

SUPPLEMENTAL MATERIAL

Data S1.

Supplemental Methods

Chemical analysis

Blood and bile samples

All blood samples were collected in fasting state. C-reactive protein (CRP), creatine kinase (CK), ApoE and safety assessment analyses (e.g. AST and ALT) in addition to biliary total bile acids were performed by certified routine assays at the Karolinska University Laboratory, Stockholm, Sweden.

For lipoprotein profiling the total and free cholesterol, and TG content in the different lipoprotein fractions corresponding to VLDL, LDL and HDL were determined after separation by size-exclusion chromatography¹. Detailed analysis of lipoprotein subclasses as well as ApoA1 and ApoB were performed with Nuclear Magnetic Resonance spectroscopy (NMR) at Nightingale Health Ltd (formerly known as Brainshake) Laboratory, Vantaa, Finland².

ApoCIII was analyzed using an enzyme-linked immunosorbent assay (Abcam, Cambridge, USA) following the manufacturer's instructions. Plasma and biliary lathosterol and campesterol were determined by isotope dilution-mass spectrometry after addition of deuterium labeled internal standards^{3, 4}. The campesterol/cholesterol and the lathosterol/cholesterol ratios were calculated as biomarkers for intestinal cholesterol absorption and total body cholesterol synthesis, respectively⁵⁻⁸. The intermediate in bile acid

synthesis 7 α -hydroxy-4-cholesten-3-one (C4) was analyzed in plasma samples by LC-MS/MS as described⁹.

Biliary lipids were extracted using the Bligh-Dyer method¹⁰. Cholesterol and phospholipid concentrations were determined by enzymatic methods with commercially available kits (Roche Diagnostics, Mannheim, Germany). Cholesterol saturation index (CSI) was calculated according to the critical tables described by Carey¹¹. The individual bile acids were analyzed using GC-MS as previously described¹². Briefly, bile was diluted with water, ethanol, KOH and hydrolyzed overnight after addition of deuterium labeled bile acids as internal standards. After acidification with HCl to pH 1 the deconjugated bile acids were extracted with diethyl ether and trimethylsilylated prior injection into GC-MS (Agilent Technologies).

PG binding

In this study we used a solid phase binding procedure that has been found reproducible for evaluation of affinity of isolated, or in plasma, apoB-lipoproteins for human arterial proteoglycans or purified proteoglycans preparations^{13, 14}. Human aortic PG were isolated from the intima-media of human aortas essentially by the method of Hurt-Camejo *et al*^{15 16}. Glycosaminoglycans were quantified by the method of Bartold and Page¹⁷, and the amounts of PG are expressed in terms of their glycosaminoglycan content. The wells of polystyrene 96-well plates (Thermo Fisher Scientific) were coated with 100 μ L of the PGs (25 μ g/mL in PBS) by incubation at 4°C overnight. Wells were blocked with 1% BSA in PBS for 1 h at 37°C. Wells without PG served as controls. One microliter of plasma was added to the wells in a buffer containing 140 mM NaCl, 2 mM MgCl₂, 5 mM CaCl₂ and 10 mM MES, pH 5.5 and incubated for 1 h at 37°C. The wells were washed with 10 mM MES-50 mM NaCl, pH 5.5, and the amount of bound cholesterol was determined using the Amplex Red cholesterol kit (Molecular Probes).

RNA extraction, cDNA synthesis and real-time RT-PCR

Total RNA was extracted and transcribed into cDNA using Omniscript (Qiagen, Sollentuna, Sweden). The mRNA expression levels were then quantified and using specific primers (Table 3), arbitrary units were calculated by linearization of the CT values and normalized to glyceraldehyde-3-phosphate dehydrogenase (*GAPDH*).

Table S1. Lipid contribution in lipoprotein fractions used in Figure 2

Data show mean \pm SEM. Plasma lipoproteins were subjected to analysis by nuclear magnetic resonance.

