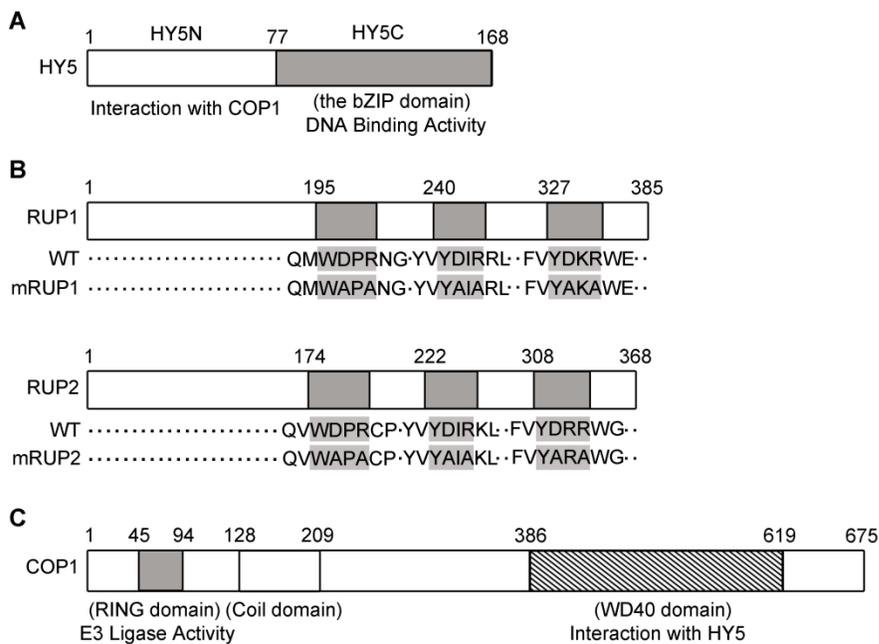


## Two E3 Ligases Antagonistically Regulate the UV-B Response in Arabidopsis

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### Supporting Information

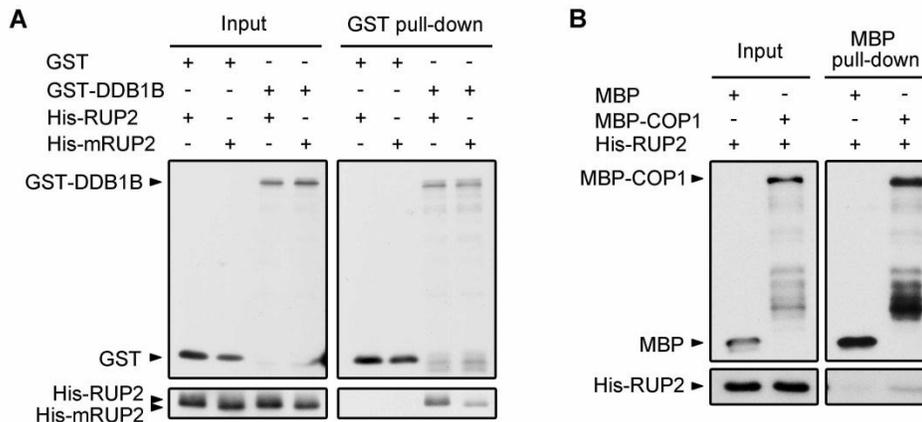


**Fig. S1. Schematic Diagrams of HY5, RUP1, RUP2 and COP1 Proteins.**

(A) Schematic diagram of the functional domains of HY5 protein. HY5N, the N-terminal domain of HY5. HY5C, the C-terminal domain of HY5. Numbers indicate the positions of amino acids.

(B) Schematic diagrams of RUP1 and RUP2 proteins. WT, RUP1/RUP2 with wild-type DWD motifs. mRUP1/mRUP2, RUP1/RUP2 with mutated DWD motifs. Numbers indicate the positions of amino acids.

