## SUPPLEMENTAL INFORMATION

## Insights into the function of the *N*-acetyltransferase SatA that detoxifies streptothricin in *Bacillus subtilis* and *Bacillus anthracis*

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Running title: Functional analysis of SatA

## SUPPLEMENTAL FIGURES



**Figure S1**. **Variant** *B. subtilis satA* alleles have no growth defect in *S. enterica*. Strains carrying wild-type (*satA*<sup>+</sup>, circles), variant *satA* alleles, or empty vector (squares) were grown in glycerol (22 mM) minimal medium (1). All strains carried *∆metE2702 ara-9* chromosomal mutations and the indicated plasmid: JE22263 (/pCV1), JE22334 (/pBsSATA1), JE23884 (/pBsSATA10), JE23887 (/pBsSATA11), JE23888 (/pBsSATA12), JE23889 (/pBsSATA13). Error bars represent one standard deviation and are present, although in many cases they are too small to be visible.



Figure S2. Mutant *B. anthracis satA* alleles have no growth defect in *S. enterica*. Strains carrying wild-type (*satA*<sup>+</sup>, circles), variant *satA* alleles, or empty vector (squares) were grown in glycerol (22 mM) minimal medium (1) with 250  $\mu$ M L-(+)-arabinose. Error bars represent one standard deviation and are present, although in many cases they are too small to be visible.



**Figure S3. Surface model of BaSatA (PDB 3PP9).** Ribbon (left) or surface (right) model of *Ba*SatA. Panels B is a 180° vertical rotation of panel A, intended to display the full surface of the enzyme. Conserved aromatic residues are colored: Y149 (purple), F154 (green), D160 (orange), and Y164 (blue), while AcCoA is shown in yellow.

	1	0 20	)	30	40	50
AAC(6')-Ig AAC(6')-Ih BaSatA BsSatA	MSLLIRELET	TNDLDNFPEID		IKPASEASLKD IMPISESQLSD LM.LSLSKVNR	WLELRNKLW WLALRCLLW RIEYTVEDV	SDSEASHLQEMH PDHEDVHLQEMR PSYEKSYLQNDN
Doodin	••••	ILINAKDI MKI M	LFFVVFGR	<u>ATTALEN</u> GVWI	*	* *
	eo	7.0	80	90	100	110
AAC(6')-Ig AAC(6')-Ih <i>Ba</i> SatA <i>Bs</i> SatA	QLLAEKYALQ QLITQAHRLQ EELVYNEYIN DDDMDVSYVE	LLAYSDHQA. LLAYTDTQQA KPNQIIY EEEGKAAF	IAMLEASI IAMLEASI IALLHNQI LYYLENNC	R F E Y VN G T E T S R Y E Y VN G T Q T S I G F I VL K K N WN I G R I K I R <mark>S N</mark> WN	PVGFLEGIY PVAFLEGIF NYAYIEDIT GYALIEDIA	VLPAHRRSGVAT VLPEYRRSGIAT VDKKYRTLGVGK VAKDYRKKGVGT
				* *	*	
	120	130	140	150	160	170
AAC(6')-Ig AAC(6')-Ih <i>Ba</i> SatA <i>Bs</i> SatA	MLIRQAEVWA GLVQQVEIWA RLIAQAKQWA ALLHKAIEWA	AKQFSCTEFAS KQFACTEFAS KEGNMPGIML KENHFCGLML	DAALDNVI DAALDNQI ETQNNNVA ETQDINISA	SHAMHRSLGFQ SHAMHQALGFH ACKFYEKCGFV ACHFYAKHHFI	ETEKVVYFS ETERVVYFK IGGFDFLVY IGAVDTMLY	KKID KNIG KGLNMTSDEVAI SNFP.TANEIAI
			**		*	
	180					
AAC(6')-Ig AAC(6')-Ih <i>Ba</i> SatA <i>Bs</i> SatA	YWYLHFDS FWYYKF					

**Figure S4. Alignment of AAC(6') and SatA proteins.** Protein sequences were aligned using Geneious software (2) (<u>https://www.geneious.com</u>) and the figure was generated using ESPript (3). Conserved residues are highlighted red while similar residues are boxed. Numbers refer to the residue number of *B. anthracis* SatA. Residues substituted in the *Ba*SatA (Y149, F154, D160, and Y164) are outlined in black. Residues of AAC(6')-Ig known to bind tobramycin are indicated with asterisks below the residue. The green asterisk corresponds to the putative active site glutamate for SatA and a reported binding site of tobramycin for AAC(6')-Ig.



Purity 90% 79% 78% 89% 79% 83% 83% 84% 90% 90% 95% 86% 92% 90% 98% 61% 86% 72% 53% 66% 60% 56% 84% 86% 89% 87% 86% Figure S5. Assessment of BsSatA and BaSatA variants purity. Samples of each variant were run on SDS-PAGE, followed by staining with Coomassie Brilliant Blue R. Protein purity was calculated by running various dilutions of proteins on a separate 12% SDS-PAGE gel and bands were quantified using Total Lab software (4). MM stands for molecular mass marker.



**Figure S6.** *Ba***SatA** *in vitro* activity is highest in HEPES buffer. Various buffers (50 mM) were used to determine the specific activity of *Ba*SatA under saturating conditions (10  $\mu$ M streptothricin and 500  $\mu$ M AcCoA) using a continuous DTNB spectrophotometric assay (5) as described in the *Material and Methods*. Abbreviations: MES (2-(*N*-morpholino)ethanesulfonic acid), Bis-Tris (Bis(2-hydroxyethyl)amino-tris(hydroxymethyl)methane), MOPS (3-(*N*-morpholino)propanesulfonic acid), HEPES (4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid), and Tris (*tris*(hydroxymethyl)aminomethane).

## REFERENCES

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