

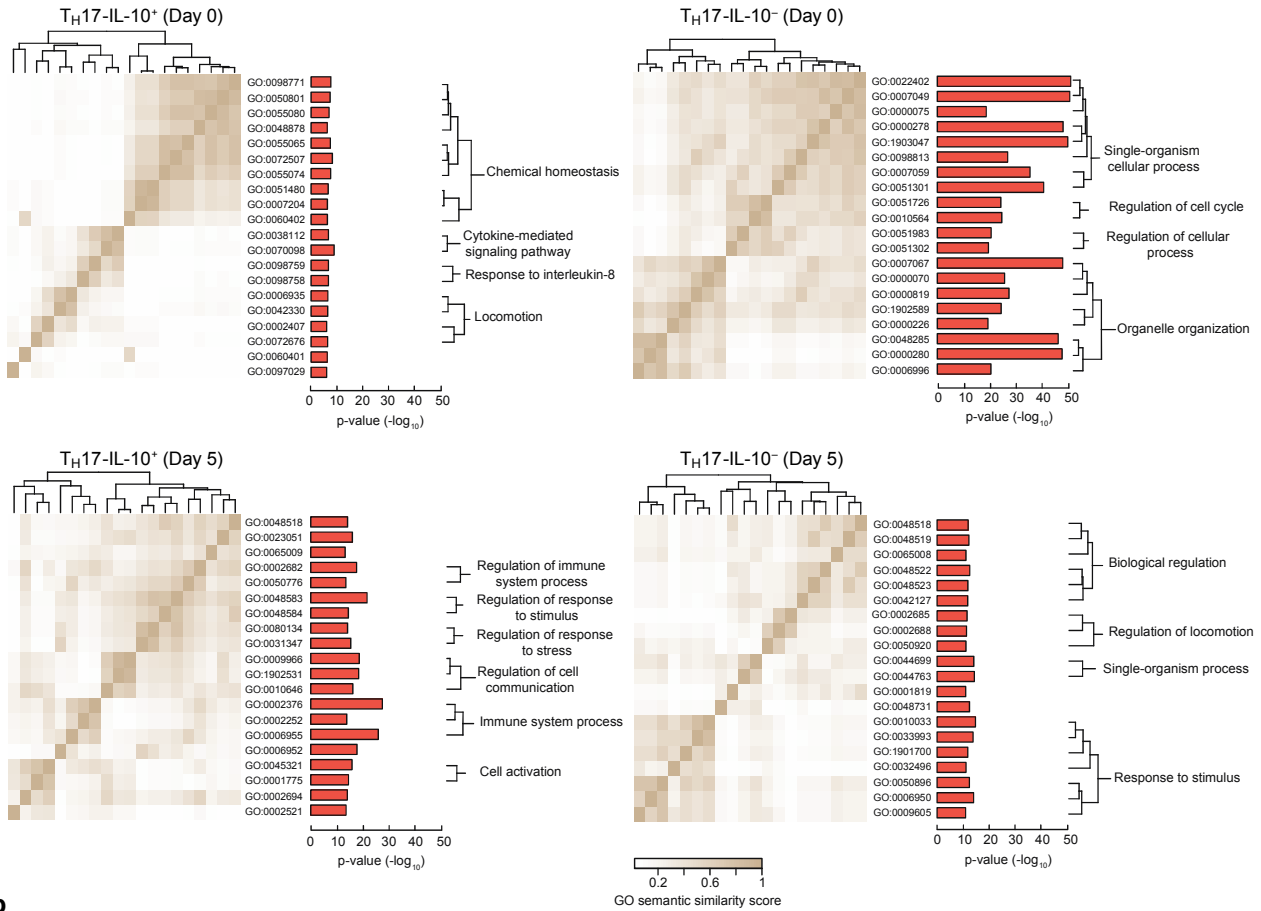
Supplementary Figure 1

Sorting strategy and cytokine production by subsets of human memory T_H17 cells

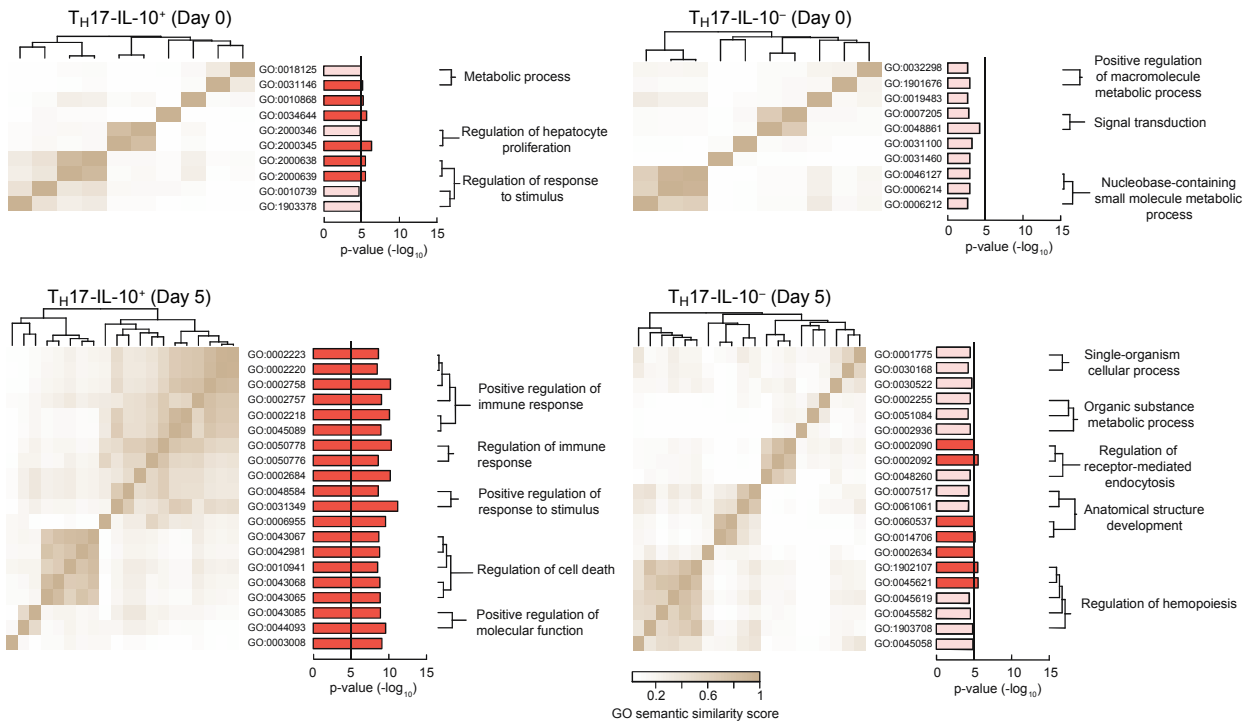
a. Sorting of human memory T_H17 cells from PBMCs based on chemokine receptor expression or cytokine secretion. MACS-enriched CD4⁺ T cells were stained with antibodies to CD45RA, CD25, CCR7, CCR6, CCR4 and CXCR3. CD4⁺ memory T_H17 cells were sorted, after exclusion of CD45RA⁺CCR7⁺ naïve T cells (T_N) and CD8⁺ CD14⁺ CD19⁺ CD56⁺ CD25⁺ cells, as CCR6⁺CCR4⁺CXCR3⁻ or, alternatively, as CCR6⁺CXCR3⁻IL-17A⁺ (a, lower panels). Dot plots from one donor, representative of more than 10 donors analyzed. **b.** Cytokine production as assessed by intracellular staining in Day 0-resting and Day 5-activated T_H17-IL10⁺ and T_H17-IL-10⁻ clone pools after 5 h stimulation with PMA plus Ionomycin. Data are represented as mean + 95% c.i., with each dot indicating a T cell clone pool from independent experiments (n ≥ 9) (note: the number of clone pools is paired and the same in T_H17-IL-10⁺ and T_H17-IL-10⁻, but can vary for different cytokines (IL-10 and IFN- γ , n = 15; IL-22, n = 13; IL-4, n = 10; GM-CSF, n = 9)). *P < 0.05; **P < 0.01; ****P < 0.0001, as determined by ratio paired *t* test.