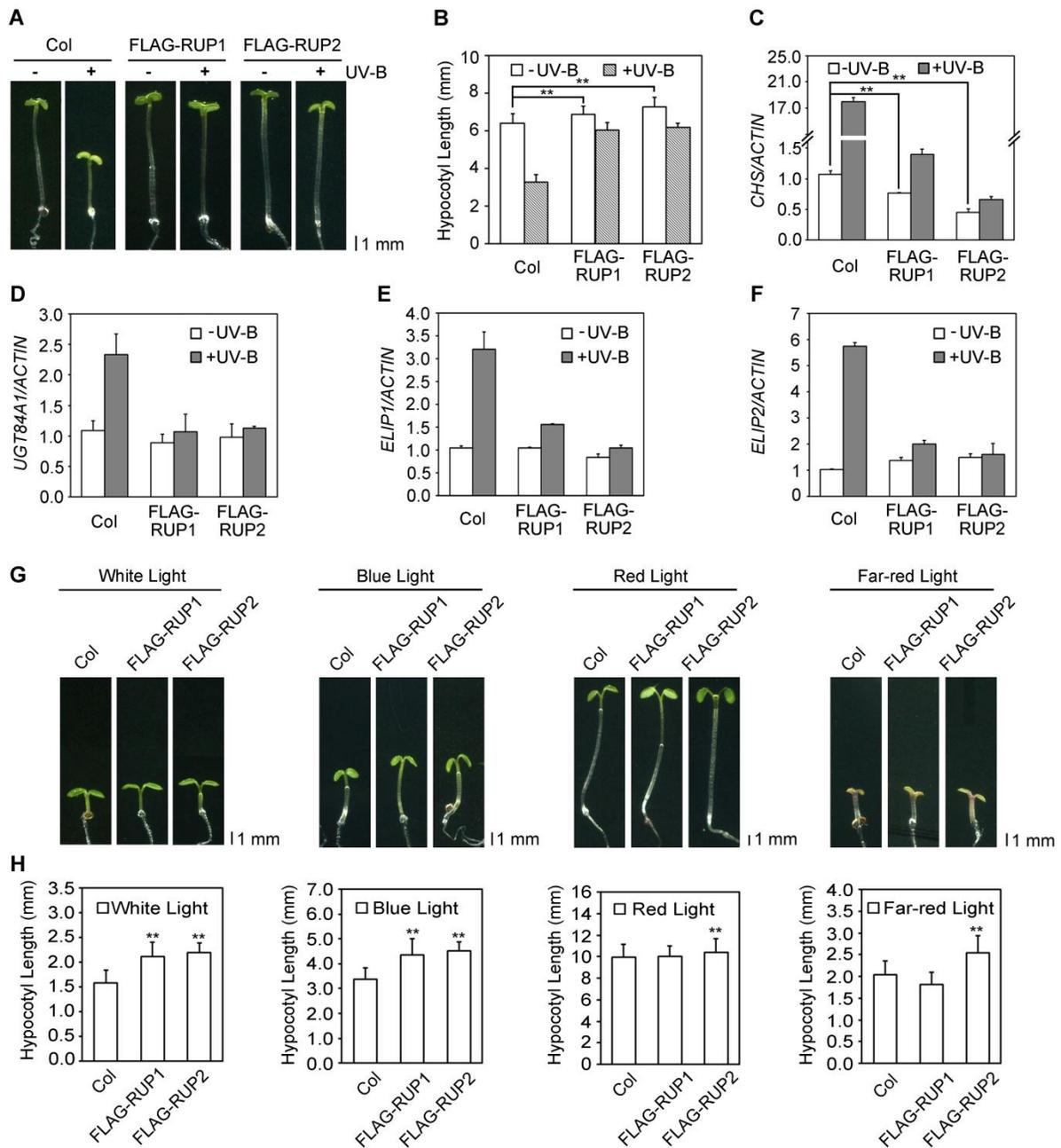
(C) Schematic diagram of the functional domains of COP1 protein. Numbers indicate the positions of amino acids.



**Fig. S2. In Vitro Pull-down Assays of GST-DDB1B/MBP-COP1 and His-RUP2.**

(A) DDB1B interacts with RUP2 in vitro. The purified GST or GST-DDB1B was incubated with His-RUP2 or His-mRUP2 before being pulled down by Glutathione Sepharose. His-RUP2 and His-mRUP2 were detected by anti-RUP2 antibodies.

(B) COP1 interacts with RUP2 in vitro. The purified MBP or MBP-COP1 was incubated with His-RUP2 before being pulled down by Amylose Resin. His-RUP2 was detected by anti-RUP2 antibodies.



