

Supplemental material

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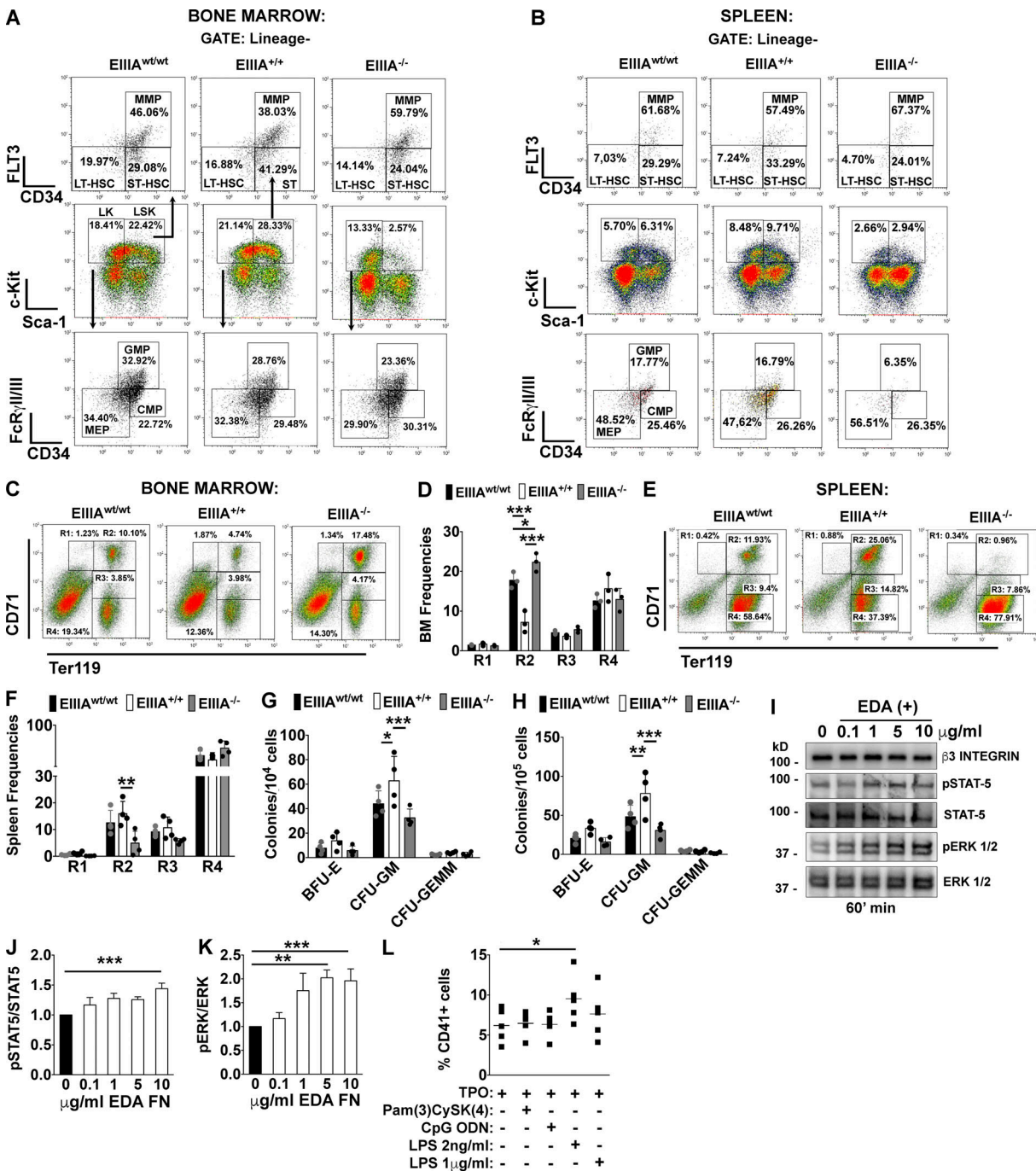


