32 (94.6%)	>32 (7.2%) >32 (19.9%)
(D)PalGR13	C0142192	8 (98.9%) 4 (97.3%)	32 (89.5%) 32 (93.2%)	4 (97.0%) 4 (98.0%)	16 (97.5%) 16 (98.7%)	2 (96.8%) 4 (95.5%)	4 (98.7%) 4 (98.9%)	2 (111.9%) 2 (131.9%)	>32 (8.2%) >32 (31.4%)
(D)PalGR9	C0142193	>32 (12.5%) 16 (98.0%)	>32 (4.7%) >32 (9.2%)	>32 (6.2%) >32 (9.9%)	>32 (19.6%) >32 (6.1%)	>32 (41.1%) 32 (94.2%)	>32 (16.5%) >32 (9.6%)	>32 (83.3%) >32 (83.1%)	>32 (12.8%) >32 (48.1%)
Pal(D)GR4	C0142194	>32 (7.3%) >32 (12.7%)	>32 (6.9%) >32 (8.6%)	>32 (2.3%) >32 (9.2%)	>32 (3.3%) >32 (4.5%)	>32 (-2.2%) >32 (12.4%)	>32 (22.8%) >32 (10.2%)	>32 (17.4%) >32 (9.2%)	>32 (14.8%) >32 (15.5%)
Pal(D)GK4	C0142195	>32 (28.1%) >32 (27.6%)	>32 (28.2%) >32 (28.7%)	32 (91.7%) 32 (93.6%)	>32 (4.9%) >32 (4.8%)	32 (99.0%) 32 (92.6%)	>32 (50.4%) >32 (31.2%)	8 (95.3%) 16 (97.3%)	>32 (23.4%) >32 (35.1%)
Pal(D)GO 4	C0142196	>32 (4.9%) >32 (13.2%)	>32 (6.6%) >32 (8.7%)	>32 (36.2%) >32 (41.8%)	>32 (7.6%) >32 (2.6%)	>32 (18.5%) >32 (20.7%)	>32 (32.8%) >32 (6.7%)	32 (91.4%) 32 (94.6%)	>32 (8.4%) >32 (25.2%)
PalHR1-18-3H3R-GR13	C0142197	>32 (39.2%) >32 (37.9%)	>32 (4.9%) >32 (9.9%)	16 (93.5%) 32 (98.3%)	>32 (7.3%) >32 (1.7%)	>32 (25.3%) >32 (38.4%)	>32 (12.3%) >32 (8.1%)	>32 (-7.5%) >32 (11.0%)	>32 (13.9%) >32 (43.9%)
HRNR136 b-1414-S-Pal	C0142198	>32 (11.0%) >32 (19.8%)	>32 (8.9%) >32 (11.7%)	>32 (3.6%) >32 (9.2%)	>32 (7.7%) >32 (2.9%)	>32 (1.9%) >32 (6.7%)	>32 (52.4%) >32 (7.0%)	>32 (2.5%) >32 (6.5%)	>32 (21.8%) >32 (34.9%)
Pal-G-Dap4	C0142199	>32 (8.8%) >32 (22.4%)	>32 (8.5%) >32 (5.0%)	32 (95.8%) 32 (99.8%)	>32 (9.1%) >32 (9.3%)	32 (95.0%) 32 (90.7%)	>32 (45.9%) >32 (24.5%)	>32 (58.8%) 16 (100.9%)	>32 (27.8%) >32 (52.9%)
Pal-GR8	C0142200	>32 (14.2%) >32 (12.9%)	>32 (4.8%) >32 (8.4%)	>32 (12.0%) >32 (12.9%)	>32 (5.0%) >32 (8.7%)	16 (83.5%) 16 (80.7%)	>32 (24.3%) >32 (62.9%)	32 (93.1%) 16 (95.1%)	>32 (8.4%) >32 (53.4%)
Pal-Om-GR13	C0142201	32 (96.1%) 32 (96.0%)	>32 (54.1%) >32 (46.5%)	32 (94.7%) 32 (95.5%)	32 (88.1%) 32 (96.1%)	32 (88.0%) 32 (92.3%)	32 (98.8%) 32 (88.8%)	32 (96.4%) 32 (108.5%)	>32 (41.1%) >32 (37.5%)