a

Protein-coding genes - GO Biological Process

**b**

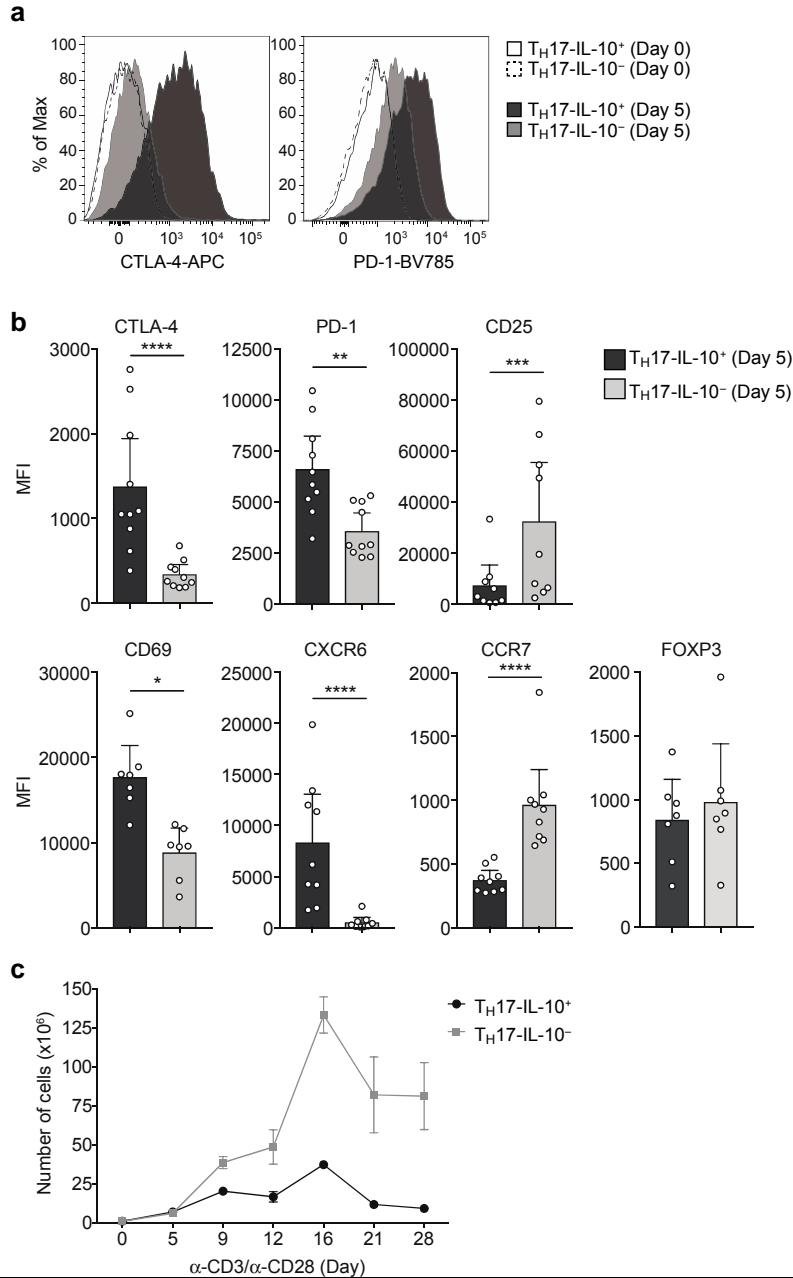
lncRNAs - GO Biological Process



Supplementary Figure 2

Gene ontology (GO) analyses of differentially expressed genes and lncRNAs in T_H17-IL-10⁺ and T_H17-IL-10⁻ cells

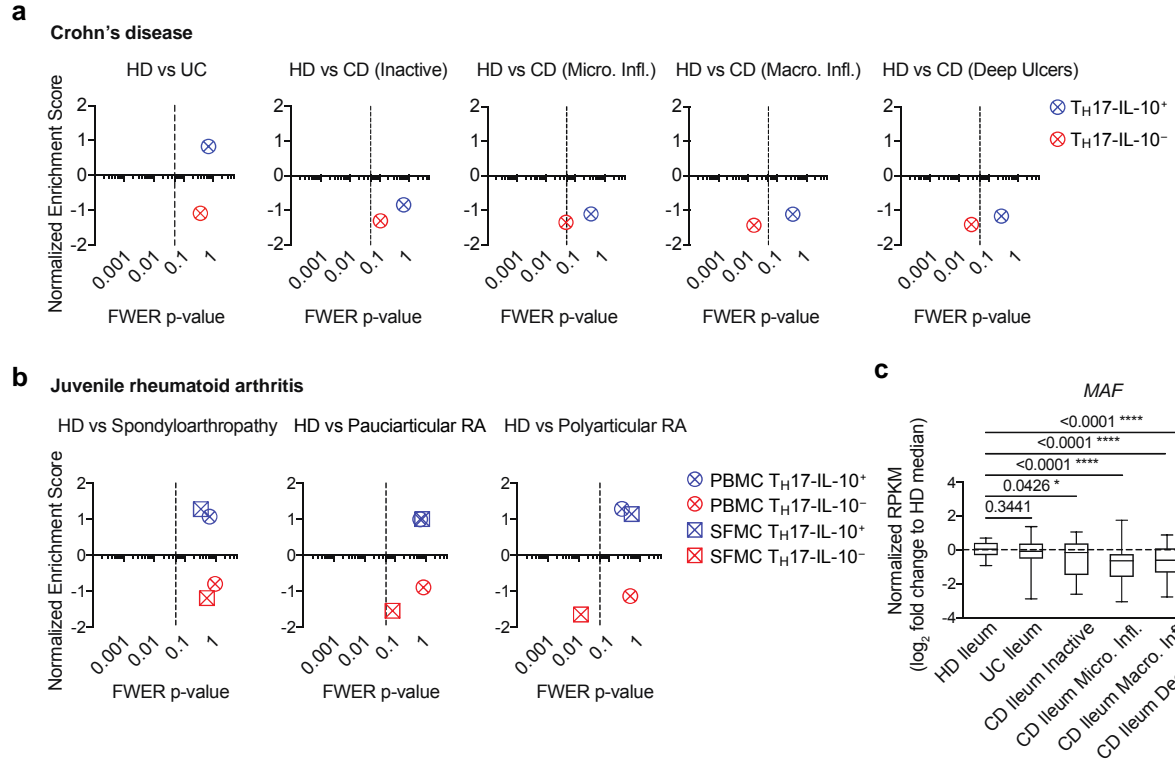
Gene ontology (GO) analyses on the biological processes associated to the differentially expressed protein-coding genes (a) or lncRNAs (b) in Day 0-resting and Day 5-activated T_H17-IL-10⁺ and T_H17-IL-10⁻ cells. GO terms are clustered by semantic similarity and a synthetic description is shown. *P*-value as determined by MetaCore based on hypergeometric distribution; *P*-value threshold (vertical line) set at 1×10^{-5} .



