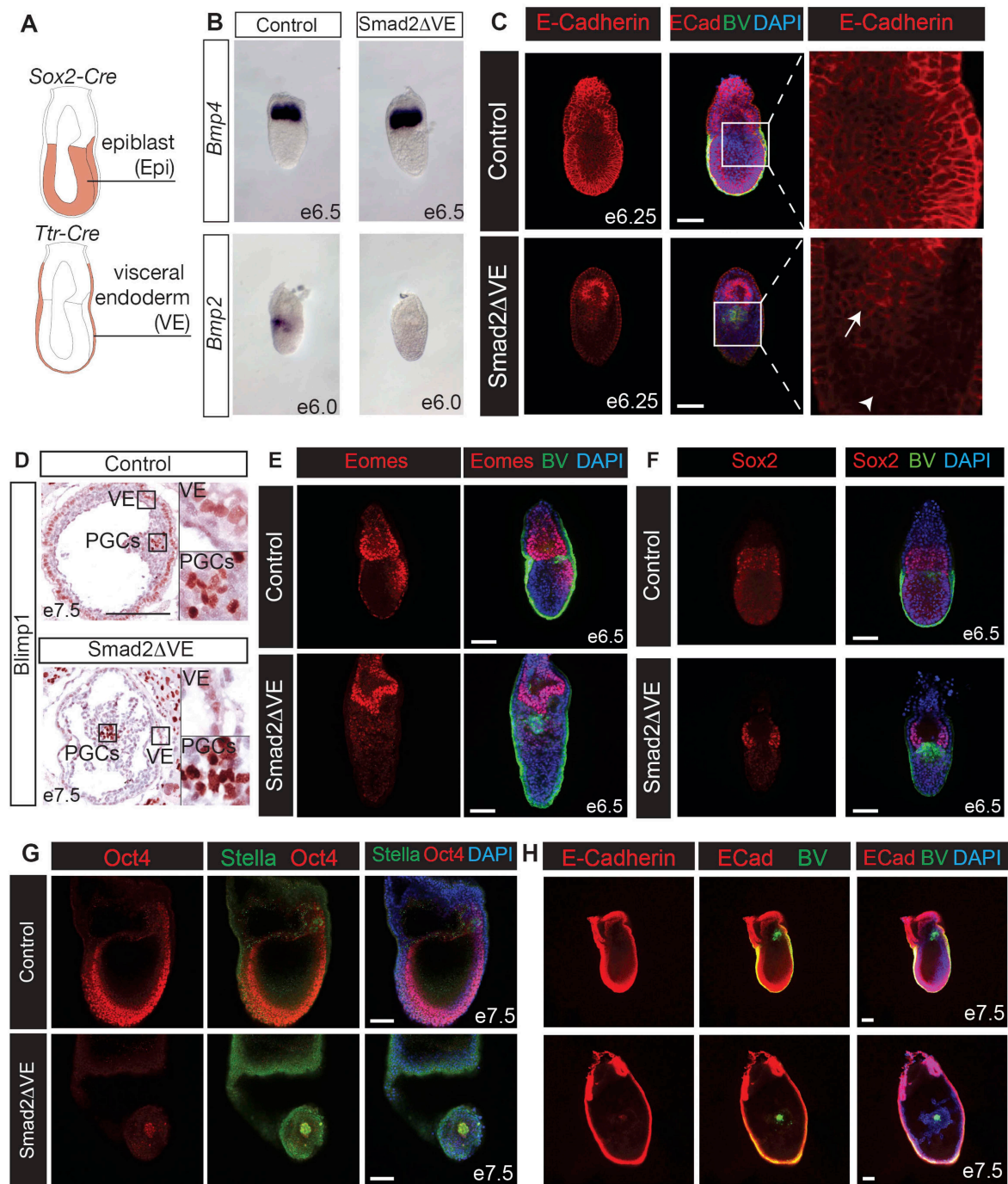


## **Supplementary Information**

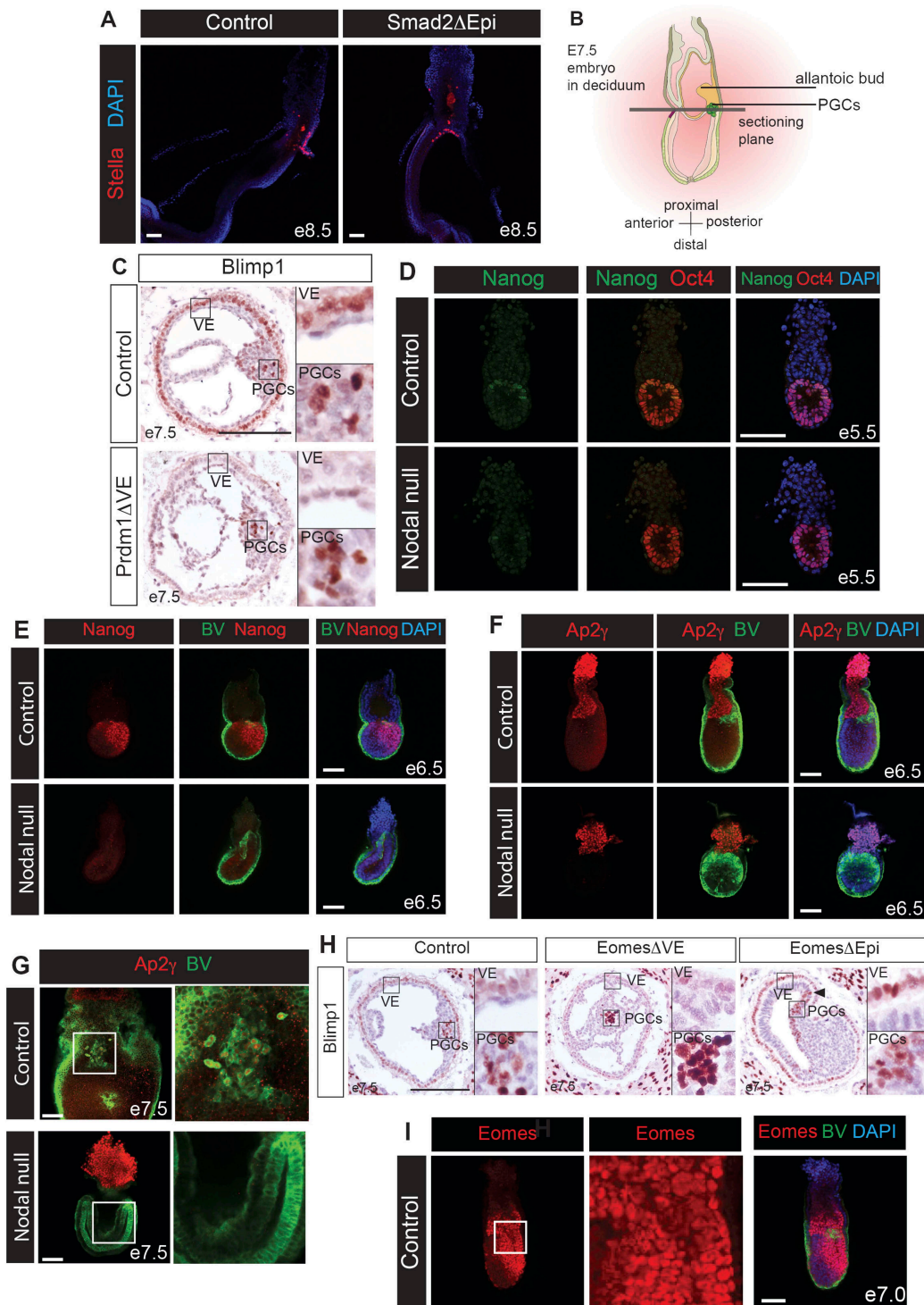
# **Genetic dissection of Nodal and Bmp signalling requirements during primordial germ cell development in mouse**

