

# Supplementary materials

## Background information on study area and species

South Africa is located at the oceanographic interface between the Atlantic and Indian Oceans, and the number of species recorded particularly along its southern coastline is greater than that of either the south-eastern Atlantic or the south-western Indian Ocean [1]. While this can be partly explained by the fact that the fauna of the two oceans overlaps in South Africa, there is also evidence for large numbers of endemic species [1,2]. Many of these are genetically subdivided and may comprise complexes of evolutionary lineages that represent cryptic or incipient species [3], suggesting that the region's true biodiversity may be much greater than is presently acknowledged. The boundaries between the ranges of many of these lineages coincide with one of the region's four temperature-defined marine provinces, namely the cool-temperate west coast, warm-temperate south coast, subtropical east coast and tropical north-east coast [3], and some species even have ranges that are limited to the transition zones between these provinces [4,5]. However, there are also numerous species in which the range of a particular evolutionary lineage spans multiple marine provinces (Fig. 1). In some species, the amount of gene flow between provinces can be considerable when conditions are favourable, but the migrants are unable to establish themselves in the long term [6,7]. Physiological studies indicate that for some species, sister lineages present in adjacent provinces exhibit distinct temperature preferences [8,9], which suggests that thermal adaptation contributes towards limiting gene flow between biogeographic regions by reducing migrant fitness and, if a conspecific sister lineage is present, by subjecting the migrants to competitive exclusion [3]. The Knysna sandgoby, *Psammogobius knysnaensis* (species 13 in Fig. 1), is one of the most common estuarine fishes in South Africa [10,11]. It has sedentary adults, so connectivity among populations is likely facilitated by its planktonic

27 larvae, which are transported out of estuaries in large numbers during ebb-tides [12] and use  
28 nearshore areas as nurseries [13]. Although the species' planktonic larval duration is not  
29 known, it is likely similar to that of other gobies, which is in the order of weeks to months  
30 [14,15]. Compared to the other candidate species with similarly wide distributions, the purity  
31 of its extracted DNA, which was extracted using the CTAB protocol [16], was much higher.  
32 Based on DNA sequence data from the mitochondrial COI gene and the nuclear S72 intron,  
33 the species exists as a single lineage whose range spans the west coast, the cool-  
34 temperate/warm-temperate transition zone on the south-west coast, the south coast and the  
35 warm-temperate/subtropical transition zone on the southern east coast [17]. Although genetic  
36 structure was identified between the west coast (site 1 in Fig. 1) and all other regions on the  
37 basis of mtDNA, this is not a result of divergent evolution (Fig. 3b), but of lower genetic  
38 diversity on the cool-temperate west coast [17].

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## 40 Supplementary methods

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### 42 Processing of raw sequences

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44 Raw sequences were processed using the `process_radtags.pl` script in STACKS 1.19 [18,19].

45 Briefly, we eliminated reads with ambiguous barcode and/or restriction site, allowing a

46 maximum of two mismatches, and all remaining reads were trimmed to 80 bp by removing

47 the barcode, the restriction site, and the last 8 bp. The dDocent 1.0 software pipeline [20] was

48 then used to remove low quality bases using a quality score limit of 30. Putative RAD reads

49 were identified *de novo* using a minimum depth of 15×, and the selected unique reads were

50 then aligned with a maximum of 8 mismatches to form reference contigs. The reference

51 contigs were then clustered based on an 80% similarity threshold. Single nucleotide

52 polymorphisms (SNPs) were called from the aligned good quality reads of multiple

53 individuals, using a Bayesian-based variant detection approach [21]. After excluding

54 individuals with >20% of missing data, the final data set comprised 109 individuals. Variants  
55 were filtered to produce a file containing only the best quality biallelic SNPs per locus, which  
56 were present in at least 80% of the individuals sequenced, with a minimum allele frequency  
57 equal to or greater than 3% (please see Table S4 for additional detail).