Figure S1. **Effects of EDA overexpression on hematopoietic progenitors in mice and concentration-dependent effects on Mk signaling pathway activation.** (A and B) Representative contour plots and gate strategy of flow cytometric analysis of LSK, LT-HSC (Lin⁻Sca-1⁺c-kit⁺CD34⁻CD135⁻), ST-HSC (Lin⁻Sca-1⁺c-kit⁺CD34⁺CD135⁻), MMP (Lin⁻Sca-1⁺c-kit⁺CD34⁺CD135⁺), CMP (Lin⁻Sca-1⁺c-kit⁺CD34⁺FcyRII/III^{low}), GMP (Lin⁻Sca-1⁺c-kit⁺CD34⁺FcyRII/III^{high}), and MEP (Lin⁻Sca-1⁺c-kit⁺CD34⁺FcyRII/III⁺) cells in the BM (A) and spleens (B) of EIIIA^{wt/wt}, EIIIA^{+/+}, and EIIIA^{-/-} mice after experimental fibrosis. (C and D) Representative dot plots of flow cytometric analysis of erythroid precursors in the BM of EIIIA^{wt/wt}, EIIIA^{+/+} and EIIIA^{-/-} mice after experimental fibrosis using surface markers CD71 and Ter119 (C). Frequencies of erythroid precursor cells are shown in bar graphs as mean ± SD (n = 3; D). (E and F) Representative dot plots of flow cytometric analysis of erythroid precursors in the spleens of EIIIA^{wt/wt}, EIIIA^{+/+}, and EIIIA^{-/-} mice after experimental fibrosis using surface markers CD71 and Ter119 (E). Frequencies of erythroid precursor cells are shown in bar graphs as mean ± SD (n = 3; F). (G and H) BM cells (10⁴; G) and spleen cells (10⁵; H) from EIIIA^{wt/wt}, EIIIA^{+/+}, and EIIIA^{-/-} mice (n = 4) were plated in methylcellulose medium supplemented with cytokines. BFU-E, CFU-GM, and CFU-GEMM colonies were scored 7 d after plating. (I) Purified Mk's were stimulated for 1 h with increasing concentration of cFN and the phosphorylation of STAT-5 and ERK 1/2 evaluated by Western blotting. β3 integrin was revealed to ensure the same number of Mk used in the experiment, while total STAT-5 and ERK 1/2 were detected to ensure equal protein loading. (J and K) Histograms showing the ratio of phosphorylated and total STAT-5 (J) and ERK 1/2 (K) proteins in mature Mk's stimulated with increasing amounts of cFN for 1 h. At least three independent experiments were performed. **, P < 0.01; ***, P < 0.001. (L) Quantification of Mk output after 3 d of culture treated only with TPO, TPO plus 1 μg/ml of Pam3Csk4, TPO plus 3 μM CpG ODN, and TPO plus LPS (2 ng/ml and 1 μg/ml, respectively). Five independent experiments were performed. *, P < 0.05.

