

**Cellular-Based Selections Aid Yeast-Display Discovery of Genuine Cell-Binding Ligands:
Targeting Oncology Vascular Biomarker CD276**

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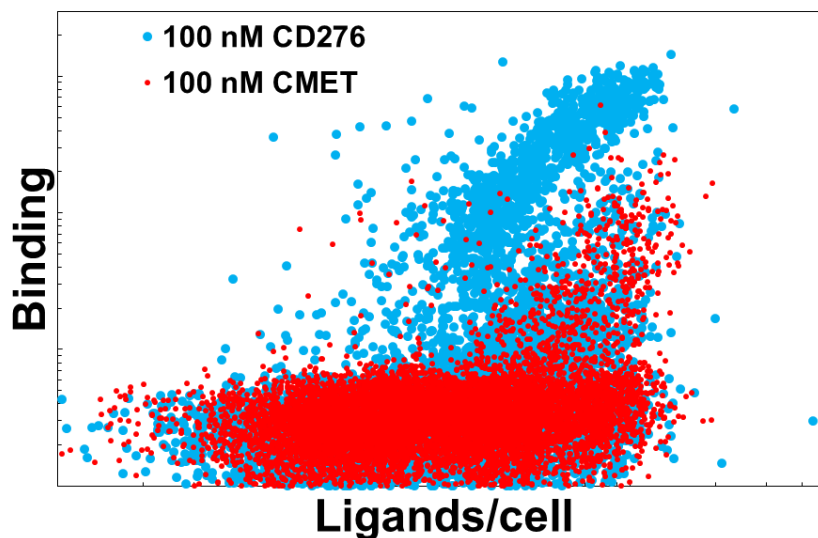
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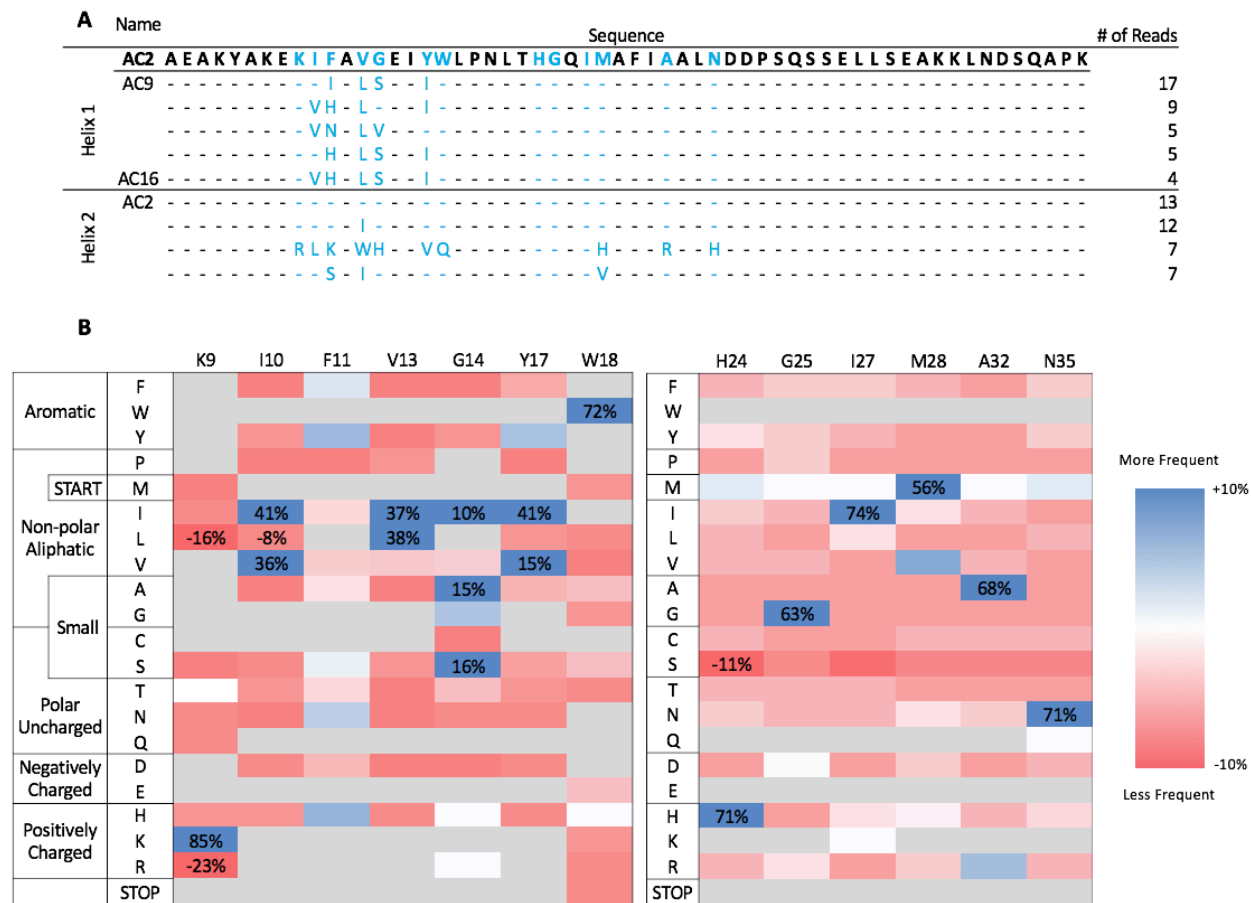
Supplemental Figures