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### 59 **Identification of loci under thermal selection and neutral loci**

60 The contribution of coastal sea surface temperature (SST) to the overall pattern of genetic  
61 differentiation was assessed using a spatially explicit generalized linear mixed model to test  
62 for direct associations between SNP allele frequencies and temperature-related variables,  
63 while controlling for the effects of spatial structure and shared population history,  
64 implemented in the R package gINLAnd [22]. Unlike  $F_{ST}$ -based outlier scans, which identify  
65 loci on the basis of population information [23], the identification of loci in in genotype-  
66 environment association methods such as gINLAnd is thus not influenced by any regional  
67 population structure [24]. Briefly, gINLAnd estimates the covariance associated with the  
68 spatial distribution of the samples and a locus-specific effect of each environment variable; it  
69 then estimates the likelihood of two competing models: a model with the environmental  
70 effect and a reduced model without the environmental effect. Finally, gINLAnd assesses the  
71 strength of genetic dependence on the environmental variable by computing a Bayes factor  
72 between the two models. To avoid false positives, we used a conservative approach in which  
73 only those loci which showed a log Bayes factor (BF)  $\geq 10$  were considered to be under  
74 selection (a log BF > 4.6 is considered decisive [25]). A plot depicting loci under selection is  
75 shown in Fig. S1. We calculated a multidimensional scaling projection of the coastal distance  
76 between sampling sites using the R package MASS 7.3 [26], which is more meaningful than  
77 using the original geographic coordinates because this would have required connecting sites  
78 via terrestrial habitat.

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80 To identify selectively neutral loci, the following approach was used. Thermal selection may  
81 be only one of a number of drivers of selection, so we used BayeScan v. 2.1 [23] to identify  
82 markers under selection on the basis of outlier scans rather than temperature data. This  
83 method was used because it has a low error rate compared to other tests for the detection of  
84 outlier loci [27]. Default settings were applied with prior odds set to 10, but a very high false  
85 discovery rate of 20% was applied to create a neutral data set with a low probability of  
86 containing any remaining loci under selection. We then excluded 304 outlier loci, together  
87 with 27 additional loci identified by gINLAnd that were not found by BayeScan, to create a  
88 data set of 8201 selectively neutral loci.

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## 91 **Temperature data**

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93 Temperature data originated from sites that are located in close proximity to the estuaries or  
94 lagoons in which samples of *P. knysnaensis* were collected [28] (Table S1). While coastal  
95 SST strongly influences thermal conditions within estuaries that are connected to the sea,  
96 many southern African estuaries become temporarily disconnected from the sea for  
97 prolonged periods of time, and their water temperatures tend to be higher [29]. Moreover,  
98 estuaries exhibit much more extensive variation in physico-chemical characteristics  
99 compared to subtidal nearshore areas, and their biota are considered to be particularly  
100 resilient to cope with this variability [30]. Adult fish in estuaries thus have very wide thermal  
101 tolerance ranges [31], and are unlikely to be under strong thermal selection. In contrast, the  
102 species' planktonic larvae, which emigrate from estuaries [13] and during their marine  
103 dispersal phase will be strongly affected by coastal SST, are likely to be more sensitive to  
104 water temperature because fish larvae have limited metabolic scope and thus limited

105 tolerance to environmental stressors [32]. This suggests that coastal rather than estuarine  
106 temperatures are particularly suitable to explore thermal selection in this species.

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108 Environmental variables run with the program gINLAnd were taken from Smit et al.[28] and  
109 included minimum and maximum temperatures, as well as the coolest and warmest 5% of  
110 temperature measurements (Table S3). We only report results for minimum and maximum  
111 temperatures as they showed the trends most clearly (Fig. S4) and differed only slightly from  
112 the other temperature readings (Table S1).

