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Supplementary Materials for

The endogenous retrovirus-derived long noncoding RNA TROJAN promotes triple-negative breast cancer progression via ZMYND8 degradation

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Published 6 March 2019, *Sci. Adv.* **5**, eaat9820 (2019) DOI: 10.1126/sciadv.aat9820

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Legend for Table S1

Other Supplementary Material for this manuscript includes the following:

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Table S1 (Microsoft Excel format). TNBC-related HERVs, basic patient information, primers, and MS data for proteins pulled down by TROJAN and SYSL1.

SUPPLEMENTARY FIGURE LEGENDS



Fig. S1. TROJAN properties. (A) Sequence homology of TROJAN among different species (UCSC genome browser database). (B) TROJAN secondary structure predicted by RNAfold (http://rna.tbi.univie.ac.at/cgi-bin/RNAWebSuite/RNAfold.cgi).
(C) The full-length sequence of TROJAN identified by RACE assay. (D) The estimated copy number of TROJAN per cell. The red dots indicate molecules per cell. Total RNA was collected from 10⁵ MDA-MB-231 LM2 and BT549 cells and analyzed by quantitative reverse transcription (qRT)-PCR. We first calculated that the

total RNA per cell was approximately 125 pg. The weight of 1 mol TROJAN (6.02×10^{23} copies) was 292544.69 g. A standard curve (blue dots) was generated by serial dilution of *in vitro* transcribed TROJAN RNA. (**E**) Validation of TROJAN expression using a pair of primers. Prior to reverse transcription, total MDA-MB-231 LM2 RNA was extracted and treated with or without deoxyribonuclease I (DNase I). N.C.: a negative control primer targeting a nontranscribed location in the genome of breast cancer cell lines (LINC01614). (**F**) Kaplan-Meier analysis of the disease-free survival based on TROJAN (CTC-260E6.4) in the TCGA data set. A log-rank test was used to determine statistical significance between the high expression group (n = 530) and the low expression group (n = 532). (**G**) TROJAN RNA expression in different subcellular fractions of MDA-MB-231 LM2 cells. Red indicates the nuclear fraction (Nu), and blue indicates the cytoplasmic (Cyto) fraction. The percentages were calculated by qRT-PCR. (**H**) Relative TROJAN expression in total RNA extracted and treated with or without poly(A) polymerase. The percentages were calculated by qRT-PCR.