Supplementary Figure 3

Differential surface marker expression in TH_H17-IL-10⁺ and TH_H17-IL-10⁻ cells

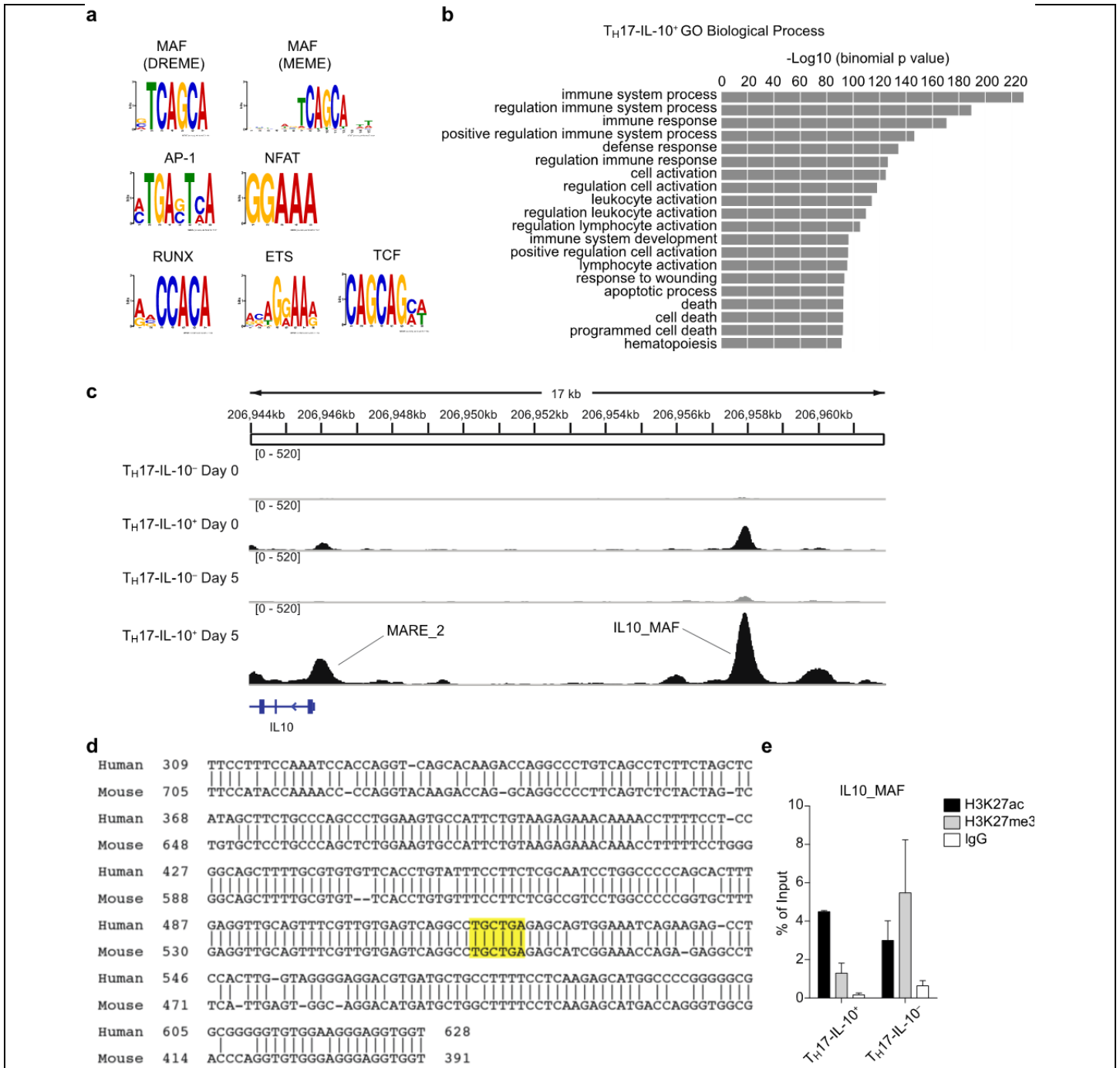
a,b. Expression of CTLA4, PD-1, CD25, CD69, CXCR6, CCR7 and FOXP3 as assessed by flow cytometry in Day 5-activated TH_H17-IL-10⁺ and TH_H17-IL-10⁻ clone pools. Shown are representative histogram plots (**a**) and cumulative data of clone pools from independent experiments (mean + 95% c.i.; CTLA-4 and PD-1, n = 10; CD25, CXCR6 and CCR7, n = 9; CD69 and FOXP3, n = 7) (**b**). **P* < 0.05; ***P* < 0.01; ****P* < 0.001; *****P* < 0.0001, as determined by ratio paired *t* test. **c.** Expansion of TH_H17-IL-10⁺ and TH_H17-IL-10⁻ cells in response to stimulation with CD3/CD28 antibodies. TH_H17-IL-10⁺ and TH_H17-IL-10⁻ cells were stimulated with plate-bound CD3/CD28 antibodies for 48 h and their proliferation was measured at the indicated time points. Shown is the average of two independent experiments (mean ± s.e.m.).



Supplementary Figure 4

Gene set enrichment analysis (GSEA) of $T_H17-IL-10^+$ and $T_H17-IL-10^-$ transcriptional signatures in autoimmune diseases

Enrichment of $T_H17-IL-10^+$ and $T_H17-IL-10^-$ -associated gene signatures in publicly available transcriptional datasets of T_H17 -mediated autoimmune diseases. Gene set enrichment in ileal biopsies from healthy donors (HD) vs. Crohn's patients (CD) (**a**) and in PBMC and SFMC from healthy donors (HD) vs. juvenile rheumatoid arthritis patients (RA) (**b**) are shown. GSEA enrichment results were reported as normalized enrichment score and familywise enrichment (FWER) P value. FWER P values smaller than 0.05 (dashed line) were considered significant. **c.** c-MAF expression in ileal biopsies from healthy donors (HD), ulcerative colitis (UC) and Crohn's disease (CD) patients, with histologically graded disease severity, is shown in a box (interquartiles, with a line indicating the median value) and whiskers (min to max values) plot. P -value as determined by Kruskal-Wallis test. Micro. Infl. = microscopic inflammation; Macro. Infl. = macroscopic inflammation.