Anna D. Senft, Elizabeth K. Bikoff, Elizabeth J. Robertson and Ita Costello



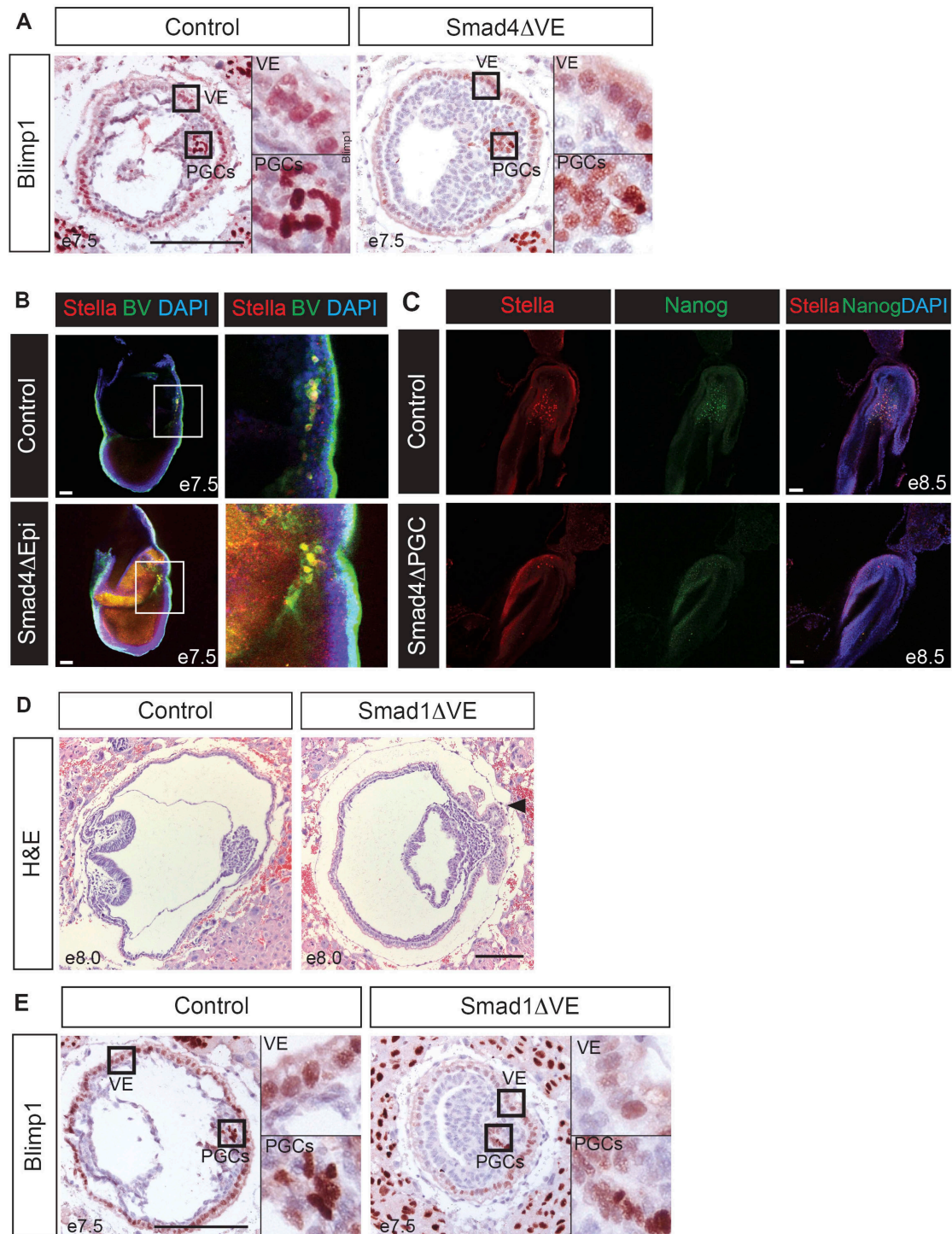
**Supplementary figure 1: Conditional Smad2 inactivation in the visceral endoderm results in expansion of the PGC niche.**

(A) Tissue specific activity of the Sox2 and Ttr Cre deleter strains. (B) Whole-mount *in situ* hybridisation analysis of *Bmp4* and *Bmp2* expression in e6.5 and e6.0 control and Smad2 $\Delta$ VE embryos. (C) E-Cadherin (ECad) staining of e6.25 control and Smad2 $\Delta$ VE BV<sup>+</sup> embryos. Arrow indicates E-Cadherin staining in BV-expressing cells. Arrowhead indicates lack of E-Cadherin staining in surrounding mesoderm-like cells. (D) Immunohistochemistry (IHC) staining of sections of e7.5 control and Smad2 $\Delta$ VE embryos using a Blimp1 antibody. Boxed regions are expanded on the right. Scale bar = 200 $\mu$ m. (E) *Eomes* expression in control and Smad2 $\Delta$ VE BV<sup>+</sup> e6.5 embryos. (F) *Sox2* expression in BV<sup>+</sup> e6.5 embryos. (G) *Stella* and *Oct4* co-localization in e7.5 control and Smad2 $\Delta$ VE embryos. (H) E-Cadherin staining of e7.5 control and Smad2 $\Delta$ VE embryos. All IF images are counterstained with DAPI. All IF scale bars = 100 $\mu$ m.