Α	NGS-F							NGS-R																	
	***	*****	*****	***	****	* ** **	*	*	** **	*	****	****	****	****	* *	***	**	* *	**	***	* 1	*****	***	**:	**
TROJAN	TCATCT	ICTTGAA	ACTGT	rc <mark>a</mark> ct	ATTGC	CACAAG	AGATG	T <mark>A</mark> TATT	AA CTT	AGT	CTTTG	CTCCC	AGGTI	ATA7	TCC	TAAA	CTTC	ACCO	AAA	T <mark>AAA</mark>	CTG	ICTAC	TTA	C <mark>a</mark> t'	TC
ZNF253-LTR70	TC <mark>A</mark> TCI	ICTTG <mark>AA</mark>	ACTGTI	rc <mark>a</mark> ct	ATTGT	C <mark>AC</mark> AGG	AGCTA	T <mark>AAA</mark> TT.	AA CTT	GAAT	CTTTG	CTCCC	AGGTI	ATA	TCC	TAAG	CTTC	ACCO	AAA	T <mark>AAA</mark>	CTG	ICT <mark>A</mark> C	TTA:	T <mark>A</mark> T:	TCP
ZNF107-LTR70	TCATC1	ICTTG <mark>A</mark> A	ACTGT	ACACI	ATTGC	CCC <mark>AA</mark> GI	AGCTA	T	AA CTT	5 <mark>AA</mark> T	CTTTG	CTCCC	AGGTI	A <mark>T</mark> AA	TTC	T <mark>AA</mark> G	TTTC	ACC	AAA	T <mark>AAA</mark>	GTG	ICT AC	TT <mark>A</mark>	TAT:	TCP
ZNF826p-LTR70-1	TCATCC	CCTTGAA	ACTGTI	rc <mark>a</mark> ct	ATTGC	C <mark>AC</mark> AAGI	AGCTA	TAAAGT.	AA CTT	5 <mark>AA</mark> T	CTTTG	CTCCC	TGGTI	ATA7	TCC	TAAG	CTT	AACO	AAA	T <mark>AAA</mark>	CTG	ICT AC	TT <mark>a</mark> :	T <mark>A</mark> T!	тст
ZNF90-LTR70	GTATCT	IGTTG <mark>A</mark> A	ACTGT	rc <mark>a</mark> ca	ATTGC	C <mark>AC</mark> AAG1	AGTTA	T <mark>AAA</mark> TT	AA CTT	GAAT	ATTTG	CTCCC	AGGTI	ATA7	TCC	TAAA	CTTO	ACCO	AAA	T <mark>AA</mark> G	CTG	ICT AC	TTA:	TAT:	TCP
ZNF486-LTR70	TC <mark>A</mark> TCI	F <mark>attg</mark> aa	ACTGTI	rc <mark>a</mark> ct	ATTGG	C <mark>AC</mark> AAG	GACTA	T <mark>AAA</mark> TT.	AA CTT	GAAT	CTTTG	CTCCC	AGGTI	ATA	CCC	TAAG	CTTO	ACCO	AAA	TAAA	CTG	ICT <mark>A</mark> C	TT <mark>a</mark> :	T <mark>A</mark> T!	TC P
ZNF826p-LTR70-2	TC <mark>A</mark> TCI	ICTTG <mark>A</mark> A	ACTGT	IC <mark>A</mark> CI	ATTGG	C <mark>AC</mark> AAGI	ACCTA	TAAGTT.	AA ATT	GAAC	CTTTG	CTCCC	AGGTI	ATA7	TCC	TAAA	CTTO	ACCO	AAA	T <mark>AAA</mark>	CTG	ICT AC	TTA:	TAT:	TCP
ZNF93-LTR70	TCATCI	ICTTGAA	ACTGTI	rc <mark>a</mark> cı	ATTGC	CACAAG1	AATTA	T <mark>AAA</mark> TT	AA CTT	GAAT	CTTTG	CTCCC	AGGTI	ATA	TCC	TAAG	CTTO	ACCO	AAG	TAAA	CTA	CT <mark>A</mark> C	TTA:	TAT:	TCP

в

A list of primers of TROJAN and the proportion of TROJAN among PCR products.

Primer Name	Sequence (5' to 3')	TROJAN %
TROJAN-1F	TGTCTCTGCAGTCTCTTAAGCA	12/12-1000/
TROJAN-1R	GGGACAGCATGCACTTTGTT	12/12-100%
TROJAN-2F	ATGTGACACTAGAGTGCTGCTG	
TROJAN-2R	TTTAGCAGCTTGAAGCCAGG	5/7=71.4%
TROJAN-3F	GCTCAGATGGAGATCTGTTTTCT	7/0-77 00/
TROJAN-3R	GGTCATACATAGTTCATCCTAAATTCAC	//9=//.8%
TROJAN-4F	ACCTGGCTTCAAGCTGCTAAA	11/11-100%
TROJAN-4R	TTGACTGGAAGTCTGAGAGGGAA	11/11=100%