LDL: low density lipoprotein, HDL: high density lipoprotein, Remnant: non-HDL and non-LDL, P: placebo; S: simvastatin, E: ezetimibe, S+E: simvastatin plus ezetimibe.

Pre-	Cholesterol				Cholesteryl ester				Triglycerides			
	P	S	E	S+E	P	S	E	S+E	P	S	E	S+E
Remnant (mmol/L)	1.4 \pm 0.2	1.5 \pm 0.2	1.2 \pm 0.1	1.4 \pm 0.1	0.9 \pm 0.1	1.0 \pm 0.1	0.8 \pm 0.1	0.9 \pm 0.1	0.9 \pm 0.1	1.2 \pm 0.1	0.9 \pm 0.1	0.9 \pm 0.1
LDL (mmol/L)	1.7 \pm 0.2	1.8 \pm 0.1	1.5 \pm 0.1	1.6 \pm 0.1	1.2 \pm 0.1	1.3 \pm 0.1	1.1 \pm 0.1	1.2 \pm 0.1	0.17 \pm 0	0.24 \pm 0	0.17 \pm 0	0.18 \pm 0.0
HDL (mmol/L)	1.6 \pm 0.1	1.4 \pm 0.1	1.4 \pm 0.1	1.3 \pm 0.1	1.2 \pm 0.1	1.1 \pm 0.0	1.1 \pm 0.1	1.0 \pm 0.0	0.14 \pm 0	0.16 \pm 0	0.13 \pm 0	0.14 \pm 0
Post-												
Remnant (mmol/L)	1.4 \pm 0.1	0.8 \pm 0.1	1.1 \pm 0.1	0.5 \pm 0.1	0.9 \pm 0.1	0.5 \pm 0.1	0.7 \pm 0.1	0.3 \pm 0.0	1.0 \pm 0.1	0.7 \pm 0.1	0.8 \pm 0.1	0.5 \pm 0.1
LDL (mmol/L)	1.7 \pm 0.2	0.9 \pm 0.1	1.4 \pm 0.1	0.6 \pm 0.1	1.3 \pm 0.1	0.6 \pm 0.1	1.0 \pm 0.1	0.3 \pm 0.1	0.18 \pm 0	0.14 \pm 0	0.15 \pm 0	0.10 \pm 0.0
HDL (mmol/L)	1.6 \pm 0.1	1.4 \pm 0.1	1.4 \pm 0.1	1.4 \pm 0.1	1.3 \pm 0.1	1.1 \pm 0.1	1.1 \pm 0.1	1.1 \pm 0.0	0.15 \pm 0.0	0.12 \pm 0.0	0.12 \pm 0.0	0.10 \pm 0.0

Table S2. Contribution of different subclasses for their respective fraction baseline.

	Percentage contribution of remnant subclasses (Non-LDL; Non-HDL) to overall remnant fraction						
	XXL-VLDL	XL-VLDL	L-VLDL	M-VLDL	S-VLDL	XS-VLDL	IDL
Particle number	0.1%	0.3%	2.0%	8.0%	14%	20%	55%
Triglyceride	2.0%	3.0%	14%	30%	25%	13%	14%
Cholesteryl ester	0.3%	0.6%	3.0%	9.0%	14%	18%	55%
	Percentage contribution of LDL subclasses to overall LDL fraction						
	L-LDL		M-LDL		S-LDL		
Particle number	36%		29%		34%		
Triglyceride	56%		27%		17%		
Cholesteryl ester	52%		30%		18%		
	Percentage contribution of HDL subclasses to overall HDL fraction						
	XL-HDL	L-HDL	M-HDL		S-HDL		
Particle number	5%	13%	30%		58%		
Triglyceride	10%	22%	32%		36%		
Cholesteryl ester	12%	24%	32%		32%		

Baseline contribution of lipoprotein subclasses for their respective fraction.

