Stem Cell Reports, Volume 12

Supplemental Information

Modeling G2019S-LRRK2 Sporadic Parkinson's Disease in 3D Midbrain

Organoids

Hongwon Kim, Hyeok Ju Park, Hwan Choi, Yujung Chang, Hanseul Park, Jaein Shin, Junyeop Kim, Christopher J. Lengner, Yong Kyu Lee, and Jongpil Kim



Figure S1. Related to Figure 1. (A) Schematic of organizing midbrain 3D organoids from hiPSCs. Timeline of various differentiation protocols for generating efficacious and optimal 3D organoids. (B) Representative immunofluorescence images of the stages of 3D organoid generation for 2 months. Scale bar = 100 µm. (C) The percentage of TUJ1-, TH-, VMAT2-, DAT- and GIRK2-positive midbrain dopaminergic neurons in midbrain 3D organoids at day 60. (D) Western blot analysis of AADC, PITX3, VMAT2 and NURR1 in immunoprecipitates of midbrain 3D organoids. (E) Representative images for mature dopaminergic neuron marker, PITX3 and AADC at 30 and 60 days. (F) Fontana-masson staining showing neuromelanin positive cells in midbrain 3D organoids at 30 and 60 days. Scale bar=50 µm. (G) Quantification of the number of neuromelanin-positive cells in midbrain 3D organoids. Data represent the mean ± SEM. ANOVA, *p<0.05, **p<0.01 (n=3 per sample). (H) Age-associated genes ANLN, GAL35ST1, FAH, MBP, ACSL1 and CLDN11 were increased in midbrain 3D organoids at 60 days. (I) Gene set enrichment analysis of the microarray data showing the considerable enrichment of human aged brain compared to fetal brain (GSM2639572 and GSM175904, http://www.brain-map.org). (J) Immunostaining of phosphorylation of histone H2AX (yH2AX) in midbrain 3D organoids at day 30 and 60. Scale bar =50 µm. (K) The percentage of gamma-H2AX/DAPI-positive cells in midbrain 3D organoids. Data represent the mean ± SEM. ANOVA, *p<0.05, **p<0.01 (n=3 per sample). (L and M) Western blot analysis shows an increase in gamma-H2AX levels at day 60. Data represent the mean ± SEM. ANOVA, *p<0.05, **p<0.01. (N) Representative immunofluorescence images of MAP2- and TH-positive dopaminergic neurons in 3D cultures and 2D cultures. Scale bar=100 µm. (O) Quantification of the number of TH- and MAP2-positive cells among all DAPI-positive cells in 3D and 2D cultures at 20 days. Data represent the mean ± SEM. ANOVA, *p<0.05, **p<0.01 (n=3 per sample). (P) Real-time qPCR analysis of the expression of the neuronal markers AADC, CHAT and MAPT and the astrocyte markers GFAP, S100B and ALDH1L1 at 20 days. Data represent the mean ± SEM. ANOVA, *p<0.05, **p<0.01 (n=3 per sample). (Q) Scatter plot of the microarray expression data comparing midbrain 3D organoids and 2D cultures. (R) Gene ontology analysis for biological processes in midbrain 3D organoids and 2D cultures. (S) Gene set enrichment analysis of the microarray data from midbrain 3D organoids compared to 2D cultures showing the considerable enrichment of human brain tissues, cerebral cortex and striatum.



Figure S2. Related to Figure 2. (A) The sequence of Cas9 targeting a locus within the human *LRRK2* gene. The single-guide RNA targeting site is marked in blue, and the 5'-NGG protospacer adjacent motif site is labeled in red. The arrow indicates the LRRK2-G2019S mutation site. (B) SURVEYOR mutation detection assay gel showing modification of the Cas9 targeting site by the single-guide RNA. (C) Representative sequences of indels in the human *LRRK2* gene targeted by CRISPR-Cas9. Deletions are indicated with a dash. (D) Schematic representation of the donor vector including the LRRK2-G2019S mutation site. Enlarged sequence represents CRISPR-Cas9induced targeting site and genetically engineered genomic locus in exon 41 of LRRK2 locus. (E) Sanger sequencing showing the G2019S mutation site in exon 41 of the *LRRK2* gene after the introduction of the Cas9 *LRRK2*-targeting and donor plasmids. (F) Representative images of PCR products using puromycin-specific primers to amplify puromycin-resistant hiPSC colonies. A CRE-transduced hiPSC colony was confirmed by the appropriate primers. (G) Representative immunofluorescence images of colonies positive for the pluripotency markers SOX2, NANOG, TRA1-60, OCT4, and SSEA1 among healthy and LRRK2-G2019S-targeted hiPSCs. Passage matched control and targeted iPSC lines was analyzed in the experiments. Scale bar=100 µm. (H) Quantitative RT-PCR analysis of healthy hiPSCs and LRRK2-G2019S hiPSCs for the pluripotency markers OCT4, SOX2, NANOG, and REX1. Data represent the mean ± SEM. ANOVA (n=3 per sample). (I) AP staining of healthy hiPSCs and LRRK2-G2019S hiPSCs. Scale bar=100 µm. (J) Quantification of the number of alkaline phosphatase-positive clones among healthy hiPSCs and LRRK2-G2019S hiPSCs. (K) Bright-field images of control and LRRK2-G2019S 3D organoids. Scale bar=1 mm. (L) Measurement of the size of control and LRRK2-G2019S 3D organoids at 45 days. (M) Real-time qPCR analysis of pluripotency marker (OCT4 and NANOG) expression in control and LRRK2-G2019S 3D organoids at 30 days. Data represent the mean ± SEM. ANOVA, *p<0.05, **p<0.01 (n=3 per sample). (N) Relative quantification of the dopaminergic neuronal markers TH, AADC, and DAT in control and LRRK2-G2019S 3D organoids using gRT-PCR. (O) Quantitative RT-PCR analysis of control and LRRK2-G2019S 3D organoids for neuronal markers (VMAT2, Synapsin1, and GAP43) and an astrocyte marker (S100B) at 60 days. Data represent the mean \pm SEM. ANOVA, *p<0.05, **p<0.01 (n=3 per sample).