С	TROJAN	
<	<<<<<<<<<<<<<<<<<<<<<<<<<<<<<<<<<<<<<<<	<<<<<<
Scale chr19: 1000_ GM78 cel pA+ - 1	500 bases - I hg19 20,331,500 I GM12878 whole cell polyA+ RNA-seq Minus signal Rep 1 from ENCODE/CSHL	20,332,500
1 1000_ H1hSC cel pA+ - 1	H1-hESC whole cell polyA+ RNA-seq Minus signal Rep 1 from ENCODE/CSHL	
K562 cel pA+ - 1	K562 whole cell polyA+ RNA-seq Minus signal Rep 1 from ENCODE/CSHL	
1000_ A549 cel pA+ - 1	A549 whole cell polyA+ RNA-seq Minus signal Rep 1 from ENCODE/CSHL	
CD20 cel pA+ - 1	CD20+ whole cell polyA+ RNA-seq Minus signal Rep 1 from ENCODE/CSHL	
HeS3 cel pA+ - 1	HeLa-S3 whole cell polyA+ RNA-seq Minus signal Rep 1 from ENCODE/CSHL	
1000 _ HeG2 cel pA+ - 1 1	HepG2 whole cell polyA+ RNA-seq Minus signal Rep 1 from ENCODE/CSHL	
1000 _ HUVEC cel pA+ - 1 1	HUVEC whole cell polyA+ RNA-seq Minus signal Rep 1 from ENCODE/CSHL	
IMR90 cel pA+ - 1 1	IMR90 whole cell polyA+ RNA-seq Minus signal Rep 1 from ENCODE/CSHL	
1000 _ MCF7 cel pA+ - 1 1	MCF-7 whole cell polyA+ RNA-seq Minus signal Rep 1 from ENCODE/CSHL	
1000 _ CD14 cel pA+ - 11	MonocytesCD14+ whole cell polyA+ RNA-seq Minus signal Rep 1 from ENCODE/CSHL	
1000_ SKNSH cel pA+ - 1 1	SK-N-SH whole cell polyA+ RNA-seq Minus signal Rep 1 from ENCODE/CSHL	
AG50 cel pA+ - 1 1	AG04450 whole cell polyA+ RNA-seq Minus signal Rep 1 from ENCODE/CSHL	
BJ cel pA+ - 1	BJ whole cell polyA+ RNA-seq Minus signal Rep 1 from ENCODE/CSHL	
1000 _ HMEC cel pA+ - 1 1	HMEC whole cell polyA+ RNA-seq Minus signal Rep 1 from ENCODE/CSHL	
1000 _ HSMM cel pA+ - 1 1	HSMM whole cell polyA+ RNA-seq Minus signal Rep 1 from ENCODE/CSHL	
1000 _ NHEK cel pA+ - 1 1	NHEK whole cell polyA+ RNA-seq Minus signal Rep 1 from ENCODE/CSHL	
1000 _ NHLF cel pA+ - 11	NHLF whole cell polyA+ RNA-seq Minus signal Rep 1 from ENCODE/CSHL	
SKRA cel pA+ - 1 Figure S2 1	SK-N-SH RA whole cell polyA+ RNA-seq Minus signal Rep 1 from ENCODE/CSHL	

Fig. S2. Expression pattern of TROJAN. (A) The compatible pair of primers covering eight expressed LTR70s used for RNA-Seq to analyze the constituent ratio of LTR70 in Fig. 1. (B) A list of primers for TROJAN and the proportion of TROJAN in the PCR products. (C) The RNA-Seq data were acquired from the UCSC genome browser database (Long RNA-seq from ENCODE/Cold Spring Harbor Lab).



Fig. S3. TROJAN phenotype. (A) Quantitative reverse transcription (qRT)-PCR analysis of relative TROJAN transcription in MDA-MB-231 LM2 and BT549 cells expressing control (Ctrl) or TROJAN short hairpin RNA (shRNA). The data are presented as the mean \pm s.d.; n = 3 independent experiments; two-tailed unpaired Student's *t*-test. (**B**) qRT-PCR detection of relative TROJAN transcription in MDA-MB-231 LM2 cells overexpressing a control vector (Vec) or TROJAN. The data are presented as the mean \pm the s.d.; n = 3 independent experiments; two-tailed unpaired Student's *t*-test. (**C**) *In vitro* growth curves of HMEC and MCF-10A cells expressing Ctrl or TROJAN shRNA. The data are presented as the mean \pm s.d.; n = 3 independent experiments. (**D**) *In vitro* transwell migration assay of MDA-MB-231 LM2 cells expressing Vec or TROJAN. The data are presented as the mean \pm s.d.; n = 3 independent experiments; two-tailed unpaired Student's *t*-test. Scale bar: 200 µm.