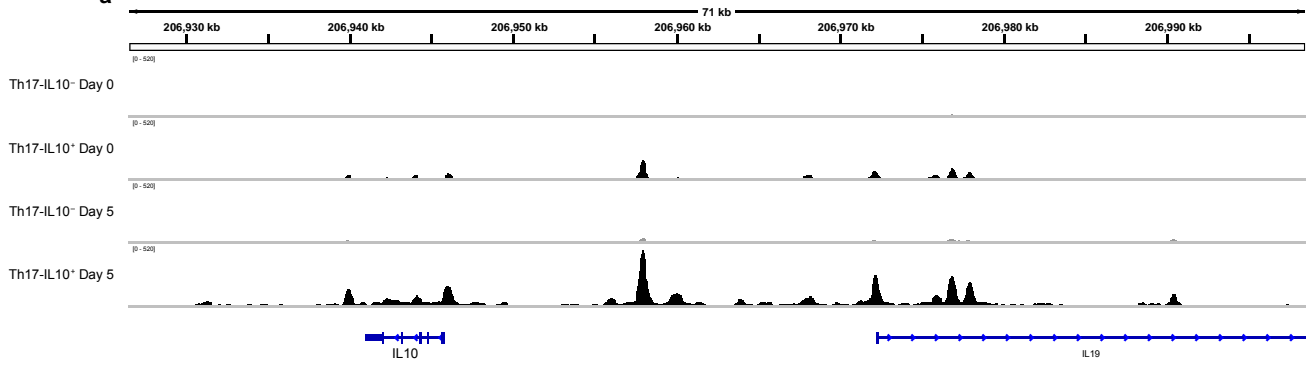
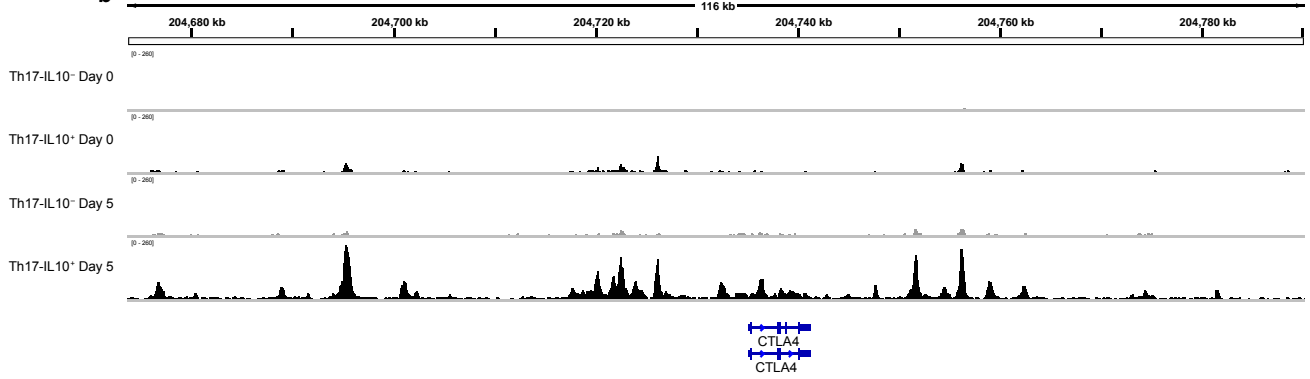
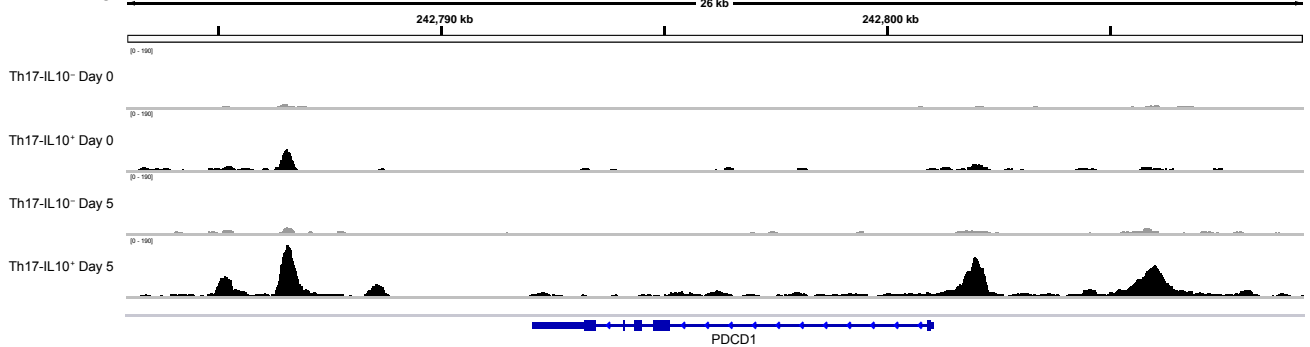
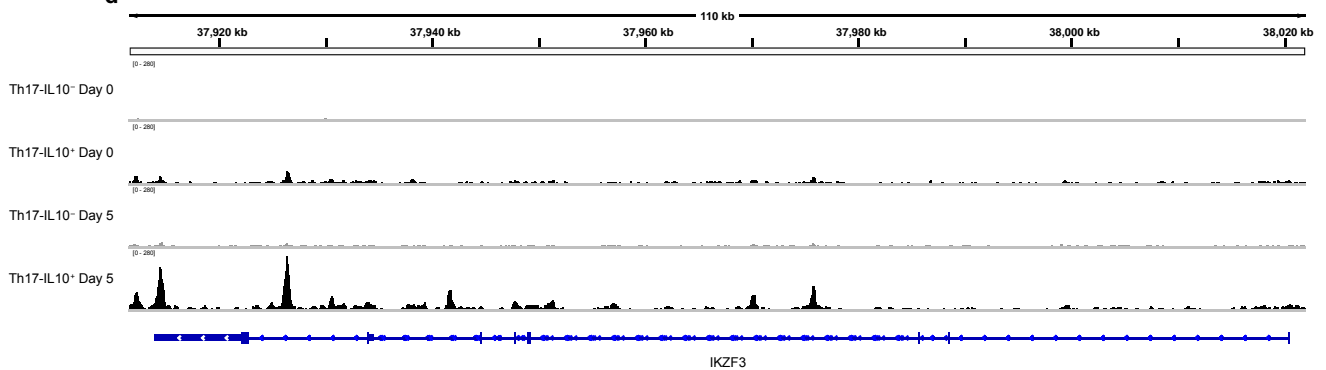
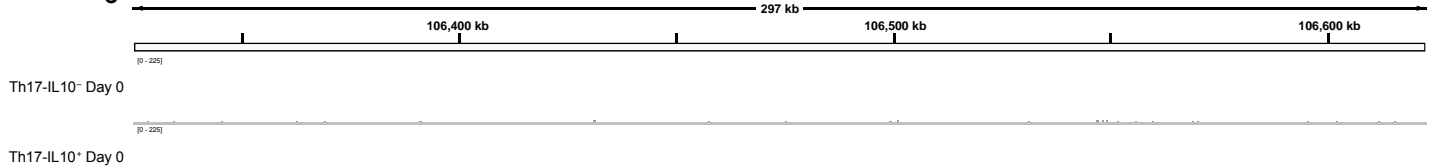
Supplementary Figure 5