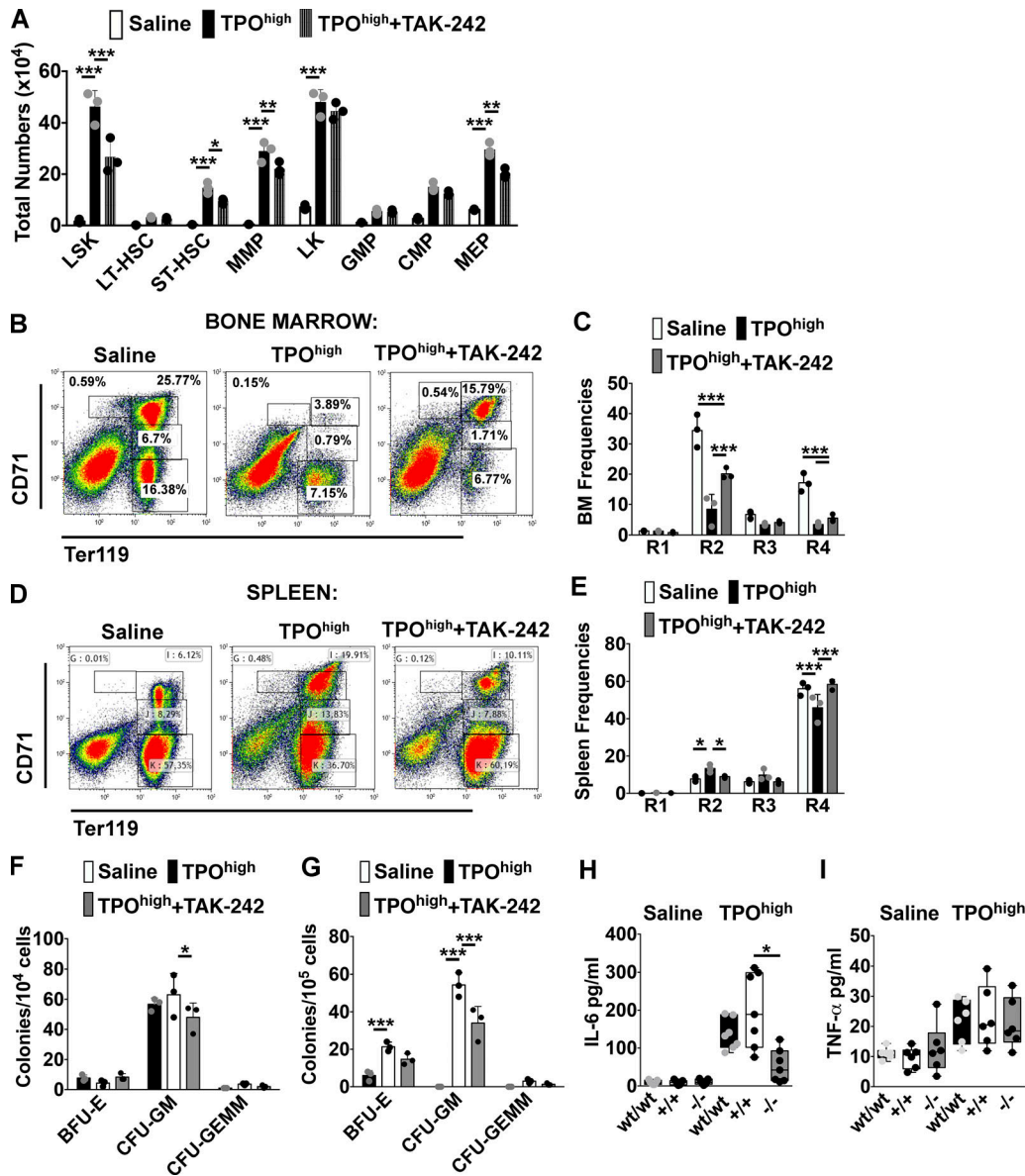


Figure S2. **Effects of TLR4 inhibition on hematopoietic progenitors in mice.** (A) Total numbers of LSK, LT-HSC, ST-HSC, MMP, CMP, GMP, and MEP cells in the spleens of saline-, TPO^{high}-, and TPO^{high} plus TAK-242-treated mice are shown in bar graphs as mean ± SD (*n* = 3). *, *P* < 0.05; **, *P* < 0.005; ***, *P* < 0.001. (B and C) Representative dot plots of flow cytometric analysis of erythroid precursors in the BM of saline-, TPO^{high}-, and TPO^{high} plus TAK-242-treated mice using surface markers CD71 and Ter119 (B). Frequencies of erythroid precursor cells are shown in bar graphs as mean ± SD (*n* = 3). (D and E) Representative dot plots of flow cytometric analysis of erythroid precursors in the BM of saline-, TPO^{high}-, and TPO^{high} plus TAK-242-treated mice using surface markers CD71 and Ter119 (D). Frequencies of erythroid precursor cells are shown in bar graphs as mean ± SD (*n* = 3); (E). (F and G) BM cells (10⁴; F) and spleen cells (10⁵; G) of saline-, TPO^{high}-, and TPO^{high} plus TAK-242-treated mice (*n* = 3) were plated in methylcellulose medium supplemented with cytokines. BFU-E, CFU-GM, and CFU-GEMM colonies were scored 7 d after plating. (H and I) Quantification of TNF-α (H) and IL-6 (*n* = 7; I) by ELISA in BM cell-free supernatants of wild-type, EIIIA^{+/-}, and EIIIA^{-/-} mice treated with saline or TPO. *n* = 7. *, *P* < 0.05.