**Fig. S3. FLAG-RUP1 and FLAG-RUP2 Exhibit Hyposensitivity to Photomorphogenic UV-**

**B.**

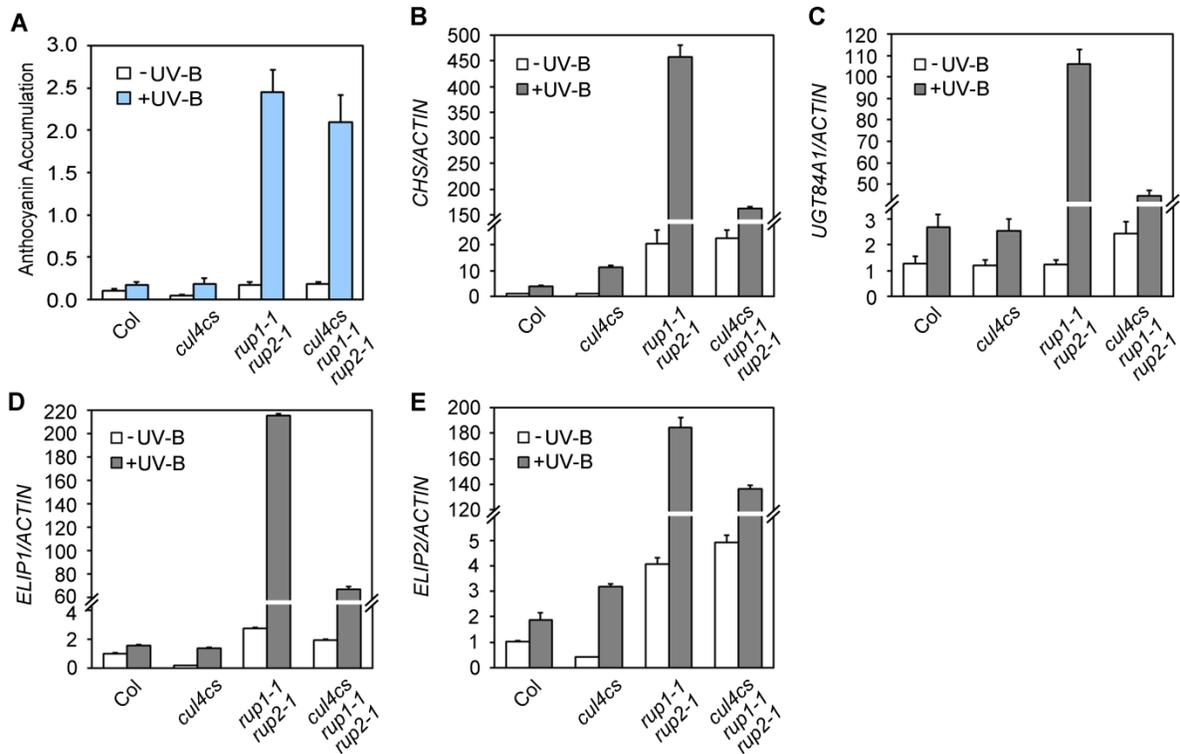
(A) Phenotypes of 4-day-old Col, FLAG-RUP1, and FLAG-RUP2 seedlings grown under -UV-B and +UV-B.

(B) Hypocotyl length of the seedlings shown in (A). Mean  $\pm$  SD,  $n \geq 30$ . The asterisks indicate significant differences by Student's t test (\*\* $p < 0.01$ ).

(C-F) *CHS* (C), *UGT84A1* (D), *ELIP1* (E), and *ELIP2* (F) mRNA levels in the seedlings shown in (A). Mean  $\pm$  SD,  $n = 3$ . The asterisks indicate significant differences by Student's t test (\*\* $p < 0.01$ ).

(G) Phenotypes of 4-day-old Col, FLAG-RUP1 and FLAG-RUP2 seedlings grown under white light, blue light, red light and far-red light conditions.

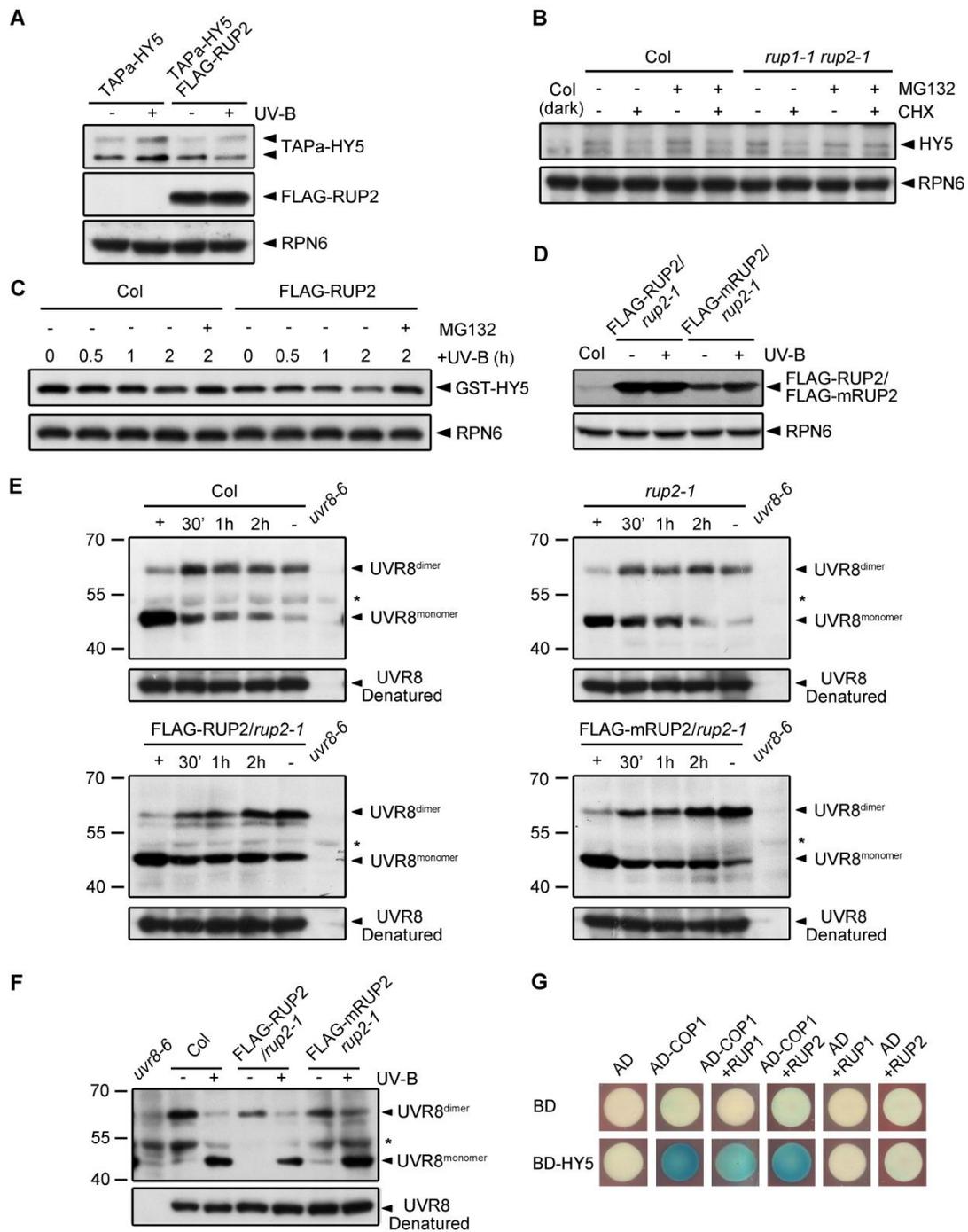
(H) Hypocotyl length of the seedlings shown in (G). Mean  $\pm$  SD,  $n \geq 30$ . The asterisks indicate significant differences by Student's t test (\*\* $p < 0.01$ ) compared to Col under each light condition.