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## 114 **Analyses of genetic structure**

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### 116 a) Discriminant Analysis of Principal Components (DAPC)

117 DAPC analyses were performed with the R package ADEGENET v. 2.1.0 [33]. DAPC defines a  
118 model with synthetic variables in which the genetic variation is maximized between clusters  
119 of individuals ( $K$ ), and minimized within clusters. We used  $k$ -means clustering and the  
120 Bayesian Information Criterion (BIC) to identify the best-supported number of clusters. We  
121 explored various combinations of maximum or minimum temperature as the environmental  
122 variable with covariance factors that included geographic distance, biogeographic boundaries  
123 and a combination of the two. A simplistic resistance matrix approach [34] was used, where  
124 geographic distance and biogeographic boundaries between pairs of sites were given a  
125 resistance value (one unit of resistance per km between sites and ten units of resistance if  
126 sites were located in different marine bioregions). We then calculated multidimensional  
127 scaling projections based on the resistance pairwise matrices, and these were used as  
128 covariance factor to control for spatial structure. We further explored the effect of using only  
129 SNP data from the coolest and the warmest marine bioregions, using geographic distance as  
130 the controlling factor (see Table S3 for detail on number of SNPs identified).

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132       b) *fastStructure* analyses

133   The program *fastStructure* uses variational Bayesian inference under a model assuming  
134   Hardy-Weinberg equilibrium and linkage equilibrium, and was primarily used here to  
135   confirm specific results based on DAPC. We used a simple prior and set all other parameters  
136   to the default value, except for the convergence criterion, which was lowered to  $10^{-8}$ . The  
137   programme was run for each value of  $K = 1-9$  independently, and each value was cross-  
138   validated 1000 times. The python script *chooseK* was used to identify an optimal range of  $K$   
139   values, and the resulting barplots were visualised with the R package *distruct2.2* [35].

140       c) BEAST analyses

141   The dataset used for discrete phylogeographic analyses in BEAST comprised data from the  
142   individual alleles of each individual, which were reconstructed in PHASE v. 2.1.1 [36] using  
143   default settings. When more than one pair of haplotypes was possible for an individual, the  
144   one with the highest probability was used. In addition to reconstructing a phylogenetic tree,  
145   this method can infer the most likely bioregion in which each ancestral node in the MCC tree  
146   was present. One hundred million generations were specified, and trees saved every 100 000  
147   generations, and the first 20% of trees were discarded as burn-in. Model and prior settings  
148   followed those recommended in the tutorial available at <http://hpc.ilri.cgiar.org>.

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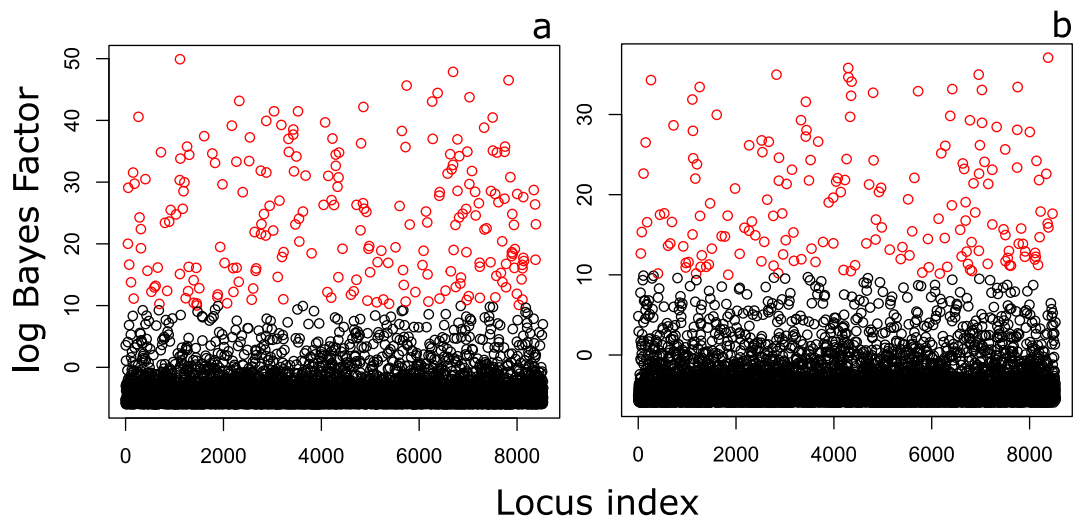
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151 **Supplementary results**

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153 A total of 405,648,596 raw reads were generated on two Illumina lanes. After demultiplexing  
154 and quality filtering, an average of 1,560,510 reads were obtained per individual, totalling  
155 224,713,440 reads. The filtered catalogue resulted in 8,532 ddRADseq loci containing 15,633  
156 SNPs. A final dataset was obtained by extracting only the SNPs with the best quality score  
157 from each polymorphic ddRADseq locus to remove SNPs that are likely in linkage  
158 disequilibrium. After removing individuals with more than 20% missing data, the final data  
159 set comprised 109 individuals genotyped for 8,532 SNPs.