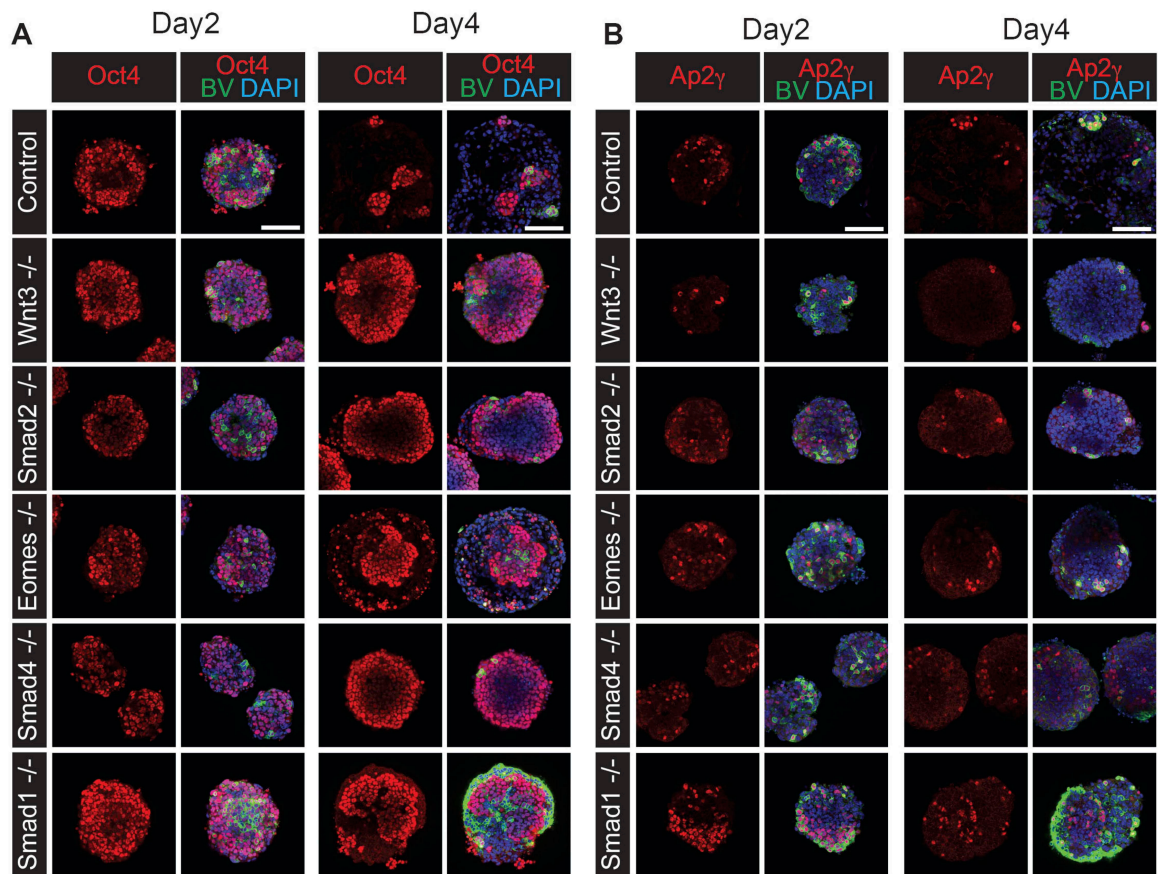
### Supplementary figure 2: Smad2, Prdm1, Nodal and Eomes functional contributions during PGC development.

(A) Stella IF staining in control and Smad2 $\Delta$ Epi e8.5 embryos. (B) Plane of section used for IHC staining. (C) Blimp1 IHC in control and Prdm1 $\Delta$ VE sections shows selective deletion of Blimp1 in the VE of Prdm1 $\Delta$ VE embryos fails to prevent PGC formation. Scale bar = 200 $\mu$ m. (D) Nanog and Oct4 staining in e5.5 control and Nodal null embryos. (E) Nanog staining in e6.5 control and Nodal null BV<sup>+</sup> embryos. (F) Ap2 $\gamma$  staining in e6.5 control and Nodal null BV<sup>+</sup> embryos (G) Ap2 $\gamma$  staining in e7.5 control and Nodal null BV<sup>+</sup> embryos. (H) Blimp1 IHC in e7.5 control, Eomes $\Delta$ VE and Eomes $\Delta$ Epi embryos. Boxed areas indicated expanded panels on the right of the image. Scale bar = 200 $\mu$ m. (I) Eomes staining in mid-late streak stage control BV<sup>+</sup> embryos. All IF scale bars = 100 $\mu$ m.