Pal-GK4	C0142202	>32 (18.5%) >32 (20.0%)	>32 (4.1%) >32 (4.0%)	>32 (45.7%) >32 (32.9%)	>32 (8.8%) >32 (9.7%)	>32 (1.2%) >32 (-22.2%)	16 (90.1%) 0.5 (97.5%)	32 (85.2%) 32 (89.6%)	>32 (32.9%) >32 (25.9%)
S-Pal-HRNR2004-2029	C0142203	>32 (15.0%) >32 (11.6%)	>32 (4.8%) >32 (4.6%)	>32 (58.2%) >32 (58.5%)	>32 (10.8%) >32 (8.4%)	>32 (18.2%) >32 (6.1%)	>32 (63.9%) 16 (81.1%)	4 (91.0%) 8 (92.9%)	22.8 (60.4%) 28.6 (56.9%)
Pal-K-GR13	C0142204	16 (103.3%) 16 (101.7%)	>32 (80.1%) >32 (48.5%)	32 (100.8%) 32 (100.1%)	32 (100.1%) 32 (100.4%)	32 (98.4%) >32 (56.6%)	32 (94.1%) 32 (98.1%)	32 (97.0%) 32 (91.3%)	>32 (22.7%) >32 (18.6%)
Pal-GR11	C0142205	8 (101.4%) 16 (99.9%)	>32 (40.4%) 32 (88.5%)	8 (97.3%) 5 (98.1%)	16 (96.8%) 16 (99.5%)	4 (99.3%) 4 (97.3%)	4 (97.7%) 8 (98.0%)	4 (95.3%) 4 (99.8%)	>32 (36.8%) >32 (29.2%)
C-Pal-GR13	C0142220	32 (84.2%) 32 (93.9%)	>32 (48.3%) >32 (55.3%)	32 (84.6%) 32 (79.4%)	32 (85.2%) 32 (84.6%)	4 (95.3%) 4 (94.5%)	2 (98.9%) 2 (98.6%)	0.5 (95.2%) 1 (99.5%)	>32 (41.8%) >32 (15.1%)
S-C-Pal-GR13	C0142221	16 (101.6%) 32 (97.4%)	32 (92.4%) 32 (97.3%)	32 (90.6%) 32 (92.7%)	32 (97.1%) 32 (97.4%)	4 (98.1%) 4 (94.8%)	4 (99.5%) 4 (98.6%)	2 (93.1%) 2 (95.9%)	>32 (35.1%) >32 (40.1%)
LCE1B 7B-88	C0142222	>32 (3.8%) >32 (17.4%)	>32 (0.4%) >32 (6.1%)	>32 (6.4%) >32 (2.9%)	>32 (9.7%) >32 (8.6%)	>32 (11.2%) >32 (11.2%)	>32 (18.3%) >32 (5.2%)	>32 (2.7%) >32 (-5.8%)	>32 (24.4%) >32 (24.9%)
LCE2A 7B-82	C0142228	>32 (14.8%) >32 (20.1%)	>32 (3.0%) >32 (6.8%)	>32 (7.6%) >32 (10.3%)	>32 (8.6%) >32 (7.3%)	>32 (16.8%) >32 (15.3%)	>32 (10.4%) >32 (9.7%)	>32 (-1.5%) >32 (-8.4%)	>32 (8.6%) >32 (6.6%)
LCE3A 5B-80	C0142229	>32 (12.7%) >32 (14.4%)	>32 (3.8%) >32 (7.5%)	>32 (6.6%) >32 (3.9%)	>32 (6.3%) >32 (4.0%)	>32 (16.0%) >32 (12.6%)	>32 (23.6%) >32 (13.6%)	>32 (-4.3%) >32 (-13.0%)	>32 (18.2%) >32 (6.6%)
LCE3C 55-66	C0142231	>32 (7.7%) >32 (11.2%)	>32 (4.5%) >32 (5.7%)	>32 (7.9%) >32 (18.0%)	>32 (13.8%) >32 (6.2%)	>32 (12.9%) >32 (5.8%)	>32 (17.8%) >32 (18.5%)	>32 (-4.0%) >32 (-5.1%)	>32 (26.9%) >32 (24.0%)
LCE3D 55-64	C0142232	>32 (1.9%) >32 (7.9%)	>32 (2.6%) >32 (2.2%)	>32 (0.3%) >32 (5.2%)	>32 (8.7%) >32 (5.2%)	>32 (9.1%) >32 (5.2%)	>32 (15.5%) >32 (7.8%)	>32 (-0.6%) >32 (-1.5%)	>32 (39.8%) >32 (23.4%)