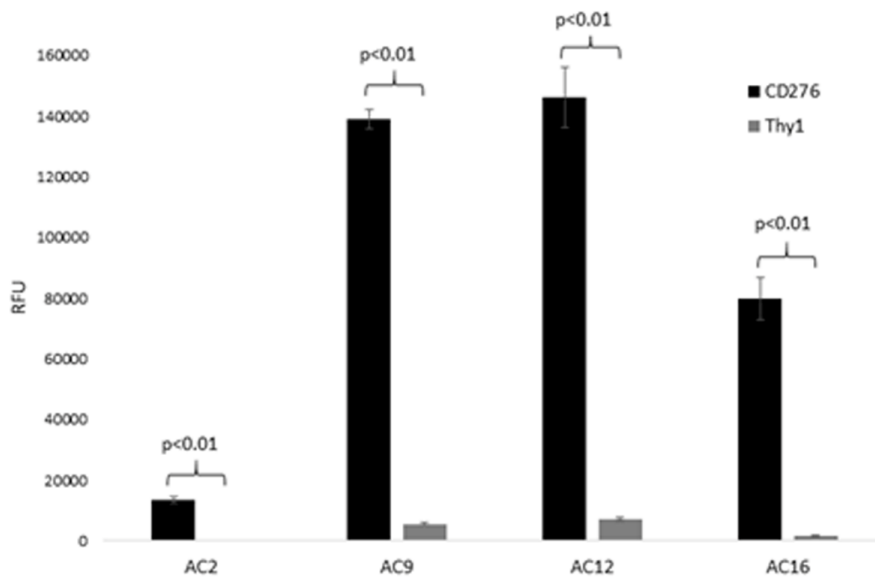
Supplemental Figure 1. *Assessment of specificity of fibronectin population against soluble CD276 extracellular domain.* The population of fibronectin domains selected by magnetic bead sorting against soluble CD276 extracellular domain was assessed for target specificity. The population was labeled by either 100 nM soluble CD276 extracellular domain (blue) or 100 nM soluble CMET extracellular domain (red).

	Library	
	Helix 1	Helix 2
K9	HIKLMNQRS	K
I10	ADFHILNPSTVY	I
F11	ADFHILNPSTVY	F
V13	ADFHILNPSTVY	V
G14	ACDFGINSTVY	G
Y17	ADFHILNPSTVY	Y
W18	AEGKLMRSTVW	W
H24	H	ACDFGHILNPRSTVY
G25	G	ACDFGHILNPRSTVY
I27	I	ACDFGHILNPRSTVY
N28	N	ACDFGHILNPRSTVY
A32	A	ACDFGHILNPRSTVY
N35	N	ACDFGHILNPRSTVY