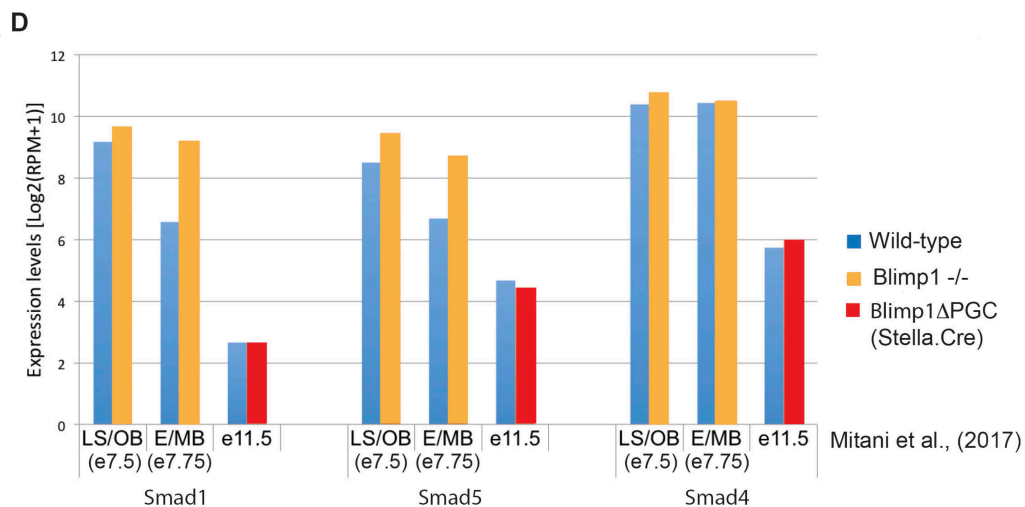
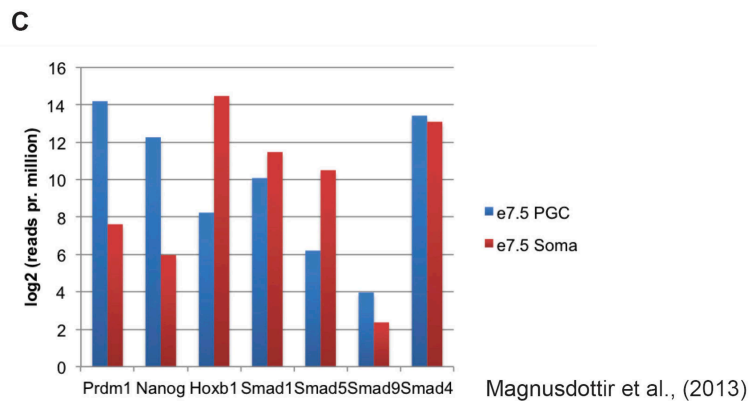
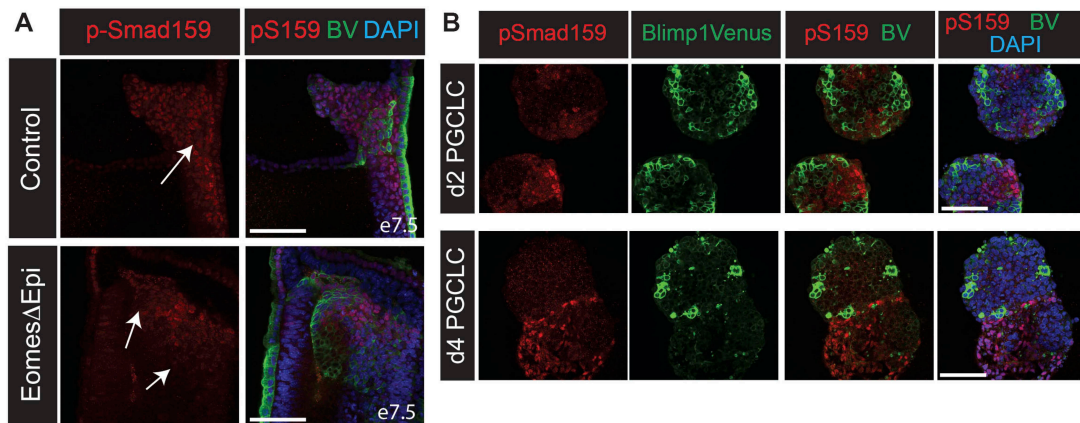
**Supplementary figure 3: Smad4 and Smad1 requirements for promoting PGC development.**

(A) Blimp1 IHC staining of e7.5 control and Smad4 $\Delta$ VE embryo sections. Boxed areas indicated expanded panels on the right of the image. Scale bar = 200 $\mu$ m. (B) Stella expression in e7.5 control and Smad4 $\Delta$ Epi BV<sup>+</sup> embryos. (C) Optical sections of the posterior region of e8.5 control and Smad4 $\Delta$ PGC embryos co-stained with Stella and Nanog antibodies, counterstained with DAPI. All IF scale bars = 100 $\mu$ m. (D) H&E staining of e8.0 control and Smad1 $\Delta$ VE embryo sections. Arrowhead indicates ruffling of the posterior VE in Smad1 $\Delta$ VE embryos. (E) Blimp1 IHC in control and Smad1 $\Delta$ VE e7.5 embryo sections. Boxed areas indicated expanded panels on the right of the image. Scale bar = 200 $\mu$ m.