**Fig. S4. CUL4 and RUP1/RUP2 Genetically Interact to Influence UV-B-induced Anthocyanin Accumulation and Gene Expression.**

(A) Anthocyanin content of 4-day-old Col, *cul4cs*, *rup1-1 rup2-1*, and *cul4cs rup1-1 rup2-1* seedlings grown under -UV-B and +UV-B. Mean  $\pm$ SD, n=3.

(B-E) *CHS* (B), *UGT84A1* (C), *ELIP1* (D), and *ELIP2* (E) mRNA levels in the seedlings shown in (A). Mean  $\pm$ SD, n=3.



**Fig. S5. The Effects of RUP1/RUP2 on HY5 Stability, UVR8 Conformation and the COP1-HY5 Interaction.**

(A) TAPa-HY5 protein levels in 4-day-old TAPa-HY5 and TAPa-HY5 FLAG-RUP2 seedlings grown under –UV-B and +UV-B. Proteins were analyzed by immunoblotting with anti-c-Myc, anti-FLAG, and anti-RPN6 antibodies. RPN6 was used as a loading control.

(B) Immunoblot analysis of HY5 proteins in 4-day-old Col and *rup1-1 rup2-1* seedlings grown under –UV-B and treated with 500 μM CHX and/or 50 μM MG132 for 3 hours. HY5 was detected with anti-HY5 antibodies. RPN6 was used as a loading and negative control.

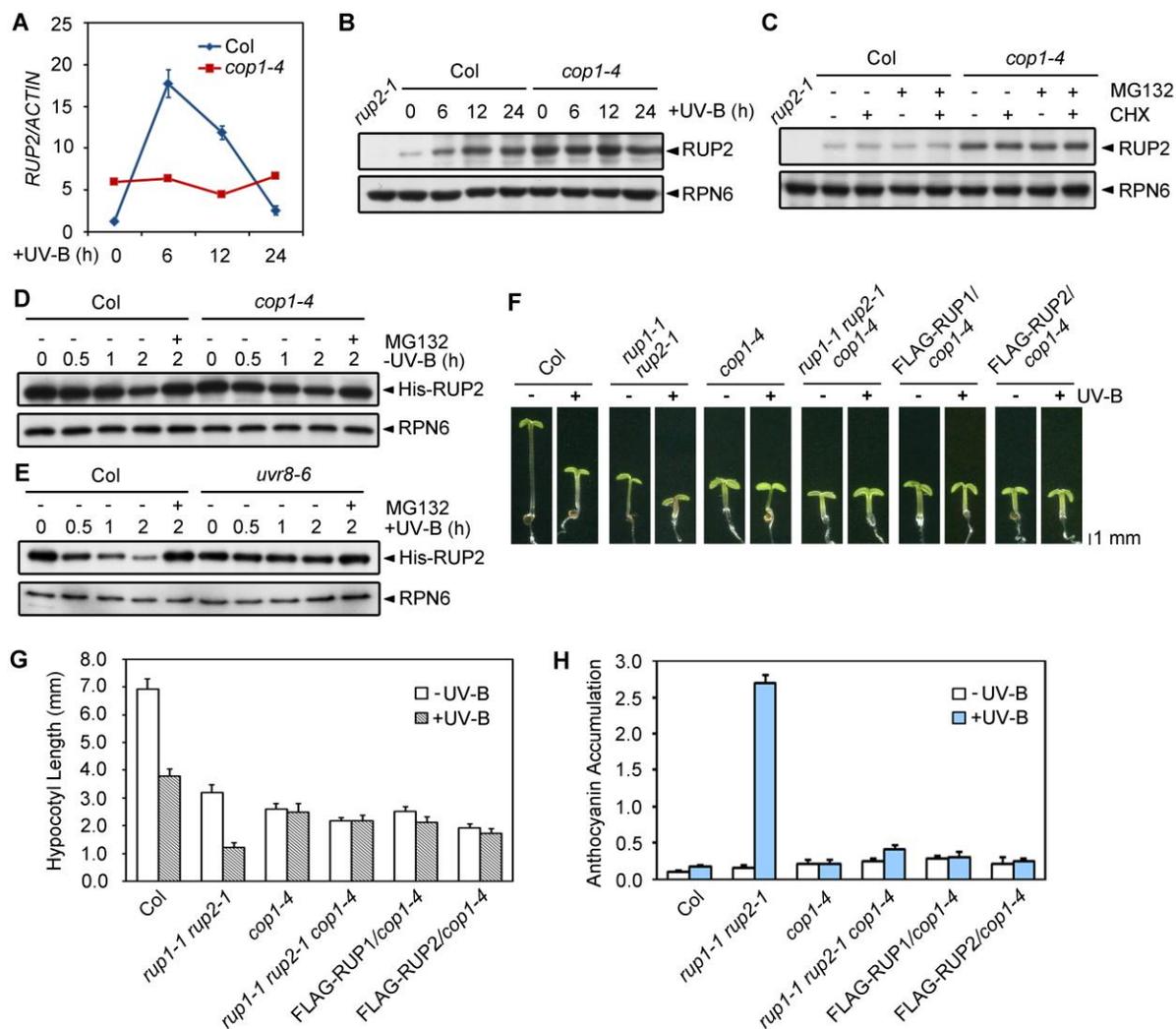
(C) The effect of FLAG-RUP2 on HY5 stability in vitro, as analyzed by cell-free degradation assays. Purified GST-HY5 was incubated with total proteins extracted from 4-day-old UV-B-grown Col and FLAG-RUP2 seedlings for 2 hours. The degradation mixture was treated with or without 50 μM MG132. GST-HY5 was detected with anti-GST antibody. RPN6 was used as a loading and negative control.