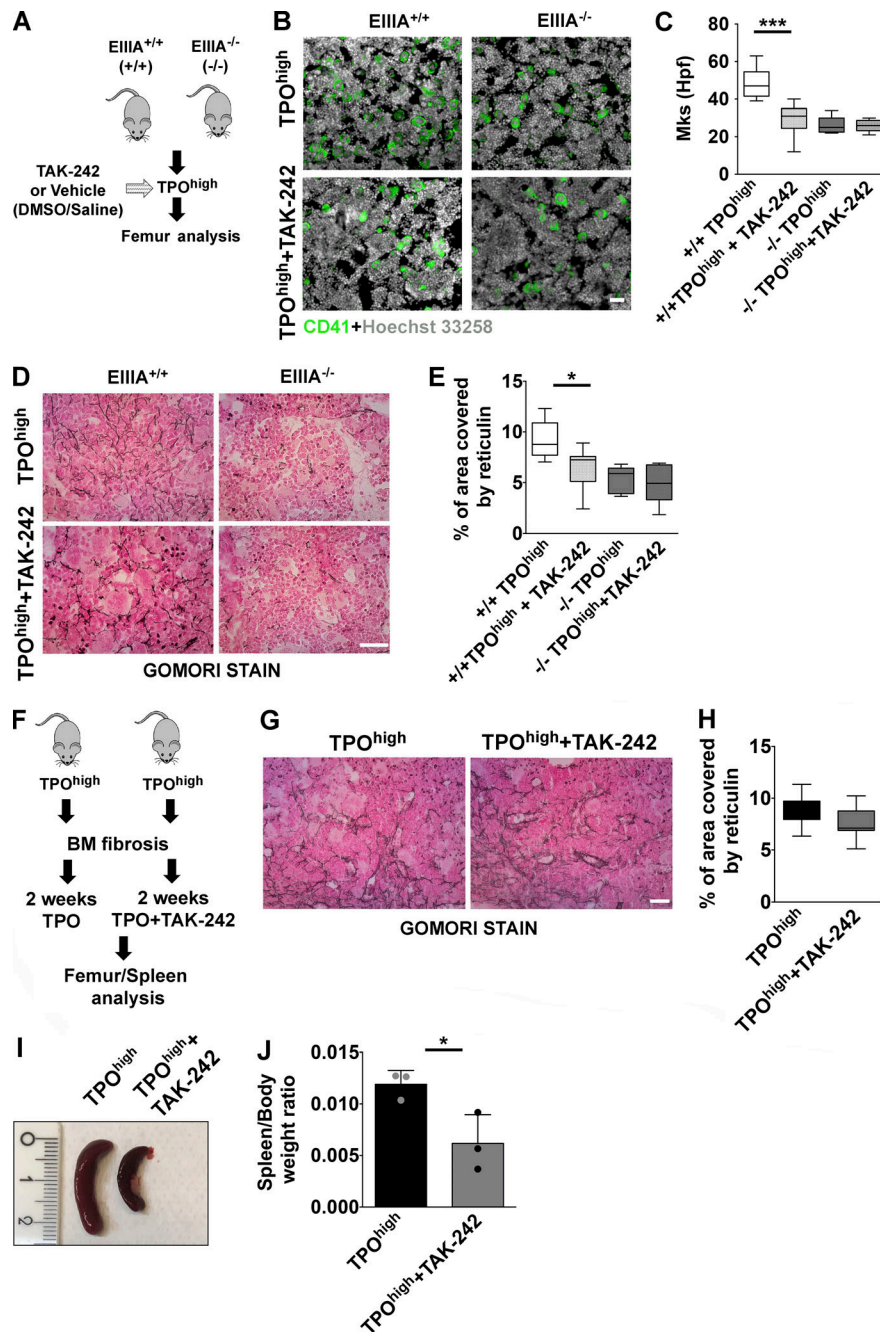


Figure S3. **TAK-242 treatment ameliorates Mk hyperplasia and BM fibrosis in EIIIA^{+/+} mice while it has null effects in EIIIA^{-/-} mice.** (A) Schematic representation of in vivo strategy for TLR4 inhibition in EIIIA^{+/+} and EIIIA^{-/-} mice during experimental fibrosis. (B) Representative immunofluorescence staining of CD41 Mks in BM sections of EIIIA^{+/+} and EIIIA^{-/-} mice treated with TPO plus vehicle or TPO plus TAK-242. Nuclei were stained with Hoechst 33258. Bar, 50 μm. (C) Quantification of CD41⁺ Mks per high power field (hpf) in BM sections of EIIIA^{+/+} and EIIIA^{-/-} mice treated with TPO plus vehicle or TPO plus TAK-242. ≥100 Mks were randomly counted from five sections of three different mice per genotype in each experimental condition. ***, P < 0.001. (D) Representative Gomori staining in BM sections of EIIIA^{+/+} and EIIIA^{-/-} mice treated with TPO plus vehicle or TPO plus TAK-242. Bar, 50 μm. Objective, 40x. (E) Assessment of reticulin deposition in EIIIA^{+/+} and EIIIA^{-/-} mice treated with TPO plus vehicle or TPO plus TAK-242. ≥15 random images from BM diaphysis per mice, with three mice per treatment and genotype, were analyzed using Imagej software. *, P < 0.05. (F) Schematic representation of in vivo strategy for TLR4 inhibition in TPO^{high} mice after the BM fibrosis is already established. (G and H) Representative Gomori staining (G) and assessment of reticulin deposition (H) in BM sections of TPO^{high} and TPO^{high} plus TAK-242-treated mice. Bar, 50 μm. Objective, 40x. (I and J) Representative spleens (I) and spleen/body weight ratios (J) were calculated in TPO^{high} and TPO^{high} plus TAK-242-treated mice.