c-MAF binds enhancer-like regions in proximity to genes involved in the immune response

a. Position weight matrix (PWM) of the top ranked DNA motifs identified by the MEME suite software in the Day 5-activated T_H17-IL-10⁺ c-MAF ChIP-seq dataset; e-values: MAF (DREME), 5.5×10^{-212} ; MAF (MEME), 6.6×10^{-175} ; AP-1, 1.7×10^{-146} ; NFAT, 3.9×10^{-145} ; RUNX, 2.6×10^{-81} ; ETS, 1.1×10^{-57} ; TCF, 1.1×10^{-20} . **b.** Gene Ontology (GO) analysis of genes associated to c-MAF-bound regions in Day 5-activated T_H17-IL-10⁺ cells. GO biological processes are ranked according to their binomial *P*-value. **c.** Graphical representation of a previously characterized (MARE_2) and a novel (IL10_MAF) c-MAF binding sites at the *IL10* locus by the Integrative Genome Viewer (IGV). **d.** The 1 kb genomic region centered on the novel putative *IL10* enhancer (IL10_MAF) was aligned to an isometric genomic region centered on a c-MAF peak localized about 9 kb upstream of *IL10* in mouse T_H17 cells¹. The core enhancer region

shown is highly conserved between human and mouse (83% identities) and includes a canonical c-MAF binding site (highlighted in yellow). **e.** The activation status of the newly identified putative *IL10* enhancer was evaluated by quantifying H3K27ac and H3K27me3 levels in Day 5-activated T_H17-IL-10⁺ and T_H17-IL-10⁻ cells by ChIP-qPCR (mean + s.e.m.; n=2).