(D) FLAG-RUP2/FLAG-mRUP2 protein levels in 4-day-old FLAG-RUP2/*rup2-1* and FLAG-mRUP2/*rup2-1* seedlings grown under –UV-B and +UV-B. Proteins were analyzed by immunoblotting with anti-FLAG and anti-RPN6 antibodies. RPN6 was used as a loading control.

(E) Immunoblot analysis of UVR8 conformation in 4-day-old Col, *rup2-1*, FLAG-RUP2/*rup2-1* and FLAG-mRUP2/*rup2-1* seedlings grown under +UV-B and then transferred to –UV-B for 2 hours or grown under continuous –UV-B. Denatured UVR8 was used as a loading control. UVR8 was detected with anti-UVR8 antibodies. The asterisks indicate nonspecific bands.

(F) Immunoblot analysis of UVR8 conformation in 4-day-old Col, FLAG-RUP2/*rup2-1* and FLAG-mRUP2/*rup2-1* seedlings grown under continuous –UV-B and +UV-B. Denatured UVR8 was used as a loading control. UVR8 was detected with anti-UVR8 antibodies. The asterisks indicate nonspecific bands.

(G) The effect of RUP1/RUP2 on the COP1-HY5 interaction in yeast. Co-transformed yeast cells were selected and then transferred to induction medium supplemented with X- $\beta$ -gal for blue color development.



**Fig. S6. The Effects of COP1 on the Expression, Stability, and Function of RUP1/RUP2.**

(A) The effect of *cop1-4* mutation on the UV-B-induced expression of *RUP2* in *Arabidopsis*. Four-day-old -UV-B-grown Col and *cop1-4* seedlings were transferred to +UV-B for the indicated time periods. Mean  $\pm$ SD, n=3.

(B) *RUP2* protein levels in 4-day-old Col and *cop1-4* seedlings grown under -UV-B and transferred to +UV-B for the indicated time periods. Proteins were analyzed by immunoblotting with anti-*RUP2* and anti-*RPN6* antibodies. *RPN6* was used as a loading control.

(C) Immunoblot analysis of RUP2 proteins in 4-day-old Col and *cop1-4* seedlings grown under –UV-B and treated with 500  $\mu$ M CHX and/or 50  $\mu$ M MG132 for 3 hours. RUP2 was detected with anti-RUP2 antibodies. RPN6 was used as a loading and negative control.