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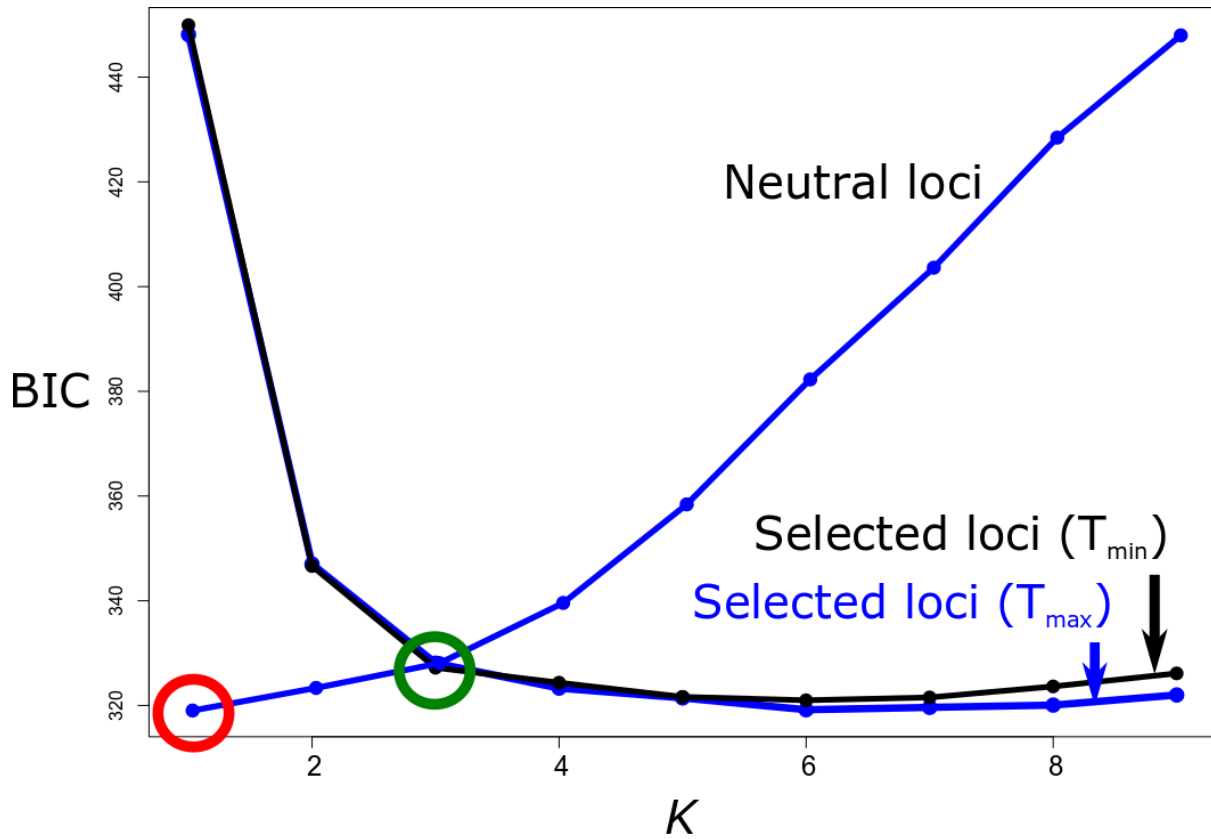
163 **Fig S1.** Examples of loci identified as being linked to genomic regions under  
164 thermal selection (red dots) while controlling for geographical distances between  
165 sites by gINLAnd analysis. Plots depict results for (a) minimum temperatures and (b)  
166 maximum temperatures. All log Bayes factors ( $\log BF$ )  $> 0$  indicate potential thermal  
167 selection, and the likelihood that these are false positives decreases with increasing  
168  $\log BF$ . In the loci shown in red ( $\log BF > 10$ ), the association between genomic and  
169 environmental variables is decisive.