* Minimal inhibitory concentration (MIC) values were determined using the broth microdilution susceptibility test according to the CLSI (Clinical and Laboratory Standards Institute) guidelines in Mueller–Hinton (MH) broth on 5×10^5 CFU/mL. They were defined as the lowest peptide concentration that prevented visible bacterial growth after incubation for 18 h at 37 °C. Results are from “Community for Open Antimicrobial Drug Discovery, CO-ADD”. Further details of the screening experiments are available from <http://www.co-add.org/file/co-add-workflowscreeningpdf-0>.

Supplementary Table 16 | Structures of CIDAMPs tested at conditions recommended by CLSI.

Name	AA-sequence
LCE-2B 74-86	SHHRPRLFRRRH
LCE-3B 56-68	SHHRCCRSRRCRR
LCE-3C 55-67	SHHRHFRSHQCRR
LCE-3C 55-75	SHHRHFRSHQCRRQRNSNSCDR
Pal-HR 1-18-HR	Pal-GRRGSGLGRSSSRGQRGSGSGR
Pal-HR-1-11-2H2R-14	Pal-RGSRSGQSSRGERR
Pal-GR13	Pal-GRRGGRGGRGRGR
Pal-GR9	Pal-GRRGGRGGR
Pal-GR4	Pal-GRGR
Pal-G-Dap4	Pal-GDapGDap
Pal-LCE-3C 55-67	Pal-SHHRHFRSHQCRR
Pal-LCE-3B 56-68	Pal-SHHRCCRSRRCRR
S-Pal-HRNR 1748-1778	SGHSSVFGQHESGSGHSSAYSQHGSGSGHFC-Pal-NH2
Pal-HR 1-18 GR13	Pal-GRRGSGLGRSSSR
Pal-HR1-18	Pal-GRHGSGLGHSSSHGQHGSGSGR
Myr-HR1-18	Myr-GRHGSGLGHSSSHGQHGSGSGR
HR-1-11-Palm	Pal-HGSRSGQSSRGERHGSSSGSSSH
Pal-HR-1-11-3H3R	Pal-RGSRSGQSSRGERRGSSSGSSSR
(D)-Pal-GR13	Pal-GR _D R _D GGR _D GGR _D GR _D GR _D
(D)-Pal-GR9	Pal-GR _D R _D GGR _D GGR _D
(D)-Pal-GR4	Pal-GR _D GR _D
(D)-Pal-GK4	Pal-GK _D GK _D
(D)-Pal-GO4	Pal-GO _D GO _D
Pal-HR-1-18-3H3R-GR13	Pal-GRRGSGLGRSSSR
HRNR 1389-1414-S-Pal	GRHGSGSGQSSGFGHHESSWQSSGC-Pal-NH2
Pal-GDab4	Pal-GDabGDab
PalGR6	Pal-GRRGGR
Pal-OrnGR13	Pal-GOOGGOGGOGGOGO
Pal-GK4	Pal-GK _D GK _D
S-Pal-HRNR2004-2029	GQHSSSGHSSSHGQHGASGQSSSC-Pal-NH2
Pal-K-GR13	Pal-GKKGGKGGKGGKGGK
Pal-GR11	Pal-GRRGGRGGRGRGR
C-Pal-GR13	GRRGGRGGRGRGR-C-Pal-NH2
8 C-Pal-GR13	GRRGGRG-C-Pal-GRGRGR
LCE1B 78-88	SHHRRRRSHCH
LCE2A 70-82	SHHRPRLFHRRRH
LCE3A 50-60	SHHRCRRSHRC
LCE3C 55-66	SHHRHFRSHQCRR
LCE3D 55-64	NHHRHHRCR

Residues cationic at pH 7, cationic only at pH < 6, and anionic at pH 7 are colored blue, green and magenta, respectively. Dab:L-2,4-Diaminobutyric acid, Dap:L-2,3-Diaminopropionic acid.