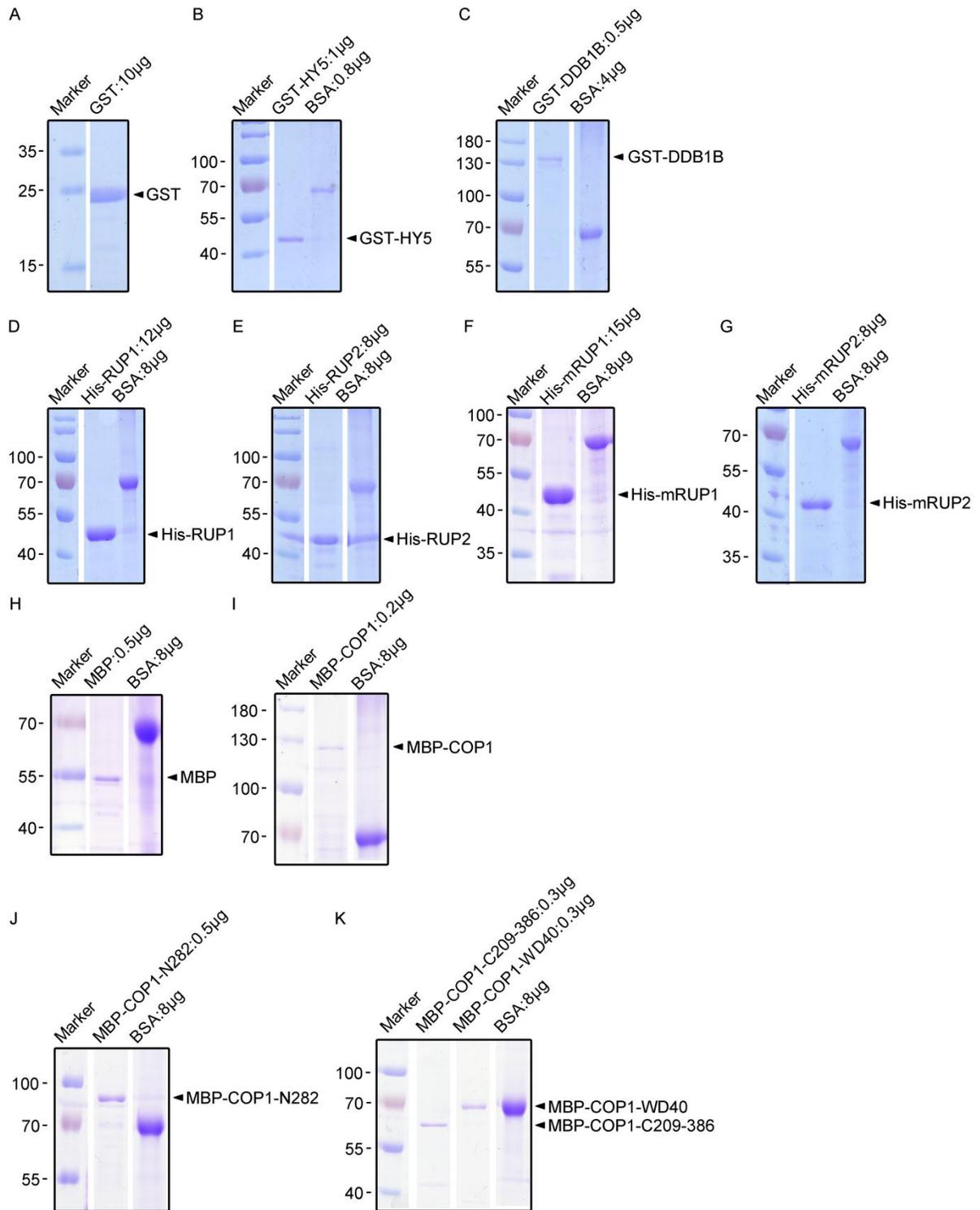
(D) The effect of COP1 on RUP2 stability in vitro under –UV-B, as analyzed by cell-free degradation assays. Purified His-RUP2 was incubated with total proteins extracted from 4-day-old Col and *cop1-4* seedlings grown under –UV-B for 2 hours. The degradation mixture was treated with or without 50  $\mu$ M MG132. His-RUP2 was detected with anti-RUP2 antibodies. RPN6 was used as a loading and negative control.

(E) The effect of UVR8 on RUP2 stability in vitro under +UV-B, as analyzed by cell-free degradation assays. Purified His-RUP2 was incubated with total proteins extracted from 4-day-old Col and *uvr8-6* seedlings grown under +UV-B for 2 hours. The degradation mixture was treated with or without 50  $\mu$ M MG132. His-RUP2 was detected with anti-RUP2 antibodies. RPN6 was used as a loading and negative control.

(F) Phenotypes of 4-day-old Col, *rup1-1 rup2-1*, *cop1-4*, *rup1-1 rup2-1 cop1-4*, FLAG-RUP1/*cop1-4*, and FLAG-RUP2/*cop1-4* seedlings grown under –UV-B and +UV-B.

(G) Hypocotyl length of the seedlings shown in (E). Mean  $\pm$  SD,  $n \geq 30$ .

(H) Anthocyanin content of the seedlings shown in (E). Mean  $\pm$  SD,  $n=3$ .