XXL-VLDL, extra extra large very low density lipoproteins; XL-VLDL, extra large-VLDL; L-VLDL, large-VLDL; M-VLDL, medium-VLDL; S-VLDL, small-VLDL; XS-VLDL, extra small-VLDL; IDL, intermediate density lipoprotein; L-LDL, large low density lipoprotein; M-LDL, medium-LDL; S-LDL, small-LDL; XL-HDL, extra-large high density lipoprotein; L-HDL, large-HDL; M-HDL, medium-HDL; S-HDL, small-HDL.

Table S3. Correlation of percentage changes of proteoglycan (PG) binding and lipoprotein particle number and composition.

	Percentage change correlation of PG binding with remnant (Non-LDL; Non-HDL) lipoprotein subclasses						
	XXL-VLDL	XL-VLDL	L-VLDL	M-VLDL	S-VLDL	XS-VLDL	IDL
Particle number (r ² ; p value)	0.52; <0.001	0.58; <0.001	0.42; <0.001	0.45; <0.001	0.51; <0.001	0.55; <0.001	0.60; <0.001
Triglyceride (r ² ; p value)	0.51; <0.001	0.55; <0.001	0.38; <0.001	0.39; <0.001	0.42; <0.001	0.43; <0.001	0.34; <0.001
Cholesteryl ester (r ² ; p value)	0.62; <0.001	0.61; <0.001	0.23; <0.01	0.59; <0.001	0.65; <0.001	0.59; <0.001	0.54; <0.001
	Percentage change correlation of PG binding with LDL lipoprotein subclasses						
	L-LDL		M-LDL		S-LDL		
Particle number (r ² ; p value)	0.49; <0.001		0.49; <0.001		0.50; <0.001		
Triglyceride (r ² ; p value)	0.32; <0.001		0.35; <0.001		0.48; <0.001		
Cholesteryl ester (r ² ; p value)	0.52; <0.001		0.49; <0.001		0.46; <0.001		
	Percentage change correlation of PG binding with HDL lipoprotein subclasses						
	XL-HDL	L-HDL		M-HDL	S-HDL		
Particle number (r ² ; p value)	0.06; NS	0.06; NS [#]		0.01; NS	0.17; <0.05 [#]		
Triglyceride (r ² ; p value)	0.33; <0.001	0.13; <0.05		0.40; <0.05	0.15; <0.05		
Cholesteryl ester (r ² ; p value)	0.06; NS	0.11; <0.05 [#]		0.04; NS [#]	0.16; <0.05		

Pearson correlation between percentage changes of PG binding and lipoprotein subclasses particle number and composition. [#] Inverse correlation

XXL-VLDL, extra extra large very low density lipoproteins; XL-VLDL, extra large-VLDL; L-VLDL, large-VLDL; M-VLDL, medium-VLDL; S-VLDL, small-VLDL; XS-VLDL, extra small-VLDL; IDL, intermediate density lipoprotein; L-LDL, large low density lipoprotein; M-LDL, medium-LDL; S-LDL, small-LDL; XL-HDL, extra-large high density lipoprotein; L-HDL, large-HDL; M-HDL, medium-HDL; S-HDL, small-HDL

Table S4. Safety assessment analyses.

	Placebo	Simvastatin	Ezetimibe	Simvastatin + Ezetimibe	P value
Alanine aminotransferase (ALT); % variation from baseline	1.68±7.26	-2.81±11.90	-6.43±11.2	21.74±16.0	NS
Gamma-glutamyltransferase (GT); % variation from baseline	5.46±13.7	-21.0±11.2	0.84±8.38	8.88±19.7	NS
Creatine Phosphokinase (CK), % variation from baseline	17.5± 14.7	22.5± 14.8	27.6± 17.5	-0.7± 12.6	NS

Data show mean ± SEM. Multi-Way ANOVA followed by LSD-test. NS: not significant.