(E) *In vitro* transwell invasion assay of MDA-MB-231 LM2 cells expressing Vec or TROJAN. The data are presented as the mean \pm s.d.; n = 3 independent experiments; two-tailed unpaired Student's *t*-test. Scale bar: 200 µm. (F) *In vitro* transwell migration assay of BT549 cells expressing Vec or TROJAN. The data are presented as the mean \pm s.d.; n = 3 independent experiments; two-tailed unpaired Student's *t*-test. Scale bar: 200 µm. (G) *In vitro* transwell invasion assay of BT549 cells expressing Vec or TROJAN. The data are presented as the mean \pm s.d.; n = 3 independent experiments; two-tailed unpaired Student's *t*-test. Scale bar: 200 µm. (G) *In vitro* transwell invasion assay of BT549 cells expressing Vec or TROJAN. The data are presented as the mean \pm s.d.; n = 3 independent experiments; two-tailed unpaired Student's *t*-test. Scale bar: 200 µm. (H) The numbers of lung metastases and metastatic nodules are shown. The data are presented as the mean \pm s.d.; two-tailed unpaired Student's *t*-test. Scale bar: 200 µm. (I) H&E staining of bone metastasis of SCP2 cells expressing Ctrl or TROJAN shRNA. Scale bar: 50 µm. The asterisks indicate bone metastasis. (J) Incidence of liver metastasis of MDA-MB-231 LM2 cells expressing Ctrl or TROJAN shRNA in the mammary fat pad xenograft model. *p < 0.05, ***p < 0.001. NS: not significant.



Fig. S4. Construction of the TROJAN knockout cell line. (**A**) Schematic diagram of the modification of the plasmid LentiCRISPRv2 and the construction of a plasmid

expressing paired guide RNAs (pgRNAs) flanking TROJAN. (B) Schematic diagram of the deletion of TROJAN. (C) PCR products generated from the genomic sequence around TROJAN showing the efficiency of targeted deletion in MDA-MB-231 LM2 cells expressing pgRNAs. TROJAN wild type (WT) or TROJAN knockout (KO). (D) Quantitative reverse transcription (qRT)-PCR analysis of relative TROJAN transcription levels in MDA-MB-231 LM2 cells with TROJAN WT or TROJAN KO. The data are presented as the mean \pm s.d.; n = 3 independent experiments; two-tailed unpaired Student's t-test. (E) In vivo growth of MDA-MB-231 LM2 cells (31 days; n = 5) with TROJAN WT, TROJAN KO or TROJAN KO plus ectopically expressed TROJAN (KO + TROJAN). Tumor volume quantification (left) and representative tumor images (right) are shown. The data are presented as the mean \pm s.d.; two-tailed unpaired Student's t-test. (F) In vivo intravenous xenograft mouse model of MDA-MB-231 LM2 cells (42 days; n = 3) with TROJAN WT, TROJAN KO or TROJAN KO plus exogenous TROJAN expression (KO + TROJAN). The relative bioluminescence intensity (BLI) indicates lung metastasis. The numbers of lung metastases and metastatic nodules are shown. The data are presented as the mean \pm s.d.; two-tailed unpaired Student's t-test. **p < 0.01, ***p < 0.001.