**Supplementary figure 4: PGC marker gene expression in wild type compared to Nodal, Bmp and Wnt signalling mutant PGCLC aggregates.**

(A) Oct4 staining of day 2 and day 4 PGCLC BV<sup>+</sup> aggregates of indicated genotypes, counterstained with DAPI. (B) Ap2 $\gamma$  staining of day 2 and day 4 PGCLC BV<sup>+</sup> aggregates of indicated genotypes, counterstained with DAPI. All scale bars = 100 $\mu$ m.



### Supplementary figure 5: PGCs selectively repress expression of Bmp/Smad signalling components.

(A) p-Smad159 staining in e6.5 control and *EomesΔEpi* BV<sup>+</sup> embryos. Arrows indicate loss of p-Smad159 signal in the BV<sup>+</sup> cell population (B) p-Smad159 staining in BV<sup>+</sup> wildtype d2 and d4 PGCLC aggregates. All IF scale bars = 100μm. (C) RNA-seq data showing expression levels [log<sub>2</sub> (reads per million)] of *Smad1*, *Smad5*, *Smad9* and *Smad4* in e7.5 PGCs (blue) compared to somatic cells (red)<sup>Ref 1</sup>. Expression levels of *Prdm1* and *Nanog* (PGC markers) and *Hoxb1* (somatic cell marker) are shown for comparison. (D) RNA-Seq data at indicated time-points showing expression [Log<sub>2</sub>(reads per million + 1)] of *Smad1*, *Smad5* and *Smad4* in BV<sup>+</sup> PGC cells isolated from wild type (blue), *Blimp1* null (orange) or embryos where *Blimp1* has been deleted from specified PGCs using a *Stella-Cre* deleter strain (*Blimp1ΔPGC*) (red)<sup>Ref 2</sup>. LS/OB, late streak/no bud stage embryo; E/MB, early/mid bud stage embryos.

**Supplementary table 1: Characterization of *Prdm1* $\Delta$ VE mice**

A. Genotypes of offspring from conditional deletion of *Prdm1* from VE

*Prdm1*<sup>-/+</sup>; *Ttr-Cre*<sup>+</sup> x *Prdm1*<sup>CA/CA</sup>

Genotype	<i>Prdm1</i> <sup>+ /CA</sup>	<i>Prdm1</i> <sup>+/<math>\Delta</math>VE</sup>	<i>Prdm1</i> <sup>m/CA</sup>	<i>Prdm1</i> <sup>m/<math>\Delta</math>VE</sup>
No of animals	2	3	3	3
Percentage	18.2	27.3	27.3	27.3
Expected %	25	25	25	25

B. Viability test of *Prdm1* $\Delta$ VE intercross

*Prdm1*<sup>m/ $\Delta$ VE</sup> x *Prdm1*<sup>m/ $\Delta$ VE</sup>

No of animals	9
---------------	---

**Supplementary table 2: Mouse strains used in this study**

Embryo name	Genotype	Phenotype Reference	Additional transgene (Ref 3)
Smad2ΔVE	<i>Smad2<sup>CA/-</sup>; Ttr-Cre<sup>+/-</sup></i>		<i>Blimp1-mVenus</i>
Smad2ΔEpi	<i>Smad2<sup>CA/-</sup>; Sox2-Cre<sup>+/-</sup></i>	Ref 4	
EomesΔVE	<i>Eomes<sup>CA/-</sup>; Ttr-Cre<sup>+/-</sup></i>	Ref 5	
EomesΔEpi	<i>Eomes<sup>CA/-</sup>; Sox2-Cre<sup>+/-</sup></i>	Ref 6	<i>Blimp1-mVenus</i>
Smad1ΔVE	<i>Smad1<sup>CA/-</sup>; Ttr-Cre<sup>+/-</sup></i>		
Smad1ΔEpi	<i>Smad1<sup>CA/-</sup>; Sox2-Cre<sup>+/-</sup></i>		<i>Blimp1-mVenus</i>
Smad1 null	<i>Smad1<sup>-/-</sup></i>	Ref 7	<i>Blimp1-mVenus</i>
Smad4ΔVE	<i>Smad4<sup>CA/-</sup>; Ttr-Cre<sup>+/-</sup></i>	Ref 8	
Smad4ΔEpi	<i>Smad4<sup>CA/-</sup>; Sox2-Cre<sup>+/-</sup></i>	Ref 9	<i>Blimp1-mVenus</i>
Smad4ΔPGC	<i>Smad4<sup>CA/-</sup>; Prdm1-Cre<sup>+/-</sup></i>		<i>Blimp1-mVenus</i>
Prdm1ΔVE	<i>Prdm1<sup>CA/-</sup>; Ttr-Cre<sup>+/-</sup></i>		
Prdm1 null	<i>Prdm1<sup>-/-</sup></i>	Ref 10	<i>Blimp1-mVenus</i>
Nodal null	<i>Nodal<sup>-/-</sup></i>	Ref 11	<i>Blimp1-mVenus</i>