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**Fig S2.** Analyses of candidate loci putatively under thermal selection and selectively neutral loci. The best value of  $K$  for selectively neutral data is based on the lowest Bayesian Information Criterion (BIC) value (red circle), whereas plots for temperature-associated loci (controlling for geographical distance) for minimum and maximum temperatures ( $T_{\min}$  and  $T_{\max}$ , respectively) have a distinct 'elbow' at  $K = 3$  beyond which further decrease in BIC is minimal (green circle). Some subsequent analyses were nonetheless conducted with 4 clusters to show differentiation between the pre-defined marine bioregions and to compare the results of DAPC analyses with those of the fastSTRUCTURE analyses.

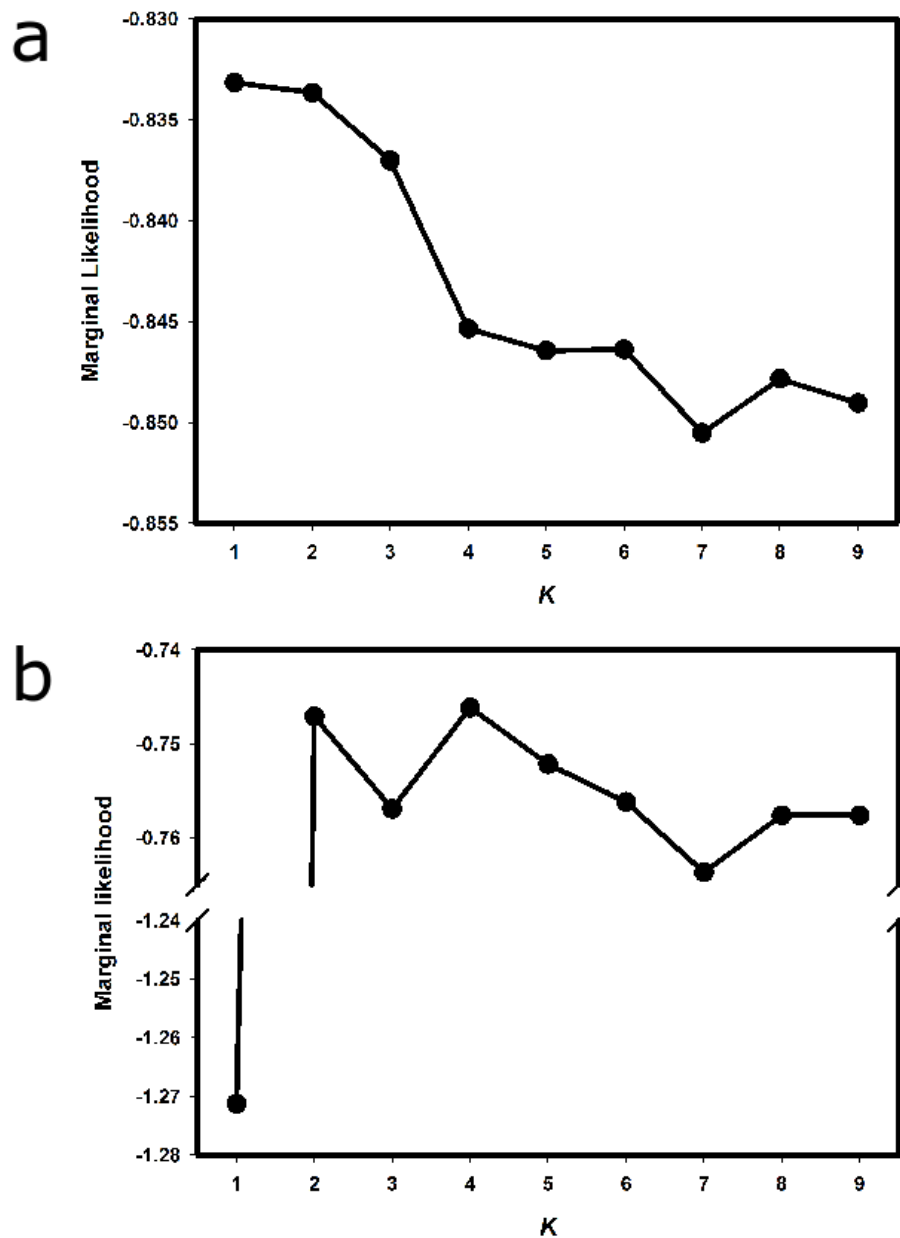
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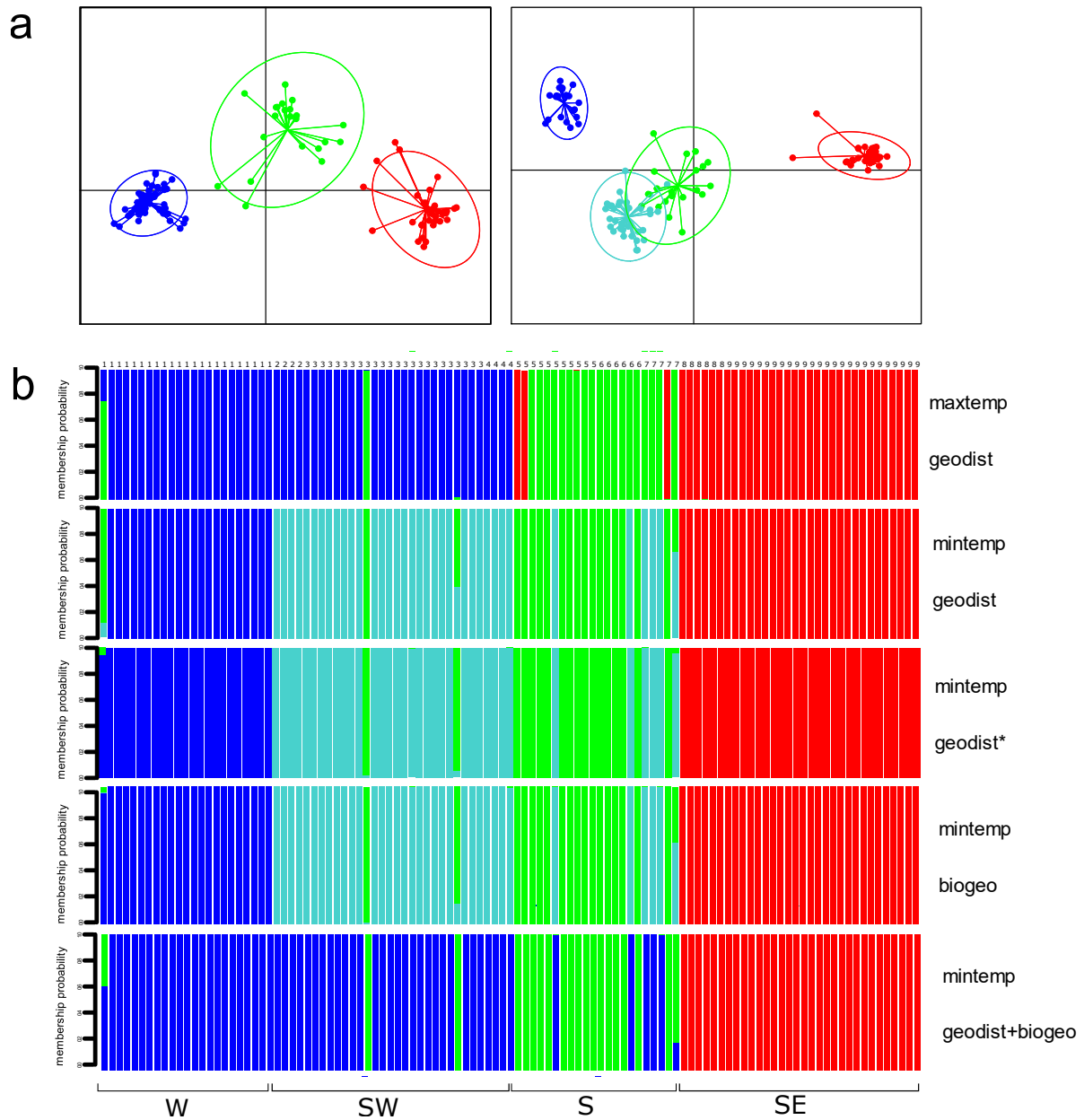