Figure S3. Related to Figure 2 and Figure 3. (A) Relative quantification of the mature neuronal marker NURR1, PITX3, EN1, TH and MAPT in control and LRRK2-G2019S 3D organoids compared to control and LRRK2-G2019S 2D cultures. Data represent the mean ± SEM. ANOVA (n=3 per sample). (B) Measurement of the neurite length of control and LRRK2-G2019S expressing dopaminergic neurons in 3D organoids from each 3 individuals. Data represent mean ± SEM (n=5 per sample). (C) Fluorescenceactivated cell sorting analysis of acridine orange/propidium idodide-positive cells from control and LRRK2-G2019S mutant 3D organoids. (D) Representative images showing colocalization analysis of EEA1-positive endosomes with pS129-α-synucleinpositive puncta. Scale bar=20 μm. (E) The percentage of pS129-α-synuclein- and EEA1-positive puncta among all EEA1-positive puncta from each 2 individuals. Data represent the mean ± SEM. ANOVA, *p<0.05, **p<0.01 (n=6 per sample). (F) Representative images showing colocalization analysis of LC3B-positive autophagy with pS129-α-synuclein-positive puncta and PARKIN-positive mitophagy. Scale bar=20 μm. (G) The percentage of pS129-α-synuclein- and PARKIN-positive puncta among all LC3B-positive puncta. Data represent the mean ± SEM. ANOVA, *p<0.05, **p<0.01 (n=4 per sample). (H) The percentage of Thioflavin T-positive cells from 2 individuals. Data represent the mean \pm SEM. ANOVA, **p<0.01 (n=5 per sample). (I) Relative intensity of phosphorylated a-synuclein oligomer levels in LRRK2-G2019S 3D organoids with and without the LRRK2 kinase inhibitor. Data represent the mean ± SEM. ANOVA, *p<0.05, **p<0.01 (n=3 per sample). (J) Relative gene expression of TH, AADC and DAT in LRRK2-G2019S 3D organoids with and without the LRRK2 kinase inhibitor. Data represent the mean ± SEM. ANOVA, *p<0.05, **p<0.01 (n=3 per sample). (K) The percentage of cleaved caspase3-positive cells from each 3 individuals. Data represent the mean ± SEM. ANOVA, **p<0.01 (n=5 per sample).



Figure S4. Related to Figure 4 and Figure 5. (A) The color of boxplots shows specific group color in Fig. 4A and yellow dotted line indicates average median of boxplots. (B) Gene set enrichment analysis of the microarray expression data from LRRK2-G2019S knock-in 2D cultures compared to wild-type 2D cultures. (C) Graph of the TXNIP protein interaction network in LRRK2-G2019S 3D organoids. Red circles indicate high expression, and blue circles represents low expression. (D and E) Western blots of P-ERK, P-P38 and TXNIP protein was determined in control and LRRK2-G2019S mutant 3D organoids from each 3 individuals. Expression of phosphorylated ERK and P38 was increased in midbrain-like 3D organoids including LRRK2-G2019S mutation. Data represent the mean ± SEM. ANOVA, *p<0.05, **p<0.01 (n=3 per sample). (F) Immunostaining of NRF2-positive cells in control and LRRK2-G2019S mutant 3D organoids. Scale bar=20 µm. (G) Chromatin immunoprecipitation analysis using NRF2 antibody (1:100, Cell signaling) in control and LRRK2-G2019S mutant 3D organoids from each 3 individuals. Immunoprecipitated DNA fragments was performed by realtime PCR using the human TXNIP promoter primers. Relative fold expression was calculated normalizing data to input. (H and I) Western blots of Thioredoxin1 protein was determined in control and LRRK2-G2019S mutant 3D organoids. (J) Thioredoxin1 activity was measured using Thioredoxin1 assay kit (Enzo life science) in control and LRRK2-G2019S mutant 3D organoids from each 3 individuals. Data represent the mean ± SEM. ANOVA, *p<0.05, **p<0.01 (n=4 per sample).



Figure S5. Related to Figure 5. (A) Immunofluorescence analysis of TXNIP protein was decreased in sh*TXNIP* treated LRRK2-G2019S mutant 3D organoids from 3 individuals. (B and C) Western blots of TXNIP protein was determined in control and sh*TXNIP* treated LRRK2-G2019S mutant 3D organoids. Data represent the mean \pm SEM. *ANOVA*, *p<0.05, **p<0.01 (n=3 per sample). (D) Relative intensity of phosphorylated α -synuclein oligomer levels in LRRK2-G2019S 3D organoids with and without *TXNIP*-shRNA. Data represent the mean \pm SEM. *ANOVA*, *p<0.05, **p<0.01 (n=3 per sample). (E) Quantification of LAMP1+ and pS129- α -synuclein+ puncta showing a reduction of pS129- α -synuclein inclusions from each 2 individuals. Data represent the mean \pm SEM. *ANOVA*, *p<0.05 (n=5 per sample).