Supplemental References:

1. Parini P, Johansson L, Broijersen A, Angelin B, Rudling M. Lipoprotein profiles in plasma and interstitial fluid analyzed with an automated gel-filtration system. *European journal of clinical investigation*. 2006;36:98-104.
2. Soininen P, Kangas AJ, Wurtz P, Suna T, Ala-Korpela M. Quantitative serum nuclear magnetic resonance metabolomics in cardiovascular epidemiology and genetics. *Circulation Cardiovascular genetics*. 2015;8:192-206.
3. Lund E, Sisfontes L, Reihner E, Bjorkhem I. Determination of serum levels of unesterified lathosterol by isotope dilution-mass spectrometry. *Scandinavian journal of clinical and laboratory investigation*. 1989;49:165-171.
4. Lutjohann D, Bjorkhem I, Beil UF, von Bergmann K. Sterol absorption and sterol balance in phytosterolemia evaluated by deuterium-labeled sterols: Effect of sitostanol treatment. *J Lipid Res*. 1995;36:1763-1773.
5. Bjorkhem I, Miettinen T, Reihner E, Ewerth S, Angelin B, Einarsson K. Correlation between serum levels of some cholesterol precursors and activity of hmg-coa reductase in human liver. *J Lipid Res*. 1987;28:1137-1143.
6. Kempen HJ, Glatz JF, Gevers Leuven JA, van der Voort HA, Katan MB. Serum lathosterol concentration is an indicator of whole-body cholesterol synthesis in humans. *J Lipid Res*. 1988;29:1149-1155.
7. Miettinen TA, Tilvis RS, Kesaniemi YA. Serum plant sterols and cholesterol precursors reflect cholesterol absorption and synthesis in volunteers of a randomly selected male population. *American journal of epidemiology*. 1990;131:20-31.
8. Stellaard F, von Bergmann K, Sudhop T, Lutjohann D. The value of surrogate markers to monitor cholesterol absorption, synthesis and bioconversion to bile acids under lipid lowering therapies. *The Journal of steroid biochemistry and molecular biology*. 2017;169:111-122.
9. Lovgren-Sandblom A, Heverin M, Larsson H, Lundstrom E, Wahren J, Diczfalusy U, Bjorkhem I. Novel lc-ms/ms method for assay of 7alpha-hydroxy-4-cholesten-3-one in human plasma. Evidence for a significant extrahepatic metabolism. *Journal of chromatography. B, Analytical technologies in the biomedical and life sciences*. 2007;856:15-19.
10. Bligh EG, Dyer WJ. A rapid method of total lipid extraction and purification. *Canadian journal of biochemistry and physiology*. 1959;37:911-917.
11. Carey MC. Critical tables for calculating the cholesterol saturation of native bile. *J Lipid Res*. 1978;19:945-955.
12. Bjorkhem I, Falk O. Assay of the major bile acids in serum by isotope dilution-mass spectrometry. *Scandinavian journal of clinical and laboratory investigation*. 1983;43:163-170.
13. Skalen K, Gustafsson M, Rydberg EK, Hulten LM, Wiklund O, Innerarity TL, Boren J. Subendothelial retention of atherogenic lipoproteins in early atherosclerosis. *Nature*. 2002;417:750-754.
14. Bancells C, Benitez S, Jauhiainen M, Ordonez-Llanos J, Kovanen PT, Villegas S, Sanchez-Quesada JL, Oorni K. High binding affinity of electronegative ldl to human aortic proteoglycans depends on its aggregation level. *J Lipid Res*. 2009;50:446-455.
15. Hurt-Camejo E, Camejo G, Rosengren B, Lopez F, Wiklund O, Bondjers G. Differential uptake of proteoglycan-selected subfractions of low density lipoprotein by human macrophages. *J Lipid Res*. 1990;31:1387-1398.
16. Oorni K, Pentikainen MO, Annala A, Kovanen PT. Oxidation of low density lipoprotein particles decreases their ability to bind to human aortic proteoglycans. Dependence on oxidative modification of the lysine residues. *The Journal of biological chemistry*. 1997;272:21303-21311.

17. Bartold PM, Page RC. A microdetermination method for assaying glycosaminoglycans and proteoglycans. *Analytical biochemistry*. 1985;150:320-324.