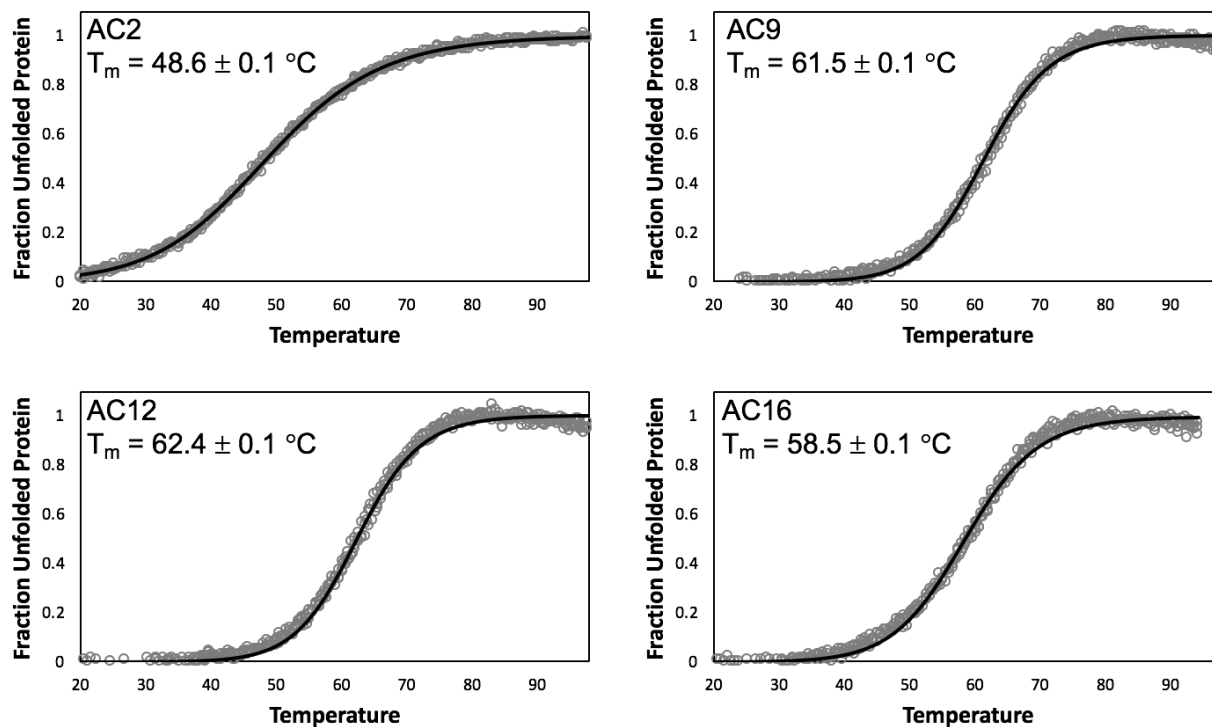
Supplemental Figure 2. *Affibody helix walking library designs.* Amino acid diversity for each site of the sub-library.