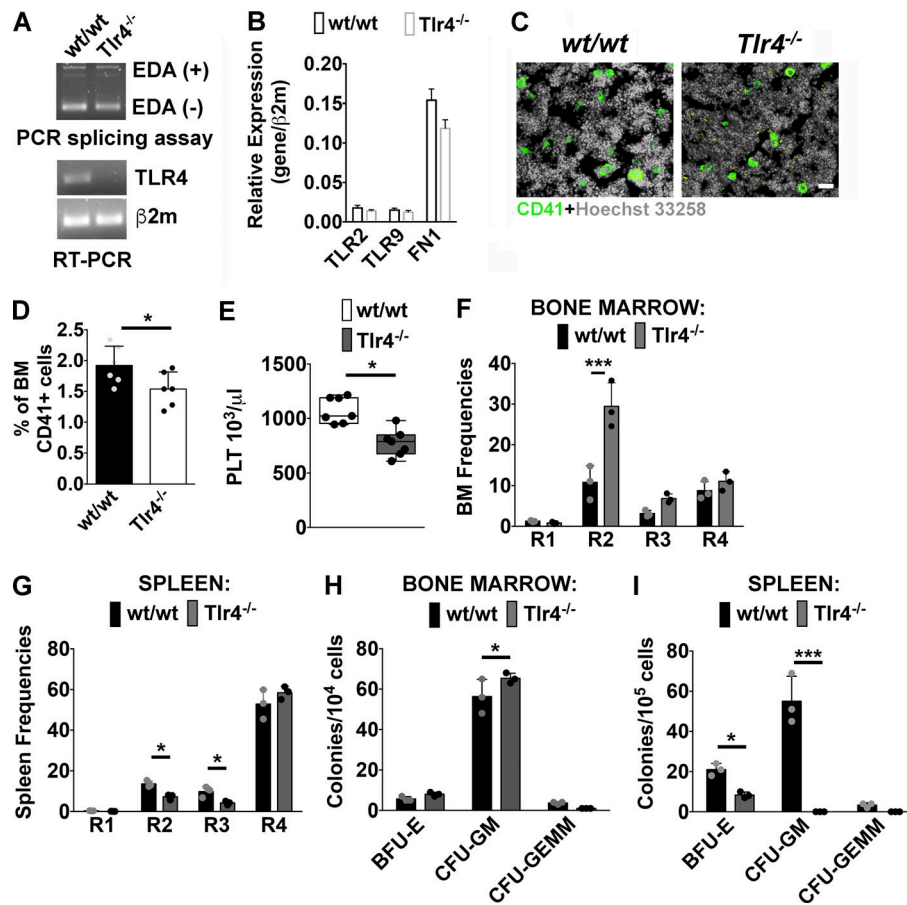


Figure S4. **TLR4 deletion in vivo induces a modest thrombocytopenia but restrains EDA FN effects on hematopoietic progenitors after experimental fibrosis.** (A) PCR splicing assay of EDA FN in BM cells recovered from untreated wild-type (*wt/wt*) and *Tlr4^{-/-}* mice. Real-time PCR products of TLR4 expression confirmed the targeted deletion of TLR4 in the knockout mice. $\beta 2$ -microglobulin was amplified for sample normalization. (B) Real-time PCR analysis of FN, TLR2, and TLR9 expression relative to $\beta 2$ -microglobulin in BM cells of wild-type and *Tlr4^{-/-}* mice. $n = 3$. (C and D) Immunofluorescence of CD41⁺ Mk's in BM sections (C) and relative quantification (D) in untreated wild-type and *Tlr4^{-/-}* mice. Bar, 50 μm . $n = 6$. Nuclei were stained with Hoechst 33258. (E) Peripheral platelet (PLT) counts in untreated wild-type and *Tlr4^{-/-}* mice. $n = 7$. (F and G) Frequencies of erythroid progenitors in BM (F) and spleens (G) of wild-type and *Tlr4^{-/-}* mice after experimental fibrosis. $n = 3$. (H and I) BM cells (10^4 ; H) and spleen cells (10^5 ; I) of wild-type and *Tlr4^{-/-}* mice after experimental fibrosis ($n = 3$) were plated in methylcellulose medium supplemented with cytokines. BFU-E, CFU-GM, and CFU-GEMM colonies were scored 7 d after plating. *, $P < 0.05$; ***, $P < 0.001$.

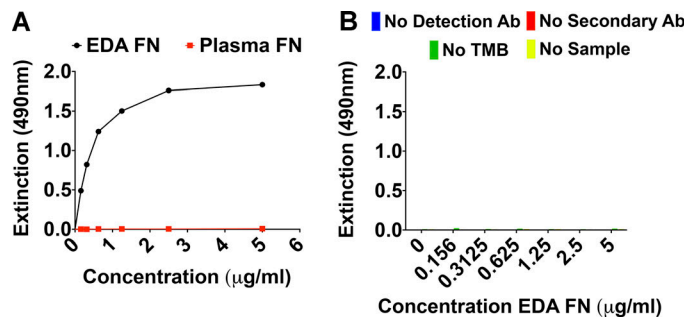


Figure S5. **Establishment of the specific ELISA for quantification of EDA FN in plasma samples.** (A) Evaluation of different EDA FN standard concentrations (black circles) and exclusion of cross-reactions to pFN (red squares). (B) Exclusion of cross-reactions to different components of the ELISA setup (no detection antibody (Ab), blue histograms; no secondary antibody, red histograms; no tetramethylbenzidine (TMB) ELISA substrate, green histograms; no sample, yellow histograms).