Fig. S5. Potential therapeutic role of TROJAN in breast cancer progression. (A) In vitro growth assay of BT549 and Hs578t cells transfected with ASO-4 targeting TROJAN or Ctrl. The data are presented as the mean \pm s.d.; n = 3 independent experiments; two-tailed unpaired Student's t-test. (B) In vitro growth assay of murine 4T1 breast cancer cells transfected with ASOs targeting TROJAN or Ctrl. CCK8 reagent was added after 72 hours, and the OD450 was determined. The data are presented as the mean \pm s.d.; n = 3 independent experiments. (C) In vitro free uptake assay in MDA-MB-231 LM2 cells. Six different concentrations of ASO were assessed $(0, 0.5, 1, 2.5, 5 \text{ and } 10 \,\mu\text{M})$. The 0 μM group was set as the control. Quantitative reverse transcription (qRT)-PCR detection of relative TROJAN expression. The data are presented as the mean \pm s.d.; n = 3 independent experiments. (**D**) qRT-PCR analysis of relative TROJAN expression in the metastatic nodules of mice treated with an ASO targeting TROJAN or PBS. The data are presented as the mean \pm s.d.; n = 3 independent experiments; two-tailed unpaired Student's *t*-test. ***p < 0.001. (E) Weights of the mice, as well as those of their livers, kidneys and spleens, in the PBS and ASO treatment groups. The data are presented as the mean \pm s.d.; two-tailed unpaired Student's t-test. (F) Serum chemistry markers of liver function (ALT, AST

and TB) and renal function (urea) in the PBS and ASO treatment groups. ALT: alanine aminotransferase; AST: aspartate aminotransferase; and TB: total bilirubin. The data are presented as the mean \pm s.d.; two-tailed unpaired Student's *t*-test. *p < 0.05, **p < 0.01 and ***p < 0.001. NS: not significant.



Figure S6

Fig. S6. TROJAN associates with the ZMYND8 protein. (A) Schematic diagram of the RNA pull-down assay combined with stable isotope labeling with amino acids (SILAC)-based quantitative proteomics and the top 10 potential TROJAN interaction proteins identified by mass spectrometry according to intensity. (B) Top 10 potential TROJAN-specific interacting proteins identified by mass spectrometry. (C) Quantitative reverse transcription (qRT)-PCR analysis of relative transcription levels from control vector (Vec), TROJAN full-length (FL) and mutant TROJAN ($\Delta 4$). The data are presented as the mean \pm s.d.; n = 3 independent experiments; two-tailed unpaired Student's t-test. (D) In vitro transwell migration assay of MDA-MB-231 LM2 cells expressing control empty vector (Vec), full-length TROJAN (FL) or mutant TROJAN (Δ 1-4). Scale bar: 200 μ m. (E) Quantification of the transwell migration assay. The data are presented as the mean \pm s.d.; n = 3 independent experiments; two-tailed unpaired Student's t-test. (F) In vivo intravenous xenograft mouse model of MDA-MB-231 LM2 cells (42 days; n = 3) expressing control empty vector (Vec), full-length TROJAN (FL) or mutant TROJAN ($\Delta 4$). The numbers of lung metastases and metastatic nodules are shown. The data are presented as the mean \pm s.d.; two-tailed unpaired Student's *t*-test. (G) Representative LTR56 genome locations (left) and RNA immunoprecipitation (RIP) assays (right) with subsequent qRT-PCR assays in HEK293T cells ectopically expressing full-length Flag-tagged ZMYND8. The relative quantification of TROJAN and LTR56 expression in RNA-protein complexes immunoprecipitated with IgG or Flag antibodies is shown. The data are presented as the mean \pm s.d.; n = 3 independent experiments; two-tailed unpaired Student's *t*-test. *p < 0.05 and ***p < 0.001. NS: not significant.



Figure S7

Fig. S7. TROJAN degrades the ZMYND8 protein. (A) Immunohistochemical staining of ZMYND8 in lungs isolated from mice administered MDA-MB-231 LM2 cells expressing short hairpin RNAs (shRNAs) targeting TROJAN or control (Ctrl) by tail vein injection. Scale bar: 50 μ m. The quantitative data are presented as the mean \pm s.d.; n = 3 independent experiments; two-tailed unpaired Student's *t*-test. (**B**)