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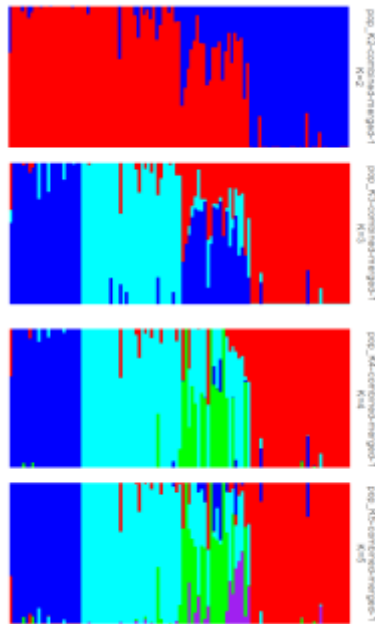
195 **Fig S3.** Support for number of clusters in fastStructure. Plots depict marginal  
196 likelihoods for number of genetic clusters ( $K$ ) for (a) neutral loci, highest likelihood at  
197  $K = 1$ ; (b) temperature-associated loci ( $T_{min}$ ), highest likelihood at  
198  $K = 4$ .

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**Fig S4.** Population genetic structure inferred for temperature-associated loci using DAPC. **a** Scatterplots with inertia ellipses representing 95% confidence intervals, with colours reflecting the dominant bioregion represented in a particular cluster (left: loci correlated with maximum temperature; right: loci correlated with minimum temperature, in both cases controlling for geographic distance); **b** DAPC compoplots indicating membership probabilities for each individual (vertical bars) within one of four genetic clusters; correlation with temperature and the controlling factor are indicated on the right (maxtemp = maximum temperature, mintemp = minimum temperature, geodist = geographic distance, biogeo = biogeography; \*indicates that only sites 1, 8 and 9 were used to find loci correlated with minimum temperature).



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215 **Fig S5.** Comparison of consensus fastStructure barplots for  $K=2-5$  based on  
216 temperature-associated loci ( $T_{\min}$ ).

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Table S1. Sampling sites, collection dates, number of high quality samples used for the analyses, and temperature data from nearby locations.

Region	Site no.	Site name	Date	GPS coordinates	N	Temperature location*	min	5%	95%	max
W	1a	Langebaan (Kraalbaai)	Feb 2016	33°08'31.32"S 18°01'26.26"E	12	Langebaan	11.9	12.1	19.6	20.2
W	1b	Langebaan (Shark Bay)	Dec 2016	33°06'42.42"S 18°02'24.47"E	11	Langebaan	11.9	12.1	19.6	20.2
SW	2	Rooiels	Oct 2015	34°17'53.48"S 18°49'11.10"E	4	Betty's Bay	12.9	13.0	18.2	18.6
SW	3	Palmiet	Dec 2016	34°20'36.33"S 18°59'39.57"E	23	Betty's Bay	12.9	13.0	18.2	18.6
SW	4	Klein	Dec 2016	34°25'08.06"S 19°18'23.78"E	4	Hermanus	12.2	13.6	19.1	19.9
S	5	Breede	Dec 2016	34°23'47.19"S 20°50'32.45"E	11	Stilbaai	12.8	13.0	23.5	23.9
S	6	Klein Brak	Oct 2015	34°05'29.82"S 22°08'45.12"E	6	Mossel Bay	12.9	13.7	23.4	23.8
S	7	Swartkops	Oct 2015	33°52'09.06"S 25°37'43.13"E	6	Humewood	13.7	14.4	22.3	22.5
SE	8	Nenga	Oct 2015	31°59'02.81"S 29°09'03.83"E	6	Cwebe	16.9	17.0	21.6	21.7
SE	9a	Bulolo	Oct 2015	31°39'02.05"S 29°30'59.14"E	6	Mzamba	18.3	18.6	24.0	24.1
SE	9b	Bulolo	May 2016	31°39'02.05"S 29°30'59.14"E	20	Mzamba	18.3	18.6	24.0	24.1