**Fig. S7. The Recombinant Proteins Used in This Study.**

**Table S1. Primers Used in This Study.**

<b>Gene name</b>	<b>Primers</b>
<b>Primers used for plant transformation</b>	
<i>RUP1-F</i>	ATAC ggtacc ATGGAGGCTT TGTTCTGCTC
<i>RUP1-R</i>	ATAC ctcgag TTAGCTTTGT TTGCCCGAGA
<i>RUP2-F</i>	ATAC gtcgac ATGAACACTC TTCATCCTCA
<i>RUP2-R</i>	ATAC gagctc CTATGGTTTT CTTTTGCCCA
<i>FLAG-mRUP2-F</i>	ATAC ctcgag CAGGATCCCCCGGGCTGCAG
<i>FLAG-mRUP2-R</i>	ATAC actagt CTATGGTTTTCTTTTGCCCA
<b>Primers used for yeast two-hybrid assays</b>	
<i>AD/BD-RUP1-F</i>	ATAC caattg ATGGAGGCTT TGTTCTGCTC
<i>AD/BD-RUP1-R</i>	ATAC ctcgag TTAGCTTTGT TTGCCCGAGA
<i>AD-RUP2-F</i>	ATAC caattg ATGAACACTC TTCATCCTCA
<i>AD-RUP2-R</i>	ATAC ctcgag CTATGGTTTT CTTTTGCCCA
<b>Primers used for <i>in vitro</i> pull-down assays</b>	
<i>His-RUP1-F</i>	ATAC caattg ATGGAGGCTT TGTTCTGCTC
<i>His-RUP1-R</i>	ATAC ctcgag TTAGCTTTGT TTGCCCGAGA
<i>His-RUP2-F</i>	ATAC caattg ATGAACACTC TTCATCCTCA
<i>His-RUP2-R</i>	ATAC ctcgag CTATGGTTTT CTTTTGCCCA
<i>MBP-COPI-F</i>	ATAC gtcgac ATGGAAGAGATTTTCGACGGA
<i>MBP-COPI-R</i>	ATAC ctcgag TCACGCAGCGAGTACCAGAA
<i>MBP-COPI-</i>	ATAC gaattc ATGGAAGAGATTTTCGACGGA

*N282-F*

*MBP-COPI-*

ATAC ctgcag ctcgag TCACGAATCTGACCCACTCAGCG

*N282-R*

*MBP-COPI-*

ATAC gaattc ATGGCTAGGGACAGATATTCTGTAAAGTT

*C209-386-F*

*MBP-COPI-*

ATAC ctgcag ctcgag TCAAAACAGCTCATCATCACGATCA

*C209-386-R*

*MBP-COPI-*

ATAC gaattc ATGGCCACTGCTGGTGTCTTAGAT

*WD40-F*

*MBP-COPI-*

ATAC ctgcag TCACGCAGCGAGTACCAGAA

*WD40-R*

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**Primers used for site-directed mutagenesis**

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*mRUP1 D196A*

GAACGGTTCAAATGTGGGCTCCAGCTAACGGAGGGACGTTAG

*R198A-F*

*mRUP1 D196A*

CTAACGTCCCTCCGTTAGCTGGAGCCCACATTTGAACCGTTC

*R198A-R*

*mRUP1 D241A*

CCGGAATGCCTACGTGTATGCCATCGCAAGACTAGTGGACCC

*R243A-F*

*mRUP1 D241A*

GGGTCCACTAGTCTTGCGATGGCATAACGTAGGCATTCCGG

*R243A-R*

*mRUP1 D328A*

CAAGTGTTTGTGTACGCTAAGGCGTGGGAGGAACCGGTTTGG

*R330A-F*

*mRUP1 D328A*

CCAAACCGGTTCCCTCCCACGCCTTAGCGTACACAAACACTTG

*R330A-R*

*mRUP2 D175A*  
GACTATGCAAGTATGGGCTCCGGCGTGTCCGCCGGAAGAATC

*R177A-F*

*mRUP2 D175A*  
GATTCTTCCGGCGGACACGCCGGAGCCCATACTTGCATAGTC

*R177A-R*

*mRUP2 D223A*  
CGGAAAGGGTACGTTTACGCTATAGCGAAACTCGTTGACCCG

*R225A-F*

*mRUP2 D223A*  
CGGGTCAACGAGTTTCGCTATAGCGTAAACGTACCCTTTCCG

*R225A-R*

*mRUP2 D309A*  
AGGGTGTTTGTGTACGCTAGGGCATGGGGGAAGCCGGTTTGG

*R311A-F*

*mRUP2 D309A*  
CCAAACCGGCTTCCCCCATGCCCTAGCGTACACAAACACCCT

*R311A-R*

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**Primers used for firefly luciferase complementation imaging**

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*RUP1-nLUC-F* ATAC ggtacc ATGGAGGCTTTGTTCTGCTC