Quantitative reverse transcription (qRT)-PCR analysis of relative ZMYND8 transcription after TROJAN knockdown using shRNAs. The data are presented as the mean \pm s.d.; n = 3 independent experiments; two-tailed unpaired Student's *t*-test. NS: not significant. (C) Western blot images showing ZMYND8 expression in MDA-MB-231 LM2 cells expressing control or TROJAN shRNA treated with DMSO (control) or chloroquine diphosphate. (D) Western blot images showing ZMYND8 expression in MDA-MB-231 LM2 cells expressing Ctrl or ZMYND8 shRNA. (E) Transwell migration assay of MDA-MB-231 LM2 cells treated with ASO-4 and/or ZMYND8 shRNA. The data are presented as the mean \pm s.d. ***p < 0.001, n = 3 independent experiments; two-tailed unpaired Student's t-test. Scale bar: 200 µm. (F) qRT-PCR analysis of relative TROJAN transcription after ZMYND8 knockdown using shRNAs. The data are presented as the mean \pm s.d.; n = 3 independent experiments; two-tailed unpaired Student's t-test. NS: not significant. (G) Western blot images showing ZMYND8 expression in MDA-MB-231 LM2 cells expressing control or ZNF592 small interfering RNA (siRNA) treated with DMSO (control) or MG132. (H) Western blot images showing ZMYND8-associated ubiquitination in control and ZNF592-downregulated HEK293T cells ectopically expressing full-length Flag-tagged ZMYND8 and treated with MG132. *p < 0.05 and ***p < 0.001. NS: not significant.

Fig. S8. TROJAN and ZMYND8 regulate the transcription of diverse target genes. (**A**) Pathway analysis of potential TROJAN target genes identified by microarray. (**B**) Quantitative reverse transcription (qRT)-PCR analysis of the relative transcription levels of the potential target genes in MDA-MB-231 LM2 and BT549 cells expressing control or TROJAN short hairpins RNAs (shRNAs). The data are

presented as the mean \pm s.d.; n = 3 independent experiments; two-tailed unpaired Student's *t*-test. (C) qRT-PCR detection of the target genes in MDA-MB-231 LM2 cells expressing TROJAN and/or ZMYND8 shRNAs. The data are presented as the mean \pm s.d.; n = 3 independent experiments. (**D**) qRT-PCR detection of the target genes in MDA-MB-231 LM2 cells with TROJAN knockout (KO) and/or ZMYND8 KO. The data are presented as the mean \pm s.d.; n = 3 independent experiments. (E) qRT-PCR analysis of ZMYND8 occupying the EGFR, VEGFA and MDM2 promoters after chromatin immunoprecipitation (ChIP) assay. (F) qRT-PCR analysis of TROJAN RNA recovery and enrichment of the promoter regions of potential target genes after chromatin isolation by RNA purification (ChIRP) assay. LacZ was set as the negative control. The data are presented as the mean \pm s.d.; two-tailed unpaired Student's *t*-test. (G) qRT-PCR analysis of KDM5C and ZMYND8 at the EGFR, VEGFA and MDM2 promoters after ChIP assay in MDA-MB-231 LM2 cells expressing control or TROJAN shRNA. (H) qRT-PCR analysis of H3K4me3 at the EGFR, VEGFA and MDM2 promoters after ChIP assay in MDA-MB-231 LM2 cells expressing control or TROJAN shRNA. The data are presented as the mean \pm s.d.; two-tailed unpaired Student's *t*-test. *p < 0.05, **p < 0.01 and ***p < 0.001. NS: not significant.

Fig. S9. TROJAN regulates breast cancer progression via ZMYND8. (A) Correlations of TROJAN with EGFR, VEGFA and MDM2 expression as determined by Pearson correlation analyses. (**B**, **C**) Kaplan-Meier analyses of the association of ZMYND8 with relapse-free survival and overall survival in FUSCC cohorts 1 (**B**) and 2 (**C**).

SUPPLEMENTARY TABLE LEGENDS AND TABLES

Table S1. TNBC-related HERVs, basic patient information, primers, and MSdata for proteins pulled down by TROJAN and SYSL1.