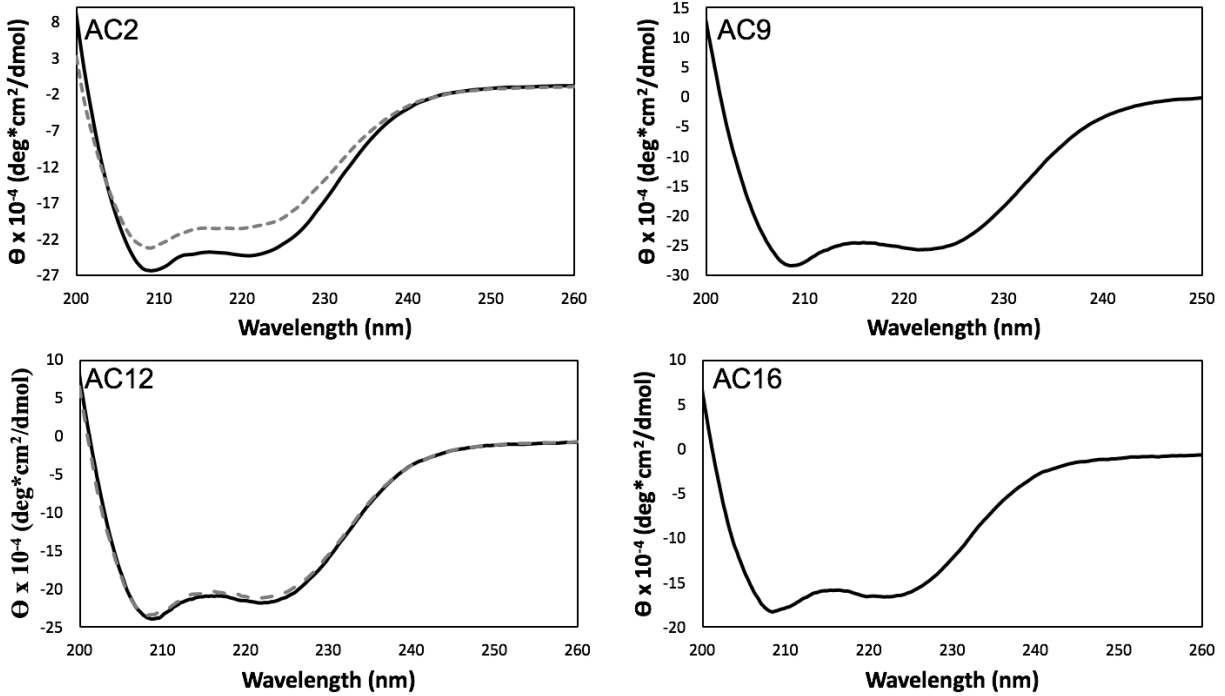
Supplemental Figure 3. Deep sequencing of sorted single-helix libraries reveals substantially improved mutants. (A) Top unique sequences of each single-helix library listed by number of reads. Parental AC2 is supplied as a reference. Blue lettering indicates diversified residues in the helix-walking libraries and dashes indicate parental amino acids at the indicated position. Clone names are supplied for sequences that were pursued for characterization. (B) Sitewise amino acid enrichments for the helix one (left) and helix two (right) libraries. Amino acid frequencies were calculated by grouping, counting, and quad-root dampening identical sequences. Values shown are change in amino acid frequency in sorted populations compared to theoretical amino acid diversity of the naïve library. Amino acids not allowed by library design are shown in grey, except in cases where they are substantially enriched or depleted.



Supplemental Figure 4. Specificity of affibody variants. Purified parental and evolved affibody variants were used to label either MS1-CD276 or MS1-Thy1 cells at saturating conditions (10 μ M for AC2, 1 μ M for all others). Binding was detected by fluorophore tagged anti-His₆ antibody via flow cytometry. Fluorescence was quantified as the difference between affibody labelled cells and control cells labelled with secondary antibody with n = 3 trials (68% confidence interval indicated). AC2 showed no significant binding to MS1-Thy1 cells relative to control. All other variants, while showing significant binding to MS1-Thy1 cells, bound significantly less strongly to MS1-Thy1 cells relative to MS1-CD276 cells.



Supplemental Figure 5. *Thermal denaturation curves of affibody variants.* Purified affibody variants were scanned at a wavelength of 220 nm during heating from 20 to 98 °C (1 °C/min). The midpoint of thermal denaturation (T_m) was calculated by linear least-squares regression using a two-state protein unfolding model.



Supplemental Figure 6. *Thermal denaturation curves of affibody variants.* Purified affibody variants were analyzed by circular dichroism spectroscopy in triplicate between 200 and 260 nm wavelengths before (solid) and after (dashed) thermal denaturation and cooling. No spectral data after cooling collected for AC9 or AC16.