*RUP1-nLUC-R* ATAC gtcgac GCTTTGT TTGCCCGAGA

*RUP2-nLUC-F* AGAC agatct ATGAACACTCTTCATCCTC

*RUP2-nLUC-R* ATAC gtcgac TGGTTTTCTTTTGCCC

*cLUC-RUP1-F* ATAC ggtacc ATGGAGGCTTTGTTCTGCTC

*cLUC-RUP1-R* ATAC gtcgac TTAGCTTTGT TTGCCCGAGA

*cLUC-RUP2-F* AGAC agatct GATGAACACTCTTCATCCTC

*cLUC-RUP2-R* ATAC gtcgac CTATGGTTTTCTTTTGCCC

*cLUC-COP1-F* ATAC ggtacc ATGGAAGAGA TTTCGACGGA

*cLUC-COPI-R* ATAC gtcgac TCACGCAGCGAGTACCAGAA  
*cLUC-COPI-N282-F* CGGTACCCGGGATCCAATGGAAGAGATTTTCGACGGA  
*cLUC-COPI-N282-R* GCTCTGCAGGTCGACTCACGAATCTGACCCACTCAGCG  
*cLUC-COPI-C209-386-F* CGGTACCCGGGATCCAATGGCTAGGGACAGATATTCTGTAAAG  
*cLUC-COPI-C209-386-R* GCTCTGCAGGTCGACTCAAACAGCTCATCATCACGATCA  
*cLUC-COPI-WD40-F* CGGTACCCGGGATCCAATGGCCACTGCTGGTGTTTCTAGAT  
*cLUC-COPI-WD40-R* GCTCTGCAGGTCGACTCACGCAGCGAGTACCAGAACT  
*cLUC-COPI- $\Delta$ RING-F1* CGGTACCCGGGATCCAATGGAAGAGATTTTCGACGGA  
*cLUC-COPI- $\Delta$ RING-R1* ACCGCCGCTTCCTCCGTCGTCA  
*cLUC-COPI- $\Delta$ RING-F2* GGAGGAAGCGGCGGTCTCGATAAGCTATTGAAGAAAACCTT  
*cLUC-COPI- $\Delta$ RING-R2* GCTCTGCAGGTCGACTCACGCAGCGAGTACCAGAACT  
*cLUC-COPI- $\Delta$ Coil-F1* CGGTACCCGGGATCCAATGGAAGAGATTTTCGACGGA

<i>cLUC-</i>	CCGAAACTGATCCAAGGGCGAT
<i>COPI<math>\Delta</math>Coil-R1</i>	
<i>cLUC-</i>	TTGGATCAGTTTCGGAAGTTGCGGATGCTCGGAGATGA
<i>COPI<math>\Delta</math>Coil-F2</i>	
<i>cLUC-</i>	GCTCTGCAGGTCGACTCACGCAGCGAGTACCAGAACT
<i>COPI<math>\Delta</math>Coil-R2</i>	
<i>cLUC-</i>	CGGTACCCGGGATCCAATGGAAGAGATTTTCGACGGA
<i>COPI<math>\Delta</math>R<math>\Delta</math>C-F1</i>	
<i>cLUC-</i>	CCGAAACTGATCCAAGGGCGAT
<i>COPI<math>\Delta</math>R<math>\Delta</math>C-R1</i>	
<i>cLUC-</i>	TTGGATCAGTTTCGGAAGTTGCGGATGCTCGGAGATGA
<i>COPI<math>\Delta</math>R<math>\Delta</math>C-F2</i>	
<i>cLUC-</i>	GCTCTGCAGGTCGACTCACGCAGCGAGTACCAGAACT
<i>COPI<math>\Delta</math>R<math>\Delta</math>C-R2</i>	

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**Primers used for ubiquitination assays in HEK293T cells**

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<i>FLAG-COP1-F</i>	AGAGAATTC ggtacc ATGGAAGAGATTTTCG
<i>FLAG-COP1-R</i>	CTTCCATGG ctcgag TCACGCAGCGAGTAC
<i>RUP2-HA-F</i>	AGAGAATTC ggatcc ATGAACACTC TTCAT
<i>RUP2-HA-R</i>	CTTCCATGG ctcgag TGGTTTTCTTTTGCC

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**Primers used for yeast three-hybrid assays**

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<i>RUP1 (Y3H)-F</i>	ATAC ggtacc ATGGAGGCTTTGTTCTGCTCT
<i>RUP1 (Y3H)-R</i>	ATAC gaattc TTAGCTTTGTTTGCCCGAGA
<i>RUP2 (Y3H)-F</i>	GTCGAATTGGGTACCATGAACACTCTTCATCCTCACAA

*RUP2 (Y3H)-R*            ACCCGGGTGGGAATTCCTATGGTTTTCTTTGCCCAC

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**Primers used for qRT-PCR**

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<i>Actin-F</i>	CAAGGCCGAGTATGATGAGG
<i>Actin-R</i>	GAAACGCAGACGTAAGTAAAAAC
<i>RUP2-F</i>	TCGGATGACGGGACT
<i>RUP2-R</i>	GACGCAACAAACAGCA
<i>CHS-F</i>	ACGTCACGTGTTGAGCGAGTATGG
<i>CHS-R</i>	GAGGAACGCTGTGCAAGACGACTG
<i>UGT84A1-F</i>	AGTCGGGTTTATCGTTCT
<i>UGT84A1-R</i>	ATCCCTTTACCTTTAGCAC
<i>ELIP1-F</i>	CGTTGCCGAAGTCACCAT
<i>ELIP1-R</i>	AATCCAACCATCGCTAAACG
<i>ELIP2-F</i>	CACCACAAATGCCACAGTCT
<i>ELIP2-R</i>	TGCTAGTCTCCCGTTGATCC

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