Sox2-Cre reference: Ref 12. Ttr-Cre reference: Ref 13.



**Supplementary table 3: Antibodies and datasets used in this study**

Antibodies	SOURCE	IDENTIFIER
Rabbit polyclonal anti-Nanog	Abcam	Cat# ab80892, RRID: AB_2150114 Lot: GR40243-12
Goat polyclonal anti-Sox2	R&D Systems	Cat# AF2018, RRID: AB_355110
Rabbit polyclonal anti-Stella	Santa Cruz	Cat# sc-67249, RRID: AB_1128372, Lot: H1809
Goat polyclonal anti-Stella	R&D Systems	Cat# AF2566, RRID: AB_2094147, Lot:UWS010811 1
Goat polyclonal anti-Oct4	Santa Cruz	Cat# sc-8628, RRID: AB_653551, Lot: F1815
Rabbit polyclonal anti-Ap2 $\gamma$	Santa Cruz	Cat# sc-8977, RRID: AB_2286995, Lot: G1112
Goat polyclonal anti-Brachyury (N-19)	Santa Cruz	Cat# sc-17743 RRID: AB_634980 Lot: A1614
Rat monoclonal anti-E-Cadherin	Sigma-Aldrich	Cat#U3254 RRID: AB_477600 Lot: 085K4798
Rabbit polyclonal anti-Eomes	Abcam	Cat#ab23345 RRID: AB_778267 Lot:GR306193-1
Rabbit monoclonal anti-phospho Smad1/5/9	Cell Signaling Technology	Cat# 13820 RRID: AB_2493181 Lot: D5810
Goat polyclonal anti-Otx2	R&D Systems	Cat# AF1979, RRID: AB_2157172, Lot: KNO0615111

Rat monoclonal anti-Blimp1	Santa Cruz	Cat# sc-130917, RRID: AB_2169704, Lot: G0913
Rabbit polyclonal anti-GFP Alexa Fluor 488	Invitrogen	Cat# A21311, RRID: AB_221477, Lot: 1891008
Chicken polyclonal anti-GFP	Abcam	Cat# Ab13970, RRID: AB_300798, Lot: GR305729-1
Goat anti-chicken Alexa Fluor 488	Invitrogen	Cat# A11039, RRID: AB_142924,
Donkey anti-rabbit Alexa Fluor 594	Invitrogen	Cat# A21207, RRID: AB_141637
Donkey anti-goat Alexa Fluor 594	Invitrogen	Cat# A11058, RRID: AB_142540
Donkey anti-rabbit Alexa 488	Molecular Probes	Cat# A21206, RRID: AB_141708
Donkey anti-rat 594	Molecular Probes	Cat# A21209 RRID: AB_2535795
Rabbit anti-rat IgG	Vector Laboratories	Cat# AI-4001 RRID: AB_2336209
<b>Used datasets</b>	<b>SOURCE</b>	<b>IDENTIFIER</b>
Transcription profiling by high throughput sequencing of BV+ primordial germ cells from e7.5 embryos and e7.5 somatic neighbors	Magnusdottir et al. 2013 (Ref 1)	E-MTAB-1178 [ <a href="https://www.ebi.ac.uk/arrayexpress/experiments/E-MTAB-1178/">https://www.ebi.ac.uk/arrayexpress/experiments/E-MTAB-1178/</a> ]
Expression profiling by high throughput sequencing of LS/OB (e7.5) and E/MB (e7.75) BV+ primordial germ cells from Blimp1 KO and BV+ wild type control embryos as well as BV+ e11.5 primordial germ cells from Stella.Cre driven Blimp1 cKO embryonic gonads and BV+ wild type controls	Mitani et al. 2017 (Ref 2)	GSE91040 [ <a href="https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE91040">https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE91040</a> ] (original data therein derived from GSE11128 and GSE74094)
Blimp1-EGFP ChIP-seq in murine d2 and d6 PGCLCs	Mitani et al. 2017 (Ref 2)	GSE91041 [ <a href="https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE91041">https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE91041</a> ]