Supplemental Table 1. Error-Prone PCR Oligonucleotide Sequences

W5	5'-CGACGATTGAAGGTAGATACCCATACGACGTTCCAGACTACGCTCTGCAG-3'
W3	5'-ATCTCGAGCTATTACAAGTCCTCTTCAGAAATAAGCTTTTGTTCGGATCC-3'
BCHPEP5	5'-CTGGAGGTTACCAACGCAACTCCGAACTCTCTGACTATTTCTTGG-3'
BCHPEP3	5'-CGGAACAGTGAATTCCTGGCTCGGGGAGTTACCACCAGTTTCGCCGTAGGTGATACGGTA-3'
DEHPEP5	5'-TACCGTATCACCTACGGCGAAACTGGTGGTAACTCCCCGAGCCAGGAATTCAGTGTCCG-3'
DEHPEP3	5'-TACAGCGTACACGGTAATGGTATAATCCTGGCCCGGTTTCAGACCGCTGATGGTCGC-3'
FGHPEP5	5'-AAACCGGGCCAGGATTATACCATTACCGTGTACGCTGTA-3'
FGHPEP3	5'-GTCGATTTTCGGTGCGATAATTGATGCTGATTGG-3'
ABY1F-b	5'-TTCTGGTGGTGGTGGTCTGCTAGCGCCGAAGCAAATAC-3'
ABY1R	5'-GGTCAGGTTTCGGCAG-3'
ABY2F	5'-CTGCCGAACCTGACC-3'
ABY2R-b	5'-TTTCTTCGCCTCAGACAGGAGTTCAGAGCTCTGGGACGGGTC-3'

Supplemental Table 2. Isolated Affibody Sequences and Frequencies

Isolated Affibody Sequences

	CD276-Targeted	Frequency
Soluble Target	AEAKYAKEKIFAVGEIYWLPNLTHGQIMAFIAALNDPPSQSSELLSEAKKLNDSQAPK	3/5
	AEAKYYKELHNAIVSIRVLPNLTVDQITAFIRALVNDPPSQSSELLSEAKKLNDSQAPK	1/5
	AEAKYSKEWFNAYVSIWGLPNLTVDQKSAFSYALDDPPSQSSELLSEAKKLNDSQAPK	1/5
Soluble Target + Lysate	AEAKYNKEWKYAYFSIVALPNLTGTQVHAFIQALHNDPPSQSSELLSEAKKLNDSQAPK	1/5
	AEAKYSKEWFTAYYQIGYLPNLTEYQRYAFVKALYDDPPSQSSELLSEAKKLNDSQAPK	2/5
	AEAKYYKELHNAIGVIRNLPNLTPIQKVAFAIALANDPPSQSSELLSEAKKLNDSQAPK	1/5
	AEAKYAKEYANAMVEIVCLPNLTLSSQGFIAALLDDPPSQSSELLSEAKKLNDSQAPK	1/5
Soluble Target + Cell Panning	AEAKYTKEKANAIQILVLPNLTVSQLHAFLSALHNDPPSQSSELLSEAKKLNDSQAPK	1/5
	AEAKYSKEWFNAYVSIWGLPNLTVDQKSAFSYALDDPPSQSSELLSEAKKLNDSQAPK	1/5
	AEAKYAKERLKAWLEIVELPNLTYTQLHAFIRALSDDPPSQSSELLSEAKKLNDSQAPK	2/5
	AEAKYNKEKFNAIASIFNLPNLTHHTQKTAFIVALNDPPSQSSELLSEAKKLNDSQAPK	1/5
Depleted Cell Panning	AEAKYTKEMYTAFDEIAQLPNLTQVQKVAFIVALWNDPPSQSSELLSEAKKLNDSQAPK	1/4
	AEAKYSKEKADAILSILLPNLTRAQVVAFMHALHNDPPSQSSELLSEAKKLNDSQAPK	1/4
	AEAKYAKEFSSALVEILTLPNLTVRQSSAFIRALHDDPPSQSSELLSEAKKLNDSQAPK	2/4
Cell Panning	AEAKYAKESSDAWHEIVQLPNLTHGQIHAFIRALHDDPPSQSSELLSEAKKLNDSQAPK	2/5
	AEAKYAKEFSSALVEILTLPNLTVRQSSAFIRALHDDPPSQSSELLSEAKKLNDSQAPK	1/5
	AEAKYSKERLRAWMEITGLPNLTKPQRIAFILALRDDPPSQSSELLSEAKKLNDSQAPK	1/5
	AEAKYYKELHNAIYSIRWLPNLTRVQKAAFLRALANDPPSQSSELLSEAKKLNDSQAPK	1/5
Cell Panning	Thy1-Targeted	
	AEAKYSKELHDAVDSIIALPNLTGHQMDAFISALINDPPSQSSELLSEAKKLNDSQAPK	1/5
	AEAKYAKEMDAAIIVILQLPNLTGYQMAAFIDALADDPPSQSSELLSEAKKLNDSQAPK	1/5
	AEAKYNKEMPTADYVIRQLPNLTLSSKQAFIHALHDDPPSQSSELLSEAKKLNDSQAPK	1/5
	AEAKYYKEQDDAIDEILSLPNLTGLQMRFAFIVALYDDPPSQSSELLSEAKKLNDSQAPK	1/5
	AEAKYNKEMDTAFDEILALPNLTGFQMDAFIYALSNDPPSQSSELLSEAKKLNDSQAPK	1/5