Supplementary Table 17 | HRNR-primer-sequences used in this study

Name	Primer Sequence	A.-T.	D.-T:
pSu-hr3-a-2576-f (Bsal)	<u>AAGGTCTCAAGGTGGCTTTGGTCACCACGAGT</u>	60°C	79,1°C
pSu-hr3-b-2591-f (Bsal)	<u>AAGGTCTCAAGGTAGTCAGCATGGGTCTGGCT</u>	60°C	77,1°C
pSu-hr3-c-2638-f (Bsal)	<u>AAGGTCTCAAGGTCAGTCTTCGGGCCACGGC</u>	62°C	82,3°C
pSu-hr3-d-2656-f (Bsal)	<u>AAGGTCTCAAGGTGGCCGACATGGGTCCGGT</u>	62°C	83,7°C
pSu-hr3-e-2658-f (Bsal)	<u>AAGGTCTCAAGGTCATGGGTCCGGTTTGGGG</u>	60°C	81,6°C
pSu-hr3-a-2707-r (BamHI)	<u>AAGGATCCTTAGTAAGCAGAGGAATGTCCTG</u>	60°C	70,8°C
pSu-hr3-b-2684-r (BamHI)	<u>AAGGATCCTTAATATGGGCCACGGCTGGAA</u>	60°C	77,4°C
pSu-hr3-d-2618-r (BamHI)	<u>AAGGATCCTTATCGTTACCCCTAGATGACTG</u>	64°C	73,5°C
pSu-hr3-e-2609-r (BamHI)	<u>AAGGATCCTTATCTAGAGCCGTGTTGTCCGT</u>	62°C	73,9°C
pSu-hr3-f-2644-r (BamHI)	<u>AAGGATCCTTAACGGGAGCCAGACCCATG</u>	60°C	77,4°C
hr3-2662G-f	<u>GTCCGGTGGCGGGCACTCCTCCAG</u>	72°C	82,1°C
hr3-2662G-r	<u>GTGCCCGCCACCGGACCCATGTCG</u>	72°C	84,9°C
hr3-2629G-f	<u>CACGGTGGTCAGCATGGGTCTGG</u>	68°C	76,7°C
hr3-2629G-r	<u>GCTGACCACCGTGGGAAGACGAACC</u>	74°C	77,6°C
hr3-2603G-f	<u>CTGGCGGCGGACAACACGGCTC</u>	68°C	80,3°C
hr3-2603G-r	<u>TGTTGTCCGCCGACAGAGGAGTG</u>	68°C	78,1°C
Su3-hr3-2684G-r (BamHI)	<u>AAAGGATCCTCAACCTGGGCCACGGCTGGAAGA</u>	62°C	83,8°C
pESu3-hrnr-3-f (FokI)	<u>AAGGGGATGAGCATGATTAGGTAAACTCCTACAAGGCGTCATC</u>	62°C	80,3°C
pESu3-hrnr-244-r (XhoI)	<u>AAGGCTCGAGTCAGTAACCAGAGGACTGCC</u>	58°C	77,8°C
pESu3-hrnr-82-f (FokI)	<u>AAGGGGATGAGCATGATTAGGTGTTGAGGCTCGTAATAAAATC</u>	58°C	79,0°C
pESu3-hrnr-245-f (FokI)	<u>AAGGGGATGAGCATGATTAGGTAGTCAGCATGGATCTGGC</u>	56°C	81,8°C
pESu3-hrnr-478-r (XhoI)	<u>AAGGCTCGAGTCAGTAACCAGAGGAATGCTCT</u>	56°C	75,1°C
HRNR-2591-f-CyBr (SbfI)	<u>AACCCTGCAGGAGATGAGTCAGCATGGGTCTGGCT</u>	74°C	83,6°C

A.-T.: Annealing temperature; D.-T.: Detection temperature. The complementary coding sequence is underlined.

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