**Supplementary table 4: qRT-PCR primers used in this study**

Gene Name	Forward Primer Sequence	Reverse Primer Sequence	Annealing temperature	Product
<i>Prdm1</i>	GGCTCCACTACCCTTATCCTG	TCCTTTTGGAGGGATTGGAGTC	55°C	171bp
<i>Tfap2c</i>	GCCGGACGCCATGTTGTGGA	ACCCCGGTGTGCGAGAGAGG	55°C	162bp
<i>Hoxb1</i>	AGCCTACGACCTCCTCTCTG	GTGGTGAAGTTTGTGCGGAG	55°C	171bp
<i>Gapdh</i>	TGCACCACCAACTGCTTAGC	GGCATGGACTGTGGTCATGAG	55°C	87bp

## References

1. Magnusdottir, E. *et al.* A tripartite transcription factor network regulates primordial germ cell specification in mice. *Nat Cell Biol* **15**, 905-915, doi:10.1038/ncb2798 (2013).
2. Mitani, T. *et al.* Principles for the regulation of multiple developmental pathways by a versatile transcriptional factor, BLIMP1. *Nucleic Acids Res* **45**, 12152-12169, doi:10.1093/nar/gkx798 (2017).
3. Ohinata, Y., Sano, M., Shigeta, M., Yamanaka, K. & Saitou, M. A comprehensive, non-invasive visualization of primordial germ cell development in mice by the Prdm1-mVenus and Dppa3-ECFP double transgenic reporter. *Reproduction* **136**, 503-514, doi:10.1530/REP-08-0053 (2008).
4. Vincent, S. D., Dunn, N. R., Hayashi, S., Norris, D. P. & Robertson, E. J. Cell fate decisions within the mouse organizer are governed by graded Nodal signals. *Genes Dev* **17**, 1646-1662, doi:10.1101/gad.1100503 (2003).
5. Nowotschin, S. *et al.* The T-box transcription factor Eomesodermin is essential for AVE induction in the mouse embryo. *Genes Dev* **27**, 997-1002, doi:10.1101/gad.215152.113 (2013).
6. Arnold, S. J., Hofmann, U. K., Bikoff, E. K. & Robertson, E. J. Pivotal roles for eomesodermin during axis formation, epithelium-to-mesenchyme transition and endoderm specification in the mouse. *Development* **135**, 501-511, doi:10.1242/dev.014357 (2008).
7. Tremblay, K. D., Dunn, N. R. & Robertson, E. J. Mouse embryos lacking Smad1 signals display defects in extra-embryonic tissues and germ cell formation. *Development* **128**, 3609-3621 (2001).
8. Li, C., Li, Y. P., Fu, X. Y. & Deng, C. X. Anterior visceral endoderm SMAD4 signaling specifies anterior embryonic patterning and head induction in mice. *Int J Biol Sci* **6**, 569-583 (2010).
9. Chu, G. C., Dunn, N. R., Anderson, D. C., Oxburgh, L. & Robertson, E. J. Differential requirements for Smad4 in TGFbeta-dependent patterning of the early mouse embryo. *Development* **131**, 3501-3512, doi:10.1242/dev.01248 (2004).
10. Vincent, S. D. *et al.* The zinc finger transcriptional repressor Blimp1/Prdm1 is dispensable for early axis formation but is required for specification of primordial germ cells in the mouse. *Development* **132**, 1315-1325, doi:10.1242/dev.01711 (2005).
11. Conlon, F. L., Barth, K. S. & Robertson, E. J. A novel retrovirally induced embryonic lethal mutation in the mouse: assessment of the developmental fate of embryonic stem cells homozygous for the 413.d proviral integration. *Development* **111**, 969-981 (1991).
12. Hayashi, S., Lewis, P., Pevny, L. & McMahon, A. P. Efficient gene modulation in mouse epiblast using a Sox2Cre transgenic mouse strain. *Mech Dev* **119 Suppl 1**, S97-S101 (2002).
13. Kwon, G. S. & Hadjantonakis, A. K. Transthyretin mouse transgenes direct RFP expression or Cre-mediated recombination throughout the visceral endoderm. *Genesis* **47**, 447-455, doi:10.1002/dvg.20522 (2009).