Supplemental Table 3. Isolated Fibronectin Domain Sequences and Frequencies

Isolated Fibronectin Domain Sequences		Frequency
	CD276-Targeted	
Soluble + Cell Panning	SSDSPRNLEVTNATPNSLTISWDAPCNTDTSGYRITYGETGGNSPSQESTVPGSNSATISGLKPGQDYTITGYAVSYGDYWWSNPISINYRTEIDKPSQ	4/9
	SSGSPRNLEVTNATPNSLTISWDAPCNTDTSGYRITYGETGGNSPSQESTVPGSNSATISGLKPGQDYTITGYAVSYGDYWWSNPISINYRTEIDKPSQ	3/9
	SSDSPRNLEVTNATPNSLTISWDAPCNTDASGYRITYGETGGNSPSQESTVPGSNSATISGLKPGQDYTITGYAVSYGDYWWSNPISINYRTEIDKPSQ	1/9
Depleted Cell Panning	SSDSPRNLEVTNATPNSLTISWDDSYDRAYYYRITYGETGGNSPSQEFTVPGTTNATISGLKPGQDYTITVYAVSYVNYAYYRSNPISINYRTEIDKPSQ	1/3
	SSDSPRNLEVTNATPNSLTISWDAPYYVYTYGYRITYGETGGNSPSQEFTVPGYNTATISGLKPGQDYTITVYAVSYHNTRYSSNPISINYRTEIDKPSQ	1/3
	SSDSPRNLEVTNATPNSLTISWDAPCRYAYGYRITYGETGGNSPSQEFTVPGNTNATISGLKPGQDYTITVYAVSNINYRYASNPISINYRTEIDKPSQ	1/3
	Thy1-Targeted	
Depleted Cell Panning	SSDSPRNLEVTNATPNSLTISWDAPYVYTFGYRITYGETGGNSPSQEFTVPGTNSATISGLKPGQDYTITVYAVSYDNYKYAHSNPISINYRTEIDKPSQ	1/1
Cell Panning	SSDSPRNLEVTNATPNSLTISWDAPDDGYTNGYRITYGETGGNSPSQEFTVPGSNTATISGLKPGQDYTITVYAVSYTSYAYYLSNPISINYRTEIDKPSQ	1/1

Supplemental Table 4. Sequences for Depletion Model System

HS	AEAKYTKEKANAIVQILVLPNLTVSQLHAFLSALHNDPSQSSELLSEAKKLNDSQAPK
HN	AEAKYAKESYASLYIGILPNLTHSQYYAFIYALQDDPSQSSELLSEAKKLNDSQAPK
LS	AEAKYNKELANAALSIVYLPNLTDQKSAFWLALQDDPSQSSELLSEAKKLNDSQAPK
LN	AEAKYAKERHRAWMEITGLPNLTRPQRIAFILALRDDPSQSSELLSEAKKLNDSQAPK
A5	AEAKYAKENFNATSEIYYLPNLTHFQRSAFSNALFDDPSQSSELLSEAKKLNDSQAPK