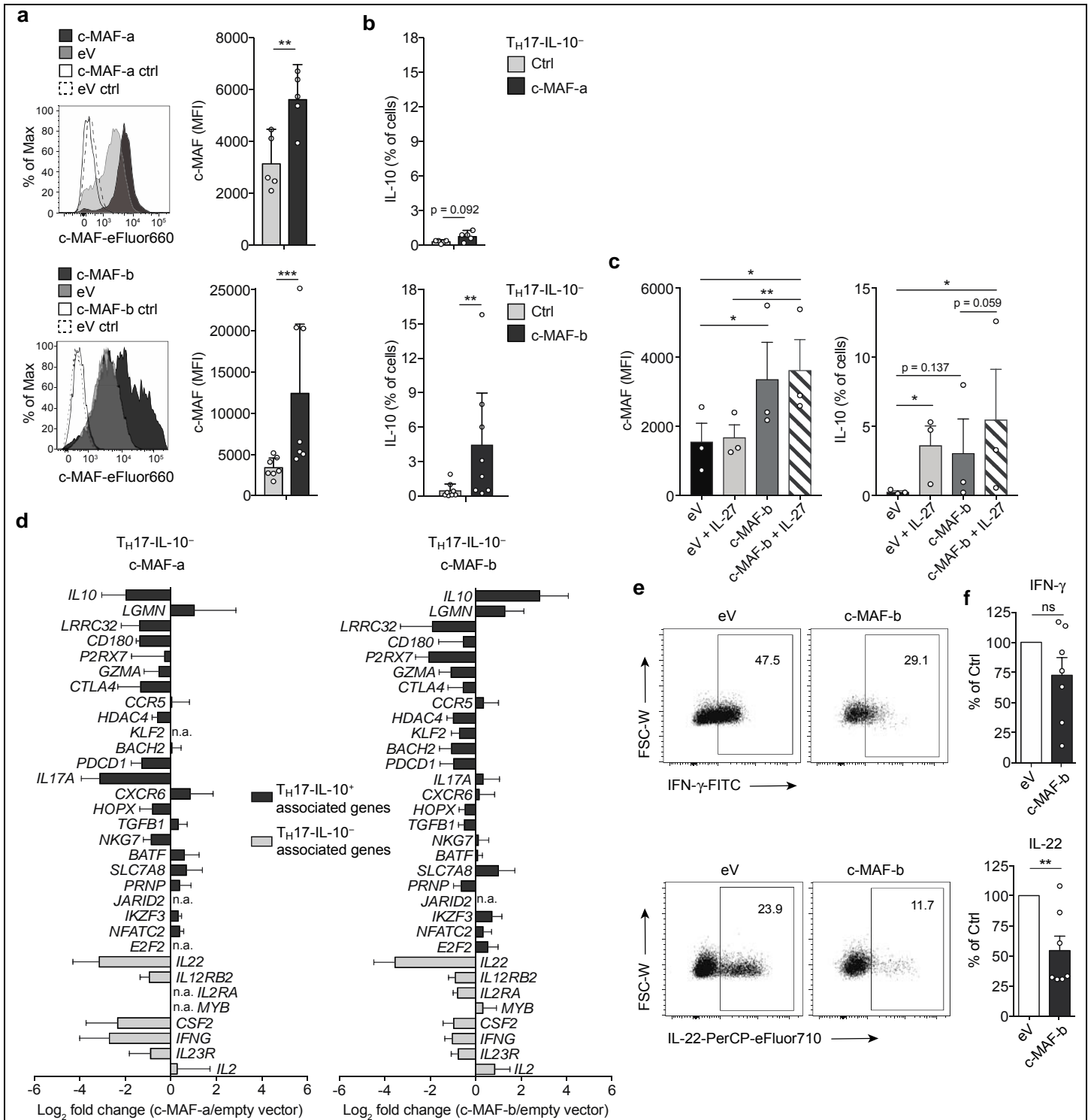
¹Ciofani, M. et al. A validated regulatory network for Th17 cell specification. *Cell* 151, 289-303 (2012).