\*Smit et al. 2013; min = minimum temperature reading; 5% = coldest 5% of temperature readings; 95% = warmest 5% of temperature readings; max = maximum temperature reading.

Table S2. Genes identified among temperature-associated loci whose activity may be directly related to temperature stress. Loci marked with an asterisk were found for  $T_{\min}$  only, those without asterisk for both  $T_{\min}$  and  $T_{\max}$ .

<b>Locus</b>	<b>Uniprot</b>	<b>Gene</b>	<b>Temperature related activity</b>
Contig_69368	A0A1S3GQV0_DIPOR	Histone H3	Biotinylation of Histone H3 is related resistance to heat stress in flies[37]. Related with control of expression of other proteins.
Contig_56520	F1QC71_DANRE	ANO5b	Part of a family of proteins (Anoctamin) that are key mediators of thermal sensing, most do not act as temperature sensors themselves but rather support the behavior in some other capacity, such as by controlling the development or function of the thermo-sensory cell or circuitry[38].
Contig_22921*	A0A1A8EID4_9TELE	SHC4	Moderate heat stress has been shown to induce maximal gene activity to promote tissue repair[39]. The gene was also indicative of adaptation to different thermal environments in <i>Anolis</i> lizards[40].
Contig_247440	G3TCM9_LOXAF	MBD2	Regulates the expression of other genes during heat stress in sea cucumbers[41].
Contig_168847	A0A147A7G3_FUNHE	14-3-3 protein gamma	Part of a protein family that has been associated with heat shock responses in gobies[42]. Increased expression in cows exposed to high temperatures, possibly to protect against liver cell apoptosis[43]. Differential expression in corals in response to a thermal challenge[44].

Contig_38433	A0A0F8AG31_LARCR	EH28_02053 (Tyrosine-protein kinase)	Interacts with Hsp90[45] and its activity increases during heat stress in a goby species[42]. Altered gene expression in response to heat stress was also recorded in the Antarctic fish <i>Trematomus bernacchii</i> [46].
Contig_93683	A0A1A8MJC6_9TELE	PRSS12	Expressed under heat stress condition in sea cucumbers[41].
Contig_330391	A0A146TQG3_FUNHE	Reverse transcriptase-like protein	Upregulated as early response to heat shock in plants[47].
Contig_351821*	S7PGQ6_MYOBR	Tubulin beta chain	Divergence on tubulin genes has been associated to adaptation to cold temperatures in Antarctic fishes[48] and protozoans[49]. It also changes expression levels during thermal acclimation in bluefin tuna[50] and is differentially expressed in response to heat shock in crustaceans[51].

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Table S3. Number of loci identified for gINLAnd analyses.

Temperature	Controlling factor	No. loci
Maximum	Geographic distance	175
Minimum	Geographic distance	226
Minimum	Geographic distance*	196
Minimum	Biogeography	239
Minimum	Geographic distance + Biogeography	169

\*Only individuals from the W coast (1) and SE coast (8, 9) were included.

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Table S4. Summary of loci obtained following raw data processing.

Region	Loci present with less than 20% missing data	Variable loci	Percentage of variable loci	$H_O$	$H_E$	Genotyped samples
W	8532	4122	48.3	0.2834	0.2703	23
SW	8532	4206	49.3	0.2849	0.2710	31
S	8532	4288	50.3	0.2592	0.2855	23
SE	8532	4251	49.8	0.2659	0.2628	32

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