a**b****c****d****e**

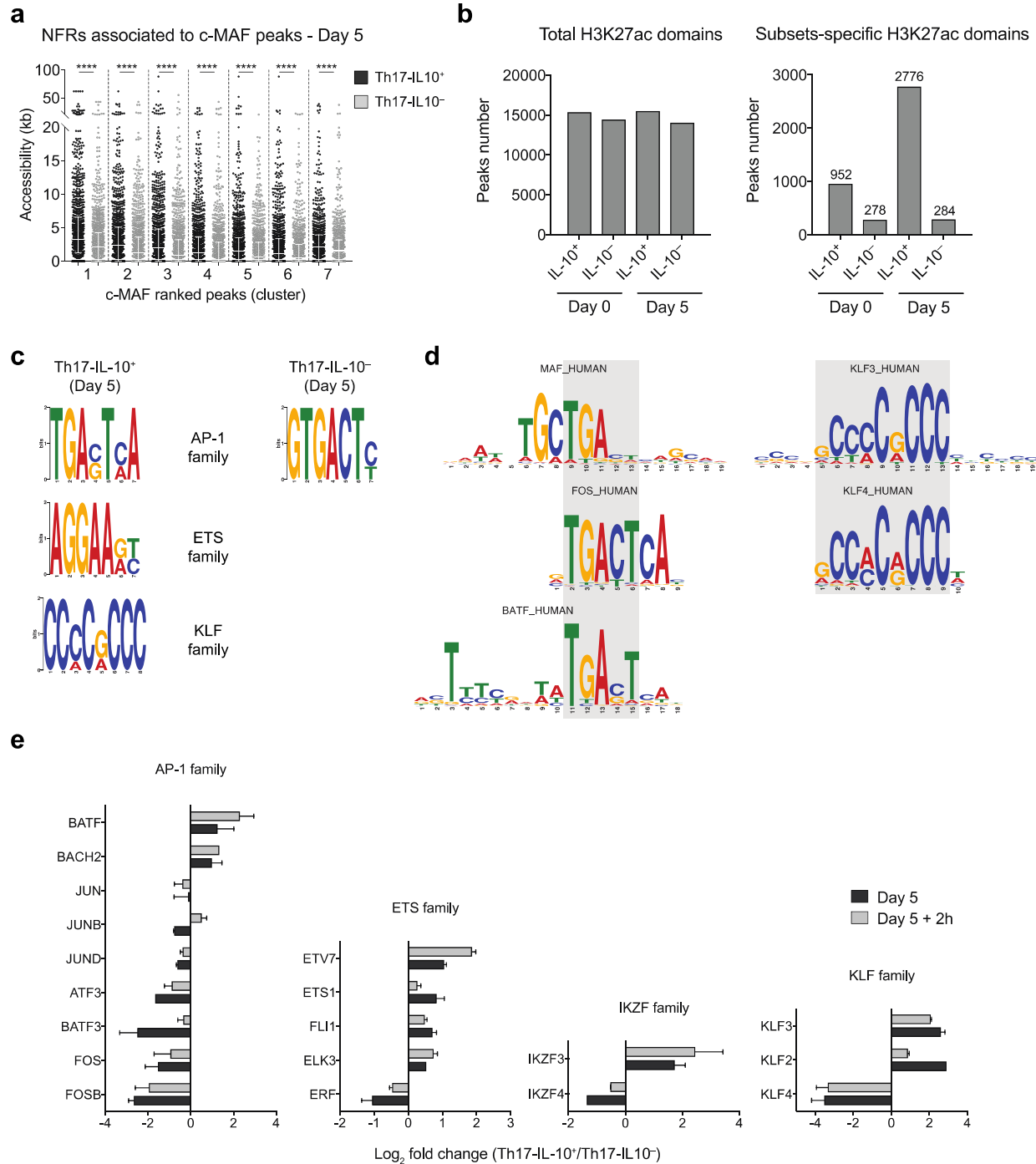
Supplementary Figure 6

Examples of c-MAF binding to immunoregulatory and tissue-residency genes loci

a-g. Graphical representation of c-MAF binding profiles at the locus of the indicated genes using the Integrative Genome Viewer (IGV).



c.i. (isoform a: n = 5, isoform b: n = 7). **c.** Expression of c-MAF and IL-10 in T_H17-IL10⁻ cells upon ectopic expression of c-MAF isoform b. Control (empty vector) cells and c-MAF-b (c-MAF isoform b) expressing T_H17-IL10⁻ cells were polyclonally stimulated with or without IL-27 (25 ng/ml) and c-MAF and IL-10 expression was measured in Day 5-activated cells by flow cytometry, after 5 h stimulation with PMA+I (mean + s.e.m.; n = 3). **d.** Expression of c-MAF-dependent genes in T_H17-IL10⁻ cells upon ectopic expression of MAF isoform a (left panel, n = 4) and b (right panel, n = 6), as measured by qPCR. Black bars indicate T_H17-IL10⁺-associated genes, grey bars indicate T_H17-IL10⁻-associated genes. Shown is the average log₂ fold change over empty vector (mean + s.e.m). n.a., not assessed. **e,f.** Representative dot plots (**e**) and cumulative data of expression of IFN- γ and IL-22 (**f**) as assessed by intracellular staining of Day 5-activated T_H17-IL10⁻ cells following ectopic expression of MAF isoform b or control empty vector (eV). Data are expressed as percentage over empty vector and represent the mean + s.e.m. (n = 7), with each dot indicating a T cell clone pool from independent experiments. ***P* < 0.01; ****P* < 0.001, as determined by ratio paired *t* test (**a-c**) and paired *t* test (**f**).



Supplementary Figure 8

Potential cooperative and antagonistic binding of transcription factors in T_H17-IL-10⁺ and T_H17-IL-10⁻ cells

a. c-MAF peaks from Day 5-activated T_H17-IL-10⁺ cells were ranked according to the fold enrichment over the input, corrected for *P*-value, and clustered in bins of 1000 peaks (cluster 7 is made of 778 peaks). c-MAF peaks-associated nucleosome-free regions (NFRs) in the same cells were identified from H3K27ac ChIP-seq data and their size was compared to the corresponding NFRs in Day 5-activated T_H17-IL-10⁻ cells. If a NFR was not detected, the size was reported as 0. Each dot in (a) represents a NFR and the white lines indicate median and interquartile ranges. *P*-values are obtained by Wilcoxon matched-pairs signed rank test. **b.** Number of total H3K27ac peaks (left panel) identified in Day 0 and Day 5 T_H17-IL-10⁺ and T_H17-IL-10⁻ cells, as assessed by ChIP-seq (MACS *P*-value

$\leq 10 \times 10^{-10}$, FDR $\leq 5\%$ and fold enrichment ≥ 5). Subset-specific H3K27ac peaks (right panel) were identified, among the total ones, as H3K27ac peaks that were specifically enriched (MACS P -value $\leq 10 \times 10^{-6}$ and fold enrichment ≥ 3) in the indicated population. Multiple H3K27ac peaks residing within 1 kb were collapsed into a single domain. **c.** Position weight matrices (PWMs) of DNA enriched motives identified in NFRs associated to recently-activated T_H17 -IL-10⁺-specific and T_H17 -IL-10⁻-specific H3K27ac domains. **d.** Consensus PWMs of the indicated transcription factors were downloaded from the Hocomoco v.11 database². Overlapping nucleotides in consensus motifs are highlighted by a light grey box in background **e.** Gene expression profile of the indicated transcription factors in Day 5-activated T_H17 -IL-10⁺ and T_H17 -IL-10⁻ cells was obtained from RNA-seq data (Fig. 4).

²Kulakovskiy, I.V. et al. HOCOMOCO: towards a complete collection of transcription factor binding models for human and mouse via large-scale ChIP-Seq analysis. *Nucleic Acids Res* 46, D252-D259 (2018).